정량분석을 통한 Eleutherococcus species의 HPLC 분석법 검증과 표준화

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Standardization of *Eleutherococcus* species and HPLC Method Validation for Quantitative Analysis

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ABSTRACT

Objective : For the standardization and quality control of eleutheroside E in *Eleutherococcus* species, HPLC analysis was performed and eleutheroside E content was compared in 23 kinds of *Eleutherococcus* species collected from Korea and China.

Methods: The content of eleutheroside E in stem bark of *Eleutherococcus* species collected from Korea and China were analyzed by HPLC. 0.5% phosphoric acid and acetonitrile was used as mobile solvent. Validation of HPLC analysis method was confirmed by analyzing specificity, linearity, precision and accuracy following ICH guideline. **Results**: Content of eleutheroside E was determined to be 1.0-1.6% and 0.5-0.8% in Korean and Chinese *E, senticosus*, respectively. Content of eleutheroside E in *E, sessiliflorus* was 0.7-1.1% and 0.2-0.4% respectively in Korean and Chinese origin. All calibration curves showed good linear regression. The method showed good precision and accuracy with intra-day and inter-day variations of 0.880-3.442% (RSD) and 0.606-3.328% (RSD), respectively, and average recovery was of 0.141-1.363% (RSD), for the eleutheroside E analyzed. **Conclusion**: These results might be used to establish a criterion of eleutheroside E in *Eleutherococcus* species.

Key words: Eleutherococcus senticosus, Method validation, Standardization, Eleutheroside E, HPLC

INTRODUCTION

Eleutherococcus species were considered as useful medicinal herbal resource in Korea, China, Japan, and Russia from the time immemorial¹⁾. In Traditional Korean Medicine (TKM), these have been used as a drug with adaptogenic activity^{2,3,4)}, anti-tumor⁵⁾, anti-stress⁶⁾, fatigue⁷⁾ and hypoglycrmic⁸⁾. Root and stem bark of *Eleutherococcus senticosus*, known as Siberian Ginseng, has been often used to treat stroke as well as tonify qi, strengthen musule and bone, tranquilize and dispel wind dampness⁹⁾. Recent researches have shown that *E*, *senticosus* exerted neuroprotective effect on amyloid beta induced neuritic

 $atrophy^{10}$. 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine induced cell death¹¹⁾, and transient focal cerebral ischemia¹²⁾.

Among various diterpenoids and triterpenoids reported from this plant, the lignan compounds, eleutheroside B (syringin) and eleutheroside E (–) syringaresinol-di-O-13-D glucoside, are known to be main active principles¹³⁾. Beside these, *E. senticosus* contains chiisanoside, daucosterin, β -sitosterol and sesamin, which are responsible for its diverse biological activities¹⁴⁾.

Among 500 herbal medicines reported in Korean Herbal Pharmacopoeia and Herbal Pharmacopoeia, only 50 herbal medicines are available forevaluationa

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quantitative analysis. On the other hand, 551 herbs listed in Chinese Herbal Pharmacopoeia, 215 herbal medicines can be possible tothrough a quantitative analysis. The standardization and the quality control of the active constituents of many herbs are still lacking¹⁵.

In this study, the content of eleutheroside E in 23 kinds of *Eleutherococcus* species collected from Korea and China weredone by HPLC analysis. It was carried out HPLC method validation and quantity standardization according to the International Conference on Harmonization (ICH) guidelines.

Material and Methods

1. Plant materials

Dried stem barks of twenty three kinds of *Eleutherococcus* species were purchased from Kyung Dong Herbal Market, Jegi-dong Seoul, Korea and Cheolwon and Yang-gu, Gangwon-do, Korea. Samples were identified by Professor Dr. Hocheol Kim, Department of Herbal Pharmacology, College of Oriental Medicine, Kyung Hee University, Seoul, Korea.

2. Chemicals and Reagents

All analytical reagents were of grade. from Acetonitrilewas purchased J Т Baker (Phililpsburg, NJ, USA). Eleutheroside E was obtained from Chromadex (purity \geq 92.3%). Water was filtered through a 0.45 µm membrane (Millipore, Bedford, MA, USA).

3. Preparation of the crude extracts

At first, the dried stem barks of *Eleutherococcus* species were cut into the pieces, and then those were extracted with 70% ethanol for 3 hours at 82 \bigcirc C in a reflux apparatus. The extracts were filtered, then the filtrate was evaporated in a rotary evaporator and the powders were lyophilized in a freeze-dryer (OperonTM, Seoul, Korea).

4. Validation for HPLC analysis

1) Preparation of standard and sample solution

Stock solution of 1.0 mg/mL was prepared in 85% methanol for eleutheroside E. A serial dilution was made on each stock solution with 85% methanol to prepare standard solutions at concentrations of 0.5, 1,

5, 10, 50, 100, and 500 μ g/mL from each of which 10 μ L was used for plotting the standard curves for eleutheroside E. The *E. senticosus* extract (ESE) sample was accurately weighed (50.0 mg), placed in 5 mL of 50% methanol in aqueous solution in an ultrasonic device for 30 sec for extraction. This ESE solution was passed through a 0.45 μ m syringe membrane filter and 10 μ L of the filtrate was injected in triplicate the HPLC system for quantitative analysis.

2) Chromatographic conditions

Analysis was performed in a Waters instrument equipped with a Waters 600 pump, a Waters 717 autosampler and a Waters 996 PDA detector using a SunFire^{**} C18 column (5 μ m; 4.6×250 mm; Ireland). The column was equilibrated with a 95:5 mixture of distilled water containing 0.5% phosphoric acid (solvent A) and acetonitrile (solvent B) at a flow rate of 1.0 mL/min. The column was eluted as follows: 0– 60 min 5–50% solvent B, 60–61 min 50–70% solvent B, 61–80 min 70–70% solvent B. Column temperature was kept constant at 25 °C. The absorbance was measured at 205 nm for detection of eleutheroside E. The 2–D HPLC chromatogram of *Eleutherococcus* species is shown in Fig. 1.

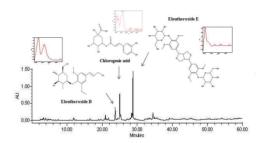


Fig. 1. 2–D HPLC Chromatogram for Standardization of *Eleutherococcus S*pecies.

Detection was performed by using a photodiode array detector. X-axis is retention time; Y-axis is wavelength, and Z-axis is absorbance unit. Analytical conditions were as follows: column, SunFire^T C18 (5µm; 4×250 mm); mobile phase, solvent A (0.5% H₃PO₄) and solvent B (CH₃CN); Gradient program, 0-60 min 5-50% B; 60-61 min 50-70% B; 61-85 min 70% B. The column temperature was kept constant at 25 °C. The flow rate was set at 1.0 mL/min and the injection volume was fixed at 10 µL. The UV wavelength was monitored at 205 nm.

3) Linearity and range

To assess the linearity of standard curve, seven different concentrations of standard were prepared and injected to HPLC system. Six replicates of eight calibration standards were analyzed for each range. The regression equation was calculated in the form of Y=AX+B, where Y and X are the area of the peak and the sample amount, respectively.

4) Specificity

The analyte was confirmed by spiking the eleutheroside E (0.1 mg/mL) with the ESE (10.0 mg/mL). UV spectra of ESE compared with the eleutheroside E. The selected monitoring wavelengths for eleutheroside E was maximum absorption wavelengths. The eleutheroside E from ESE was identified by comparing to the retention times and UV spectra of the eleutheroside E.

5) Precision and accuracy

Intra-day and inter-day variations were determined the precision were estimated by analyzing six replicates containing the standard compound at three different concentration (25, 50, 100 μ g/mL) in a single day and for six days, respectively. Variations were expressed by the relative standard deviations (R.S.D.).

$$\begin{aligned} \text{Recovery (\%)} &= \frac{\text{Amount(bright)} - \text{Amount(Original)}}{\text{Amount (splited)}} \times 100\%, \\ \text{R.S.D. (\%)} &= \frac{\text{S. D.}}{\text{Mean}} \times 100\% \end{aligned}$$

The average recovery was calibrated by formula.

Where $amount_{(found)}$ is the determined total amount of each analyte, $amount_{(Original)}$ is the original amount of each analyte in ESE measured above, and $amount_{(spiked)}$ is the spiked amount of each analyte.

6) Limit of detection and limit of quantification (LOD and LOQ)

The LOD was defined as the concentration of the standard solution with a signal-to-noise (S/N) ratio \rangle 3.3. The LOQ was defined as the concentration of standard solution with a S/N \rangle 10. LOD and LOQ for

eleutheroside E were also shown in Table 1.

The LOD and LOQ were calibrated by formula.

- $LOD = 3.3 \times (S.D. \text{ of the response / slope of the calibration curve})}$
- $LOQ = 10 \times (S.D. \text{ of the response / slope of the calibration curve})}$

7) Quantitative analysis

The concent of eleutheroside E in the final solution was determined by using a calibration curve of concentration versus peak area and expressed as percentages.

$$C (\%) = (C_{spl} \times V) / M \times 1000$$

Where C_{spl} is the concentration of eleutheroside E in the final solution ($\mu g/mL$), V is the total volume of the final solution (mL) and M is the ESE taken for extraction (g).

8) Statistical Methods

Data were expressed as percentages, and recorded as means \pm standard deviation(S.D) of triplicate measurements. And *n* represents the number of replicates. For thestatistical analysis Excel 2007[®] (Microsoft) software was used.

Results

1. Method validation

1.1 Linearity

The calibration curves were linear in the tested concentration ranges. The peak area ratio of the eleutheroside E was linear in the range of 0.5-500 μ g/mL. The correlation coefficients were all greater than 0.99, indicating high correlation and good linearity of the method. The slopes, y-intercepts, and correlation coefficients (r²) obtained from regression analysis are shown in Table 1.

Table 1. Calibration curves, LOD and LOQ for the elentheroside E standards, ^aThe notation for analyte (Eleutheroside E), ^bLOD refers to the limits of detection, ^cLOQ refers to the limits of quantification,

Analyte ^a	Calibration curve	Correlation coefficient (r^2)	Linear range (µg/mL)	LOD ^b (mg/mL)	LOQ ^c (mg/mL)
1	y = 38,668,439.209x - 59,326.819	0.999			
2	y = 37,696,822.099 x + 187,360.296	0.999			
3	y = 41,940,407.048x + 419,938.529	0.995	0 5 500	0.179	0 590
4	y = 36,901,753.862 x + 124,709.293	0.999	0.5-500	0.173	0.526
5	y = 33,120,802.473 x + 86,798.450	0.999			
6	y = 38,892,847.629 x + 309,993.436	0.997			

1.2 Specificity

The eleutheroside E from ESE was identified by comparing the retention times and UV spectra of the

eleutheroside E. Typical chromatogram of eleutheroside E and ESE recorded at 205 nm are depicted in Fig. 2.

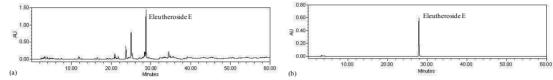


Fig. 2. HPLC Chromatograms of (a) E. senticosus extract and (b) Eleutheroside E (at 205 nm).

1.3 Precision and Recovery

The relative standard deviation (RSD) values for inter-day precision were 0.88-3.44% and those of intra-day were 0.61-3.33%. The average recovery was

81.26-07.75% (RSD 0.14-1.36%). All of these values are within the acceptance limits of ICH. The results of intra-day and inter-day accuracy and precision analyses of ESE are shown in Table 2 and Table 3.

Table 2. Intra-day and Inter-day variation of eleutheroside E in Esenticosus. (R.S.D., n=6)

		Precis	sion	
Analyte	Intra-da	у	Inter-da	у
	Mean (µg/mL)	RSD (%)	Mean (µg/mL)	RSD (%)
1	4.842	0.880	4.187	0.915
2	6.075	1.477	6.023	1.406
3	4.725	2.542	4.687	3.328
4	4.560	2.623	4.475	2.490
5	4.537	2.396	4.581	2.649
6	5.660	3.442	5.577	0.606

Table 3. Validation for the Recovery of eleutheroside E in *E. senticosus*, (Recovery (%), *n*=3)

Added		Eleutheroside	εE	
Added	Recovery (%)	Mean (%)	SD (%)	RSD(%)
	107.73			
100%	108.74	107.75	0.976	0.906
	106.79			
	81.18			
75%	81.22	81.26	0.115	0.141
	81.40	-		
	82.17	-		
50%	83.50	83.37	1.136	1.363
	84.43	-		

1.4 Limits of Detection and Limits of Quantification

The LOD and LOQ were 0.173 mg/mL and 0.526 mg/mL, respectively. These results indicate that the method provided adequate sensitivity (Table 1).

2. Analysis of eleutheroside E in different Eleutherococcus species and standardization

2.1. Comparison of eleutheroside E in Eleutherococcus species according to cultivation region

The developed HPLC method was applied to determine the content of eleutheroside E in 23 kinds

of *Eleutheorococcus* speices collected from Korea and China. The content of eleutheroside E was found to be 0.52-2.44% in *E. senticosus*, and 0.24-1.61% in *E. sessiliflorus* of Korean origin. Also, in Chinese origin, the content of eleutheroside E was found to be 0.13-0.98% in *E. senticosus*, and 0.12-1.61% in *E. sessiliflorus*. Content of the eleutheroside E in Korean species was found to be 2 times higher than that of Chinese species. The quantitative analytical results of eleutheroside E in 23 kinds of *Eleutherococcus* species of Korean and Chinese origin are shown in Table 4 to Table 7. A typical chromatogram of the crude extract and eleutheroside E is shown in Fig. 1.

Sample No.	Cultivation region	Content of eleutheroside E		
Sample No.		Extract bias ^a (%, n=3)	Raw material bias ^b (%, n=3)	
EK1	Korea, Cheolwon	1.69 ± 0.04	0.050	
EK2	Korea, Cheolwon	0.92 ± 0.02	0.128	
EK3	Korea, Cheolwon	2.44 ± 0.04	0.090	
EK4	Korea, Cheolwon	0.52 ± 0.17	0.051	
EK5	Korea, Yang-Gu	1.01 ± 0.01	0.053	
EK6	Korea, Cheolwon	1.37 ± 0.01	0.045	
I	Average		0.070	
D	80%	1.06	0.056	
Range	120%	1.59	0.083	
S	tandard	1.02-1.64	0.011-0.183	

Table 4. Contents of Eleutheroside E among 6 kinds of Korean *E, senticosus* (^a Data are given as Mean±S,D, ^bRefer to dry weight (g) of *E, senticosus*.).

Table 5. Contents of eleutheroside E among 6 kinds of Chinise *E, senticosus*, (^a Data are given as Mean±S.D. ^bRefer to dry weight (g) of *E, senticosus*)

Converto No	Cultivation region	Content of eleutheroside E		
Sample No.		Extract bias ^a (%, n=3)	Raw material bias ^b (%, n=3)	
EC1	EC1 China, Unknwon		0.050	
EC2	EC2 China, Unknwon		0.061	
EC3	China, Unknwon	0.29 ± 0.05	0.017	
EC4	China, Unknwon	0.49 ± 0.05	0.045	
EC5	EC5 China, Unknwon		0.035	
EC6	China, Chuancai	0.13 ± 0.01	0.011	
I	Average	0.622 ± 0.034	0.037	
D	80%	0.42	0.029	
Range	120%	0.63	0.053	
S	tandard	0.39-0.67	0.006-0.096	

2.2 Comparison of eleutheroside E content according to Eleutherococcus species

Eleutheroside E content was compared in 12 kinds of E. senticosus and 11 kinds of E. sessiliflorus collected from Korea and China. Content of eleutheroside E was found to be 0.13-2.44% in E. senticosus and 0.12-1.61% in *E. sessiliflorus*. The quantitative analytical results of eleutheroside E in 23 kinds of *Eleutherococcus* species are shown in Table 4 to Table 7.

2.3 Variation of the eleutheroside E content in E. senticosus according to harvesting time

The content of eleutheroside E was found to be varied according to the harvesting time of E, senticosus. The eleutheroside E content in E, senticosus of 2–3 years old was 0.92%, that of 5–6 years old one was 2.44%, and that of 6–9 years old one was 1.69%. The content of eleutheroside E of 5–6 years old was found to be highest. (Table 8).

Table 6. Contents of eleutheroside E among 4 kinds of Korean *E, sessilitlorus*. (^aData are given as Mean±S,D. ^bRefer to dry weight (g) of *E, sessilitlorus*)

Sample No.	Cultivation region	Content of eleutheroside E		
Sample No.		Extract bias ^a (%, <i>n</i> =3)	Raw material bias ^b (%, $n=3$)	
EsK1	EsK1 Korea, Yangsan		0.042	
EsK2	EsK2 Korea, Unknown		0.095	
EsK4	Korea, Yeongju	0.30 ± 0.02	0.020	
EsK5	Korea, Unknown	0.23 ± 0.00	0.020	
	Average	0.86 ± 0.02	0.042	
Range	80%	0.69	0.033	
Nange	120%	1.03	0.050	
S	tandard	0.68-1.05	0.003-0.096	

Comple No		Content of eleutheroside E		
Sample No.	Cultivation region —	Extract bias ^a (%, n=3)	Raw material bias ^b (%, <i>n</i> =3)	
EsC1	China, Unknown	0.72 ± 0.02	0.055	
EsC2	EsC2 China, Unknown		0.033	
EsC3	China, Unknown	0.66 ± 0.01	0.046	
$\mathrm{EsC4}$	China, Unknown	0.13 ± 0.00	0.008	
EsC5	China, Unknown	0.12 ± 0.01	0.013	
EsC6	China, Unknown	0.16 ± 0.00	0.019	
Average		0.56 ± 0.01	0.024	
Range	80%	0.25	0.019	
Nange	120%	0.38	0.029	
S	tandard	0.25-0.39	0.004-0.063	

Table 7. Contents of eleutheroside E among 6 kinds in Chinese origin *E, sessiliflorus,* (^a Data are given as Mean±S,D, ^b Refer to dry weight (g) of *E, sessiliflorus*).

Table. 8, Contents of eleutheroside E among 3 kinds of Korean *E, senticosus* according to harvesting time, (^a Data are given as Mean±S,D. ^bRefer to dry weight (g) of *E, senticosus*.)

Sample No.	No. Cultivation region(Harvest time) —	Content of eleutheroside E		
Sample No.		Extract bias ^a (%, n=3)	Raw material bias ^b (%, n=3)	
EK2	Korea, $(2 \sim 3)$	0.92 ± 0.02	0.050	
EK3	Korea, $(5 \sim 6)$	2.44 ± 0.04	0.128	
EK1	Korea, $(6 \sim 9)$	1.69 ± 0.04	0.090	
	Average		0.089	
Dongo	80%	1.347	0.014	
Range	120%	2.021	0.236	
	Standard	1.32-2.06	0.07-0.10	

2.4 Standardization of eleutheroside E content in Eleutherococcus species

Finally, quality standardization was determined by 80-120% range of average value of eleutheroside E content. Content of eleutheroside E was found to be 1.02 to 1.64% and 0.39 to 0.67% in Korean and Chinese *E. senticosus*, respectively. Eleutheroside E content was 0.68 to 1.05% and 0.25 to 0.39% in *E. sessiliflorus* of Korean and Chinese origin.

Discussion

Quantity of photochemical in plant might be affected by the geographical variation, climatic condition, soil condition, environmental factors, etc. This leads to the variation in the quantity of active constituents depending upon seasonal and environmental factors¹⁶.

The method was developed and validated in compliance with the ICH guidelines and is suitable for the simultaneous determination of eleutheroside E in *Eleutherococcus* species. Calibration curves show the linearity of 0.995–0.999%, LOD of 0.173 mg/mL, LOQ of 0.526 mg/mL, inter-days precision of 0.880–3.442% (RSD), and intra-day precision of 0.606–3.328% (RSD). The average recovery values were of 0.141–1.363% (RSD) for the method

validation.

In this study, quantitative analysis of eleutheroside E in 23 kinds of *Eleutherococcus* species collected from Korea and China was done by HPLC method and their contents were compared. Content of eleutheroside E was found to be 0.52-2.44% in E. senticosus, and 0.24-1.61% in E. sessiliflorus of Korean origin. Also, in Chinese origin, eleutheroside E content was found to be 0.13-0.98% in E. senticosus. and 0.12-1.61% in E. sessiliflorus. Content of the eleutheroside E in Korean species was found to be 2 times higher than that of Chinese species. Eleutheroside E content was compared in 12 kinds of E. senticosus and 11 kinds of E. sessiliflorus collected from Korea and China. Content of eleutheroside E was found to be 0.13-2.44% in E. senticosus and 0.12-1.61% in E. sessiliflorus. Eleutheroside E content was found to be 2 times higher in E. senticosus than that in E. sessiliflorus. It was consist with previous report which contents of eleutheroside E was 3 times higher in *E. senticosus* than in *E. sessiliflorus*¹⁷. This result clearly shows that content of phytochemicals are various depending upon the cultivation region and origin. Also, the content of eleutheroside E was found to be varied according to the harvest time of E. senticosus. The eleutheroside E content in Esenticosus of 2-3 years old was 0.92%, that of 5-6 years old one was 2.44%, and that of 6-9 years old

one was 1.69%. At the optimum age of the plant, the active constituent will be in highest concentration and rest of the time its amount is altered¹⁸. Here, it was found that the optimum age of *E. senticosus* harvest time was 5–6 years. Finally, quality standardization was determined by 80–120% range of average value of eleutheroside E content.

In summary, harvesting time and the age of the plant play vital role for the presence of the active constituents in appropriate amount. So, one should be very careful and have sound knowledge about the plant material in order to ensure high quantity of active constituents. Constantly, active constituent should be compared according to the cultivation region and harvesting time for quantity standardization and management. Among the various methods for quality standardization of the herbal medicine, the developed HPLC method is efficient standardization methods for the identification and quantification of *Eleutherococcus* species. Therefore, this result might be useful resource for Korea Food & Drug Administration (KFDA) to identify the quality standardization criteria of herbal medicine and furthermore, helpful to the related food industrial company for setting QC guidelines.

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