

Ultrastructure of *Sarcocystis grueneri*-like Sarcocysts from Cardiac Muscle of Red Deer (*Cervus elaphus*) in Korea

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Abstract: *Sarcocystis grueneri*-like sarcocysts were found from the cardiac muscles of a rearing red deer (*Cervus elaphus*) carcass in Korea. In the light microscopical examination of sarcocysts, they were oval to spherical cysts and 90-170 × 110-380 μ m in size. However, there was no inflammation and myofiber degeneration. In the transmission electron microscope, these cysts were located within the sarcoplasm of the host cell and filled with merozoites. The sarcocysts were enclosed by a very thin wall (0.45-0.6 μ m thick) that consists of protrusions and ground substance. The primary cyst wall formed numerous strip-like protrusions which were 0.2-0.3 μ m wide and up to 4.2 μ m long. The protrusions were running in parallel with the surface of the cyst. A characteristic of the cyst wall was absent of fibrils inside the protrusions. Merozoites in the compartment measured about 15 × 4 μ m. The merozoite consisted of four regions: micronemes and rhoptries, amylopectin granule, nucleus, and amylopectin granules. The number of rhoptry was counted in 7-13.

Key words: Sarcocystis grueneri, Ultrastructure, Cervus elaphus, Korea.

Introduction

The Genus *Sarcocystis* Lankester, 1882 is related to protozoa and the largest genera of the phylum Apicomplexa that can cause mortality in many species of domestic and wild animals (30). The taxonomy of *Sarcocystis* was the list of 122 species with their synonyms and hosts given. Both definitive and intermediate hosts are known for only 56 species. The fine structure of the sarcocyst wall may change with age and not considered necessarily satisfactory for separating species (21).

Sarcocysts are detected in muscles of cervids (*i.e.* red deer, roe deer, mule deer, white-tailed deer, wapiti, moose). The cervids act as intermediate host of *Sarcocystis*. They have been intensively studied (2,4,5,6,8,9,19,29), and several species or cyst types are commonly found in the same host (1,24). Morphological studies of sarcocysts from different cervids have suggested that they harbour *Sarcocystis* spp. with highly similar cyst wall morphology. The descriptions of species have been hitherto based in part on cyst morphology, particularly at the ultrastructural level, and on life-cycle studies of this obligatory two-host parasite (7). Although a lot of papers have described the cyst with morphology and the identification of the final host, in some cases neither of these have been established. There are several named species of *Sarcocystis* in cervids. The roe deer has been known as an intermediate host of *S. gracilis* Ratz, 1909 for a long time. Erber *et al* (12) described two new species, *S. capreolicanis* and unnamed *Sarcocystis* sp. The unnamed *Sarcosystis* sp. of Erber was tentatively classified as *Sarcosystis* cf. *hofimanni* by Sedlaczek and Wesemeier (28), due to the similar morphology of the sarcocysts such as *S. hofimanni* of badgers (26), *S. wapiti* in wapiti (29) and *S. sybillensis* have been reported from North American elk (7).

The present paper describes the ultrastructural morphology of *Sarcocystis grueneri*-like sarcocysts from a naturally infected rearing red deer in Korea.

Materials and Methods

A rearing red deer carcass from a farm in Gyeonggi province was examined for *Sarcocystis* infections. The samples from heart, liver, lung were fixed in 10% neutral buffered formaline, embedded in paraffin, sectioned at 4 μ m, and stained with hematoxylin and eosin for examination by light microscopy.

For electron micriscopy, pieces of heart muscle containing cysts were fixed with 2.5% glutaradehyde solution in 0.1 M phosphate buffer (pH 7.4) for 4 hrs. After washing with buffer, the specimens were post-fixed with 1% osmium tetroxide at 4°C for 4 hrs. Afterward, the specimens were dehydrated in a

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graded ethyl alcohol series and two changes of propylene oxide, and embedded in epon mixture. The ultrathin sections were stained with uranyl acetate and lead citrate and examined in a Hitachi H-600 electron microscope at 75 kV.

Results

Grossly, sarcocysts were easily recognized by their whitish appearance within the dark-red coloured tissue in the heart. In histopathologic examination, the several cysts were found in the heart muscle. They were oval to spherical shaped and measured $90-170 \times 110-380 \ \mu m$ in diameter. There was no inflammation and myofiber degeneration but many sarcocysts were detected (Fig 1).

The general organization of the cyst wall is shown in Fig 2-A. These cysts were located within the sarcoplasm of the host cell and were filled with merozoites. A layer of fibrous material appeared on the outside of sarcoplasm. The sarco-cysts were enclosed by a very thin wall that consists of protrusions and ground substance. The protrusions of the cyst wall



Fig 1. (A) Light micrographs of sarcocysts (arrows) in the cardiac muscle of *Cervus elaphus*. Sarcocysts initiated no tissue reaction. Scale bar = $100 \ \mu m$. (B) Higher magnification of the sarcocyst in A, showing many merozoites. H-E stain, Scale bar = $25 \ \mu m$ (B).



Fig 2. Transmission electron micrographs of cyst of *Sarcocystis grueneri* from the heart of *Cervus elaphus*. (A) The sarcocyst is enclosed by a thin wall that consist of protrusions and ground substance. The protrusions of the cyst wall adjacent to the sarcoplasm of host cells. (B) The cross sectioned anterior region of merozoite containing several rhoptries. (C) The single merozoite (circle). The merozoites contain numerous amylopectin granules. (D) The merozoites in a component (circle). The component separated by thin septa (arrow) of ground substance. *A*, amylopectin granule; *CW*, cyst wall; *F*, fibrous material; *GS*, ground substance; *MN*, micronemes; *N*, nucleus; *PR*, protrusion; *R*, rhoptry; *S*, septa; *SP*, sarcoplasm. Scale bar = 1 μ m.

adjacent to the sarcoplasm (parasitophorous vacuole) of host cells. The primary cyst wall (0.45-0.6 µm thick) was made up of the former parasitophorous vacuole membrane with a border of elongated protrusions that folded over. The primary cyst wall formed extremely thin, strip-like configuration of the cyst wall protrusions which were 0.2-0.3 µm wide and up to 4.2 µm long. The protrusions were running in parallel with the surface of the cyst. The cyst wall was very thin, electrondense, and bears flattened protrusions that did not contain fibrils. The ground substance located around the base of the projections was 0.25-0.3 µm thick and formed by a concentration of electron-dense granules which were also scattered irregularly through the ground substance including the septa (Fig 2-A). The cysts were separated by thin septa into compartments and all sarcocysts had prominent septa (Fig 2-D). A characteristic feature of the cyst wall was absence of fibrillar elements inside the protrusions. Merozoites in the compartment were elongated with a pointed anterior end and a rounded posterior end, measuring about $15 \times 4 \mu m$. The merozoites consisted of four regions: an anterior region containing rhoptries and a large number of micronemes, two fourth region with abundant amylopectin granules, three fourth region containing the nucleus, and a posterior region containing abundant amylopectin granules (Fig 2-B,C). The several rhoptries appeared in cross sectioned anterior region of merozoite. The number of rhoptry was counted in 7-13 (Fig 2-B).

Discussion

This report described the ultrastructure of sarcocyst from red deer in Korea. The cysts found in heart tissue appear thin-walled in the light and electron microscope. All the cysts studied appeared to be in the same developmental stage. The structure of the cysts was typical for *Sarcocystis* spp. in that they have a primary cyst wall, beneath which was a ground substance layer which was continuous with septa dividing the interior of the cysts into compartments (22).

Micrograph has that all Sarcocystis species (S. cervicanis, Sarcocystis sp., S. wapiti and S. sybillence) hitherto reported in the genus Cervus share a common morphological pattern (6,10,18,29). However, there are considerable differences in structure and thickness of cyst walls among the sarcocysts of cervid. The cysts are described as thick-walled (> $10 \mu m$) or thin-walled (1-3 μ m) by the thickness (6,9,10,22). A hypothesis was proposed that the thin-walled cysts of Sarcocystis with band like villar protrusions detected in cervids populating all the Holarctic belong to the same species and named this species S. cf. grueneri by Wesemeier and Sedaczek (31). The name was confirmed as S. grueneri. Also, they reported two more species in red deer, which were named S. cf. capreolicanis and S. cf. hofmanni. There were distinct hair and finger like villar of protrusions, respectively. The finger like protrusions were found in S. gracilis, S. hominis, S. hirsuta, etc. (7,10,13, 27).

Kutkiene (20) reported that three types of cysts differentiated

under a light microscope from red deer were identified as belonging to the species S. cf. capreolicanis, S. cf. hofmanni and Sarcocystis sp. (S. cf. grueneri?). Dahlgren et al (3) reported that the five species were found by the light and scanning electron microscopy or DNA amplification and sequencing at the small subunit rRNA gene. The three sarcocyst types were newly named, *i.e. S. alces, S, ovalis,* and S. scandinavica. He concluded that molecular methods are necessary for unequivocal species identification, as different cervid hosts harbour morphologically indistinguishable sarcocysts.

The characteristic protrusions of primary cyst wall in this study were very similar to those of S. grueneri from reindeer (16). The protrusions of cyst wall with strip-like folds were very thin and measured 0.25-0.3 µm in thickness. Based on their cyst wall structure and location in cardiac muscle, the cysts in this paper belong to as described by Gjerde (15) and Gjerde (16). Gjerde (16) asserted that the name S. grueneri should not be used for a Sarcocystis sp. of the red deer. Because the original descriptions of S. grueneri were mainly concerned with sarcosporidia of reindeer. But the synonym of S. grueneri, S. alceslatrans, S. cervicanis and S. wapiti is very probable (6,16,24,25,28,31,32). In fact, the host was C. elaphus of S. cervicanis by Hernndez-Rodríguez et al (18), Sarcocystis sp. by Entzeroth et al (11) in red deer, S. wapiti by Speer et al (29) in wapiti and S. sybillensis by Dubey et al (6) in North American elk.

The final host in this study was not determined, but it is possible that these deer intermediate hosts may harbour a common species of *Sarcocystis* on the basis of other reports (11,18,29). In general, *Sarcocystis* species are specific for their intermediate hosts. But the occurrence of unusual *Sarcocystis* species has been reported (14,17,23). This paper is the first report of *S. grueneri*-like sarcocysts in red deer in Korea. It will be investigated the future studies to the identification and distribution of this species of sarcocyst in the flocks in Korea.

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한국산 Red Deer (Cervus elaphus)의 심근에서 관찰된 Sarcocystis grueneri 양 포낭의 투과전자현미경 소견

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요 약 : 한국산 양축 엘크시슴 (*Cervus elaphus*)의 심장 조직에서 *Sarcocystis grueneri* 양 포낭을 광학 및 투과전자현 미경으로 관찰하였다. 심장조직의 병리소견에서는 다수의 포낭이 관찰되었으며, 염증 및 근섬유의 변성은 관찰되지 않 았다. 염증 및 근섬유의 변성은 관찰되지 않았으며 다수의 포낭이 관찰되었다. 포낭은 난형에서 난원형 이었으며, 크 기는 90-170 × 110-380 µm로 계측되었다. 투과전자현미경 소견에서 포낭은 숙주세포 유래의 sarcoplasm에 싸여 있었 고 0.45-0.6 µm 두께의 매우 얇은 포낭벽을 갖고 있었으며, 포낭벽은 protrusion 및 ground substance로 구성되어 있었 다. 다수의 길고 가느다란 protrusion 층의 높이는 0.2-0.3 µm, protrusion의 길이는 최장 4.2 µm로 계측되었다. Protrusion은 포낭의 표면과 평행하게 주행하였다. 포낭은 중격에 의하여 소실로 나뉘어져 있었고 소실에는 크기가 약 15 × 4 µm인 다수의 merozoites가 존재하였다. 관찰된 모든 포낭은 같은 발육기를 갖고 있었다. Merozoite는 4개 구역 (micronemes and rhoptries, amylopectin granules, 핵 및 amylopectin granules)으로 구분되었으며 7-11 개의 rhoptry 가 충체의 전방에 분포하였다.

주요어 : Sarcocystis grueneri, 포낭, Cervus elaphus, 투과전자현미경.