Inhibitory effect of Plant Essential Oils on Malassezia pachydermatis

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Received June 8, 2010; Accepted September 6, 2010

Effect of the plant essential oils on the growth of Malassezia pachydermatis was evaluated and the essential oils of Ocimum basilicum L., Melaleuca alternifolia (Maid. & Bet.) Cheel, and Rosa damascene Mill. were the most active against M. pachydermatis and their activity were high than that of itraconazole at 2 mg/mL. The major constituents of the three oils by GC-MS analysis were linalool (21.83%) and estragole (74.29%) for O. basilicum, ã-terpinolene (17.96%) and terpinen-4-ol (45.54%) for M. alternifolia, and â-citronellol (59.98%) and geraniol (27.58%) for R. damascene. Results showed that these selected three oils could be effective toward controlling M. pachydermatis opportunistic infections.

Keywords: anti-Malassezial activity, essential oil, GC-MS analysis, Malassezia pachydermatis

The yeasts of the *Malassezia* genus are common commensal organisms of the skin of humans, birds and many domestic and wild animal species [Crespo *et al.*, 2002; Coutinho *et al.*, 2006]. Today 10 lipid-dependent species are known (i.e. *M. dermatis, M. equi, M. furfur, M. globosa, M. japonica, M. nana, M. obtusa, M. restricta, M. slooffiae and M. sympodialis*), together with *M. pachydermatis* which is not dependent on lipid supplementation for *in vitro* growth [Hirai *et al.*, 2004]. In humans, *M. pachydermatis* was the responsible agent of infection both in immunocompetent [Ming Fan *et al.*, 2006], and in immunocompromised subjects [Midgley, 2000].

Yeasts are considered to be normal constituents of the feline ear microflora [Claudia *et al.*, 2005]. However, the pathogenic role of these yeasts has been recognized in various animals, mainly otitis externa and dermatitis disorders [Guillot and Bond, 1999; Crespo *et al.*, 2002].

Azoles, such as ketoconazole and itraconazole, are antifungal agents available in clinical practice, but they are highly toxic and expensive to use in prolonged treatments [Martinez Fernandez *et al.*, 1998]. Therefore, the development of more economical antifungal agents is required.

In this communication, we have analysed the antifungal properties of plant essential oils on *Malassezia pachydermatis*. The most active *Chamomilla recutita* (L.) Rauschert and *Artemisia dracunculus* L. essential oils were phytochemically examined by GC-MS analysis, and its main constituents were identified.

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doi:10.3839/jabc.2010.033

In our experiments, 87 plant essential oils were used for antifungal activity tests, and they were purchased from UNIQ F&F Co., Ltd. (Seoul, Korea). The detailed information of these essential oils is shown in Table 1. Itraconazole was purchased from Sigma-Aldrich (St. Louis, MO, USA). All other chemicals were a reagent grade.

Malassezia pachydermatis (KCCM 50374) was obtained from the Korean Culture Center of Microorganisms (Seoul, Korea). This strain was grown on Sabouraud Dextrose Broth (SDB) or Sabouraud Dextrose Agar (SDA) (Difco, Sparks, MD, USA) supplemented with 1% (v/v) of pure olive oil (Yakuri Pure Chemicals, Kyoto, Japan), following incubation at 37°C during 2-7 days. Malassezia strains were maintained on the same medium described previously, at 4°C, with subcultures being carried out on a monthly basis. The same medium was used in all the experiments. Inoculum suspensions were prepared by the method as described previously [Rukayadi et al., 2006]. One milliliter of 48 h culture was centrifuged (3000 g at 4°C for 1 min), followed by washing the pellets twice with 1 mL of phosphate buffered saline (PBS). Clusters of Malassezia cells were formed upon preparation of inoculum suspensions. The washing of these suspensions with PBS promotes single-cell status and more accurate turbidity measurements.

The antifungal activity of the plant essential oils was carried out by the disk diffusion method [Anesini and Perez, 1993] using $100~\mu L$ of suspension containing 5×10^6 CFU/mL of *M. pachydermatis*. The disks (Whatman, 6 mm in diameter) which impregnated with 2, 1.5, 0.5, and 0.1 mg/mL of essential oil were placed on the inoculated agar. Control disk containing only ethanol employed to dissolve the essential oil showed no inhibition. Itraconazole of four concentrations were used as positive reference standards. The antifungal activity was

evaluated by measuring the inhibition-zone diameter observed after 48 h of incubation.

The analysis was performed by gas chromatography-mass spectrometry (6890, Aglient Co., Palo Alto, CA, USA). Helium as carrier gas for individual constituents was averaged. The amount of the samples injected was $0.2~\mu L$ in split mode (400:1). The injector temperature was set at $270^{\circ}C$. The GC column was DB- 5MS stationary phase (60 m×0.32 mm i.d., 0.25 μ m film thickness, J&W Scientific, Folsom, CA, USA). The GC oven temperature was initially maintained at 60°C for 2 min and then programmed to 5°C/min to 300 and maintained for 5 min. $0.2~\mu L$ of sample dissolved in CH_2Cl_2 (1:100 v/v) was injected. Essential oil samples were analyzed and the relative peak areas for individual constituents averaged. Quantification

was computed as the percentage contribution of each compound to the total amount present. The percentage composition of the oils was computed by the normalization method from the GC peak areas. GC-MS analysis of the essential oil with the GC (6890 Plus, Agilent Co.) coupled with 5973 mass selective detector quadrupole mass spectrometer. The mass spectrometer was run in the electron impact (EI) mode with electron energy at 70 eV. The mass spectrometer was operated in full scan mode between 35 and 700 amu. The components of essential oil were tentatively identified by comparison of mass spectra of each peak with those of authentic samples in the NIST MS library.

Antifungal activity of 86 plant essential oils was found when the essential oil was assayed at 2 mg/mL (Table 1). The results showed the inhibitory effects of 11 oils [Ocimum basilicum L.,

Table 1. Antifungal activities against Malassezia pachydermatis of 86 plant essential oils

Oil	Family	Species	Part	Antifungal activity (cm)
Cananga	Annonaceae	Cananga odorata Hook fil. et Thomp.	flower	-
Ylang ylang	Annonaceae	Cananga odorata Hook. f. et Thomson	flower	-
Anise	Apiaceae	Pimpinella anisum L.	fruit	-
Aniseed	Apiaceae	Pimpinella anisum L.	fruit	-
Carrot seed	Apiaceae	Daucus carota L.	seed	-
Celery seed	Apiaceae	Apium graveolens L.	seed	-
Coriander	Apiaceae	Coriandrum sativum L.	flower	-
Fennel	Apiaceae	Foeniculum vulgare Mill.	seed	-
Galbanum	Apiaceae	Ferula galbaniflua Boiss.et Buhse	root	-
Lovage root	Apiaceae	Levisticum officinale L. Koch	fruit	-
Parsley herb	Apiaceae	Petroselinum crispum (Mill.) Nyman	whole plant	-
Parsley seed	Apiaceae	Petroselinum crispum (Mill.) Nyman	seed	-
Star anise	Apiaceae	Illicium verum L.	fruit	-
Armoise	Asteraceae	Artemesia vulgaris L.	whole plant	
Chamomile blue	Asteraceae	Chamomilla recutita (L.) Rauschert	flower	-
Chamomile roman	Asteraceae	Chamaemelum nobil (L.) All.	flower	_
Davana	Asteraceae	Artemisia pallens Wall. Ex DC	whole plant	-
Helichrysum	Asteraceae	Helichrysum angustifolium DC	flower	-
Estragon	Asteraceae	Artemisia dracunculus L.	leaf	-
Wormwood	Asteraceae	Artemisia absinthium L.	flower	-
Frankincense	Burseraceae	Boswellia thurifera Roxburgh	root	-
Myrrh	Burseraceae	Commiphora myrrha var. molmol Engl.	stem	.
Tarragon	Asteraceae	Artemisia dracunculus L.	stem	-
Yarrow	Asteraceae	Achillea millefolium L.	flower	-
Cade	Cupressaceae	Juniperus oxycedrus L.	wood	-
Cedarleaf	Cupressaceae	Thuja occidentalis L.	leaf	-
Cedarwood	Cupressaceae	Juniperus irginiana L.	bark	-
Cedarwood Chinese	Cupressaceae	Juniperus funebris Endl.	bark	-
Cedarwood Texas	Cupressaceae	Juniperus mexicana Spring.	bark	-
Cypress	Cupressaceae	Cupressus sempervirens L.	twig	-
Juniperberry	Cupressaceae	Juniperus communis L.	berry	<u></u>
Wintergreen	Ericaceae	Gaultheria procumbens L.	leaf	-
Cascarilla bark	Euphorbiaceae	Croton eleuteria Bennett	bark	-
Basil	Lamiaceae	Ocimum basilicum L.	flower	1.5
Basil sweet	Lamiaceae	Ocimum basilicum L.	whole plant	.
Clary sage	Lamiaceae	Salvia sclarea L.	flower	-
Hyssop	Lamiaceae	Hyssopus officinalis L.	leaf	=
Lavender	Lamiaceae	Lavaendula officinalis (Chaiz.)	flower	-
Lavender 10/42	Lamiaceae	Lavandula angustifolia Mill.	flower	_

Table 1. Continued.

Oil	Family	Species	Part	Antifungal activity (cm)
Marjoram	Lamiaceae	Thymus mastichina L.	leaf	-
Melissa	Lamiaceae	Melissa officinalis L.	leaf	•
Patchouly	Lamiaceae	Pogostemon cablin (Blanco) Benth.	leaf	1.2
Peppermint	Lamiaceae	Mentha piperita L.	flower	<u>.</u>
Rosemary	Lamiaceae	Rosmarinus officinalis L.	flower	-
Sage	Lamiaceae	Salvia officinalis L.	whole plant	-
Sage Dalmatian	Lamiaceae	Salvia officinalis L.	leaf	-
Sage Spanish	Lamiaceae	Salvia lavandulaefolia Vahl.	leaf	**
Spearmint	Lamiaceae	Mentha spicata L.	flower	-
Cassia especial	Lauraceae	Cinnamomum cassia Bl.	bark	-
Cinnamon bleached	Lauraceae	Cinnamomum zeylanicum Garc. Ex Blume Nees	bark	•
Cinnaom leaf oil terpenes	Lauraceae	Cinnamomum zeylanicum Garc. Ex Blume Nees	leaf	-
Rosewood	Lauraceae	Aniba roseadora var. amazonica Ducke	wood	-
Nutmeg	Myristicaceae	Myristica fragrans Houtt.	seed	
Eucalyptus	Myrtaceae	Eucalyptus globulus Labill.	leaf	-
Eucalyptus 80/85	Myrtaceae	Eucalyptus globulus Labill.	leaf	-
Myrtle	Myrtaceae	Myrtus communis L.	leaf	-
Niaouli	Myrtaceae	Melaleuca viridiflora Sol. Ex Gaertn.	leaf	-
Tea tree	Myrtaceae	Melaleuca alternifolia (Maid. & Bet.) Cheel	leaf	1.5
Jasmin absolute	Oleaceae	Jasminum grandiflorum L.	flower	. -
Pine	Pinaceae	Pinus sylvestris L.	needle	<u></u>
Pine needle	Pinaceae	Pinus sylvestris L.	needle	· .
Black pepper	Piperaceae	Piper nigrum L.	fruit	
Geranium	Poaceae	Pelargonium graveolens L.	flower	0.9
Lemongrass	Poaceae	Cymbopogon citratus (DC) Stapf.	whole plant	1,2
Vetiver Haiti	Poaceae	Vetiveria zizanioides L.	root	-
Bergamot	Rutaceae	Citrus bergamia Risso	peel	-
Buchu	Rutaceae	Agathosma crenulata (L.) Pillans	leaf	
Buchu leaf	Rutaceae	Agathosma betulina (Berg.) Pillans	leaf	-
Grapefruit	Rutaceae	Citrus paradisi Macfadyen	fruit	0.9
Orange	Rutaceae	Citrus sinensis (L .) Osbeck	peel	-
Lemon	Rutaceae	Citrus limonum L.	peel	-
Lemon 10F	Rutaceae	Citrus limonum L.	peel	-
Lime dis 5F	Rutaceae	Citrus aurantifolia Swing.	peel	-
Mandarine	Rutaceae	Citrus reticulata Blanco	peel	-
Neroli	Rutaceae	Citrus aurantium L.	flower	= .
Petitgrain	Rutaceae	Citrus aurantium L. subp. amara	leaf	-
Tangerine	Rutaceae	Citrus reticulata Blanco	peel	-
Sandalwood	Santalaceae	Santalum album L.	wood	0.9
Valerian	Valerianaceae	Valeriana officinalis L.	rhizome	-
Rose	Rosaceae	Rosa damascene Mill.	flower	1.8
Chamomile blue	Asteraceae	Chamomilla recutita (L.) Rauschert	flower	1.4
Estragon	Asteraceae	Artemisia dracunculus L.	leaf	1.2
Tamanu	Clusiaceae	Calophyllum inophyllum L.	fruit	- · · · · · · · · · · · · · · · · · · ·
Xanthoxylum	Rutaceae	Zanthoxylum armatum	seed	-
Eucalyptus	Myrtaceae	Eucalyptus citriodora	leaf	0.9
Ginger	Zingiberaceae	Zingiber officinale Roscoe	rhizome	- -
Itraconazole		G		1.5

Samples treated with concentration of 2 mg. '-' expressed no activity.

Pogostemon cablin (Blanco) Benth., Melaleuca alternifolia (Maid. & Bet.) Cheel, Pelargonium graveolens L., Cymbopogon citratus (DC) Stapf., Citrus paradisi Macfadyen, Santalum album L., Rosa damascene Mill., Chamomilla recutita (L.) Rauschert, Artemisia dracunculus L., and Eucalyptus citriodora]

on *M. pachydermatis* at 2 mg/mL. Among the 11 active oils, the oils of *Ocimum basilicum* L., *Melaleuca alternifolia* (Maid. & Bet.) Cheel, and *Rosa damascene* Mill. showed strong inhibitory activity at a dose dependant manner and the activity were higher than itraconazole at the same concentration (Fig. 1).

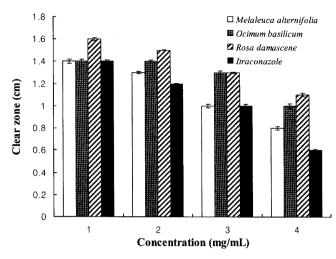


Fig. 1. Antifungal activity of essential oil of *Ocimum basilicum*, *Melaleuca alternifolia*, and *Rosa damascene* against *Malassezia pachydermatis*. Values represent the means of 3 independent experiments.

Table 2. Chemical composition of Melaleuca alternifolia essential oil

No.	Compound	RI ^a	Relative composition ratio, %
1	α-thujene	921	0.65
2	α-pinene	928	2.50
3	β-pinene	967	0.59
4	β-myrcene	981	6.44
5	α-terpinene	1,007	7.25
6	p-cymene	1,010	5.32
7	1,8-cineole	1,018	3.72
8	Limonene	1,019	0.99
9	γ-terpinene	1,049	17.96
10	Terpinolene	1,077	2.81
11	Terpinen-4-ol	1,159	45.54
12	α-terpineol	1,170	3.26
13	Armodendrene b	1,435	1.02
14	Cadinene ^b	1,514	0.88
Total			98.93

^aRetention indices.

Table 3. Chemical composition of Ocimum basilicum

No.	Compound	RI ^a	Relative composition ratio, %
1	Linalool	1,084	21.83
2	Estragole	1,173	74.29
3	$lpha$ -Humulene $^{ extsf{b}}$	1,447	2.17
Total			98.29

aRetention indices.

The three oils were analysed by GC-MS. The chemical compositions of O. basilicum oil are shown in Table 2~4. Altogether 3 compounds were identified, representing 98.29% of the total oil constituents. The major constituents of the oil were linalool (21.83%) and estragole (74.29%). M. alternifolia oil composed of major constituents of the γ -terpinolene

Table 4. Chemical composition of Rosa damascene

No.	Compound	RI a	Relative composition ratio, %
1	Linalool	1,084	3.56
2	Roseoxide	1,096	1.72
3	β-Citronellol	1,211	59.98
4	Nerol ^b	1,214	1.75
5	Geraniol	1,236	27.58
6	2-Phenylethyl acetate b	1,242	2.04
7	Geranyl acetate	1,360	0.75
8	Methyl eugenol	1,370	1.24
	Nonadecane b	1,872	1.39
Total			100.0

^aRetention indices

(17.96%) and terpinen-4-ol (45.54%). The major constituents of *R. damascene* were β -citronellol (59.98%) and geraniol (27.58%).

Consequently, we demonstrated that the 11 oils among the 86 plant essential oils had inhibitory activity against *M. pachydermatis* at first screening. The highest antifungal activity was found in the essential oils of *O. basilicum*, *M. alternifolia*, and *R. damascene*. This activity was dose-dependant and higher than that of itraconazole. Although, further studies are needed, the use of essential oils of *O. basilicum*, *M. alternifolia*, and *R. damascene* against microbial growth seems a valuable alternative as antifungal compound, especially in the cases of anti-*Malassezia* resistance.

References

Anesini C and Perez C (1993) Screening of plants used in Argentine folk medicine for antimicrobial activity. *J Ethnopharmacol* **39**, 119-128.

Claudia C, Sabrina G, Gioia C, and Domenico O (2005)
Occurrence and population size of Malassezia spp. in the external ear canal of dogs and cats both healthy and with otitis.

Mycopathologia 160, 143-149.

Coutinho SD, Fedullo JD, and Corrêa SH (2006) Isolation of Malassezia spp. from cerumen of wild felids. Med Mycol 44, 383-387

Crespo MJ, Abarca ML, and Cabañes FJ (2002) Occurrence of Malassezia spp. in the external ear canals of dogs and cats with and without *otitis externa*. *Med Mycol* **40**, 115-121.

Crespo CMJ, Abarca ML, and Cabañes FJ (2002) Occurrence of *Malassezia* spp. in horses and domestic ruminants. *Mycoses* **45**, 333-337.

Guillot J and Bond R (1999) Malassezia pachydermatis: a review. *Med Mycol* **37**, 295-306.

Hirai A, Kano R, and Makimura K (2004) *Malassezia nana* sp. nov, a novel lipid-dependent yeast species isolated from animals. *Int J Syst Evol Microbiol* **54**, 623-627.

Martinez Fernandez CA, Carnot Uría J, De Castro Arenas R, Muñío Perureral J, and Torres Ilibar W (1998) Tratamiento de las micosis profundas. *Estado actual Acta med* **8**, 80-85.

^bTentatively identified by mass library.

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Ming Fan YM, Huang WM, Li SF, Wu GF, Lai K, and Chen RY (2006) Granulomatous skin infection caused by *Malassezia pachydermatis* in a dog owner. *Arch Dermatol* **142**, 1181-1184. Midgley G (2000) The lipophilic yeasts: state of the art and

prospects. *Med Mycol* **38**, 9-16. Rukayadi Y, Yong D, and Hwang JK (2006) In vitro anticandidal

activity of xanthorrhizol isolated from Curcuma xanthorrhiza Roxb. *J Antimicrob Chemother* **57**, 1231-1234.