

Anti-*Helicobacter pylori* Activity and Structure-Activity Relationships of Berberine Derivatives

Yeong-Hwan Han, Kap-Duk Lee,[†] and Dong-Ung Lee^{*}

Division of Bioscience, Dongguk University, Gyeongju 780-714, Korea. *E-mail: dulee@dongguk.ac.kr

[†]Department of Nanomaterial Chemistry, Dongguk University, Gyeongju 780-714, Korea

Received October 22, 2009, Accepted November 23, 2009

Key Words: *Helicobacter pylori*, Berberine analogs, Urease, Structure-activity relationship

In 1983, Warren and Marshall found that gastritis and most stomach ulcers are caused by *Helicobacter pylori* infection, and not by stress or spicy food, as had been previously assumed.¹ This bacteria is known to produce a toxin that may be potentiated by ammonia generated from urease.² Since that time, many types of anti-*H. pylori* agents focusing on inhibition of *H. pylori* growth and/or anti-urease activity have been chemically synthesized³ or isolated from natural sources⁴ for treatment of patients with gastroduodenal disorders. For example, administration of a mixture of such synthetic compounds as bismuth subsalicylate, tetracycline, and metronidazole for eight weeks was found to inhibit growth of *H. pylori*,⁵ but with adverse side-effects, due to long treatment periods. Natural products, such as decurcin, decurcinol angelate, berberine, and magnolol have been reported to demonstrate moderate inhibition of *H. pylori* growth.⁴

Among the natural compounds tested for anti-*H. pylori* activity to date, we focused on berberine, owing to its extensive anti-bacterial activity including anti-*H. pylori* activity. Berberine is a major constituent of *Coptis chinensis* and *Phellodendron amurense*, which are used in the Orient for treatment of diarrhea and other gastrointestinal diseases. Although berberine was known to inhibit the growth of *H. pylori*,⁶ its derivatives have not thoroughly been investigated yet. We have previously discussed the antimicrobial,⁷ antimalarial,⁸ and anticancer activities⁹ of the synthetic protoberberine-type alkaloids. In the present study, we prepared berberine derivatives in order to evaluate their inhibitory activity against *H. pylori* and urease, which play important roles in pathogenesis of gastric ulcers. Nine synthetic protoberberine alkaloids (four 8-alkyl-dihydroberberines, two 8-alkyl-berberines, and three 12-bromo-8-alkyl-berberines) (Scheme 1), including three unreported analogs, and natural berberine and palmatine were tested for elucidation of both their activity and their structure-activity relationships.

The effect of growth inhibition by berberine (1) against *H. pylori* was much stronger than that of palmatine (2) (Fig. 1), proving that activity of the methylenedioxy group is more significant than that of the methoxy group. Dihydroberberines (5 or 6) showed weaker activity than the corresponding berberines (7 or 8), suggesting that ring C should be aromatic. Inhibitory activity was increased in proportion to the number of carbons at the C-8 position, suggesting that greater lipophilicity is an indication of stronger activity (e.g., 3 < 4 < 5); however, the longer hexyl side chain resulted in decreased activity (6 or 8). Among nine berberine analogs tested, five compounds showed stronger activity than berberine itself. Introduction of an *n*-butyl group at the C-8 position of berberine resulted in significantly increased activity in comparison with other alkyl groups (com-

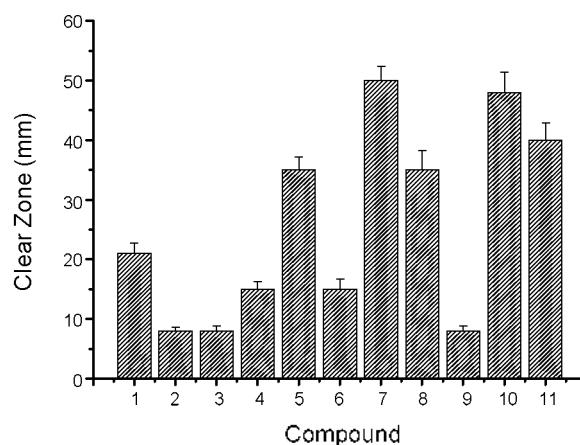
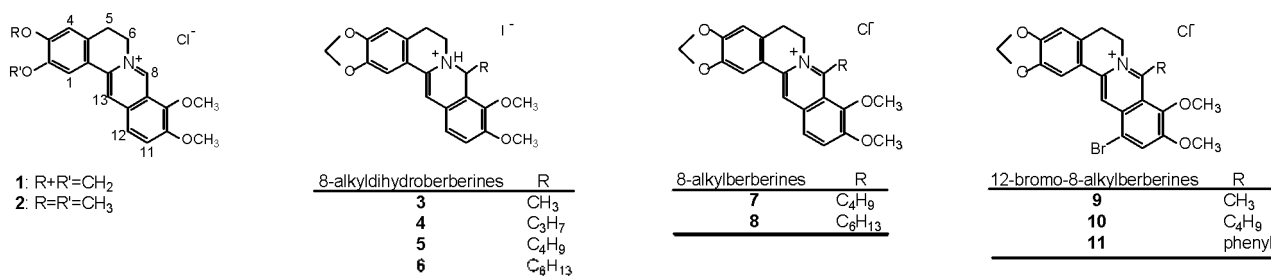


Figure 1. Inhibitory effect of berberine (1), palmatine (2) and synthetic analogs against *Helicobacter pylori* at a final concentration of 1.25 mg/mL. Data represent the mean \pm S. D. ($n = 3$).



Scheme 1. Chemical structures of natural protoberberines (1 and 2) and synthetic compounds tested

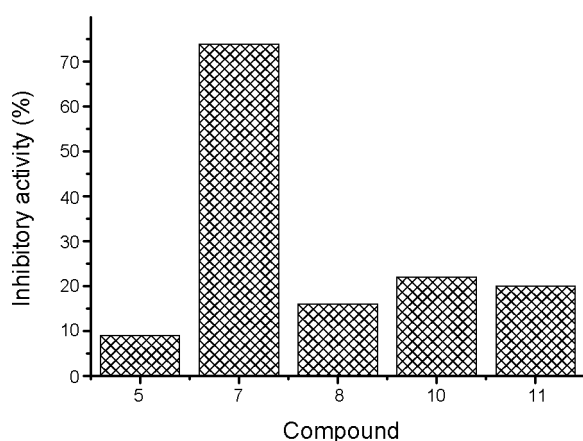


Figure 2. Urease inhibitory effect of some synthetic compounds having stronger anti-*Helicobacter pylori* activity than that of berberine (**1**) at a final concentration of 1.0 mg/mL. $n = 2$.

pare **5** with **3**, **4**, **6**, or **7** with **8**, or **10** with **9**, **11**). Replacement of H-12 of **7** by a bromine atom did not affect activity much. As a result, 8-butyl-substituted berberine derivatives (**7** and **10**) exhibited the most effective anti-*H. pylori* activity.

Our previous reports indicated that the antibacterial and antifungal activities of C-8-substituents increased as the aliphatic chain lengthened,⁷ supporting the above results. However, although an antibacterial activity of **7** was much weaker than that of **10**, anti-*H. pylori* activity of both compounds were nearly the same.

Next, compounds **5**, **7**, **8**, **10**, **11**, which suppress growth of *H. pylori* more than berberine, were investigated for their inhibitory activity against urease. Urease is considered to play a key role in growth of *H. pylori*, because the bacteria can survive in the strongly acidic conditions of the stomach through neutralization of acid with ammonia forming from urease.¹⁰ Among the tested compounds, 8-butylberberine (**7**) revealed potent anti-urease activity, with 73.9% inhibition, at a concentration of 1.0 mg/mL, and the others showed relatively weak activity (Fig. 2). DMSO used as a solvent showed no antibacterial and urease inhibitory activity.

In conclusion, 8-butylberberine (**7**) would be acceptable as a superior anti-*H. pylori* agent, based on its suppressing effect on bacterial growth as well as its inhibitory property on urease activity.

Experimental Section

General. Melting point was measured on an Electrothermal IA9100 apparatus (Thermo Scientific, Pittsburgh, PA, USA), and was uncorrected. NMR spectra were recorded on a Varian VXR-500 (500 MHz). Liquid secondary ion mass spectra (LSIMS, positive ion) were determined on a Hitachi M-4100 instrument, using glycerol as a matrix. Berberine and palmatine were obtained from Sigma-Aldrich (St. Louis, MO, USA). All other compounds tested, with the exception of compounds **6**, **8**, and **9**, were previously prepared.^{6,8}

Preparation of 8-hexyldihydroberberine iodide (6): According to procedures for compounds **3-5**,⁷ 8-hexyl derivative was

prepared using Grignard reagent (mixture of Mg turning and *n*-hexyl iodide in anhydrous ether). Recrystallization with 80% MeOH yielded a dark brown powder. Yield 45%. mp. 146 - 157 °C. ¹H-NMR (DMSO-*d*₆) δ 0.95 (CH₃, t, $J = 7.0$ Hz, 3H), 1.35-1.80 and 3.43 (5 \times CH₂, m), 3.17 (H-5, t, $J = 6.0$ Hz), 4.08 (9-OMe, s), 4.13 (10-OMe, s), 4.80 (H-6, t, $J = 6.0$ Hz), 5.32 (H-8, s), 6.20 (OCH₂O, s), 7.11 (H-4, s), 7.52 (H-1, s), 7.80 (H-12, d, $J = 9.0$ Hz), 8.03 (H-11, d, $J = 9.0$ Hz), 8.52 (H-13, s). PI-LSIMS m/z : 422 [M-I]H⁺ (mw. 549 for C₂₆H₃₂NO₄I).

Preparation of 8-hexylberberine chloride (8): Hydroiodide **6** was dehydrogenated, then converted into the corresponding chloride in the same manner as that of other alkyl substituents,⁷ furnishing **8** as a yellow powder. Yield 52%. mp. 185 - 191 °C. ¹H-NMR (DMSO-*d*₆) δ 0.92 (CH₃, t, $J = 7.0$ Hz, 3H), 1.38-1.82 and 3.49 (5 \times CH₂, m), 3.18 (H-5, t, $J = 6.0$ Hz), 4.05 (9-OMe, s), 4.11 (10-OMe, s), 4.81 (H-6, t, $J = 6.0$ Hz), 6.18 (OCH₂O, s), 7.14 (H-4, s), 7.56 (H-1, s), 7.78 (H-12, d, $J = 9.0$ Hz), 8.09 (H-11, d, $J = 9.0$ Hz), 8.54 (H-13, s). PI-LSIMS m/z 420 [M-Cl]⁺ (mw. 455 for C₂₆H₃₀NO₄Cl).

Preparation of 12-bromo-8-methylberberine chloride (9): **9** was prepared according to the method applied to synthesis of related compounds,⁷ yielding a brown powder. Yield 35%. mp. 204 - 211 °C. ¹H-NMR (DMSO-*d*₆) δ 3.41 (CH₃, s, 3H), 3.27 (H-5, t, $J = 6.0$ Hz), 4.12 (9-OMe, s), 4.22 (10-OMe, s), 4.95 (H-6, t, $J = 6.0$ Hz), 6.13 (OCH₂O, s), 6.98 (H-4, s), 7.68 (H-1, s), 8.43 (H-11, s), 8.61 (H-13, s). PI-LSIMS m/z 428 and 430 [M-Cl]⁺ (mw. 464 for C₂₁H₁₉NO₄BrCl).

Bacteria and growth conditions. *H. pylori* (KCCM 41351) was obtained from the Korean Culture Center of Microorganisms. The bacterium was grown on brucella agar containing 10% fetal bovine serum (FBS) under microaerophilic conditions (5% O₂, 10% CO₂, 10% H₂, and 75% N₂) in anaerobic jars with gas mixture systems at 37 °C for 3 - 5 days.

Bacterial growth inhibition assay. Growth inhibition was performed using the Kirby-Bauer method on brucella agar containing 10% FBS under microaerophilic conditions at 37 °C: 100 μ L of this cell suspension was smeared over the entire surface of a brucella agar plate containing 10% FBS. Berberine derivatives (final concentration 1.25 mg/mL) were dropped onto paper discs, and the latter were placed on agar plates. Diameter of inhibition zones were measured after 3 days of incubation at 37 °C under microaerophilic conditions.

Preparation of urease. Broth cultured cells were harvested by centrifugation (15,000 \times g, 10 min). Harvested *H. pylori* cells were washed with 5 μ L of 20 mM citrate buffer (pH 5.0), then resuspended in the same buffers. Cells were disrupted using a French press (Fred S. Carver Inc., 15,000 psi), centrifuged at 15,000 \times g for 10 min, and supernatants were used for the urease activity assay.

Urease activity assay. Urease activity was assayed according to the known method.¹¹ Reaction mixtures comprising 100 μ L of enzyme solution, 300 μ L of buffer containing 400 mM urea, and 50 μ L of berberine derivatives (final concentration 1.0 mg/mL) were incubated at 37 °C for 30 min, after which 100 μ L 1 N H₂SO₄ was added. For ammonia determination using the indophenol method, 2.5 mL each of phenol reagent (1% w/v phenol and 0.005% w/v sodium nitroprusside) and an alkali reagent (5.5% w/v Na₂HPO₄·12H₂O, 0.5% w/v NaOH, and 0.1%

active chloride NaOCl) were added to each reaction mixture. Following incubation at 65 °C for 20 min, absorbance was measured at 630 nm. Inhibition ratio was calculated as a percentage of that in the control experiment, in which 50 µL of DMSO was added instead of the test compound solution.

References

1. Marshall, B. J.; Warren, J. R. *Lancet*. **1984**, *323*, 1311.
2. Ysujii, M.; Kawano, N.; Tsukii, S.; Fusamoto, H.; Kamada, T.; Sato, N. *Gastroenterol.* **1992**, *102*, 1881.
3. Cheng, N.; Xie, J. S.; Zhang, M. Y.; Shu, C.; Zhu, D. X. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 2703.
4. Mahady, G. B.; Pendland, S. L.; Stoia, A.; Chadwick, L. R. *Phytother. Res.* **2003**, *17*, 217.
5. Hentschel, E.; Brandstatter, G.; Dragsics, B.; Hirschil, A. M.; Nemeč, H.; Schutze, K.; Taufer, M.; Wurzer, H. *N. Engl. J. Med.* **1993**, *328*, 308.
6. Bae, E. A.; Han, M. J.; Kim, N. J.; Kim, D. H. *Biol. Pharm. Bull.* **1998**, *21*, 990.
7. Iwasa, K.; Lee, D. U.; Kang, S. I.; Wiegrebe, W. *J. Nat. Prod.* **1998**, *61*, 1150.
8. Iwasa, K.; Kim, H. S.; Wataya, Y.; Lee, D. U. *Eur. J. Med. Chem.* **1998**, *33*, 65.
9. Iwasa, K.; Moriyasu, M.; Yamori, T.; Turuo, T.; Lee, D. U.; Wiegrebe, W. *J. Nat. Prod.* **2001**, *64*, 896.
10. Celli, J. P.; Turner, B. S.; Afdhal, N. H.; Keates, S.; Ghiran, I.; Kelly, C. P.; Ewoldt, R. H.; McKinley, G. H.; So, P.; Erramilli, S. *Proc. Natl. Acad. Sci. U. S. A.* **2009**, *106*, 14321.
11. Pervez, H.; Iqbal, M. S.; Tahir, M. Y.; Nasim, F. U. H.; Choudhary, M. I.; Khan, K. M. *J. Enzym. Inhib. Med. Chem.* **2008**, *23*, 848.