

Cordyceps militaris Enhances MHC-restricted Antigen Presentation via the Induced Expression of MHC Molecules and Production of Cytokines

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Background: *Cordyceps militaris* water extract (CME) has been reported to exert antitumor and immunomodulatory activities in vivo and in vitro. However, the therapeutic mechanism has not yet been elucidated. In this study, we examined the effects of CME on the antigen presenting function of antigen presenting cells (APCs). **Methods:** Dendritic cells (DCs) were cultured in the presence of CME, and then allowed to phagocytose microspheres containing ovalbumin (OVA). After washing and fixing the efficacy of OVA, peptide presentation by DCs were evaluated using CD8 and CD4 T cells. Also, we confirmed the protein levels of proinflammatory cytokines through western blot analysis. **Results:** CME enhanced both MHC class I and class II-restricted presentation of OVA in DCs. In addition, the expression of both MHC class I and II molecules was enhanced, but there was no changes in the phagocytic activity of exogenous OVA. Furthermore, CME induced the protein levels of iNOS, COX-2, proinflammatory cytokines, and nuclear p65 in a concentration-dependent manner, as determined by western blot. **Conclusion:** These results provide an understanding of the mechanism of the immuno-enhancing activity of CME on the induction of MHC-restricted antigen presentation in relation to their actions on APCs.

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INTRODUCTION

The interest of researchers in medicinal plants as natural sour-

ces of active principles has markedly increased over the past decades with particular attention paid to the polysaccharide compounds of various traditional Asian medicines (1,2). For example, *Cordyceps militaris* (CM) has been used as a folk medicine in Korea, China, and Japan for a long time. Its chemical components, including cordycepin (3,4), nucleoside, and various polysaccharides (5,6), have been widely studied. However, little is known about its immune-enhancing effects or the effects of its mechanism on murine APCs.

Antigen presenting cells, especially Dendritic cells (DCs), play a critical role in the initiation of immune responses and the induction of immune tolerance. DCs are the most important accessory cells for the activation of naïve T cells and the generation of primary T cell responses (7). They can acquire and process antigens in the periphery, and migrate to secondary lymphoid tissues where they prime primary T cell responses. Since T cells can only recognize antigens presented on MHC molecules, modulation of MHC-restricted antigen processing pathways may provide novel pharmacological targets for the regulation of T cell responses.

In the present study, we examined the effects of *Cordyceps militaris* water extract, CME, on the in vitro function of DCs with the goal of characterizing the mechanism of CME as an immune-enhancing agent. We used OVA as an exogenous antigen in conjunction with CME, and then compared the change in cross presentation of CME-related DCs to that of a control group along with the level of MHC class I and II

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molecules. Further, we examined the immune-enhancing activity of CME on peritoneal macrophages. We found that CME activated several parameters of macrophage activation such as nitric oxide and cytokine expression and antigen presenting to T cells. Ultimately, CME may explain some of the therapeutic efficacies of CM, which has been used in folk medicine to treat various diseases including cancer.

MATERIALS AND METHODS

Reagents

Lipopolysaccharide (LPS) was purchased from Sigma (St. Louis, USA). Cell culture media DMEM, antibiotic-penicillin/streptomycin solution, and fetal bovine serum (FBS) (HyClone, Logan, USA) were used for the cell culture. *Cordyceps militaris* water extract (CME) were supplied by CM Biotec. (Kangnung, Korea) (6).

Cell culture

T cell hybridomas, CD8OVA1.3 and DOBW (8), were kindly provided by Dr. Clifford V. Harding (Case Western Reserve University, Cleveland, OH). The DC cell line (DC2.4) was obtained from the Dana-Farber Cancer Institute, Boston, MA, USA (9). CD8OVA1.3, DOBW, and DC2.4 cells were maintained in DMEM medium supplemented with 10% FBS, 100 U/ml of penicillin, 100 μ g/ml of streptomycin, and 2 mM L-glutamine. Bone marrow-derived DCs were generated from BALB/C mice bone marrow precursor cells (10). Bone marrow cells obtained from femurs of mice were cultured in a 6-well plates in a culture medium supplemented with 200 U/ml rmGM-CSF. At days 3 and 4 after initiation of the culture, nonadherent cells were discarded by replacing the culture medium with fresh medium containing cytokines, after gentle shaking. DCs were harvested by gentle pipetting at day 6. Primary macrophages were collected from the peritoneal cavities of mice (8-week-old male ICR mice) after an intraperitoneal (i.p.) injection of 3 ml of 3% thioglycolate broth (Sigma, St. Louis, MO) 4 days before harvesting. The peritoneal macrophages were then washed with 1 \times PBS (Ca²⁺- and Mg²⁺-free) and plated with DMEM containing 10% FBS, 100 U/ml of penicillin, and 100 U/ml of streptomycin at 37°C in a 5% CO₂ incubator.

Preparation of microencapsulated OVA

OVA-containing microspheres were prepared by a solvent-evaporation method (11). Briefly, albumin from chicken egg

white was dissolved in 3% polyvinyl alcohol (4 mg/ml), and poly (DL-lactide-co-glycolide) (PLGA; lactide : glycolide=50 : 50; Sigma-Aldrich, St.Louis, MO, USA) was dissolved in a mixture of acetone and ethanol (9 : 1) (5%). The concentration of OVA was determined using a micro-bicinchoninic acid assay kit (Pierce, Rockford, IL) according to the manufacturer's instructions after lysing the microspheres in a lysis buffer containing 0.1% SDS and 0.1 N NaOH. For the phagocytosis assay, microspheres containing both OVA and fluorescein isothiocyanate (FITC) were prepared by adding FITC (final, 5 mg/ml) to a mixture of acetone and ethanol (9 : 1) together with PLGA (final, 5%).

MHC class I-restricted presentation assay of exogenous OVA

DCs were cultured in the presence of different concentrations of CME for 2 hrs in 96-well plates (1×10^5 cells/ml), and then combined with OVA-microspheres (6.25 μ g/min as OVA). After 6 hrs incubation at 37°C, the plate was washed twice of pre-warmed PBS, and then fixed with ice-cold 1% paraformaldehyde for 5 min at room temperature. The plate was washed and CD8OVA1.3 cells (1×10^5 cells/ml) were added. After overnight incubation at 37°C, the plate was centrifuged at 1,800 rpm, and the culture supernatant was collected and assayed for IL-2 content using an IL-2 ELISA kit (eBioscience, San Jose, CA, USA).

MHC class II-restricted presentation assay of exogenous OVA

DCs and peritoneal macrophages were cultured in the presence of different concentration of CME for 2 hrs in 96-well plates (1×10^5 cells/ml), and then combined with OVA-microspheres (6.25 μ g/min as OVA). After 6 hrs incubation at 37°C, the plate was washed twice of pre-warmed PBS, and then fixed with ice-cold 1% paraformaldehyde for 5 min at room temperature. The plate was washed and DOBW cells (1×10^5 cells/ml) were added. After overnight incubation at 37°C, the plate was centrifuged at 1,800 rpm, and the culture supernatant was collected and assayed for IL-2 content using an IL-2 ELISA kit.

Phagocytosis assay

DCs were cultured in the presence of different concentrations of CME for 2 hrs in 6-well plates (2×10^6 cells/well), and then combined with microspheres (average diameter, 300 nm) containing both OVA and FITC. After 2 hrs, unphagocytosed

microspheres were removed by washing with pre-warmed PBS. The plate was chilled on ice for 20 min, and subsequently, the cells remaining on the bottom of plate wells were harvested by rough pipetting with cold PBS, washed with 10 ml of cold PBS to remove the extra particular OVA, and then fixed with 1% paraformaldehyde. Flow cytometric analysis was performed on a FACS Canto flow cytometer

(Beckman coulter, Brea, USA).

Phenotype analysis

DCs were cultured in the presence of different concentrations of CME for 2 hrs in 6-well plates (2×10^6 cells/well). The plate was chilled on ice for 20 min, and subsequently, the cells remaining on the bottom of plate wells were harvested by

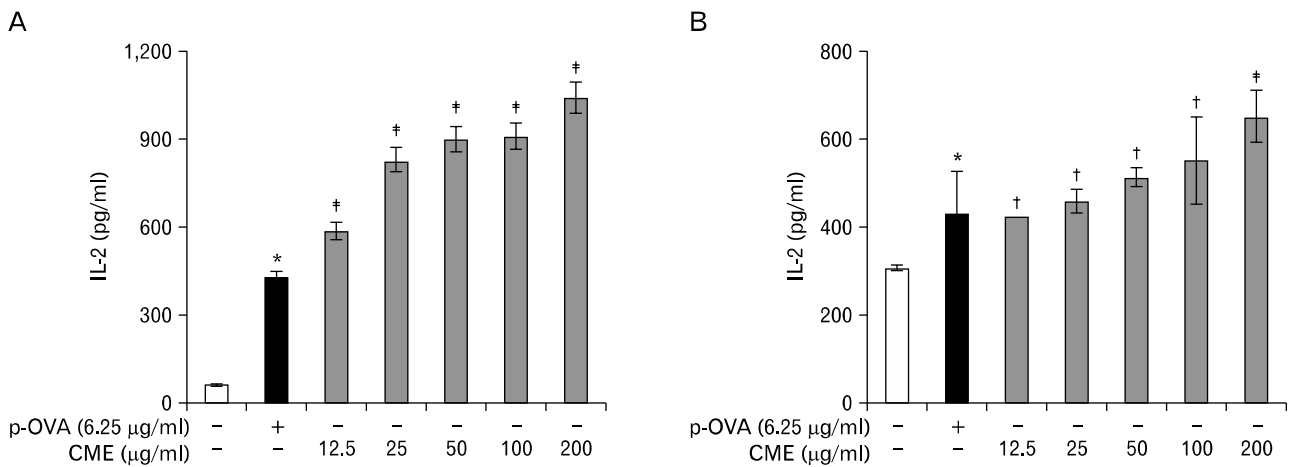


Figure 1. Effects of CME on the cross-presentation of exogenous OVA in DCs. (A) DC2.4 cells and (B) BM-DCs were incubated with the indicated amounts of CME for 2 hrs, and then combined with OVA-microspheres. After 6 hrs incubation, the cells were washed, fixed, and the amounts of OVA peptides presented on MHC class I molecules were assessed using OVA-specific CD8 T cell hybridoma, CD8OVA1.3. The amounts of IL-2 produced from OVA-specific CD8 T cells were assayed by a commercial IL-2 ELISA kit. Data have been presented as means \pm S.D. of three independent experiments. * $p < 0.05$ vs. cells only; † $p < 0.05$, ‡ $p < 0.01$ vs. OVA only.

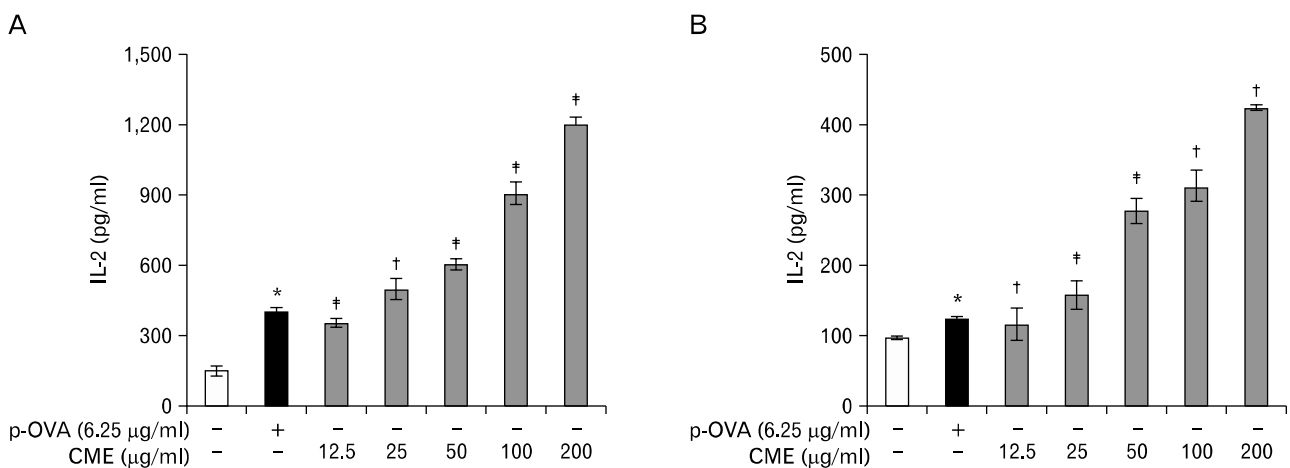


Figure 2. Effects of CME on the MHC class II-restricted presentation of exogenous OVA in APCs. (A) BM-DCs and (B) peritoneal macrophages were incubated with the indicated amounts of the CME for 2 hrs, and then combined with OVA-microspheres. After 6 hrs incubation, the cells were washed, fixed, and the amounts of OVA peptides presented on MHC class II molecules were assessed using OVA-specific CD4 T cell hybridoma, DOBW. The amounts of IL-2 produced from OVA-specific CD4 T cells were assayed by a commercial IL-2 ELISA kit. Data have been presented as means \pm S.D. of three independent experiments. * $p < 0.05$ vs. cells only; † $p < 0.05$, ‡ $p < 0.01$ vs. OVA only.

rough pipetting with cold PBS. The cells were stained with monoclonal antibodies recognizing murine cell surface molecules after blocking of FcR-binding anti-CD16/CD32 monoclonal antibody, and flow cytometry analysis was performed on a FACS Canto flow cytometer. The monoclonal antibodies, anti-H-2K^b, anti-I-A^b, and isotype-matched control antibodies were purchased from BD Biosciences (San Jose, USA).

Preparation of nuclear extracts

After cultured cells were collected and washed twice with cold PBS, they were resuspended in hypotonic buffer (10 mM HEPES, pH 7.9, 10 mM KCl, 1.5 mM MgCl₂, 0.2 mM PMSF, 0.5 mM DTT, 10 μg/ml aprotinin). After 15 min incubation on ice, the cells were lysed by the addition of 0.1% NP-40

and vigorous vortexing for 1 min. The nuclei were pelleted by centrifugation at 12,000×g for 1 min at 4°C and resuspended in high salt buffer (20 mM HEPES, pH 7.9, 25% glycerol, 400 mM KCl, 1.5 mM MgCl₂, 0.2 mM EDTA, 0.5 mM DTT, 1 mM NaF, 1 mM sodium orthovanadate). The supernatant fluid was stored in aliquots at -70°C.

Western blot analysis

Peritoneal macrophages were washed with PBS and lysed by lysis buffer (1% SDS, 1.0 mM sodium vanadate, 10 mM Tris-Cl buffer, pH 7.4) for 5 min. 20 μg of protein from the cell lysates was applied to 8~12% SDS-polyacrylamide gels and then transferred to nitrocellulose membranes. The membranes were blocked with 5% skim milk in PBST solution for

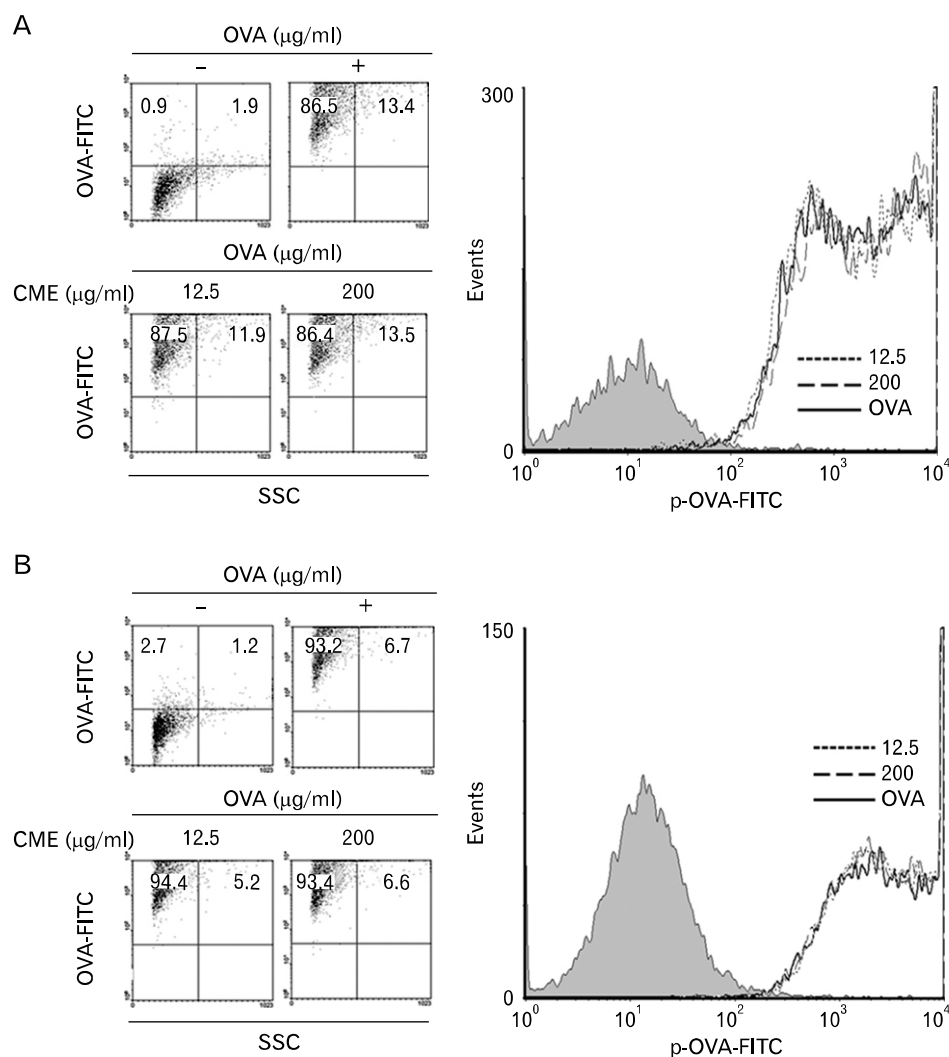


Figure 3. Effects of CME on the phagocytic activity. (A) DC2.4 cells and (B) BM-DCs were cultured with CME for 2 hrs, and then combined with microspheres containing both OVA and FITC (6.25 μg/ml). After 2 hrs incubation, unphagocytosed microspheres were washed, and the cells were harvested by gentle pipetting, and then analyzed by flow cytometry.

1 hr. They were then incubated with anti-IL-1 β , anti-IL-6, anti-TNF- α , anti-i-NOS, anti-COX-2, and anti-NF- κ B monoclonal antibodies (Santa Cruz Biotechnology Inc., Santa Cruz, California, USA) for 2 hrs and washed 3 times with PBST. After incubation with an alkaline phosphatase-labeled secondary antibody (Abcam, Cambridge, USA) for 2 hrs, the bands were visualized using a Western Blot Kit with an alkaline phosphatase substrate (Vector, Burlingame, USA).

Data analysis

Data were expressed as mean \pm standard deviation. Statistical significance between the groups was determined by a paired t-test and one-way ANOVA for repeated measures. Results of $p < 0.05$ were considered statistically significant. Data were

assessed using SPSS (version 15.0, SPSS Inc., Chicago, Illinois).

RESULTS

CME enhanced MHC-restricted presentation of exogenous OVA

The effects of CME on MHC class I-restricted presentation of exogenous OVA in DC2.4 cells and BM-DCs were examined. DCs were treated with CME for 2 hrs, after which micro-encapsulated OVA were added into cultures for phagocytosis to examine whether exogenous antigen presentation pathways could be affected by CME. The cells were washed and then an MHC class I-restricted presentation assay was per-

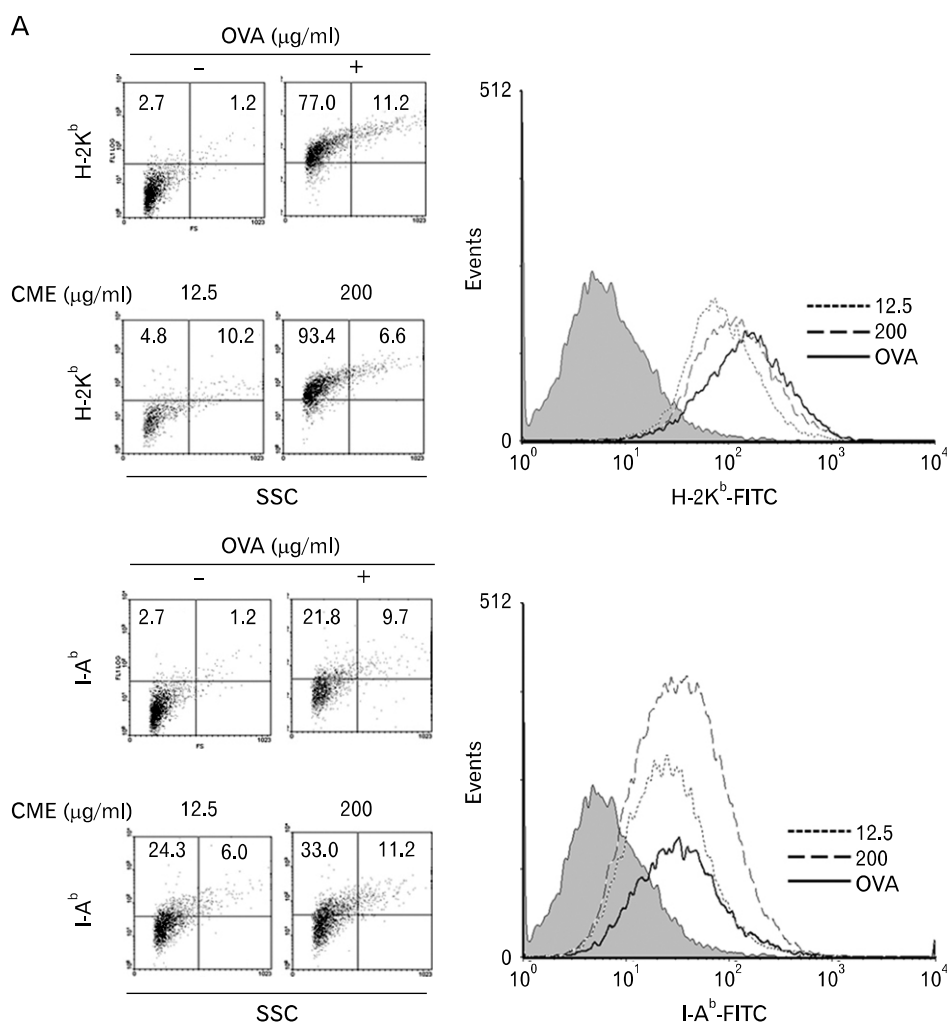
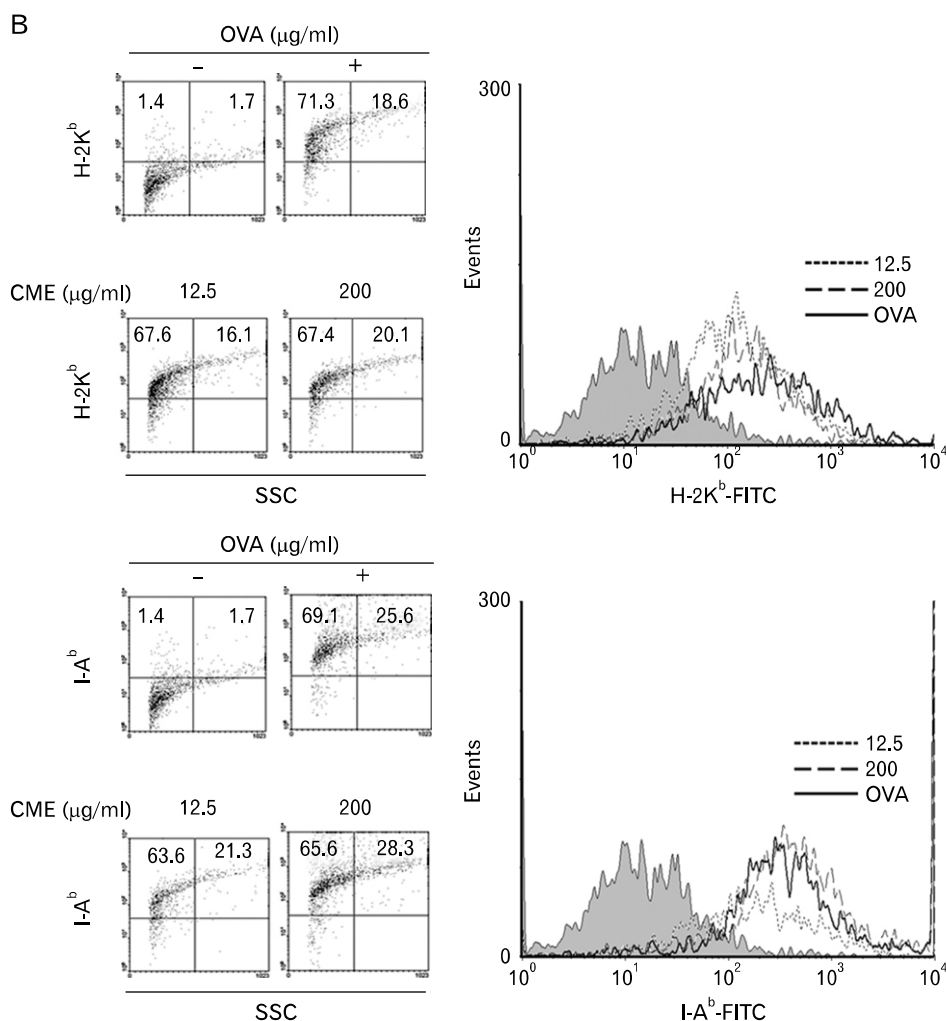


Figure 4. Effects of CME on the expression of MHC molecules. (A) DC2.4 cells and (B) BM-DCs were cultured with CME for 2 hrs, and then the cells were harvested by gentle pipetting. The expression levels of class I and class II MHC molecules were assessed using anti-H-2K^b and anti-I-A^b monoclonal antibodies.



formed with CD8OVA1.3 cells. After 18 hrs incubation at 37°C and 5% CO₂, the supernatant was collected, and then assayed for IL-2 content using an IL-2 ELISA kit. As shown in Fig. 1, CME enhanced MHC class I-restricted OVA presentation. The effects of CME on the class II presentation of exogenous OVA in both BM-DCs and peritoneal macrophages were examined. DCs were generated from bone marrow cells of BALB/C mice and peritoneal macrophages were elicited by injection of thioglycolate into mouse peritoneum. Subsequently cells were treated for 2 hrs with CME, allowed to phagocytose OVA-microspheres for 6 hrs, and then unphagocytosed OVA microspheres and CME were removed by washing, after which a class II MHC-restricted presentation assay was performed with DOBW cells. After 18 hrs incubation

at 37°C and 5% CO₂, the supernatant was collected, and then assayed for IL-2 content using an IL-2 ELISA kit. As shown in Fig. 2, MHC class II-restricted presentation of exogenous OVA was significantly increased by CME in both BM-DCs and peritoneal macrophages.

CME did not change the phagocytic activity of DCs

To prove that the augmentation of antigen presentation by CME was not due to increased phagocytic activity of DCs, FITC-conjugated OVA microspheres were prepared according to the method described above. DCs were incubated with CME for 2 hrs. Then microspheres containing both OVA and FITC were added. After 2 h, DCs were washed and then harvested. The level of phagocytosis was determined by a

flow cytometry. As shown in Fig. 3, flow cytometric analysis of the harvested cells neither of the drugs induced the changes in the phagocytic activity of DCs.

CME enhanced the expression of MHC molecules

To measure the effects on the expression level of total MHC molecules of DCs which were pre-treated with the indicated concentration of CME, DCs were stained with anti-H-2K^b (MHC I molecule) and anti-I-A^b (MHC II molecule). As shown in Fig. 4, CME increased the expression of MHC class I and II molecules.

CME increased pro-inflammatory cytokines and cytokine related enzymes via NF- κ B activation

To further investigate the important role of CME on the im-

mune response, inducing peritoneal macrophages for overnight with LPS or CME resulted in the production of pro-inflammatory cytokines. As shown in Fig. 5, CME concentration-dependently induced the expression of IL-1 β , IL-6, and TNF- α protein (Fig. 5A) as well as cytokine-related enzymes such as, iNOS and COX-2 protein (Fig. 5B). Also, to investigate whether CME could affect nuclear translocation of NF- κ B, a western blot analysis for NF- κ B p65 was carried out with cell lysate in macrophages (Fig. 5C). The amount of NF- κ B p65 markedly increased upon exposure to LPS alone; CME also increased NF- κ B p65.

DISCUSSION

Cordyceps militaris is a herbal ingredients, which has been used as a medicinal product for a long time in Oriental medicine for patients suffering from cancer (12) Some immunomodulatory and antitumor activities of CM have been established (13,14). Also, CM has been shown to inhibit tube formation of endothelial cells in vitro and in vivo and to repress the growth of melanoma cells in mice (12). However, little is known about its immunomodulatory effects or the mechanism of its effects on DCs.

This is the first report of which we are aware concerning the effects of CME on the both DCs and peritoneal macrophages. In this study, we have shown that DCs exposed to CME are enhanced with respect to MHC-restricted exogenous antigen presenting capability. Further, CME increased MHC class I and class II-restricted exogenous antigen presenting capability of DCs leading to induced expression of total MHC molecules. However, CME did not affect phagocytic activity of exogenous antigen.

Since T cells can only recognize antigens presented on MHC molecules, the impact of the inhibition of MHC-restricted antigen presentation must be far-reaching. DCs, a professional APCs, play key role in the initiation of primary immune responses (7) and are capable of processing cell-associated antigens for presentation on MHC class I molecules (15). In vitro, several forms of antigens can access the exogenous pathway for cross presentation, apparently through phagocytic or nonphagocytic mechanisms (16). Further, DCs can acquire and process antigens in the periphery and migrate to secondary lymphoid tissues where they prime primary T cell responses. The classical paradigm of antigen presentation by APCs is that endogenous antigens are presented via MHC class I molecules to CD8 T cells, whereas exogenous

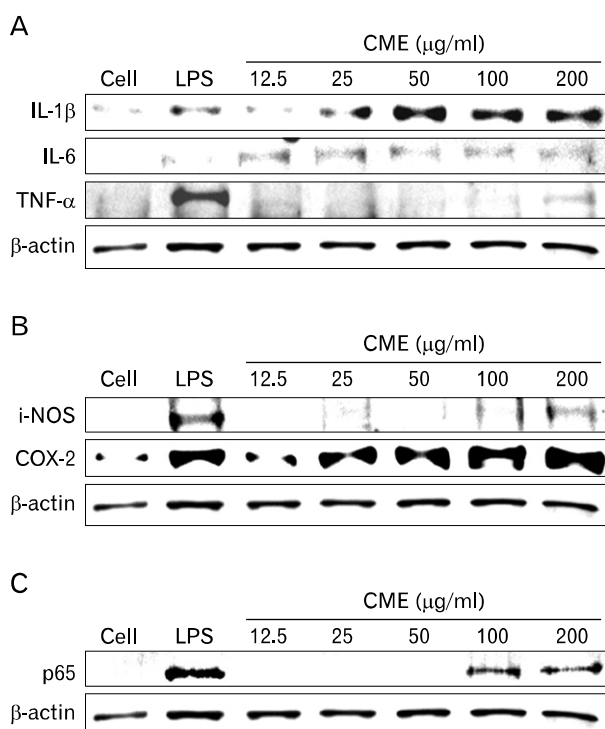


Figure 5. Effects of CME on the expression of pro-inflammatory cytokines, cytokine-related enzymes, and NF- κ B protein. Cells were incubated overnight with various concentrations of CME (12.5~200 μ g/ml) in the absence or presence of LPS (100 ng/ml). The expression of (A) pro-inflammatory cytokines such as IL-1 β , IL-6, and TNF- α , (B) cytokine-related enzymes, iNOS and COX-2, and (C) NF- κ B protein was examined. Total cellular proteins (20 μ g) were resolved by SDS-PAGE, transferred to nitrocellulose membranes, and detected with specific antibodies, as described in the Materials and Methods. β -actin was used as a control. The experiment was repeated in triplicate and similar results were obtained in all three.

antigens are presented via CD4 T cells (17). However, APCs, process exogenous antigens for presentation by MHC class I molecules to CD8 T cells. This process, termed cross presentation, may be a mechanism by which naïve T cells can be primed to antigens that are present in nonprofessional APCs (18,19). The results presented in this paper demonstrate that short-term exposure of DCs to CME significantly enhances both MHC class I and II-restricted antigen presentation capabilities of DCs. Based on the importance of DCs in the initiation of T cell responses and the current finding, which demonstrated that short-term exposure to CME induced MHC-restricted antigen presentation, we are tempted to speculate that the therapeutic efficacy of CME is due, at least in part, to the enhancement of the antigen presenting capability of DCs.

Also, macrophages are unique cells in the immune system in that they not only can initiate immune responses, but also can serve as effector cells. Furthermore, activated macrophages become more efficient antigen presenting cells because they express increased levels of class II MHC molecules and co-stimulatory molecules (6,20,21).

Thus, based on the results of the present study, which showed that CME can regulate MHC class I and class II-restricted presentation of exogenous antigens through an increase in the expression of proinflammatory cytokine proteins, but do not affect phagocytosis in DCs, we are tempted to speculate that some of therapeutic efficacies such as immune-enhancer agents for immunodeficiency diseases.

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CONFLICTS OF INTEREST

The authors have no financial conflict of interest.

REFERENCES

1. Zhang Y, Kiyohara H, Matsumoto T, Yamada H: Fractionation and chemical properties of immunomodulating polysaccharides from roots of *Dipsacus asperoides*. *Planta Med* 63:393-399, 1997
2. Matsumoto T, Sun XB, Hanawa T, Kodaira H, Ishii K, Yamada H: Effect of the antiulcer polysaccharide fraction from *Bupleurum falcatum* L. on the healing of gastric ulcer induced by acetic acid in rats. *Phytother Res* 16:91-93, 2002

3. Huang LF, Liang YZ, Guo FQ, Zhou ZF, Cheng BM: Simultaneous separation and determination of active components in *Cordyceps sinensis* and *Cordyceps militaris* by LC/ESI-MS. *J Pharm Biomed Anal* 33:1155-1162, 2003
4. Shin S, Moon S, Park Y, Kwon J, Lee S, Lee CK, Cho K, Ha NJ, Kim K: Role of cordycepin and adenosine on the phenotypic switch of macrophages via induced anti-inflammatory cytokines. *Immune Netw* 9:255-264, 2009
5. Yu R, Wang L, Zhang H, Zhou C, Zhao Y: Isolation, purification and identification of polysaccharides from cultured *Cordyceps militaris*. *Fitoterapia* 75:662-666, 2004
6. Shin S, Kwon J, Lee S, Kong H, Lee S, Lee CK, Cho K, Ha NJ, Kim K: Immunostimulatory effects of *cordyceps militaris* on macrophages through the enhanced production of cytokines via the activation of NF-kappaB. *Immune Netw* 10:55-63, 2010
7. Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, Liu YJ, Pulendran B, Palucka K: Immunobiology of dendritic cells. *Annu Rev Immunol* 18:767-811, 2000
8. Harding CV, Collins DS, Kanagawa O, Unanue ER: Liposome-encapsulated antigens engender lysosomal processing for class II MHC presentation and cytosolic processing for class I presentation. *J Immunol* 147:2860-2863, 1991
9. Shen Z, Reznikoff G, Dranoff G, Rock KL: Cloned dendritic cells can present exogenous antigens on both MHC class I and class II molecules. *J Immunol* 158:2723-2730, 1997
10. Lee JK, Lee MK, Yun YP, Kim Y, Kim JS, Kim YS, Kim K, Han SS, Lee CK: Acemannan purified from *Aloe vera* induces phenotypic and functional maturation of immature dendritic cells. *Int Immunopharmacol* 1:1275-1284, 2001
11. Lee YH, Lee YR, Im SA, Park SI, Kim KH, Gerelchuluun T, Song S, Kim K, Lee CK: Calcineurin inhibitors block MHC-restricted antigen presentation in vivo. *J Immunol* 179:5711-5716, 2007
12. Yoo HS, Shin JW, Cho JH, Son CG, Lee YW, Park SY, Cho CK: Effects of *Cordyceps militaris* extract on angiogenesis and tumor growth. *Acta Pharmacol Sin* 25:657-665, 2004
13. Liu J, Yang S, Yang X, Chen Z, Li J: Anticarcinogenic effect and hormonal effect of *Cordyceps militaris* Link. *Zhongguo Zhong Yao Za Zhi* 22:111-113, 1997
14. Zhao-Long W, Xiao-Xia W, Wei-Ying C: Inhibitory effect of *Cordyceps sinensis* and *Cordyceps militaris* on human glomerular mesangial cell proliferation induced by native LDL. *Cell Biochem Funct* 18:93-97, 2000
15. Brossart P, Bevan MJ: Presentation of exogenous protein antigens on major histocompatibility complex class I molecules by dendritic cells: pathway of presentation and regulation by cytokines. *Blood* 90:1594-1599, 1997
16. Larsson M, Fonteneau JF, Bhardwaj N: Dendritic cells resurrect antigens from dead cells. *Trends Immunol* 22:141-148, 2001
17. Guermontprez P, Valladeau J, Zitvogel L, Théry C, Amigorena S: Antigen presentation and T cell stimulation by dendritic cells. *Annu Rev Immunol* 20:621-667, 2002
18. Bevan MJ: Cross-priming for a secondary cytotoxic response to minor H antigens with H-2 congenic cells which do not cross-react in the cytotoxic assay. *J Exp Med* 143:1283-1288, 1976

19. Harding CV: Phagocytic processing of antigens for presentation by MHC molecules. Trends Cell Biol 5;105-109, 1995
 20. Adams DO, Hamilton TA: The cell biology of macrophage activation. Annu Rev Immunol 2;283-318, 1984
 21. Fujiwara N, Kobayashi K: Macrophages in inflammation. Curr Drug Targets Inflamm Allergy 4;281-286, 2005
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