

Communications

Barbituric Acid Derivatives as Protein Tyrosine Phosphatase Inhibitors

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Barbiturates infer derivatives of barbituric acid. Commonly used barbiturates like phenobarbital and secobarbital have substituents at the 5 position on this basic skeleton. Barbiturates slow down central nervous system (CNS) functions and have been used to treat medical conditions such as epileptic seizures and as anesthesia for surgical procedures.¹ Beside the effects as CNS depressants, barbiturates have been shown to exhibit wide variety of other biological activities, including anti-tumor activities, immuno-modulating activities, herbicidal or insecticidal activities, PPAR γ agonistic activities, and inhibitory activities against mucosal addressin cell adhesion molecule-1 interactions.²⁻⁶ Notably, barbiturates, including phenobarbital and secobarbital, were also found to inhibit the phosphatase activity of calcineurin, a protein Ser/Thr phosphatase.⁷ In view of this observation, we considered the barbituric acid moiety as a candidate scaffold for the design of protein tyrosine phosphatase (PTP) inhibitors.

PTPs, together with protein tyrosine kinases, are key regulators of the phosphorylation of proteins involved in cellular signal transduction pathways. The importance of the regulated protein phosphorylation is evidenced by the recognition of a wide variety of diseases accompanied by the disturbances in the phosphorylation states of cellular proteins. Suggested as a strategy

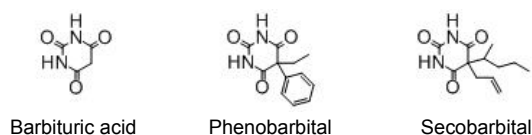
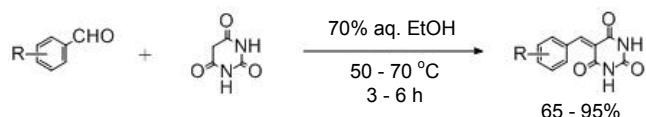


Figure 1. Barbituric acid and its derivatives used as CNS depressants.

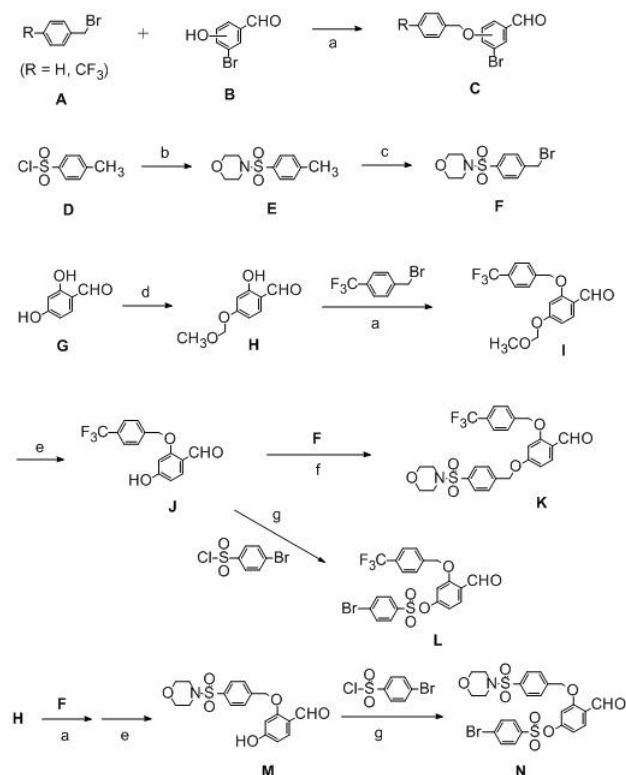


Scheme 1. Synthetic strategy for the synthesis of barbituric acid derivatives

for the treatment of such diseases, inhibition of PTPs constitutes a major theme of medicinal chemistry as well as of this study.⁸

In this study, barbituric acid derivatives were synthesized by condensation of appropriate benzaldehyde derivatives and barbituric acid with minor modifications of a reported procedure (Scheme 1).⁹ The desired products were obtained in > 90% yields except a few cases where the yields were 65 - 80%. The benzaldehyde derivatives for the synthesis were purchased or prepared as shown in Scheme 2.

The barbituric acid derivatives were assayed for their inhibitory activity against PTP1B, a human PTP. Among those,

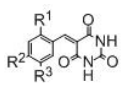


Scheme 2. Synthesis of benzaldehyde derivatives. Reagents and conditions: (a) K₂CO₃, DMF, rt, 1 - 4 h, > 85%, (b) morpholine, Et₃N, THF, rt, 3 h, 95%, (c) N-bromosuccinimide, CCl₄, reflux, 4 h, 56%, (d) CH₃OCH₂Cl, K₂CO₃, acetone, 0 °C → rt, 4 h, 53%, (e) 6 M HCl, THF, 50 °C, 2 h, >80%, (f) K₂CO₃, acetone, reflux, 2 h, 68%, (g) *N,N*-diisopropylethylamine, DMF, 0 °C → rt, 2 h, > 95%

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Table 1. Inhibitory activity of the barbituric acid derivatives against PTP1B


Compounds	R ¹	R ²	R ³	IC ₅₀ (μM) ^a
1		H	Br	340 ± 30
2	H		Br	200 ± 10
3	H		Br	47 ± 2
4			H	10 ± 1
5			H	27 ± 2
6			H	44 ± 3
7	H	OH	OCH ₃	> 1000
8			H	27 ± 3
9			H	52 ± 2
10		H	Br	88 ± 15

^aValues are the mean ± standard deviations of two or more experiments. PTP1B assay was performed as previously described using *p*-nitrophenyl phosphate (*p*NPP) as the substrate.¹⁰ IC₅₀ values were determined by measuring the *p*NPP hydrolase activity in a range of different inhibitor concentrations. Kinetic data were analyzed using GraFit 5.0 program (Erithacus Software).

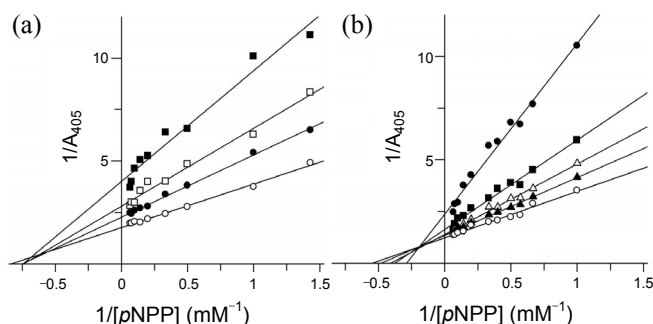
Table 2. Inhibition of PTP1B and other PTPs by compound 4^a

Compound	IC ₅₀ (μM) ^b		
	PTP1B	YTP1	YOP
4	10 ± 1	12 ± 1	15 ± 1

^aYTP1 were expressed in *E. coli* expression systems and purified as previously described.¹⁰ YOP was purchased from New England Biolabs, Inc. (Beverly, MA, U.S.A.). The assay condition was the same for all PTPs except the enzyme concentrations, which were 40 nM for PTP1B, 15 nM for YTP1 and 50 units (manufacturer's definition)/mL for YOP. ^bValues are mean ± standard deviations of two or more experiments.

compound 4 with trifluoromethylbenzyloxy substituents at both R¹ and R² was proved most potent with an IC₅₀ value of 10 μM (Table 1). The inhibitory activity of 4 was also evaluated against two other PTPs, YTP1 and YOP (Table 2). YTP1 is from *Saccharomyces cerevisiae* and YOP is the Yop51 gene product of *Yersinia enterocolitica*, that contains the C235R mutation (Yop51*). Compound 4 exhibited similar inhibitory potencies against all three PTPs.

To investigate the mode of inhibition, steady-state kinetic experiments of PTP1B and YTP1 were performed for compound 4. The mode of inhibition was determined by the Lineweaver-Burk plot analysis of the results of the kinetic experiments (Figure 2). The compound 4 inhibited PTP1B and YTP1 in a noncompetitive and a mixed-type noncompetitive fashion, respectively. These results indicate that the binding site for the compound 4 is different from that for the substrate. The noncompetitive inhibition of PTP1B might be explained by the possible binding of the barbituric acid moiety in the secondary aryl

**Figure 2.** Lineweaver-Burk plot analysis. Phosphatase activity was measured against *p*NPP for (a) PTP1B in the presence of 50 μM (■), 40 μM (□), 30 μM (●) or none (○) of compound 4 and (b) YTP1 in the presence of 35 μM (●), 25 μM (■), 20 μM (Δ), 15 μM (▲) or none (○) of compound 4.

phosphate-binding site, present near the active site of PTP1B.¹¹ On the other hand, the mixed-type noncompetitive inhibition of YTP1 by 4 could hardly be explained on the same basis, because no aryl phosphate-binding site distinct from the active site has been identified on YTP1. Nondiscriminative inhibition of the three structurally diverse PTPs by 4 also implicates the possible similarity of the inhibitor binding sites on these PTPs. Explanation of these apparently conflicting observations and considerations requires additional studies including X-ray crystallographic structure determination of the enzyme-inhibitor complex. Enzyme-inhibitor interaction yet to be explained at the molecular level, the barbiturate moiety was proven to be a promising scaffold for the design of PTP inhibitors.

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