

Evaluation of Magnetic and Thermal Properties of Ferrite Nanoparticles for Biomedical Applications

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Magnetic nanoparticles can potentially be used in drug delivery systems and for hyperthermia therapy. The applicability of Fe₃O₄, CoFe₂O₄, MgFe₂O₄, and NiFe₂O₄ nanoparticles for the same was studied by evaluating their magnetization, thermal efficiency, and biocompatibility. Fe₃O₄ and CoFe₂O₄ nanoparticles exhibited large magnetization. Fe₃O₄ and NiFe₂O₄ nanoparticles exhibited large induction heating. MgFe₂O₄ nanoparticles exhibited low magnetization compared to the other nanoparticles. NiFe₂O₄ nanoparticles were found to be cytotoxic, whereas the other nanoparticles were not cytotoxic. This study indicates that Fe₃O₄ nanoparticles could be the most suitable ones for hyperthermia therapy.

Keywords : magnetic nanoparticle, drug delivery systems, hyperthermia, cytotoxicity

1. Introduction

Nanoparticles have several potential biomedical applications, since they are compatible in size. Protein molecules are (5-50 nm) and segmented DNA molecules (2 nm × (10-100 nm)) [1]. Nanoparticles are particularly promising as carriers in drug delivery systems for treating cancer because of the enhanced permeability and retention (EPR) effect. The EPR effect refers to the phenomenon of enhanced extravasations of macromolecules from tumor blood vessels and the retention of these macromolecules in tumor tissues; the same is not observed in normal vasculature [2]. Magnetic nanoparticles can be utilized not only as carriers for drug delivery systems but also as heat sources for hyperthermia therapy. Hyperthermia therapy is a heat treatment for cancer that poses a low risk to the body and causes few side effects as compared to surgery, chemotherapy, and radiation therapy. Hyperthermia therapy can also be administered repeatedly. Magnetic nanoparticles have attractive properties such as magnetic guidance and induction heating in an ac magnetic field. Thus, magnetic nanoparticles enable combination therapy involving drug delivery systems and hyperthermia. Ferrite nanoparticles are suitable materials

for biomedical applications because of their wide range of size, diversity, and chemical stability as compared to metal nanoparticles [3]. Fe₃O₄ and γ -Fe₂O₃ nanoparticles have been explored extensively from the viewpoint of their biocompatibility [4]; however, for other ferrite nanoparticles there have been only a few reports on their induction heating ability and cytotoxicity [5, 6]. The size dependence of the magnetic and thermal property and the ac magnetic property of NiFe₂O₄ nanoparticles (7.7 nm and 242 nm, respectively) have been reported [7, 8]. In this study, the applicability of various ferrite nanoparticles as drug delivery systems and for hyperthermia therapy is investigated by evaluating their magnetic characterization, thermal properties, and cytotoxicity.

2. Experiments

Fe₃O₄, CoFe₂O₄, MgFe₂O₄, and NiFe₂O₄ nanoparticles having various sizes were prepared in a dried powder state. Fe₃O₄ and NiFe₂O₄ nanoparticles (particle size: 20-30 nm) were purchased from Nanostructured & Amorphous Materials, Inc.; other nanoparticles were synthesized using the newly modified sol-gel method [9]. The mean size and deviation of the nanoparticle are listed in Table 1. The samples except NiFe₂O₄ (20-30 nm) was supplied to magnetic and thermal characterization studies. The dc magnetization curve of the major loop was measured

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Table 1. The average particle sizes and distributions of the particles were determined by a high resolution transmission electron microscopy (HRTEM) and a field emission electron microscopy (FESEM).

Material	Size [nm]
Fe ₃ O ₄	20-30
CoFe ₂ O ₄	26.5 ± 6.1, 32.7 ± 7.3, 146.1 ± 39.3
MgFe ₂ O ₄	27.4 ± 9.2, 34.4 ± 16.1, 130.2 ± 40.7
NiFe ₂ O ₄	20-30, 24.8 ± 6.3, 32.8 ± 8.6, 130.7 ± 44.9

using a vibrating sample magnetometer (VSM) at room temperature with a maximum field of 10 kOe. The thermal property of the particles was measured by applying an ac magnetic field. The field strength was varied from 50 to 500 Oe, and the frequency was fixed at 10 kHz. In the magnetic and thermal characterization studies, 60 mg of each sample was used in the dried state. A cytotoxicity study of Fe₃O₄ (20-30 nm), CoFe₂O₄ (26.5 ± 6.1 nm), MgFe₂O₄ (27.4 ± 9.2 nm), and NiFe₂O₄ (20-30 nm) nanoparticles was conducted on human cervical carcinoma cells (HeLa cells). HeLa cells were cultured in Dulbecco's modified eagle medium (DMEM; GIBCO) with 10% fetal bovine serum (Equitec-bio, Inc.) and 1% penicillin streptomycin (GIBCO); they were then incubated at 37°C in a 5% CO₂ atmosphere. HeLa cells were seeded at a density of 2 × 10⁴ cells/well in a 24-well plate and incubated at 37°C in a 5% CO₂ atmosphere. After 24 hours of incubation, HeLa cells were exposed to 200 μg/ml of each nanoparticle dispersed in the medium. The cell viability was measured for 4 days. The relative ratio of the cells treated with and without magnetic nanoparticles was calculated by the following equation:

$$\text{Relative ratio [\%]} = \frac{\text{number of cells treated with nano-}}{\text{number of cells treated without nano-}}$$

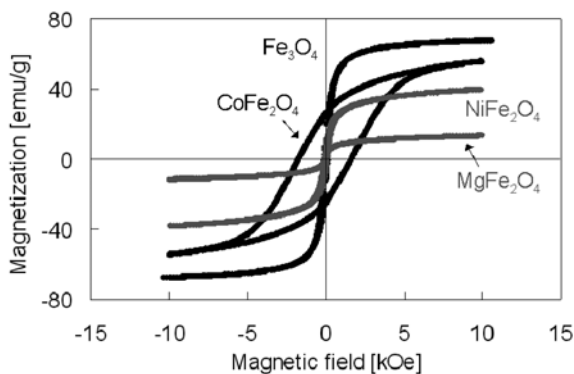


Fig. 1. Hysteresis loops of Fe₃O₄ (20-30 nm), CoFe₂O₄ (26.5 ± 6.1 nm), MgFe₂O₄ (27.4 ± 9.2 nm), and NiFe₂O₄ (24.8 ± 6.3 nm) measured by a vibrating sample magnetometer at room temperature with the maximum applied field of 10 kOe.

particles [cells]/number of cells treated without nanoparticles [cells]

3. Results and Discussion

3.1. Magnetic properties

Fig. 1 shows the hysteresis curves of Fe₃O₄ (20-30 nm), CoFe₂O₄ (26.5 ± 6.1 nm), MgFe₂O₄ (27.4 ± 9.2 nm), and NiFe₂O₄ (24.8 ± 6.3 nm) as measured using a VSM at room temperature. The magnetic properties of the samples

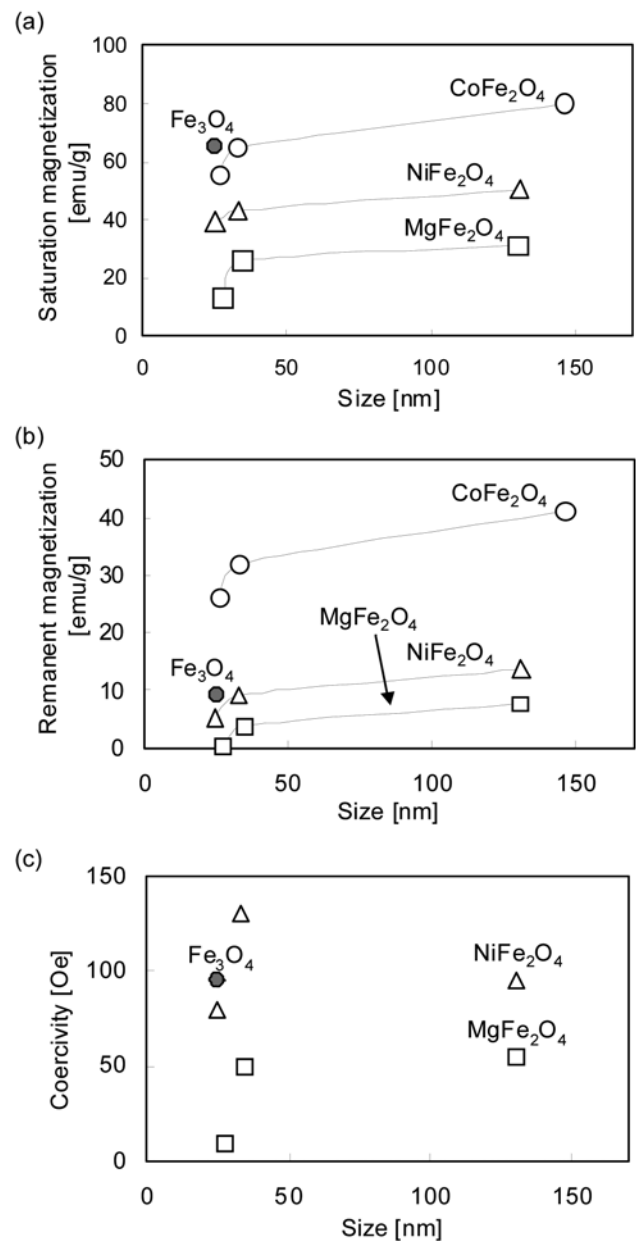


Fig. 2. Magnetic characterization of samples measured by a vibrating sample magnetometer at room temperature with the maximum applied field of 10 kOe. (a) saturation magnetization, (b) remanent magnetization, and (c) coercivity.

clearly exhibited material dependence. The saturation and remanent magnetizations of all samples listed in Table 1 are shown in Figs. 2(a) and (b), respectively. The coercivity of all samples except for the CoFe_2O_4 nanoparticles is shown in Figs. 2(c). The coercivity of CoFe_2O_4 at 26.5 nm, 32.7 nm, and 146.1 nm was 1720 Oe, 1570 Oe, and 1030 Oe, respectively. Fig. 2(a) shows that the smaller particles exhibited smaller saturation magnetization. Similar reduced saturation magnetization of various ferrite nanoparticles has been reported in some papers [7, 10-12]. This reduced magnetization of magnetic nanoparticles can be explained by the existence of a nonmagnetic layer as a surface dead layer [13]. The saturation magnetization of CoFe_2O_4 is comparable to that of Fe_3O_4 and is greater than that of the other nanoparticles. Figs. 2(b) and (c) show that CoFe_2O_4 exhibits much larger remanent magnetization and coercivity than the other nanoparticles because of its large anisotropy. The coercivity largely depends on the anisotropy of the material. The coercivity of the samples was shown to be strongly size-dependent. In multi-domain particles, magnetization reversal occurs because of domain wall movement. As domain walls move through a particle, they are pinned at grain boundaries; additional energy is required for the domain walls to continue moving. Therefore, reducing the particle size creates more pinning sites

and increases the coercivity of multi-domain nanoparticles [14]. In single-domain particles, the higher coercivity of nanoparticles can be attributed to the demagnetization caused by domain rotation, which requires greater energy than the domain wall movement of multi-domain particles [12]. Therefore, the coercivity of single-domain nanoparticles increases with their size. The coercivities of MgFe_2O_4 and NiFe_2O_4 nanoparticles correspond to these reports.

3.2. Thermal properties

The temperature rise of all samples listed in Table 1 in an ac magnetic field is shown in Fig. 3. The temperature rise of the samples is indicated by $\Delta T/\Delta t$ at $t \approx 0$, the initial slope for time dependence, where T and t are the measured temperature and time, respectively. Only hysteresis loss was assumed to contribute to the heat generated by the nanoparticles because of the low frequency (10 kHz). Fig. 3(a) shows that Fe_3O_4 (20-30 nm) nanoparticles exhibited the highest temperature rise as compared to the other nanoparticles because it had the highest magnetization. Fig. 3(b) shows that CoFe_2O_4 nanoparticles exhibited the lowest temperature rise and that the slope increased exponentially with the magnetic field strength. This was because of the high coercivity of

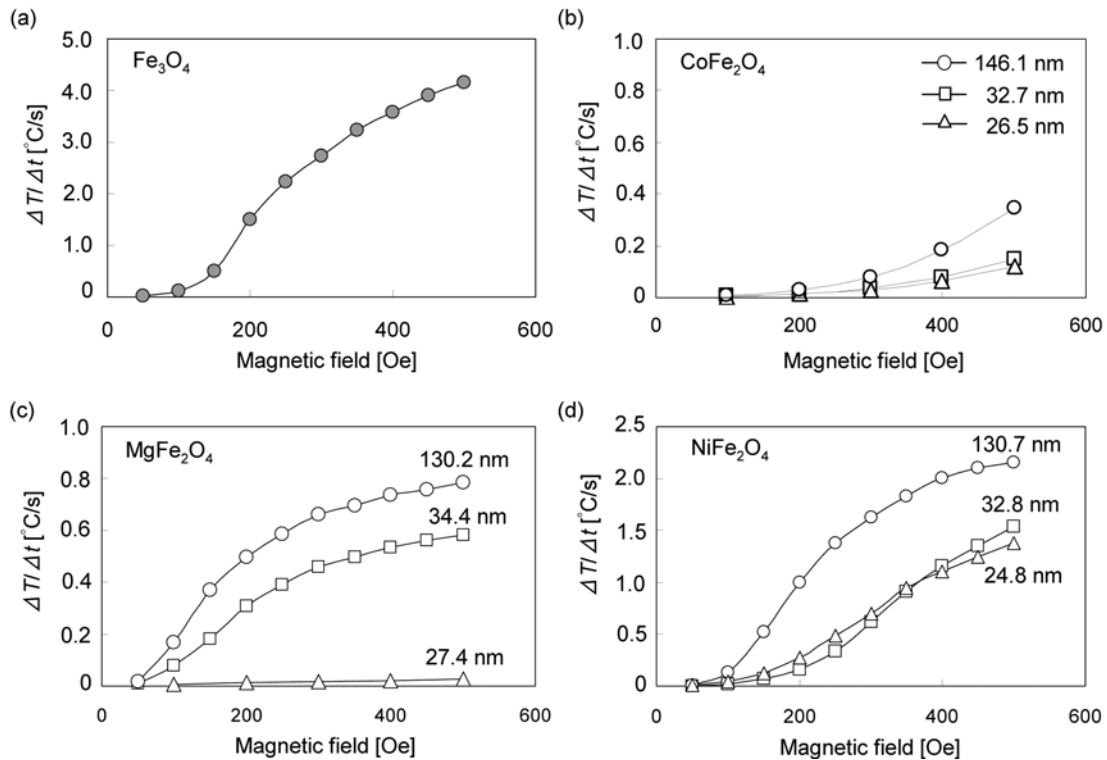


Fig. 3. Magnetic field strength dependence of induction heating. The induction heating was indicated by $\Delta T/\Delta t$ at $t \approx 0$, its initial slope for time dependence, where T and t are the measured temperature and time, respectively. The ac field frequency was 10 kHz and amplitude was varied from 50 to 500 Oe. (a) Fe_3O_4 , (b) CoFe_2O_4 , (c) MgFe_2O_4 , and (d) NiFe_2O_4 .

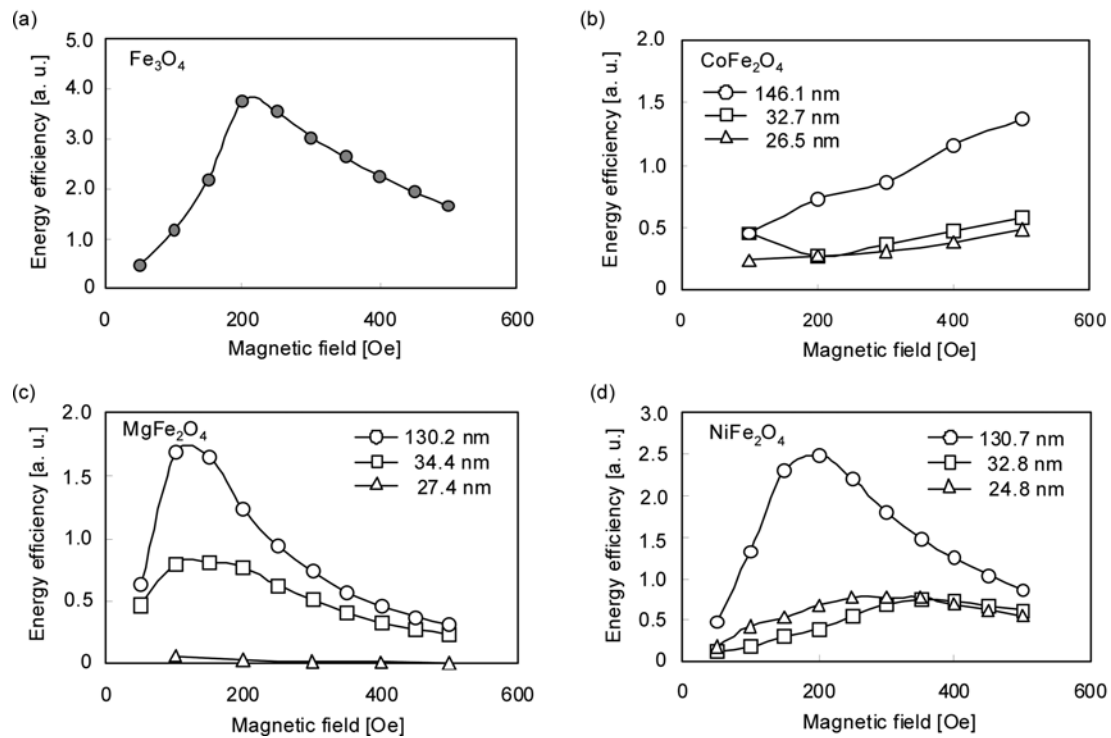


Fig. 4. Efficiency of applied energy utilized to generate induction heating, plotted as a function of applied magnetic field. The efficiency is defined by temperature rise divided by $H^2 \times f$, where H and f are the amplitude and frequency, respectively, of the applied ac magnetic field. (a) Fe_3O_4 , (b) CoFe_2O_4 , (c) MgFe_2O_4 , and (d) NiFe_2O_4 .

the CoFe_2O_4 nanoparticles (larger than 1000 Oe). Fig. 3(c) shows that the temperature rise of MgFe_2O_4 (130.2 nm) was higher than that of the 34.4 nm and 27.4 nm samples because of its higher magnetization and coercivity. In contrast, MgFe_2O_4 (27.4 nm) exhibited a lower temperature rise because of its extremely low coercivity (10 Oe). Fig. 3(d) shows that the temperature rise of NiFe_2O_4 (130.7 nm) was higher than that of the 32.8 nm and 24.8 nm samples, as in the case of MgFe_2O_4 . The temperature rise of the 32.8 nm NiFe_2O_4 was lower than that of the 24.8 nm NiFe_2O_4 when the field strength was less than 400 Oe and higher than that of the 24.8 nm NiFe_2O_4 when the field strength was greater than 400 Oe. This was because of the different coercive forces of 32.8 nm and 24.8 nm NiFe_2O_4 at 80 Oe and 130 Oe, respectively. The hysteresis losses of magnetic nanoparticles depend on their coercive forces. Energy efficiency of applied energy to generate induction heating is shown in Fig. 4. The energy efficiency is determined by the temperature rise divided by $H^2 \times f$, where H and f are the amplitude and frequency, respectively, of the applied ac magnetic field. The energy applied to generate a magnetic field is proportional to the product of H^2 and f . Fig. 4 shows that the optimum field strength of Fe_3O_4 (20-30 nm) is 200 Oe; 27.4 nm and 34.4 nm MgFe_2O_4 is 100-

150 Oe; and 24.8 nm, 32.8 nm, and 130.7 nm NiFe_2O_4 is 250-300 Oe, 350 Oe, and 150-200 Oe, respectively. The optimum field strength of CoFe_2O_4 samples could not be determined because the temperature rise of these samples continued within a field strength of 50-500 Oe. Nanoparticles with lower coercivity, such as Fe_3O_4 , 34.4 nm MgFe_2O_4 , and 130.7 nm NiFe_2O_4 , exhibited a high temperature rise at lower field strengths.

3.3. Cytotoxicity study

The viability of HeLa cells exposed to 200 $\mu\text{g}/\text{ml}$ of Fe_3O_4 (20-30 nm), CoFe_2O_4 (26.5 ± 6.1 nm), MgFe_2O_4 (27.4 ± 9.2 nm), and NiFe_2O_4 (20-30 nm) nanoparticles is shown in Fig. 5. The viability of HeLa cells exposed to NiFe_2O_4 nanoparticles is clearly shown to be extremely low as compared to that of cells exposed to the other nanoparticles. The relative ratio of HeLa cells exposed to NiFe_2O_4 nanoparticles for 2 days and 4 days is 52% and 8%, respectively. It is also found that only HeLa cells exposed to NiFe_2O_4 exhibited time dependence in terms of their cell number. The cells exposed to NiFe_2O_4 continued to decrease in number because the cell number did not increase, although the number of cells exposed to other nanoparticles remained unchanged for 4 days. Fe_3O_4 has been reported to be a biocompatible material [4, 15].

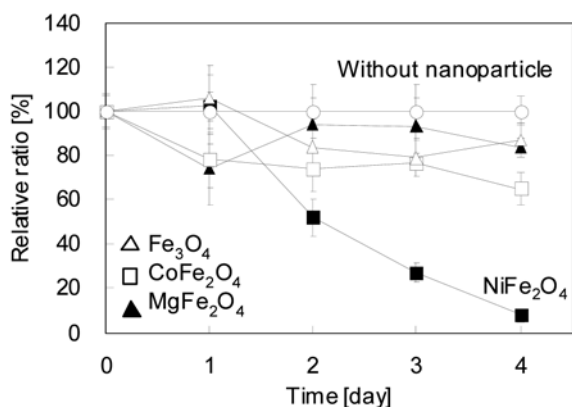


Fig. 5. Relative ratio of HeLa cells treated with and without magnetic nanoparticles at concentration of 200 $\mu\text{g/ml}$ for Fe_3O_4 (20-30 nm), CoFe_2O_4 (26.5 ± 6.1 nm), MgFe_2O_4 (27.4 ± 9.2 nm), and NiFe_2O_4 (20-30 nm) nanoparticles.

The high viability of the cells exposed to Fe_3O_4 conforms to these reports. There have been a few reports on the cytotoxicity of NiFe_2O_4 nanoparticles. Uncoated nickel ferrite particles (10 nm and 150 nm) did not affect the cell viabilities of mouse neuroblastoma cells [16]. However, in our study, NiFe_2O_4 nanoparticles greatly reduced the cell viability of HeLa cells. This discrepancy may have been caused by the difference in cell lines.

4. Conclusion

In this study, the magnetic characterization, thermal properties, and cytotoxicity of Fe_3O_4 , CoFe_2O_4 , MgFe_2O_4 , and NiFe_2O_4 nanoparticles were evaluated. Fe_3O_4 nanoparticles exhibited a high temperature rise and good biocompatibility. Thus, these nanoparticles are suitable for hyperthermia therapy. CoFe_2O_4 nanoparticles can be used as carriers for drug delivery systems because of their large magnetization, which is comparable to that of Fe_3O_4 particles, although they are not suitable for hyperthermia therapy because of their poor induction heating ability. MgFe_2O_4 nanoparticles are not suitable for biomedical applications because of their low magnetization compared to the other nanoparticles. NiFe_2O_4 nanoparticles exhibited a high temperature rise; however, they must be coated

with biocompatible materials because of their cytotoxicity to HeLa cells.

References

- [1] Q. A. Pankhurst, J. Connolly, S. K. Jones, and J. Dobson, *J. Phys. D: Appl. Phys.* **36**, R167 (2003).
- [2] H. Maeda, G. Y. Bharate, and J. Daruwalla, *Eur. J. Pharm. Biopharm.* **71**, 409 (2009).
- [3] H. Nathani and R. D. K. Misra, *Mater. Sci. Eng.* **94**, 228 (2004).
- [4] N. Sadeghiani, L. S. Barbosa, L. P. Silva, R. B. Azevedo, P. C. Morais, and Z. G. M. Lacava, *J. Magn. Magn. Mater.* **289**, 466 (2005).
- [5] T. Maehara, K. Konishi, T. Kamimori, H. Aono, H. Hirazawa, T. Naohara, S. Nomura, H. Kikkawa, Y. Watanabe, and K. Kawachi, *J. Mater. Sci.* **40**, 135 (2005).
- [6] S. Bae, S. W. Lee, and Y. Takemura, *Appl. Phys. Lett.* **89**, 252503 (2006).
- [7] A. Tomitaka¹, H. Kobayashi¹, T. Yamada¹, M. Jeun, S. Bae, and Y. Takemura, *J. Phys.: Conf. Ser.* **200**, 122010 (2010).
- [8] H. Kobayashi, A. Hirukawa, A. Tomitaka, T. Yamada, M. Jeun, S. Bae, and Y. Takemura, *J. Appl. Phys.* **107**, 09B322 (2010).
- [9] S. W. Lee, S. Bae, Y. Takemura, E. Yamashita, J. Kunisaki, S. Zurn, and C. S. Kim, *IEEE Trans. Magn.* **42**, 2833 (2006).
- [10] X. Huang and Z. Chen, *J. Magn. Magn. Mater.* **280**, 37 (2004).
- [11] V. Sepelak, I. Bergmann, D. Menzel, A. Feldhoff, P. Heitjans, F. J. Litterst, and K. D. Becker, *J. Magn. Magn. Mater.* **316**, e764 (2007).
- [12] A. Pardeep, P. Priyadharsini, and G. Chandrasekaran, *J. Magn. Magn. Mater.* **320**, 2774 (2008).
- [13] S. Balakrishnan, M. J. Bonder, and G. C. Hadjipanayis, *J. Magn. Magn. Mater.* **321**, 117 (2009).
- [14] M. Ma., Y. Wu, J. Zhou, Y. Sun, Y. Zhang, and N. Gu, *J. Magn. Magn. Mater.* **268**, 33 (2004).
- [15] P. Pradhan, J. Giri, G. Samanta, H. D. Sarma, K. P. Mishra, J. Bellare, R. Banerjee, and D. Bahadur, *J. Biomed. Mater. Res. Part B: Appl. Biomater.* **81B**, 12 (2007).
- [16] H. Yin, H. P. Too, and G. M. Chow, *Biomaterials* **26**, 5818 (2005).