Investigation of Ectomycorrhizal Fungal Colonization in Pinus thunbergii Seedlings at a Plantation Area in Gangneung, using Morphotyping and Sequencing the rDNA Internal Transcribed Spacer Region

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Abstract: The status of ectomycorrhizal (ECM) fungal colonization in *Pinus thumbergii* seedlings was investigated 2 years after planting in an eastern coastal area of Korea. We established three 10×10 m plots at a *P. thumbergii* plantation in Gangneung and sampled lateral roots from 10 seedlings in each plot. ECMs were classified into morphological groups and the number of root tips of each morphotype was counted. In total, 8 ECM morphotypes were observed and fungal species that form each morphotype were identified by sequencing of the internal transcribed spacer (ITS) region of the nuclear rDNA. *Suillus granulatus* was the most abundant species (44.1-65.7% of relative abundance) in all plots, followed by *Tomentella ellisii* (14.0-37.8%) and unidentified fungus belonged to Atheliaceae (10.6-20.1%). These 3 fungal species accounted for almost all of the ECM abundance in each plot (94.9-99.8%). The remaining 5 fungal species were uncommon and rare. There was no clear difference in ECM fungal communities among plots. Community structure of ECM fungi in the young *P. thunbergii* plantation was simple and composed of fungal species that were also observed in mature coastal pine forests.

Key words: Ectomycorrhizal fungi, Pinus thunbergii, plantation, Seedling, Suillus

Introduction

Coastal forests carry out significant functions for human activity such as alleviating damage from salt and sand dispersion to inland areas and providing place for recreation (Konta, 2001). Pinus thunbergii Parl., which grows naturally in Korea, has excellent abilities to tolerate high salinity and drought stress and grows well in the coastal areas. Therefore, P. thunbergii has been used for afforestation on many coastal dunes (Lee et al., 2005) and became one of the major constituents of the coastal forests in Korea (Choi, 1986; Park et al., 2002). Recently, however, much coastal pine forests have been cut down and have not maintained properly due to the recent increase of the developments for tourism or military purposes. In recent years, several studies related to afforestation such as recovery and maintenance of coastal pine forest were started in Korea (Chun et al., 2008).

Mycorrhizal associations are common in terrestrial

forest ecosystems and play a significant role in plant establishment including promoting nutrient and water uptake of host plants and also enhancement of tolerance towards stressful situations encountered by their hosts (Smith and Read, 1997). Therefore, the status of mycorrhizal colonization in planted seedlings is one of the important factors for their successful establishment. It is known that the effect of ectomycorrhizal (ECM) associations on host plant growth differs among different species of ECM fungi (Baxter and Dighton, 2001). Therefore, it is important to clarify what kinds of fungal species comprise a given community in plantation sites.

It is known that ECM fungi extensively colonize the fine roots of *P. thunbergii*. Several studies have been done to describe the ECM fungal community in *P. thunbergii* in coastal pine forests using sporocarps survey and/or molecular tools. Recorded various ECM fungal partners include basidiomycetes *Amanita*, *Laccaria*, *Lactarius*, *Inocybe*, *Pisolithus*, *Rhizopogon*, *Russula*, *Suillus*, *Tomentella* and ascomycetes *Cenococcum geophilum* Fr. (Taniguchi *et al.*, 2007; Matsuda *et al.*, 2009; Obase *et al.*, 2009a). However, there are no reports on the status of ECM fungal colonization

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in P. thunbergii plantation in coastal areas.

In this study, we investigated the status of ECM fungal colonization in *P. thunbergii* seedlings at a plantation area in Gangneung, Korea. ECMs were classified into morphological groups and the number of root tips of each morphotype was counted. Fungal species were identified from ECM root tips by sequencing the rDNA internal transcribed spacer (ITS) region.

Materials and Methods

1. Study site

For developing diversity-rich coastal forests that provide significant functions such as prevention of sand and salt dispersion, scenic values and recreation, a plantation was established in Gangneung, Korea (37° 23' N, 129° 13' E). Wood fences were constructed along the coast from a distance of about 40 m from shorelines for protecting seedlings against strong wind from sea and promoting of initial seedling growth. At an inland side from the fences, 3600 P. thunbergii, 2400 Rosa rugosa Thumb., 700 Chionanthus retusus Lindl. et Paxton, 900 Koelreuteria paniculata Laxm. and 700 Vitex rotundifolia L. seedlings were planted at interval of 1 m in 0.9 ha area in 2006. Seedling age of P. thunbergii was 6 when they were planted in the plantation site. A preliminary study revealed that only P. thunbergii seedlings grew well and other plants frequently died or showed symptoms of poor growth. Plant height and stem diameter (at the ground level) of P. thunbergii seedlings (377 individuals) were 136.7±21.1 and 2.6±0.6 cm in average ± standard deviation in 2008, respectively. The soil consisted entirely of maritime sand and shallow inland soils and has no litter and humus layers. In 2008, climatic data for Gangneung from the Korea Meteorological Administration indicated annual precipitation of 1343 mm, ranging between 0 and 330.6 mm (February and August, respectively) and annual temperature of 14.0°C, ranging between 1.1 and 26.7°C (January and July, respectively).

To understand ectomycorrhizal (ECM) fungal community in the plantation, we established three 10×10 m plots, parallel to fences and at intervals of c.a. 50 m. The plant community of this plantation includes herbaceous plants such as *Cassia nomame* (Sieb.) Honda, *Phragmites australis* (Cav.) Trin. ex Steud., *Artemisia* spp. and a fern, *Equisetum arvense* L. We sampled three soil cores (5 cm in diameter and 5 cm in depth) from each plot in September 2008 and determined soil water content, averaged 5.1 to 5.5% (w/w).

2. Sampling methods and observation of ECM colonization

We sampled lateral roots that were included in soil

blocks (within a radius of 30-40 cm from a stem and 15-20 cm in depth) from 10 seedlings in each plot. All samples were stored in plastic bags at 4°C until further analysis. Roots were separated from adhering soil by soaking and washing carefully with tap water using 0.5 mm sieve. All root segments were viewed under dissecting microscope after cutting into 5-10 cm segments. Viability of ECMs was interpreted based on color, texture of surface and degree of development of mantle layer. We excluded non-vital ECMs that looked shrunken, discolored and brittle to a piece. ECMs from each sample were classified into morphological groups based on such as ramification system, color, shape, texture, organization and abundance of emanating hyphae or cystidia, rhizomorphs and hyphal arrangement of mantle surface according to Agerer (1995) and Ingleby et al. (1990).

3. Identification of ECM fungal species

ECM fungal DNA was extracted from one ECM root tip of each morphotype using the DNeasy Plant Mini kit (QIAGEN, USA) according to the manufacturer's instructions. DNA amplification of the ITS regions, including the 5.8S rDNA, was performed on My Cycler thermalcycler (BioRad, USA) by using Takara Ex Taq (TAKARA, Japan) with a specific primer for higher fungi ITS1-f (Gardes and Bruns, 1993) and basidiomycete ITS4b (Gardes and Bruns, 1993). We used following PCR amplification conditions; 94°C (3 min), followed by 30 cycles of 94°C (30 s), 50°C (30 s) and 72°C (2 min), and final hold at 72°C (10 min). We determined the quality and quantity of the PCR products by agarose gel electrophoresis using QA-Agarose TM (MP Biomedicals, USA).

Each PCR product arising from different ECM morphotypes was purified using the QIA quick PCR purification kit (QIAGEN, USA) according to the manufacturer's instructions, and sequenced with primer ITS1f and ITS4. Sequencing reactions were entrusted to Solgent Co., Ltd. (Daejeon, South Korea). Fungal sequences were compared with the GenBank database at the National Center for Biotechnology Information (NCBI: http://www.ncbi. nlm.nih.gov) and UNITE (http://unite.ut.ee), using the nucleotide-nucleotide basic local alignment search tool algorithm (BLAST program). Sequences were aligned with closely matched reference sequences and submitted to neighbor-joining analysis at NCBI for taxonomic interpretation at the species, genus or family level. Sequences were considered to be identified at the species level when more than 97% identities with reference sequences derived from sporocarps were obtained.

4. Estimation of mycorrhizal colonization

ECM abundance indicating the number of ECM root tips of each morphotype was recorded separately for

each sample. Relative ECM abundance was calculated as an average value of a ratio of abundance of a given morphotype to total ECM abundance present in a given sample. The frequencies of ECM colonization that indicated the number of samples that contained a given ECM morphotype were also recorded.

Species-accumulation curves were drawn by plotting the mean of the accumulated number of expected species in pooled samples after 50 randomizations without replacement using EstimateS program version 8.0 (Colwell, 2005) for estimating whether sampling effort was enough to describe ECM fungal flora in plots. Estimated measurement of ECM fungal species richness using bootstrap, Jackknife 1 and Jackknife 2 estimators and similarity of fungal flora using Morishita-Horn index between plots were calculated by using EstimateS program version 8.0 (Colwell, 2005). ECM fungal diversity (Shannon's and Simpson's index) was also estimated using EstimateS program version 8.0 (Colwell, 2005).

Results

1. Identification of ECM fungi

We observed 7228 ectomycorrhizal (ECM) root tips (range 1628-2923 root tips in a plot) and classified them into 8 groups by morphological characteristics (Table 1, Figure 1). Each morphotype was formed by different ECM fungal species (Table 1). We identified 4 (Suillus

granulatus (L.) Roussel, Suillus luteus (L.: Fr.) S.F. Gray, Thelephora terrestris Fr. and Tomentella ellisii (Sacc.) Jülich & Stalpers) and 3 fungal species (Inocybe sp., Tomentella sp. 1 and Tomentella sp. 2) to species and genus level by matching sequences with those of known sporocarps. One fungal species could not be matched with sequences of known sporocarps but matched with those of estimated fungal species in other reports; it could be identified to the family level (Atheliaceae).

2. Status of ectomycorrhizal colonization

All samples contained ECMs in almost all vital root tips. The number of fungal species observed in each plot ranged 4 to 6 (Table 2). Average species richness per sampling ranged 2.6-3.3 and did not differ among plots. Total estimated species richness by bootstrap, Jackknife 1 and Jackknife 2 estimators were 9.3, 10.9 and 12.8, respectively. The species accumulation curve for ECM fungi was ascending, indicating that the probability of detection of rare fungal species is still high as sample size increases (Figure 2). Three fungal species (S. granulatus, T. ellisii and unidentified species belonged to Atheliaceae) were common among plots. These three common species accounted for 94.9 to 99.8% of total abundance of ECMs in each plot. Similarity of species composition of ECM fungi between plots were 0.88 to 0.97 using the Morishita-Horn index.

Table 1. Morphological characteristics and possible identities of ectomycorrhizal (ECM) fungal species that formed each ECM type observed in the roots of *Pinus thunbergii* seedlings in a plantation in a coastal area in Gangneung, Korea.

Possible identity (Accession No. of the highest similarity	Morphological characteristics of an ECM							
	Color Shape		Texture	Mantle (outer layer)	Emanating hyphae			
<i>Inocybe</i> sp. EU554696 (556/557; 99%)	Pale yellow to yellow	Unbranched to forked-branched	Smooth	Net synenchyma	Absent to infrequent			
Suillus granulatus AB284447 (594/597; 99%)	White and partially brown	Monopodial-pinnate to dichotomous	Densely wooly	Net synenchyma	Abundant, rhizomorphs were infrequently observed			
Suillus luteus DQ068969 (616/616; 100%)	Reddish brown	Dichotomous	Smooth and shiny	nd *	Infrequent			
Thelephora terrestris FN393122 (619/625; 99%)	Pale brown	Unbranched to dichotomous	Smooth	Net synenchyma	Absent to infrequent			
<i>Tomentella ellisii</i> FJ013069 (604/606; 99%)	Pale yellow to yellow	Unbranched to forked-branched	Loosely wooly	Net synenchyma	Absent to infrequent			
<i>Tomentella</i> sp. 1 EF619788 (564/587; 96%)	Reddish brown	Unbranched to dichotomous	Smooth and shiny	Non interlocking irregular synen- chyma	Absent			
<i>Tomentella</i> sp. 2 AB253522 (544/546; 99%)	White (very tip) to brown (older part)	Dichotomous	Smooth	Net synenchyma	Infrequent			
Unidentified species belonged to Atheliaceae AB253524 (531/533; 99%)	Pale yellow to yellow	Unbranched to dichotomous	Cottony	Net synenchyma	Abundant			

^{*}nd; could not determined.

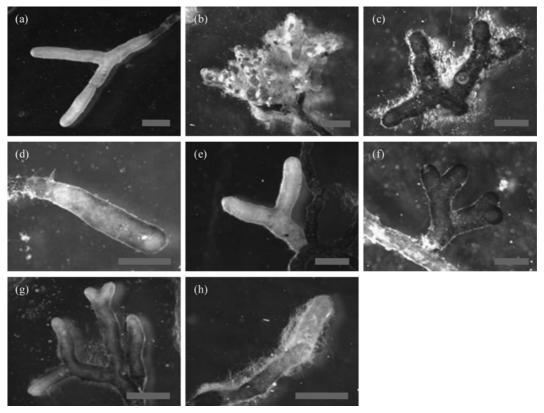


Figure 1. Appearance of ectomycorrhizas observed in the roots of *Pinus thunbergii* seedlings an costal plantation area in Gangneung, Korea; (a) *Inocybe* sp., (b) *Suillus gramulatus*, (c) *Suillus luteus*, (d) *Thelephora terrestris*, (e) *Tomentella ellisii*, (f) *Tomentella* sp. 1, (g) *Tomentella* sp. 2, (h) unidentified species belonged to Atheliaceae. Bars indicated 1 mm.

Table 2 Relative abundance (RA) and frequencies (F) of each ectomycorrhizal (ECM) fungi in roots of *Pinus thunbergii* seedlings in a plantaion in a coastal area in Gangneung, Korea.

ECM funcial amosics	Plot 1		Plot 2		Plot 3		Total	
ECM fungal species	RA (%)	F (/10)	RA (%)	F (/10)	RA (%)	F (/10)	RA (%)	F (/30)
Suillus granulatus	44.1	9	65.7	8	58.5	10	54.3	27
Tomentella ellisii	37.8	10	14.0	9	25.8	10	28.0	29
Unidentified species belonged to Atheliaceae	16.4	10	20.1	8	10.6	9	15.1	27
Tomentella sp. 1					3.8	2	1.4	2
Tomentella sp. 2	1.5	2	0.2	1			0.7	3
Thelephora terrestris					1.3	1	0.5	1
Inocybe sp.	0.2	1					0.1	1
Suillus luteus	0.1	1					< 0.1	1
Total No. ECM root tips observed	2923		1628		2677		7228	
Mean No. species per sampling*	3.3 ± 0.5		2.6 ± 0.7		3.2 ± 0.4		3.0 ± 0.6	
Observed species richness	6		4		5		8	
Mean estimated species richness								
Bootstrap	6.8		4.4		5.5		9.3	
Jackknife 1	7.8		4.9		5.9		10.9	
Jackknife 2	8.7		5.7		6.0		12.8	
Shannon's index	1.1		0.9		1.1		1.1	
Simpson's index	2.8		2.0		2.4		2.5	

^{*}Averages and standard deviations were represented.

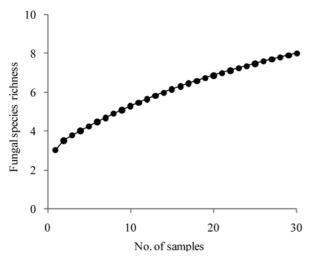


Figure 2. Species-accumulation curve for ectomycorrhizal fungi observed in roots of *Pinus thunbergii* seedlings in a plantation in a coastal area in Gangneung, Korea.

Suillus granulatus was the most abundant ECM fungal species observed in all the plots and accounted for 44.1 to 65.7% in relative abundance. *T. ellisii* and unidentified species belonged to Atheliaceae were the second or third most abundant fungal species and accounted for 14.0 to 37.8% and 10.6 to 20.1% in relative abundance, respectively. These 3 fungal species were observed in 27 to 29 out of 30 soil samples. The remaining 5 fungal species were uncommon and showed low relative abundance (less than 0.1 to 3.8%) and frequencies (1 to 3 out of 30 soil samples).

Discussion

We investigated abundances and frequencies of ectomycorrhizal (ECM) fungi colonized roots of *P. thunbergii* seedlings planted in an eastern coastal area and showed ECM fungal communities. However, it appeared that sampling efforts such as number of seedlings observed and replicated sampling locations was not enough to clarify ECM fungal community in *P. thunbergii* plantations; it means our results are relevant only to this particular locations and cannot be extrapolated to *P. thunbergii* plantations in general. Further investigations involving more intense sampling efforts are needed for obtaining general information about ECM fungal community in *P. thunbergii* plantations.

Our results showed a species-poor and simple ECM fungal community in the *P. thunbergii* plantation. No more than 3 ECM fungal species frequently colonized seedlings and occupied in the range of 10.6 to 65.7% in relative abundance, and the remaining fungal species were detected from the samples of small amount of root tips. Few dominant fungi and the majority of rare fungi in species richness is a common pattern as described in

previous surveys conducted in several forest ecosystems (Jonsson et al., 1999; Matsuda and Hijii, 2004; Valentine et al., 2004). Suillus granulatus was the most dominant in abundance and frequency in the plantation. This fungus is possibly one of the most widespread species of Suillus associated with pine species in Asia (Yamada and Katsuya, 1995; Sims et al., 1997; Dunstan et al., 1998), Europe (Pera and Alvarez, 1995; Manian et al., 2001; Wiemken and Boller, 2006), America (Jacobson et al., 1993; Kretzer et al., 1996) and South Africa (Dunstan et al., 1998). This fungus has also been observed in P. thunbergii coastal forests in Japan (Taniguchi et al., 2007) and Korea (Obase et al., 2009a). Sporocarps of this fungus often observed in mature and young pine stands. Cullings et al. (2003) showed dominance of this fungus in roots of young pine stands that had undergone stand-replacing fire and as a result, has very little resident litter and organic matter. Wiemken and Boller (2006) also showed dominance of this fungus in young pine trees established on edge of forests and grassland. Obase et al. (2009a) detected this fungus in roots of 0-10 year old P. thunbergii seedlings and also c.a. 50 year old mature trees in coastal pine forests. It appears that this fungus has characteristics as an early stage fungus and is able to colonize young P. thunbergii plantations.

Tomentella ellisii was the second dominant species. Recent reports showed that Tomentella spp. were common ECM symbionts in several boreal and temperate forests (Horton and Bruns, 1998; Taylor and Bruns, 1999), including P. thunbergii coastal forests (Obase et al., 2009a). On the other hands, T. ellisii was also detected in seedlings established on disturbed areas under initial vegetation recovery. We have detected this fungus in P. densiflora seedlings established on exposed granite slopes and in surrounding forestlands (data not shown). Obase et al. (2007) detected T. ellisii in roots of Salix, Populus and Betula seedlings established on volcanic debris where organic matter was absent (Assigned name as Thelepholaceae 1). Obase et al. (2009b) showed that this fungus promote the growth of aspen seedlings on the volcanic debris.

The third dominant species was unidentified species belonged to Atheliaceae. This fungus has been detected mainly in roots of pine trees. Nara and Hogetsu (1996) isolated this fungus from roots of *P. densiflora* grown in a nursery and assigned name as unidentified species Tanashi 01. We also observed this fungus abundantly in roots of *P. densiflora* seedlings established on exposed granite slopes (data not shown). Taniguchi *et al.* (2007) and Kataoka *et al.* (2008) observed this fungus in roots of *P. thunbergii* seedlings in coastal pine forests. In view of these observations, the ECM fungal community in the plantation was composed of ECM fungal species that

were also observed in mature coastal pine forests. These fungi seem to have an excellent ability to immediately invade and colonize roots of seedlings growing in nurseries and plantation.

Obase et al. (2009a) investigated ECM fungal community in roots of 0-10 year old regenerating P. thunbergii seedlings and mature trees in coastal pine forests and showed that Cenococcum geophilum Fr. was the most dominant species both in seedlings and mature trees. Matsuda et al. (2009) investigated abundance of ECM fungal species in four stands of P. thunbergii coastal forests and showed that C. geophilum is common and the most probable dominant fungal species in coastal areas of Japan. We also observed ECMs of C. geophilum abundantly and isolated its strains from P. thunbergii coastal forests in Gangneung, Taean and Incheon, Korea (data not shown). So, it appears that C. geophilum is a major colonizer in P. thunbergii coastal forests in Korea. It is uncertain why C. geophilum was absent in the plantation, however our study revealed the possibility that some ECM fungal species, including even the most common ones in coastal areas, could not invade immediately after plantation establishment.

This study revealed that seedlings established in the plantation were associated with several naturally occurring ECM fungi. Cram et al. (1999) showed that naturally occurring ECM fungi could increase survival and growth of forest trees better than artificial inoculation. So, the survival and adaptation of P. thunbergii seedlings in a coastal area may be partly due to their symbiotic associations with these symbionts. However, it is possible that the status of ECM fungal colonization in outplanted seedlings is variable among stand locations because it is affected by environmental conditions such as soil properties, intensity of salt and/or drought stress and presence or absence of enough mycorrhizal propagules in the site or surrounding areas. Some effective indigenous ECM fungi like C. geophilum in coastal areas can not immediately invade and colonize planted seedlings possibly due to lack of dispersal ability. Therefore, it should be advantageous to inoculate site-adaptable and effective fungal strains on seedlings before planting in plantation site in coastal areas. Further investigations on the effects of naturally inhabiting ECM fungi on the establishment of P. thunbergii seedlings in coastal areas are needed for understanding what kinds of ECM fungal strains are effective for establishment of seedlings and applicable in reforestation program in coastal areas.

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Literature Cited

- Agerer, R. 1995. Anatomical characteristics of identified ectomycorrhizas: an attempt towards a natural classification. In: Varma, A.K. and Hock, B. (Eds.), Mycorrhiza: Structure, function, molecular biology and biotechnology. Springer, Berlin Heidelberg New York, 685-734.
- 2. Baxter, J.W. and Dighton, J. 2001. Ectomycorrhizal diversity alters growth and nutrient acquisition of grey birch (*Betula populifolia*) seedlings in host-symbiont culture conditions. New Phytologist 152:139-149.
- Choi, M.G. 1986. Characteristics of salt tolerance in tree species. Journal of Korean Forestry Society 29(2): 1-8. (In Korean with English abstract)
- Chun, K.W., Lee, J.H., Kim, K.N., Yeom, K.J., Seo, J.C., Ma, H.S., Park, M.S. and Ezaki, T. 2008. The present status and future task of seaside protection forest in Korea. Journal of the Japanese Society of Coastal Forest 7(2): 21-25. (In Japanese with English abstract)
- 5. Colwell, R.K. 2005. EstimateS: Statistical estimation of species richness and shared species from samples. Version 7.5. [http://purl.oclc.org/estimates].
- Cram, M.M., Mexal, J.G. and Souter, R. 1999. Successful reforestation of south Carolina sandhills is not influenced by seedling inoculation with *Pisolithus tinctorius* in the nursery. Southern Journal of Applied Forestry 23: 46-52.
- Cullings, K.W., New, M.H., Makhija, S. and Parker, V.T. 2003. Effects of litter addition on ectomycorrhizal associates of a lodgepole pine (*Pinus contorta*) stand in Yellowstone National Park. Applied and Environmental Microbiology 69: 3772-3776.
- 8. Dunstan, W.A., Dell, B. and Malajczuk, N. 1998. The diversity of ectomycorrhizal fungi associated with introduced *Pinus* spp. in the Southern Hemisphere, with particular reference to Western Australia. Mycorrhiza 8: 71-79.
- 9. Gardes, M. and Bruns, T.D. 1993. ITS primers with enhanced specificity for basidiomycetes: application to the identification of mycorrhizae and rusts. Molecular Ecology 2: 113-118.
- 10. Horton, T.R. and Bruns, T.D. 1998. Multiple host fungi are the most frequent and abundant ectomycorrhizal types in a mixed stand of Douglas fir (*Pseudotsuga* menziesii (Mirb.) Franco) and bishop pine (*Pinus muri*cata D. Don). New Phytologist 139: 331-339.
- 11. Ingleby, K., Mason, P.A., Last, F.T. and Fleming, L.V.

- 1990. Identification of ectomycorrhizas. ITE Research Publication No. 5, HMSO, London.
- 12. Jacobson, K.M., Miller, O.K.Jr. and Turner, B.J. 1993. Randomly amplified polymorphic DNA markers are superior to somatic incompatibility tests for discriminating genotypes in natural populations of the ectomycorrhizal fungus *Suillus granulatus*. Proceeding of the National Academy of Sciences of the United States of America 90: 9159-9163.
- 13. Jonsson, L., Dahlberg, A., Nilsson, M.C., Kårén, O. and Zackrisson, O. 1999. Continuity of ectomycorrhizal fungi in self-regenerating boreal *Pinus sylvestris* forests studied by comparing mycobiont diversity on seedlings and mature trees. New Phytologist 142: 151-162.
- 14. Kataoka, R., Taniguchi, T., Ooshima, H. and Futai, K., 2008. Comparison of the bacterial communities established on the mycorrhizae formed on *Pinus thunbergii* root tips by eight species of fungi. Plant and Soil 304: 267-275.
- 15. Konta, F. 2001. The present conditions and functions of the coastal forests in Japan. Journal of the Japanese Society of Coastal Forest 1(1): 1-4. (In Japanese with English abstract)
- Kretzer, A., Li, Y., Szaro, T. and Bruns, T.D. 1996.
 Internal transcribed spacer sequences from 38 recognized species of *Suillus* sensu lato: Phylogenetic and taxonomic implications. Mycologia 88: 776-785.
- Lee, C.Y., Jeong, Y.H. and Kim, J.H. 2005. Actual condition and management plan for the coastal prevention forest. International Symposium proceeding of the Japanese Society of Coastal Forest and the Korea Society of Coastal Forest: 54-55. (In Korean)
- Manian, S., Sreenivasaprasad, S., Bending, G.D. and Mills, P.R. 2001. Genetic diversity and interrelationships among common European *Suillus* species based on ribosomal DNA sequences. FEMS Microbiology Letters 204: 117-121.
- Matsuda, Y. and Hijii, N. 2004. Ectomycorrhizal fungal communities in an *Abies firma* forest, with special reference to ectomycorrhizal associations between seedlings and mature trees. Canadian journal of Botany 82: 822-829.
- 20. Matsuda, Y., Hayakawa, N. and Ito, S. 2009. Local and microscale distributions of *Cenococcum geophilum* in soils of coastal pine forests. Fungal Ecology 2: 31-35.
- Nara, K. and Hogetsu, T. 1996. Effects of inoculation with ectomycorrhizal fungus to seven *Pinus* species including several foreign pines. Transactions of the Japanese Forestry Society 107: 227-228. (In Japanese)
- 22. Obase, K., Tamai, Y., Yajima, T. and Miyamoto, T.

- 2007. Mycorrhizal associations in woody plant species at the Mt. Usu volcano, Japan. Mycorrhiza 17: 209-215.
- 23. Obase, K., Cha, J.Y., Lee, J.K., Lee, S.Y., Lee, J.H. and Chun, K.W. 2009a. Ectomycorrhizal fungal communities associated with *Pinus thunbergii* in the eastern coastal pine forests of Korea. Mycorrhiza 20: 39-49.
- Obase, K., Tamai, Y., Yajima, T. and Miyamoto, T. 2009b. Mycorrhizal synthesis of four ectomycorrhizal fungi in potted *Populus maximowiczii* seedlings. Mycoscience 50: 143-145.
- 25. Park, W.G., Yi, M.J., Chun, K.W. and Ezaki, T. 2002. A study on stand structure of *Pinus thunbergii* Parl. Forests in eastern part of Kangwon-do, Korea. Journal of the Japanese Society of Coastal Forest 1(2): 13-18. (In Japanese with English abstract)
- 26. Pera, J. and Alvarez, I.F. 1995. Ectomycorrhizal fungi of *Pinus pinaster*. Mycorrhiza 5: 193-200.
- Sims, K., Watling, R., De la Cruz, R. and Jeffries, P. 1997. Ectomycorrhizal fungi of the Philippines: a preliminary survey and notes on the geographic biodiversity of the Sclerodermatales. Biodiversity and Conservation 6: 45-58.
- 28. Smith, S.E. and Read, D.J. 1997. Mycorrhizal symbiosis, 2nd edn. London, UK, Academic Press.
- 29. Taniguchi, T., Kanzaki, N., Tamai, S., Yamanaka, N. and Futai, K. 2007. Does ectomycorrhizal fungal community structure vary along a Japanese black pine (*Pinus thunbergii*) to black locust (*Robinia pseudoacacia*) gradient? New Phytologist 173: 322-334.
- 30. Taylor, D.L. and Bruns, T.D. 1999. Community structure of ectomycorrhizal fungi in a *Pinus muricata* forest: minimal overlap between the mature forest and resistant propagule communities. Molecular Ecology 8: 1837-1850
- 31. Valentine, L.L., Fiedler, T.L., Hart, A.N., Petersen, C.A., Berninghausen, H.K. and Southworth, D. 2004. Diversity of ectomycorrhizas associated with *Quercus garryana* in southern Oregon. Canadian Journal of Botany 82: 123-135.
- 32. Wiemken, V. and Boller, T. 2006. Delayed succession from alpine grassland to savannah with upright pine: Limitation by ectomycorrhiza formation? Forest Ecology and Management 237: 492-502.
- Yamada, A. and Katsuya, K. 1995. Mycorrhizal association of isolates from sporocarps and ectomycorrhizas with *Pinus densiflora* seedlings. Mycoscience 36:315-323.