pISSN 1598-2629 eISSN 2092-6685

Stimulatory Effect of β -glucans on Immune Cells

Hyung Sook Kim, Jin Tae Hong, Youngsoo Kim and Sang-Bae Han*
College of Pharmacy, Chungbuk National University, Cheongju 361-763, Korea

 β -Glucans are naturally occurring polysaccharides that are produced by bacteria, yeast, fungi, and many plants. Although their pharmacological activities, such as immunomodulatory, anti-infective and anti-cancer effects, have been well studied, it is still unclear how β -glucans exert their activities. However, recent studies on the β -glucan receptors shed some light on their mechanism of action. Since β -glucans have large molecular weights, they must bind surface receptors to activate immune cells. In this review, we summarize the immunopharmacological activities and the potential receptors of β -glucans in immune cells.

[Immune Network 2011;11(4):191-195]

CHEMISTRY OF β -GLUCANS

 β -Glucans are heterogeneous polysaccharides of glucose polymer, consisting of a backbone of β -(1-3)-linked β -D-glucopyranosyl units with β -(1-6)-linked side chains of varying distribution and length. The activity of β -glucan depends on the molecular structure, size, branching frequency, structural modification, conformation, and solubility. It appears that the most active forms of β -glucans contain β -(1-3)(1-6) linkages (1). The structure of several biologically active β -glucans has been reported. β -Glucan from many mushrooms has a β -(1-3) backbone with shorter β -(1-6) linked branches, while β -glucan from Alcaligenes faecalis contains only β -(1-3)-glucosidic linkages (2). Schizophyllan from Schizophyllum commune and scleroglucan from Sclerotium glucanicum both have a β -(1 \rightarrow 3) linked backbone with one β -(1 \rightarrow 6)-glucose substitution every three backbone residues (3). Lentinan from Lentinus edodes has a β -(1 \rightarrow 3) linked backbone and two β -(1 \rightarrow 6) side chains every five residues (4). β -Glucan from oat and barley are linear with β -(1-4) linkage with shorter stretches of β -(1-3) (3).

Biologically active β -glucans usually have a large molecular weight. However, it is unclear whether β -glucans having intermediate or small molecular weight have biological activities, although some of them are active *in vivo*. Short β -glucans below 5,000-10,000 Da of molecular weight are generally inactive (5). The optimal branching frequency is suggested as 0.2 (1 in 5 backbone residues) to 0.33. Although unbranched β -glucan curdlan showed proper biological activity, chemical addition of β -(1-6) glucose residues to the curdlan backbone led to an increase in antitumor activity (6), as highly branched β -glucan has higher affinity for cognate receptors (7). Furthermore, soluble β -glucans appear to be stronger immunostimulators than insoluble ones. When insoluble scleroglucan is modified by sulfation or carboxymethylation, the antitumor activity increases (8).

Further study is still required before we will fully understand the structure-activity relationship in β -glucans. Orally administered β -glucans may be modified to smaller oligosaccharides *in vivo* (3). Thus, the actual β -glucans binding to the immune cell surface receptors *in vivo* may in fact be these smaller ones. However, there is no information on this topic to date. If we can use standardize smaller β -glucans, the biological data might be fruitful,

IMMUNOPHARMACOLOICAL ACTIVITY OF β -GLUCANS

 β -Glucans, generally called biological response modifiers, are now recognized as anti-tumor and anti-infective drugs. The most popular β -glucan is lentinan, which is isolated

Received on July 8, 2011. Revised on July 14, 2011. Accepted on July 18, 2011.

Keywords: β -glucan, Receptors, Immune cells

[©] This is an open access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

^{*}Corresponding Author. Tel: 82-43-261-2815, Fax: 82-43-268-2732; E-mail: shan@chungbuk.ac.kr

from fruiting bodies of *Lentinus edodes* is the most popular β -glucan and is a well-known drug with anti-tumor and anti-infective activities (9). β -Glucan has been shown to protect against infection by bacteria, viruses, and pathogenic microorganisms (10). β -Glucan also prevents cancer promotion and progression and has synergistic antitumor effects with monoclonal antibodies and cancer chemotherapeutics (11). β -Glucan promotes antibody-dependent cellular cytotoxicity through a biological pathway involved in carcinogenesis (12). However, lentinan might not directly affect cancer cells or infectious microorganisms. There are no reports on the direct effect of lentinan on these cells. Instead, it is believed that lentinan shows these biological activities through activation of host immune systems,

The effects of β -glucans on immune cells are well established. Traditionally, macrophages and dendritic cells are considered the main target cells of β -glucans, although neutrophils, B cells, T cells, and natural killer cells are also known to be activated by β -glucan. The immunomodulatory activities of β -glucans are usually studied with regard to the activation of macrophages. Lentinan enhances cytotoxic activity and inflammatory cytokines of primary macrophages and RAW264.7 cell lines (13). It can also enhance the phenotypic and functional maturation of dendritic cells with significant IL-12 production (14). Stimulatory effects of lentinan on T cells have also been reported. Lentinan enhances the virus-specific T cell functions induced by DNA vaccine, acts as a vaccine adjuvant (15), and increases T cell functions in tumor-bearing mice (16) and malaria-infected mice (10). In addition, lentinan is reported to enhance T cell functions in cancer patients (17). However, there is no report showing that lentinan directly activates T cells in vitro. It has been demonstrated that lentinan indirectly activates T cells via IL-12 and IFN- γ produced by lentinan-activated macrophages and dendritic cells (18). However, lentinan might be an indirect activator of T cells and T cell activation might be observed only under in vivo conditions with mixed immune cell subsets. It has been reported that lentinan increases NK cell-mediated killing of Yac-1 cells both in vitro and in vivo (19). However, this does not necessarily mean that lentinan directly activates NK cells, since total spleen cells were used in this experiment. The only clear fact is that β -glucan directly activates macrophages and dendritic cells, but the effect of β -glucan on other immune cells remains controversial. Further in vitro studies with purified immune cell subsets are required to clarify whether β -glucan directly activates these cells.

B-GLUCAN RECEPTORS

Macrophages and dendritic cells have typical cell surface receptors called pattern recognition receptors (PRRs) that detect innately non-self molecules including pathogen-associated molecular patterns (PAMPs) (20). β -Glucans might act as PAMPs and are recognized by PRRs, since β -glucans cannot directly penetrate cell membrane due to their large molecular size (3). The major PRRs for β -glucans might be dectin-1 and the roll-like receptor (TLR). Upon binding with β -glucan, dectin-1 and TLR might inducing signaling cascade and activate immune cells. Other receptors, such as complement receptor 3 (CR3), scavenge receptors (SR), and lactosylceramide (LacCer), might be involved (20).

DECTIN-1

Dectin-1 is a lectin that consists of four components: an extracellular carbohydrate-recognition domain, a short stalk region, a single transmembrane region, and a short 40 amino acid intracellular cytoplasmic tail (21). Dectin-1 consists of 244 amino acids and has six cysteine residues. In particular, two amino acids (Trp221 and His223) located near the fourth cysteine residue appear to be critical for binding of β -glucans (22). Dectin-1 specifically recognizes β -(1-3)(1-6) glucans from fungi, plants, and bacteria (23). However, it is not reactive toward β -(1-4) glucans or α -mannan (24).

Binding of dectin-1 with β -glucans induces several signaling pathways to activate innate immune responses, such as phagocytosis, ROS production, and inflammatory cytokine production (25). The cytoplasmic tail of dectin-1 has an immunoreceptor tyrosine-based activation (ITAM)-like motif (YxxxI/Lx7YxxL) to activate tyrosine kinases (26). Upon ligand binding, tyrosines in the ITAM sequences are phosphorylated by Src family kinases, providing a docking site for Syk (spleen tyrosine kinase) by interacting with the two SH2 (Src homology 2) domains of Syk (26). The spacing between the YxxxL sequences is important to engage both SH2 domains of Syk family kinase, thus contributing to enzyme activation (27). Activated phospholipase C γ (PLC γ) produces inositol trisphosphate and diacylglycerol (DAG) (28). Also, Syk activates the PI3K/Akt pathway, MAPK, NFAT, and NF- κ B (29).

TLRs

TLRs are expressed on macrophages, dendritic cells, B cells, T cells and endothelial cells and are type I transmembrane receptors of a novel protein family. At least 13 members of this family exist in human. TLRs can recognize diverse microbes including fungi, bacteria, viruses and protozoa. Several ligands have been shown to bind TLRs: TLR2-zymosan, TLR3-dsRNA, TLR4-LPS, and TLR5-Flagellin, etc. (3). Binding of specific ligands to TLRs induces several signaling pathways, such as My88-mediated signaling and TRIF-mediated signaling. TLR signaling usually results in activation of NF- κ B and MAPK signalings (30). There are many β -glucans that can bind to TLRs. For example, zymosan binds to TLR2/4 of macrophages and increases the cytokine production such as TNF- α and IL-12 via NF- κ B signaling (31). β -Glucans isolated from plants, such as Sparassis crispa, Phellinus linteus, Platycodon grandiflorum, Cordyceps millitaris, and Angelica gigas Nakai, induce dendritic cell maturation through binding to TLR4 (32-37). Signalings downstream from TLRs or dectin-1 might cross-talk with each other (38). Zymosan can bind both dectin-1 and TLR2, and both dectin-1/Syk and TLR/Myd88 signalings are required to fully induce the translocation of NF- κ B subunits to the nucleus (39).

OTHER RECEPTORS

First identified over 25 years ago, CR3 can recognize carbohydrates (40). CR3 acts as an opsonic receptor for the complement component and as a nonopsonic receptor for a variety of exogenous ligands. CR3 is a heterodimeric transmembrane integrin consisting of CD11b ($\alpha_{\rm m}$) and CD18 ($\beta_{\rm 2}$) chains. CD11b has two binding sites. One for β -glucan is located within the C terminus, while the other for iC3b (cleaved component 3 fragment of serum complement system) is located within the N-terminus (40). Binding of β -glucan to the C-terminal lectin domain increases adhesion to microbial cells and activates iC3b pathways causing tumor cytotoxicity (41). Ligand binding of CR3 is known to mediate intracellular signaling and induce a variety of cellular responses, including adhesion, cytotoxicity, phagocytosis and migration (41). However, whether CR3 directly binds to β -glucan is still unclear. CR3-deficiency in NK cells reduced cytotoxicity, but CR3-deficient leukocytes are still capable of recognizing and responding to β -glucan (42). The β -glucan of small molecular size binds to CR3 in NK cells and that of large molecular size binds dectin-1 and TLRs in macrophages and dendritic cells

LacCer (CDw17 and Gal4Glc1Cer) is expressed on neutrophils and endothelial cells. LacCer recognizes a variety of microbes and pathogens, including fungi, such as *Candida albicans*, *Cryptococcus neoformans*, and *Saccharomyces cerevisiae* (43). It consists of a hydrophobic ceramide and a hydrophilic sugar moiety and is identified as a receptor for β -glucan (44). The interaction of β -glucan with LacCer induces a number of cellular responses *in vitro*. In alveolar neutrophils, β -glucan from *Pneumocystis carinii* can induce the production of macrophage inflammatory protein-2 (MIP-2) and TNF- α via NF- κ B and PKC signaling. Blocking antibodies to LacCer or CR3 are also reported to inhibit β -glucan binding to and the activation of human neutrophils (45).

SR are expressed on epithelial cells, endothelial cells and myeloid cells and comprise a family of proteins that are structurally diverse and have a range of cellular functions (46). SR were initially described in cultured macrophages in which they mediate cholesterol uptake. Based on their structures, SR can be categorized into class A, B and C. SR-A has a collagen-like domain, which is essential for ligand binding (47). SR recognizes a variety of ligands including LDL (low-density lipoprotein), HDL (high-density lipoprotein), selected polyanionic molecules, and a number of microbes (47). SR has also been implicated in the recognition of β -glucan. Lentinan can bind to SR and activate multiple signals, such as PI3K, Akt kinase, and MAPK (47).

SUMMARY

 β -Glucans are potent immunomodulators that have multiple activities such as anti-tumor and anti-infective activities. However, how β -glucan exerts these diverse biological activities is still unknown. The first step mediating β -glucan action might be immunostimulation. In particular, binding of β -glucan to specific receptors in macrophages and dendritic cells can induce the production of various cytokines, indirectly activating other immune cells such as T cells and B cells under *in vivo* conditions. Systemic immunostimulation might be the main route in preventing the growth of cancer cells and infective microorganisms in the host. Several β -glucan receptors in macrophages and dendritic cells, such as dectin-1 and TLRs, might play a key role in the recognition of β -glucans, but the exact signaling pathways downstream

from the respective receptors and the cross-talk between them is unclear to date. If we can understand these issues in greater detail, β -glucans might be widely used in the therapy of cancer and infectious diseases.

ACKNOWLEDGEMENTS

This work was supported by the research grant of the Chungbuk National University in 2010.

CONFLICTS OF INTEREST

The authors have no financial conflict of interest.

REFERENCES

- Demleitner S, Kraus J, Franz G: Synthesis and antitumour activity of sulfoalkyl derivatives of curdlan and lichenan. Carbohydr Res 226;247-52, 1992.
- McIntosh M, Stone BA, Stanisich VA: Curdlan and other bacterial (1-->3)-beta-D-glucans. Appl Microbiol Biotechnol 68; 163-73, 2005.
- Chen J, Seviour R: Medicinal importance of fungal beta-(1->3), (1->6)-glucans. Mycol Res 111;635-52, 2007.
- Suzuki M, Takatsuki F, Maeda YY, Hamuro J, Chihara G: Antitumor and immunological activity of lentinan in comparison with LPS. Int J Immunopharmacol 16;463-8, 1994.
- Zhang L, Li X, Xu X, Zeng F: Correlation between antitumor activity, molecular weight, and conformation of lentinan. Carbohydr Res 340;1515-21, 2005.
- Jamois F, Ferrières V, Guégan JP, Yvin JC, Plusquellec D, Vetvicka V: Glucan-like synthetic oligosaccharides: iterative synthesis of linear oligo-beta-(1,3)-glucans and immunostimulatory effects. Glycobiology 15;393-407, 2005.
- Mueller A, Raptis J, Rice PJ, Kalbfleisch JH, Stout RD, Ensley HE, Browder W, Williams DL: The influence of glucan polymer structure and solution conformation on binding to (1-->3)-beta-D-glucan receptors in a human monocyte-like cell line. Glycobiology 10;339-46, 2000.
- Wang Y, Zhang L, Li Y, Hou X, Zeng F: Correlation of structure to antitumor activities of five derivatives of a beta-glucan from Poria cocos sclerotium. Carbohydr Res 339;2567-74, 2004.
- Chihara G, Hamuro J, Maeda Y, Arai Y, Fukuoka F: Fractionation and purification of the polysaccharides with marked antitumor activity, especially lentinan, from Lentinus edodes (Berk.) Sing. (an edible mushroom). Cancer Res 30;2776-81, 1970.
- Zhou LD, Zhang QH, Zhang Y, Liu J, Cao YM: The shiitake mushroom-derived immuno-stimulant lentinan protects against murine malaria blood-stage infection by evoking adaptive immune-responses, Int Immunopharmacol 9;455-62, 2009.
- 11. Harada K, Itashiki Y, Takenawa T, Ueyama Y: Effects of lentinan alone and in combination with fluoropyrimidine anti-

- cancer agent on growth of human oral squamous cell carcinoma in vitro and in vivo. Int J Oncol 37;623-31, 2010.
- Sier CF, Gelderman KA, Prins FA, Gorter A: Beta-glucan enhanced killing of renal cell carcinoma micrometastases by monoclonal antibody G250 directed complement activation. Int J Cancer 109;900-8, 2004.
- Kerékgyártó C, Virág L, Tankó L, Chihara G, Fachet J: Strain differences in the cytotoxic activity and TNF production of murine macrophages stimulated by lentinan. Int J Immunopharmacol 18;347-53, 1996.
- Chan WK, Law HK, Lin ZB, Lau YL, Chan GC: Response of human dendritic cells to different immunomodulatory polysaccharides derived from mushroom and barley. Int Immunol 19;891-9, 2007.
- Wang J, Dong S, Liu C, Wang W, Sun S, Gu J, Wang Y, Boraschi D, Qu D: beta-Glucan oligosaccharide enhances CD8(+) T cells immune response induced by a DNA vaccine encoding hepatitis B virus core antigen. J Biomed Biotechnol 2010;645213, 2010.
- McCormack E, Skavland J, Mujic M, Bruserud Ø, Gjertsen BT: Lentinan: hematopoietic, immunological, and efficacy studies in a syngeneic model of acute myeloid leukemia. Nutr Cancer 62;574-83, 2010.
- Yoshino S, Tabata T, Hazama S, Iizuka N, Yamamoto K, Hirayama M, Tangoku A, Oka M: Immunoregulatory effects of the antitumor polysaccharide lentinan on Th1/Th2 balance in patients with digestive cancers. Anticancer Res 20;4707-11, 2000
- Murata Y, Shimamura T, Tagami T, Takatsuki F, Hamuro J: The skewing to Th1 induced by lentinan is directed through the distinctive cytokine production by macrophages with elevated intracellular glutathione content. Int Immunopharmacol 2:673-89, 2002.
- Vetvicka V, Vetvickova J, Frank J, Yvin JC: Enhancing effects of new biological response modifier beta-1,3 glucan sulfate PS3 on immune reactions. Biomed Pharmacother 62;283-8, 2008
- Brown GD, Gordon S: Immune recognition of fungal beta-glucans. Cell Microbiol 7;471-9, 2005.
- Ariizumi K, Shen GL, Shikano S, Xu S, Ritter R 3rd, Kumamoto T, Edelbaum D, Morita A, Bergstresser PR, Takashima A: Identification of a novel, dendritic cell-associated molecule, dectin-1, by subtractive cDNA cloning. J Biol Chem 275; 20157-67, 2000.
- Adachi Y, Ishii T, Ikeda Y, Hoshino A, Tamura H, Aketagawa J, Tanaka S, Ohno N: Characterization of beta-glucan recognition site on C-type lectin, dectin 1. Infect Immun 72;4159-71, 2004
- 23. Palma AS, Feizi T, Zhang Y, Stoll MS, Lawson AM, Díaz-Rodríguez E, Campanero-Rhodes MA, Costa J, Gordon S, Brown GD, Chai W: Ligands for the beta-glucan receptor, Dectin-1, assigned using "designer" microarrays of oligosaccharide probes (neoglycolipids) generated from glucan polysaccharides. J Biol Chem 281;5771-9, 2006.
- Brown GD, Taylor PR, Reid DM, Willment JA, Williams DL, Martinez-Pomares L, Wong SY, Gordon S: Dectin-1 is a major beta-glucan receptor on macrophages. J Exp Med 196;407-12, 2002.

- Grünebach F, Weck MM, Reichert J, Brossart P: Molecular and functional characterization of human Dectin-1. Exp Hematol 30;1309-15, 2002.
- 26. Rogers NC, Slack EC, Edwards AD, Nolte MA, Schulz O, Schweighoffer E, Williams DL, Gordon S, Tybulewicz VL, Brown GD, Reis e Sousa C: Syk-dependent cytokine induction by Dectin-1 reveals a novel pattern recognition pathway for C type lectins. Immunity 22;507-17, 2005.
- 27. Hatada MH, Lu X, Laird ER, Green J, Morgenstern JP, Lou M, Marr CS, Phillips TB, Ram MK, Theriault K, Zoller MJ, Karas JL: Molecular basis for interaction of the protein tyrosine kinase ZAP-70 with the T-cell receptor. Nature 377; 32-8. 1995
- Xu S, Huo J, Lee KG, Kurosaki T, Lam KP: Phospholipase Cgamma2 is critical for Dectin-1-mediated Ca2+ flux and cytokine production in dendritic cells. J Biol Chem 284;7038-46, 2009.
- Shah VB, Ozment-Skelton TR, Williams DL, Keshvara L: Vav1 and PI3K are required for phagocytosis of beta-glucan and subsequent superoxide generation by microglia. Mol Immunol 46;1845-53, 2009.
- 30. Takeda K, Akira S: TLR signaling pathways. Semin Immunol 16;3-9, 2004.
- Lebron F, Vassallo R, Puri V, Limper AH: Pneumocystis carinii cell wall beta-glucans initiate macrophage inflammatory responses through NF-kappaB activation. J Biol Chem 278; 25001-8, 2003.
- 32. Kim HS, Kim JY, Ryu HS, Shin BR, Kang JS, Kim HM, Kim YO, Hong JT, Kim Y, Han SB: Phenotypic and functional maturation of dendritic cells induced by polysaccharide isolated from Paecilomyces cicadae. J Med Food 14;847-56, 2011.
- 33. Kim HS, Kim JY, Kang JS, Kim HM, Kim YO, Hong IP, Lee MK, Hong JT, Kim Y, Han SB: Cordlan polysaccharide isolated from mushroom Cordyceps militaris induces dendritic cell maturation through toll-like receptor 4 signalings. Food Chem Toxicol 48;1926-33, 2010.
- 34. Han SB, Lee CW, Kang MR, Yoon YD, Kang JS, Lee KH, Yoon WK, Lee K, Park SK, Kim HM: Pectic polysaccharide isolated from Angelica gigas Nakai inhibits melanoma cell metastasis and growth by directly preventing cell adhesion and activating host immune functions. Cancer Lett 243;264-73, 2006.
- 35. Han SB, Yoon YD, Ahn HJ, Lee HS, Lee CW, Yoon WK, Park SK, Kim HM: Toll-like receptor-mediated activation of B cells and macrophages by polysaccharide isolated from cell culture of Acanthopanax senticosus. Int Immunopharmacol 3;1301-12, 2003.
- Han SB, Park SK, Ahn HJ, Yoon YD, Kim YH, Lee JJ, Lee KH, Moon JS, Kim HC, Kim HM: Characterization of B cell membrane receptors of polysaccharide isolated from the root of Acanthopanax koreanum. Int Immunopharmacol 3;683-91, 2003.

- 37. Han SB, Park SH, Lee KH, Lee CW, Lee SH, Kim HC, Kim YS, Lee HS, Kim HM: Polysaccharide isolated from the radix of Platycodon grandiflorum selectively activates B cells and macrophages but not T cells. Int Immunopharmacol 1;1969-78, 2001.
- 38. Brown GD: Dectin-1: a signalling non-TLR pattern-recognition receptor. Nat Rev Immunol 6;33-43, 2006.
- 39. Dennehy KM, Ferwerda G, Faro-Trindade I, Pyz E, Willment JA, Taylor PR, Kerrigan A, Tsoni SV, Gordon S, Meyer-Wentrup F, Adema GJ, Kullberg BJ, Schweighoffer E, Tybulewicz V, Mora-Montes HM, Gow NA, Williams DL, Netea MG, Brown GD: Syk kinase is required for collaborative cytokine production induced through Dectin-1 and Toll-like receptors, Eur J Immunol 38;500-6, 2008.
- Thornton BP, Větvicka V, Pitman M, Goldman RC, Ross GD: Analysis of the sugar specificity and molecular location of the beta-glucan-binding lectin site of complement receptor type 3 (CD11b/CD18). J Immunol 156;1235-46, 1996.
- 41. Xia Y, Vetvicka V, Yan J, Hanikýrová M, Mayadas T, Ross GD: The beta-glucan-binding lectin site of mouse CR3 (CD11b/CD18) and its function in generating a primed state of the receptor that mediates cytotoxic activation in response to iC3b-opsonized target cells, J Immunol 162;2281-90, 1999.
- Li B, Allendorf DJ, Hansen R, Marroquin J, Ding C, Cramer DE, Yan J: Yeast beta-glucan amplifies phagocyte killing of iC3b-opsonized tumor cells via complement receptor 3-Syk-phosphatidylinositol 3-kinase pathway. J Immunol 177; 1661-9, 2006.
- Jimenez-Lucho V, Ginsburg V, Krivan HC: Cryptococcus neoformans, Candida albicans, and other fungi bind specifically to the glycosphingolipid lactosylceramide (Gal beta 1-4Glc beta 1-1Cer), a possible adhesion receptor for yeasts. Infect Immun 58:2085-90. 1990
- 44. Zimmerman JW, Lindermuth J, Fish PA, Palace GP, Stevenson TT, DeMong DE: A novel carbohydrate-glycosphingolipid interaction between a beta-(1-3)-glucan immunomodulator, PGG-glucan, and lactosylceramide of human leukocytes. J Biol Chem 273;22014-20, 1998.
- Wang J, Gigliotti F, Maggirwar S, Johnston C, Finkelstein JN, Wright TW: Pneumocystis carinii activates the NF-kappaB signaling pathway in alveolar epithelial cells. Infect Immun 73;2766-77, 2005.
- 46. Acton SL, Scherer PE, Lodish HF, Krieger M: Expression cloning of SR-BI, a CD36-related class B scavenger receptor. J Biol Chem 269;21003-9, 1994.
- 47. Assanasen C, Mineo C, Seetharam D, Yuhanna IS, Marcel YL, Connelly MA, Williams DL, de la Llera-Moya M, Shaul PW, Silver DL: Cholesterol binding, efflux, and a PDZ-interacting domain of scavenger receptor-BI mediate HDL-initiated signaling. J Clin Invest 115;969-77, 2005.