

유근피 추출물의 피부개선효과

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Anti-Wrinkle Effect of *Ulmus davidiana* Extracts

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ABSTRACT : The bark of the root and stem of *Ulmus davidiana* var. *japonica* has been used as a traditional Korean medicine to treat inflammatory disorders. This plant reportedly shows antioxidant, anticancer, and anti-inflammatory effects. In this study, we investigated the protective effects of *Ulmus davidiana* var. *japonica* ethanolic extract (UDE) on UVB irradiation-induced wrinkle in hairless mice. We evaluated for their free radical-scavenging activities against 1,1-diphenyl-2-picrylhydrazyl (DPPH) and the anti-elastase activities, and for their anti-matrix metalloproteinase-1 (MMP-1) activity in human skin fibroblast cells. In the wrinkle measurement and image analysis of skin replicas, the results showed that UDE significantly inhibited wrinkle formation caused by chronic UVB irradiation. These results suggest that UDE has anti-wrinkle activity.

Key Words : Anti-Wrinkle, *Ulmus davidiana*, UVB, DPPH, MMP-1

INTRODUCTION

Many skin changes are mere cutaneous senescence and cumulative environmental insults (Zimble *et al.*, 2001). The relationship observed between sun exposure and decreased skin elasticity (Takema *et al.*, 1998). Available evidence suggests that there are at least two types of elastases in the skin, neutrophil elastase and skin fibroblast elastase (Godeau *et al.*, 1988). The overproduction of elastases induced by ultraviolet (UV) irradiation affects the elastic-fiber network. The fact that the exposure of animal skin to UV light at less than a suberythral dose also causes wrinkles, despite the lack of inflammatory cell infiltration including neutrophils (Learn *et al.*, 1991).

When the skin is exposed to UV or visible light, it leads to wrinkle formation are recognized as major environmental factors deleterious skin (Ha *et al.*, 2010). It's changes in the dermal elastic fibers attributed to a loss of linearity and curling

(Imokawa *et al.*, 1995). UV irradiation-induced injury to the skin can be photodamage (Farkas *et al.*, 2002).

The effects of UV on the skin is the use of antioxidants scavenging and quenching reactive oxygen species (ROS) (Bae *et al.*, 2009). Oxidative stresses can be generated in the connective tissues and the skin cells by photodamage and inflammatory processes (Jackson *et al.*, 1999)

Excessive MMP activity, which causes the collapse of the meshwork in the extracellular matrix, produces UV irradiation-like skin damage, including wrinkling, loss of elasticity, and dilation of surface microcapillary vessels (Bologna, 1993)

Collagen and elastin provide suppleness and elasticity to the skin and reinforce the fibers of the two fundamental elements which constitute the supporting capacity of the cutaneous layer. Collagen and elastin contribute to form a network under the epidermis and significantly reduce the lines and wrinkles (Berthod *et al.*, 2001). Elastase is an enzyme associated with the

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deterioration of collagenous fibers, keratins and proteoglycans, important components of the skin matrix (Wiedow *et al.*, 1990). Its activity may be negatively regulated by a family of naturally occurring tissue inhibitors (Murphy *et al.*, 1992). Therefore, inhibition of elastase activity could be an excellent method for protection against skin aging (Jeong *et al.*, 2009).

UDE is a deciduous tree which is widely distributed in Korea and has been used for treatment of edema, mastitis, gastric cancer and inflammation in oriental medicine (Jin *et al.*, 2006). The UDE also has analgesic and antileukocyte migration (Hong *et al.*, 1990) and antioxidative activity (Han *et al.*, 2006).

Then, we hypothesized that UDE is a skin care cosmetics agent because of having antioxidative activity. Therefore, the main purpose of this study was to determine the biological activities such as antioxidant, anti-elastase, and anti-wrinkle activity of the UDE *in vivo* for further industrial applications.

MATERIALS AND METHODS

1. Preparation of UDE

UD (*Ulmus davidiana*) was collected from Kunsan in Korea (August 2004). A voucher specimen (HPR-207) was deposited at the herbarium of Herbal Crop Research Institute (Eumsung, Republic of Korea). *Ulmus davidiana* extract (UDE) was prepared as dried powder from the consecutive extractions with diluted ethanol, and provided by Kangwon National university. Specifically, UD (100 g) was extracted with 600 mL of 55% ethanol at 50-53°C for 24 h with stirring. The extraction procedure was repeated three times. The pooled filtrates were subjected to vacuum evaporation at 50-60°C to yield 50 g of solid content, which was further purified with 500 mL of 88% ethanol at 5-10°C overnight and centrifuged at 5000 × g for 10 min. The supernatants were evaporated and spray dried to yield 17 g of UDE.

2. Cell Culture

Human dermal fibroblasts (HDFs, derived from newborn skin) were purchased from American Type Culture Collection (ATCC). Fibroblast cells were grown in Dulbecco's modified Eagle's medium (DMEM; Hyclone) supplemented with 10% fetal bovine serum (FBS; Gibco) and antibiotics (100 U/ml of penicillin and 100 µg/mL of streptomycin). Cells were plated in 75 cm² culture flasks and placed in an incubator at 37°C with a humidified atmosphere containing 5% carbon dioxide. When the cells reached 80-90% confluence, they were subcultivated to 60

mm culture dishes.

3. Determination of Anti-oxidant Activity

The DPPH assay was performed as described previously (Lee *et al.*, 2006). The antioxidant reaction was carried out in 99.8% ethanol containing 0.1 mM/L DPPH and UDE. The scavenging effect against DPPH radical was assessed at room temperature for 10 min. The change in the absorbance at 520 nm as measured in a 96-well reader.

4. Determination of MMP-1 Secretions by ELISA

HDFs were seeded in 100 mm culture dishes at density of 2×10^6 cells per dish, and then irradiated with UVB (25 mJ/cm²). Following 24 h of incubation, the culture supernatants were collected and centrifuged at 10,000 × g for 5 min to remove the particulate matter, and stored at -80°C in fresh tubes. In supernatants, the protein concentration was determined using the Bradford method. The active MMP-1 in culture supernatants were quantified by fluorescent assay, using the Fluorokine E Human Active MMP-1 Fluorescent Assay Kit (R&D Systems, Minneapolis, USA) in the cell culture supernatants was then determined using Quantikine ELISA kits (R&D Systems, Minneapolis, USA), according to the manufacturers protocol.

5. Elastase Inhibition Activity

Elastase inhibition activity was determined by the method of Tschesche and colleagues (Tschesche *et al.*, 1992). In detail, 0.1 mL of a 0.2 M Tris-HCl buffer (containing 1% albumin), 0.025 mL of a substrate solution [10 mM MAAPVN(*N*-(methoxysuccinyl)-ala-ala-pro-val 4-nitroanilide)], and 0.05 mL of a sample were mixed, and then 0.025 mL of elastase (3 units/mL, Green Elastase Assay Kit) was added. The reaction mixtures were incubated in a 25°C water bath for 20 minutes, and the inhibition rate was measured by an enzyme-linked immunosorbent assay (ELISA) reader (Molecular Devices Toronto, Canada). Inhibition rate (%) = $[1 - (C - D) / (A - B)] \times 100$, where A indicates the absorbance at 410 nm without a test sample after incubation, B indicates the absorbance at 410 nm without a test sample before incubation, C indicates the absorbance at 410 nm with a test sample after incubation, and D indicates the absorbance at 410 nm with a test sample before incubation.

6. Mice

Male SKH-1 hairless mice (20 g each, SLC, Japan,) were acclimated for 1 week under standard laboratory conditions at room temperature of 22-26°C and humidity of 45-55% under a

12-h light/12-h dark cycle. The mice had free access to tap water and to a commercial standard mice chow throughout the experimental period. This study was conducted according to the “Guiding Principles for the Care and Use of Laboratory Animals”, and all procedures were approved by the Animal Care and Use Committee of Kyung Hee University Medical Center.

7. UV Irradiation

UVB was supplied by an array of five G5T5 Sankyo Denki sunlamps containing 30% UVA (Kanagawa, Japan). UVB radiation was applied to the backs of the mice three times a week for 8 weeks. The amount of irradiation was progressively increased, from 100 mJ/cm² per exposure at week 1 (1 minimal erythematous dose = 100 mJ/cm²) to 400 mJ/cm² at week 8.

8. UDE Treated

The mice were divided into three groups (n=10) using a randomized block design in accordance with body weight. Each mouse in the group was topically applied for 300 mg/ml of UDE daily for 8 weeks. The control group was treated with 300 µL distilled water.

9. Wrinkle Measurement

After 8 weeks, impressions were made of the back skin of nine unrestrained mice, using Aphrodite-III (Enhanced Image Technologies, LLC, Dallas, USA). We set the impression of wrinkles on the sample stand so that the measurement surface was wrinkled. Dorsal skin wrinkling caused by chronological suberythemal dose UVB exposure was graded each week, as described (Bissett et al., 1990). As follows: grade 0, no coarse wrinkles; grade 1, a few shallow coarse wrinkles; grade 2, some coarse wrinkles; grade 3, deep coarse wrinkles. Image analysis of wrinkles was performed on a 10 X10 mm area. The percentage of area of wrinkles in the image analysis area was then calculated as previously described (Takema et al., 1997). That percentage has been established to be closely related to wrinkle scores (Imokawa et al., 1993).

10. Statistical Analysis

The wrinkle grading score and wrinkle area (%) were expressed as mean±SD which were made in triplicate experiments. The paired t-test was used for comparisons between control group and UDE treated group. All analyses were performed using an SPSS 16 system (SPSS institute, Chicago, IL, USA).

RESULTS AND DISCUSSION

In this study, topical application of UDE, which inhibits skin fibroblast-derived elastase, decreased UVB-induced wrinkle formation (evaluated by visible score and replica image) in mouse dorsal skin.

1. Free radical scavenging activity

It has been reported that free radical scavenging capacity ageing (Guanglong et al., 2004). Assays of the free radical scavenging capacity were carried out by the DPPH method. The results being shown in Table 1. The free radical scavenging capacity is expressed as SC₅₀, the concentration needed to reduce 50% of DPPH radical. UDE had the radical scavenging activity (SC₅₀=8.6 µg/ml). This extract showed very high free radical scavenging activity compared to BHT (di-*t*-hydroxytoluene: SC₅₀=28.6 µg/ml) which was used as a positive control.

2. Inhibition of the elastase activity

Table 2 showing the results of elastase activities. UDE was found to have the elastase inhibition activity (IC₅₀=54.7 µg/ml) compared to oleanolic acid (IC₅₀=85.4 µg/ml) which was used as a positive control. This result suggested that UDE would have potential as an anti-wrinkle agent for use in cosmetic products.

3. UDE inhibites the UVB- induced MMP-1 expression and secretions in HDFs

UVB activates the secretion of MMPs, which is the hallmark

Table 1. Free radical scavenging activity of UDE.

	DPPH radical scavenging activity (%)				SC ₅₀ (µg/ml)*
	100 µg/ml	50 µg/ml	10 µg/ml	1 µg/ml	
UDE	–	92.53±0.79	75.4±14.31	9.16±0.6***	8.55±1.19****
BHT ^b	86.56±6.66	73.07±7.87*	35.75±4.82***	9.4±1.97***	7.1±4.0***

*SC₅₀ indicates the concentration (µg/ml) at which the percentage inhibition of DPPH radical scavenging activity was 50%. BHT^b (di-*t*-butyl hydroxyl toluene). **Those with different superscript letters are significantly at *p < 0.05, *** p < 0.001.

Table 2. Elastase inhibition activity of UDE.

	Inhibiton (%)			IC ₅₀ ($\mu\text{g}/\text{ml}$)*
	100 $\mu\text{g}/\text{ml}$	50 $\mu\text{g}/\text{ml}$	10 $\mu\text{g}/\text{ml}$	
UDE	78.57 \pm 6.09	61.25 \pm 16.72	10.33 \pm 0.47*,**	54.6 \pm 23.27
Oleanolic acid	64.6 \pm 9.87	48.55 \pm 16.91*	6.73 \pm 3.79***	85.42 \pm 8.44

*IC₅₀ indicates the concentration ($\mu\text{g}/\text{ml}$) at which the percentage inhibition of elastase activity was 50%. **Those with different superscript letters are significantly at *p < 0.05. ***p < 0.001.

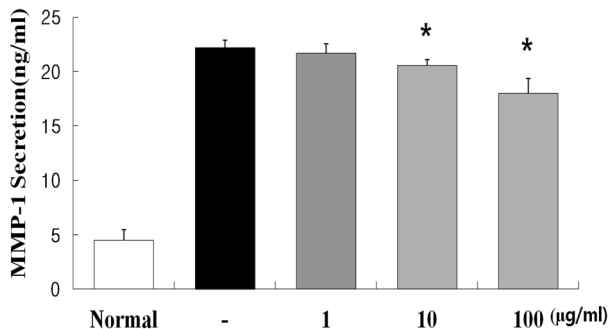


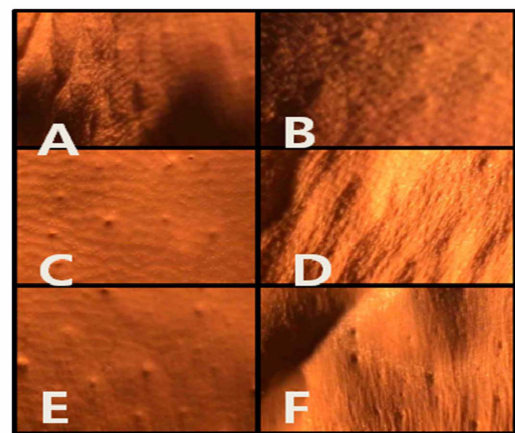
Fig. 1. Effect of UDE on UVB-induced MMP-1 expressions in HDFs. Cells were stimulated with UVB (25 mJ/cm²) and the indicated concentrations of UDE for 24 h. The presence of MMP-1 in the cell-free culture supernatants was measured using a commercially available ELISA kit as described in Materials and Methods. Each value represents the mean \pm SD of three independent experiments. *p < 0.05 (vs. control control + UDE)

of skin aging (Pillai *et al.*, 2005). We examined effects of UDE on UVB-induced MMP expression. UDE reduced MMP-1 protein expression in irradiation of HDFs with UVB (25 mJ/cm²). We also determined the effect of UDE on UVB-induced MMP secretion by ELISA. UVB irradiation of HDFs resulted in an increase in the secretion of MMP-1, and UDE significantly diminished the UVB-induced MMP-1 secretions (Fig. 1). UDE itself had no effects on expression and secretion of MMP-1 in HDFs. This result indicates that UDE inhibits UVB-induced MMP-1 expressions and secretions in HDFs.

4. Inhibitory activity of UDE on wrinkle formation

As early as 4 weeks after UV irradiation there were visible signs of wrinkling on the dorsal skin of hairless mice, and the wrinkles became distinct at week 8 of irradiation in contrast to the absence of wrinkle formation in the age-matched unirradiated controls (Fig. 2A, B). When UDE was topically applied daily for 8 weeks at a concentration of 300 mg/ml to dorsal skins of hairless mice immediately after each suberythral UV irradiation, the formation of wrinkles was obviously diminished compared with the water-treated controls (Fig. 2A). Comparison

A



B.

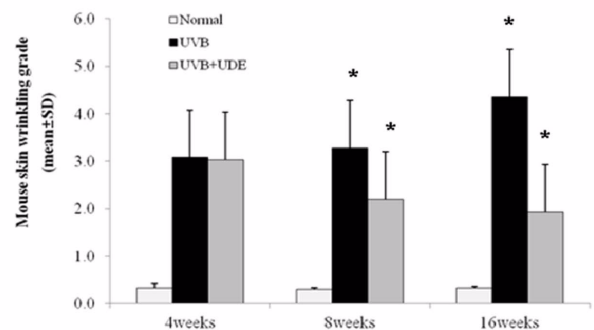


Fig. 2. Effects of UDE treated mice with UV B on skin wrinkles. (A) Photographs after UVB irradiation (three times a week for 8 weeks.) of hairless mouse skin at last day after UVB irradiation. A, C, E : UVB (8 weeks) +UDE (topically application for 8 weeks) , B, D, F : UVB irradiation (16 weeks, No application of UDE). (B) Visual scoring (scale 1/4 0-3) of wrinkles during UVB exposure of mice. The scoring was performed by the method of Bissett *et al.*, after UVB irradiation. Data represent mean \pm SD. *p < 0.05 (vs. control control + UDE)

of wrinkle scores revealed that UDE significantly decreased wrinkle formation by 13-16 weeks of irradiation compared with the water-treated controls (Fig. 2B). UVB radiation is one of the most environmental hazardous effects that can cause acute and chronic response in human skin (Bissett *et al.*, 1990) and

stimulate synthesis of elastin, which is highly related to the elasticity and renewal of skin, and induces wrinkles and a lack of elasticity (Schwartz *et al.*, 1995). The degeneration of elastic fibers is estimated to be due to an increase in fibroblast elastase, an enzyme produced and secreted by dermal fibroblasts that degrades elastin (Tsukahara *et al.*, 2001). The connective tissue elements collagen and elastin as known to be degraded by collagenase and elastase produced by inflammatory cells (Hastly *et al.*, 1982) and fibroblast (Welgus *et al.*, 1982). Moloney *et al.* (1992) reported that visible signs of wrinkling were occur after approximately six weeks of UVB irradiation and were very obviously after ten weeks of irradiation. The progress of wrinkle formation noticed in our experiment was similar to that in the above report. The timing of the inhibition of wrinkles seemed to be similar. In the present study, comparison of skin properties between UDE treated group after UVB irradiation and sham-operated hairless mice without UVB irradiation demonstrated that UDE treated group at week 4 appeared no difference in skin elasticity compared with sham-operated group. The reduced skin elasticity in the UVB irradiation group is accompanied by a significant increase in the activity of elastase in the skin. In addition, during the experiments with UVB irradiation, redness and exfoliation were observed in the skin of many mice only in the UVB irradiation.

Our study demonstrated that elastase activity in UVB-irradiated mouse skin, was not changed following 4 weeks of UVB irradiation, but was markedly increased following 8 weeks of UV irradiation. Our study observations demonstrated that fine elastic fibers were markedly decreased after 16 weeks of UVB irradiation (Fig. 2A.).

Photoaging (UV irradiation) causes human skin aging through activation of MMPs, which are responsible for the degradation of collagen. It is conceivable that the degeneration of elastic fibers observed in the UVB exposed skin is mainly due to the increased activity of skin fibroblast elastase. In this report, the wrinkle formation by the elastase inhibitor UDE was accompanied by a marked decrease in degradation of the dermal fine elastic-fiber network.

In Korea, UDE has been used to treat various skin problems, including atopy, skin aging, and pimples. Eom *et al.* (2006) studied that UDE recovered from photo-induced damage after UVA irradiation (3 J/cm²) with 2 times higher than that of positive control and UDE showed approximately 48% of the increased cell viability of the control. UDE has been popular in cosmetic compositions and found to have beneficial functions similar to

those investigated in this study. However, no side effects were reported until now. Therefore, the UDE, a natural agent for human skin, would have more benefits than chemical treatments.

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