A Short Path to the 1,3-cis-Substituted Core Skeleton of Tetrahydroisoquinolines

Le Anh Tuan^{†,‡} and Guncheol Kim^{†,*}

Department of Chemistry, College of Natural Science, Chungnam National University, Daejon 305-764, Korea *E-mail: guncheol@cnu.ac.kr

Finstitute of Chemistry, Vietnamese Academy of Science and Technology (VAST), Hanoi, Vietnam Received August 31, 2009, Accepted September 5, 2009

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The family of tetrahydroisoquinoline alkaloids has attracted considerable attention because of their biological activity and novel structures. Natural alkaloids, such as saframycins, renieramycins, and ecteinascidin 743. which are shown in Figure 1 possess potent antitumor, antibiotic, and antimicrobial activities. A number of methods to synthesize these alkaloids have been published recently. 16,5

Efforts to develop strategies to synthesize these alkaloids have encountered the key stereochemical issue of installing the common *cis* relationship at the C1 and C3 positions of the core structures. Several interesting approaches include radical cylization. 6 the addition of organometallic compounds followed by ionic hydrogenation, 7 and intermolecular 8 or intramolecular Pictet-Spengler reaction. 9

We were interested in inducing a *cis* relationship for the related piperidine rings in the course of alkaloid synthesis under reductive cyclization. ¹⁰ and we wanted a new way that uses the existing C3 stereocenter of a natural product to induce the desired stereochemistry between C1 and C3 by stereoselective hydrogenation (Scheme 1). For this approach, we used *L*-dopa

Figure 1. Structures of saframycins, renieramycins, and ecteinascidin 743.

Ecteinascidin 743

as the starting material and prepared an oxalate amide 3. The oxalate amide moiety would provide a cyclic imine ester 2 *via* the Bischler-Napieralski reaction, which could be reduced stereoselectively to afford the required *cis* relationship, and then azide-ester cyclization would afford lactam 1 containing a [3.3.1] ring system.

Preparation of the substrate 5 was accomplished by the known procedures starting with L-dopa¹¹ and the product was converted to acetate 6 in quantitative yield (Scheme 2). Deprotection of the Boc group of 6 was carried out using AlCl₃, and the corresponding amine product was treated with ethyl oxalyl chloride to yield oxalate amide 7 in 81% yield. The Bischler-Napieralski reaction of 7 with P₂O₅ in CHCl₃ under refluxing condition afforded the cyclic imine ester 8 in 79% yield. However, use of POCl₃, the common reagent, under various solvents resulted in decomposition only. Reduction of 8 under hydrogen atmosphere with Pd/C provided a ca. 1:4 mixture of tetrahydroisoquinoline isomers 9a and 9b in 18% and 74% yields. The major product 9b was converted to 10 by protection with methyl chloroformate (81%). Then hydrolysis of 10 to alcohol (60%), reaction with diphenylphosphoryl azide (DPPA) to give the corresponding azide (65%). 12 and Staudinger reaction with this azide provided the desired lactam 11 in 51% yield. The other azide isomer obtained by the identical method from 9a did not afford any lactam moiety under the same reaction conditions.

To find a more practical route toward the lactam 11, we tried a sequential reductive process to obtain 11 from 13 (Scheme 3). Compound 5 was transformed to 12, using DPPA to furnish the azide functional group. This was followed by a three-step sequence from 12, deprotection of the Boc group

Scheme 1

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Scheme 2. Reagents and conditions (a) Ac₂O, Et₃N, DMAP/CH₂Cl₂, 99%; (b) AlCl₃, CH₂Cl₂; (c) ClCOCO₂Et, NaHCO₃, 81% for 2 steps; (d) P₂O₅, CHCl₃, reflux, 79%; (e) H₂, Pd/C, 74%; (f) ClCO₂Me, pyridine/CH₂Cl₂, 81%; (g) K₂CO₃, MeOH, 60%; (h) DPPA, DEAD, PPh₃, 65%; (i) PPh₃, THF-H₂O, 51%.

Scheme 3. Reagents and conditions (a) DPPA, DEAD, PPh₃, 70%; (b) AlCl₃, CH₂Cl₂; (c) ClCOCO₂Et, NaHCO₃, 61% for 2 steps; (d) P₂O₅, CHCl₃, reflux, 79%; (e) H₂, MeOH, Pd/C, 54%; (f) ClCO₂Me, pyridine/CH₂Cl₂, quantitative yield.

with AlCl₃, oxalate formation with ethyl oxalyl chloride, and the Bischler-Napieralski reaction with P₂O₅ to provide cyclic imine ester 13 in 39% total yield.

Under hydrogen atmosphere with Pd/C (Scheme 3), a lactam product was obtained as a single isomer in 54% yield and the protection of the lactam-amine product with chloromethylformate yielded the lactam 11 in quantitative yield, confirming the desired reductive sequential cyclization from compound 13. It seemed that the single isomer formation would be attributed to the formation of lactam followed by hydrogenation of double bond.

We have developed a short path to the 1,3-cis-substituted core skeleton of tetrahydroisoqunolines, using the Bischler-Napieralski reaction followed by reductive cyclization. We are applying this short approach to synthesize natural tetrahydroisoqunoline compounds.

Experimental Section

(S)-2-(tert-Butoxycarbonylamino)-3-(3,4-dimethoxyphenyl) propyl acetate (6). To a solution of (S)-2-(tert-butoxycarbonyl-

amino)-3-(3,4-dimethoxyphenyl)propyl alcohol 5 (311 mg, 1 mmol) in anhydrous CH₂Cl₂ (10 mL) were added Ac₂O (0.95 mL, 10 mmol), Et₃N (1.4 mL, 20 mmol) and DMAP (10 mg). After stirred for 2 h at rt. the reaction solution was dissolved in water and extracted with CH_2Cl_2 (20 mL × 3), dried over anhydrous MgSO₄ and concentrated in vacuo. The resulting residue was purified by silica gel column chromatography (n-hexane/EtOAc 2:1) to give the product 6 (350 mg. 99%) as a white solid. H-NMR (400 MHz, CDCl₃) δ 6.81 (1H. d, J= 8.8 Hz), 6.72-6.70 (2H, m), 4.78 (1H, b, NH), 4.07 (1H, m), 4.05 (2H. m). 3.87 (3H, s, OMe). 3.85 (3H, s, OMe). 2.81 (1H, m), 2.73 (1H, dd, J = 7.2, 14 Hz), 2.09 (3H, s), 1.42 (9H, s). ¹³C-NMR (100 MHz, CDCl₃) δ 170.7, 155.1, 148.8, 147.7, 129.6, 121.2, 112.2, 111.2, 79.4, 65.0, 55.8, 55.5, 50.5, 37.3, 28.2, 20.7. EI-HRMS calcd for C₁₈H₂₇NO₆ 353.1838, found: 353.1831.

(S)-2-(tert-Ethyloxalylamino)-3-(3,4-dimethoxyphenyl) propyl acetate (7). To a solution of 6 (2.83 g. 8.02 mmol) in anhydrous CH₂Cl₂ (40 mL) was added AlCl₃ (2.14 g, 16.04 mmol), followed by the addition of COCICOOEt (1.78 mL, 16.04 mmol) after 1 h. The reaction mixture was stirred for 30 min, and solid NaHCO₃ (2.02 g, 24.06 mmol) was added. After 1 h, the reaction mixture was then treated with aqueous saturated NaHCO3 solution slowly and water and extracted with CH_2Cl_2 (70 mL × 3). The organic phases were washed with brine, dried over anhydrous MgSO₄ and concentrated in vacuo. The residue was purified by silica gel column chromatography (n-hexane/EtOAc 1 : 1) to afford 7 (2.29 g. 81%) as a white solid. H-NMR (400 MHz, CDCl₃) δ 7.31 (1H, d, J = 8.8 Hz, NH), 6.80 (1H, d, J = 8.8 Hz), 6.72 (1H, m), 4.43 (1H, m), 4.34(2H, q, J = 7.2 Hz), 4.12 (2H, m), 3.87 (3H, s, OMe), 3.86 (3H, s)s. OMe). 2.85 (2H, m). 2.11 (3H, s). 1.39 (3H, t, J = 7.2 Hz). ¹³C-NMR (100 MHz, CDCl₃) δ 170.7, 160.4, 156.1, 149.0, 147.9, 128.6, 121.1, 112.0, 111.2, 64.1, 63.2, 55.8, 55.7, 50.1, 36.6, 20.7, 13.8, EI-HRMS calcd for C₁₇H₂₃NO₇ 353.1475, found: 353.1478.

(S)-Ethyl-3-(acetoxymethyl)-6,7-dimethoxy-3,4-dihydroisoquinoline-1-carboxylate(8). A mixture of 7 (1.67 g. 4.73 mmol) and P_2O_5 (9.4 g, 33.12 mmol) in anhydrous CHCl₃ (50 mL) was refluxed at 80 °C for 12 h. Then, the reaction mixture was cooled by an ice-water bath, slowly treated with NaOH 2M solution until pH = 9 and extracted with CH_2Cl_2 (50 mL × 3). The combined organic layer was dried over anhydrous Na₂SO₄ and concentrated *in vacuo*. The resultant residue was purified by silica gel column chromatography (n-hexane/EtOAc 1:1) to yield the product 8 (1.25 g. 79%) as a reddish liquid. ¹H-NMR (400 MHz, CDCl₃) δ 7.23 (1H. s), 6.72 (1H. s), 4.53-4.33 (4H, m), 3.93 (3H, s, OMe), 3.89 (3H, s, OMe), 3.83 (1H, m), 2.75 (1H, dd, J = 5.6, 16 Hz), 2.63 (1H, dd, J =14, 28.4 Hz), 2.10 (3H, s), 1.43 (3H, t, J = 7.2 Hz). ¹³C-NMR (100 MHz, CDCl₃) δ 171.0, 165.0, 159.5, 151.9, 147.6, 130.7, 118.7, 110.4, 110.1, 66.9, 62.0, 56.4, 56.1, 56.0, 27.8, 21.0, 14.2. EI-HRMS calcd for $C_{17}H_{21}NO_6$ 335.1369, found: 335.1361.

Ethyl-3-(acetoxymethyl)-6,7-dimethoxy-1,2,3,4-tetrahydro-isoquinoline-1-carboxylate isomers (9). Compound 8 (652 mg, 1.95 mmol) in MeOH (15 mL) was hydrogenated with catalytic amount of Pd/C 10% (105 mg) for 2 h under H₂ gas atmosphere in a balloon. After filtration through a Celite layer.

the filtrate was evaporated *in vacuo* and the resulting residue was purified by silica gel column chromatography (n-hexane/EtOAc 1 : 1 to 1 : 2) to give the product 9a as a colerless liquid (131 mg. 18%) and the product 9b (485 mg, 74%) as a yellowish liquid. Compound 9a: 1 H-NMR (400 MHz, CDCl₃) δ 6.94 (1H. s), 6.58 (1H, s), 4.65 (1H, s), 4.32 (1H. m), 4.19 (2H. m), 4.00 (1H. m), 3.86 (3H. s, OMe), 3.85 (3H. s, OMe), 3.57 (1H. m), 2.60 (1H. m), 2.13 (3H. s), 1.29 (3H. t, J = 7.2 Hz). Compound 9b: 1 H-NMR (400 MHz, CDCl₃) δ 6.89 (1H, s), 6.59 (1H. s), 4.83 (1H. s), 4.35-4.16 (4H. m), 3.86 (3H. s, OMe), 3.84 (3H, s, OMe), 3.16 (1H. m), 2.65 (1H. m), 2.12 (3H. s), 1.34 (3H. t, J = 7.2 Hz). 1 3C-NMR (100 MHz, CDCl₃) δ 172.3, 171.0, 148.2, 147.5, 126.9, 123.8, 112.0, 108.4, 67.6, 61.5, 60.1, 55.83, 55.82, 51.1, 31.2, 20.9, 14.3. EI-HRMS calcd for C_{17} H₂₃NO₆ 337.1525, found: 337.1521.

Cyclic lactam (11). To a solution of 9b (72 mg, 0.21 mmol) in anhydrous CH₂Cl₂ (10 mL) were added anhydrous pyridine (35 μL, 0.43 mmol) and CICOOMe (33 μL, 0.43 mmol) successively. After 3 h, the solvent was removed under reduced pressure and the resultant residue was purified by silica gel column chromatography (n-hexane/EtOAc 1:1) to yield the protected amine (67 mg. 81%) as a colorless liquid. H-NMR (400 MHz, CDCl₃) δ 7.06 (1H. s), 6.23 (1H. s), 5.54 (0.55H. s), 5.45 (0.45H, s), 4.81 (0.45H, m), 4.60 (0.55H, m), 4.35-4.17 (3H, m), 4.05 (1H, m), 3.87 (3H, s, OMe), 3.86 (3H, s, OMe), 3.79 (1.7H, s), 3.76 (1.3H, s), 3.00 (1H, dd, J = 6, 16 Hz), 2.80 (1H, m), 2.04 (1.7H, s), 2.00 (1.3H, s), 1.30 (3H, m). ¹³C-NMR (100 MHz. CDCl₃) δ 170.8, 170.7, 170.4, 156.3. 155.8, 148.5, 147.7, 147.6, 123.7, 123.4, 120.8, 120.0, 111.5, 111.3, 109.1, 64.8, 64.2, 61.4, 61.3, 56.9, 56.7, 55.7, 55.6, 52.9, 52.8, 48.1, 47.7, 29.8, 29.6, 20.6, 13.9, EI-HRM\$ calcd for C₁₉H₂₅NO₈ 395.1580, found: 395.1585. Anhydrous K₂CO₃ (19 mg, 0.135 mmol) was added to the solution of the protected amine above (107 mg, 0.27 mmol) in anhydrous MeOH (10 mL). After stirred for 1 h at rt, the reaction mixture was treated with water and extracted with EtOAc (20 mL ×3). dried over anhydrous Na2SO4 and concentrated in vacuo. The resulting residue was purified by silica gel column chromatography (n-hexane/EtOAc 1 : 2) to yield the alcohol (55 mg. 60%) as a colorless liquid. ¹H-NMR (400 MHz, CDCl₃) 8 7.07 (0.65H, s), 7.01 (0.35H, s), 6.63 (0.35H, s), 6.61 (0.65H, s), 5.65 (0.65H, s). 5.60 (0.35H, s), 4.64 (1H, m), 3.95-3.73 (12H, m), 3.75-3.44 (2H, m), 3.02-2.97 (1H, m), 2.80-2.60 (1H. m). To a solution of the resultant alcohol (22 mg, 0.065) mmol) in anhydrous THF (2 mL) at 0 °C were added PPh₃ (38 mg, 0.143 mmol), DEAD (23 μL, 0.143 mmol). After 5 min, DPPA (31 µL, 0.143 mmol) was added and the mixture was stirred at rt for 2 h. The reaction mixture was treated with H₂O (0.4 mL), then evaporated in vacuo and purified by silica gel column chromatography (n-hexane/EtOAc 1 : 1) to afford azide 10 (15.4 mg, 65%) as a colorless liquid. H-NMR (400 MHz, CDCl₃) δ 6.98 (0.6H, s), 6.96 (0.4H, s), 6.66 (1H, s). 5.58 (0.6H, s), 5.49 (0.4H, s), 4.60 (0.4H, m), 4.43 (0.4H, m), 3.88 (3H, s), 3.86 (3H, s), 3.8 (3H, s), 3.79 (1.6H, s), 3.77 (1.4H, s), 3.70-3.65 (1H, m), 3.35-3.29 (1H, m), 3.05-2.95 (1H. m), 2.87-2.79 (1H. m). The mixture of azide 10 (10 mg. 0.027 mmol), PPh₃ (15 mg. 0.055 mmol) in THF/H₂O (8 mL/0.2 mL) was stirred overnight at rt. After evaporation in reduced pressure, the resultant residue was purified by silica gel column chromatography (EtOAc/MeOH 20 : 1) to yield lactam **11** (4.2 mg, 51%) as a white solid. ¹H-NMR (400 MHz, CDCl₃) δ 6.86 (1H, s), 6.60 (1H, s), 6.50 (0.3H, s, NH), 6.36 (0.7H, s, NH), 5.42 (0.3H, s), 5.31 (0.7H, s), 4.98 (0.7H, bm), 4.85 (0.3H, bm), 3.88 (3H, s, OMe), 3.84 (3H, s, OMe), 3.74 (3H, s), 3.38 (1H, dd, J = 7.2, 16.4 Hz), 3.20 (1H, dd, J = 1.2, 12 Hz), 2.70 (1H, d, J = 17.6 Hz). ¹³C-NMR (100 MHz, CDCl₃) δ 169.8, 154.7, 149.0, 147.6, 124.7, 124.4, 111.4, 109.6, 56.5, 56.0, 55.9, 53.1, 47.1, 42.5, 32.6, EI-HRMS calcd for C₁₅H₁₈ N₂O₅ 306.1216, found: 306.1217.

(S)-2-(tert-Ethyloxalylamino)-3-(3,4-dimethoxyphenyl)propyl azide (12). To a solution of 5 (623 mg. 2 mmol) in anhydrous THF (20 mL) at 0°C were added PPh₃ (577 mg. 4.4 mmol). DEAD (693 μ L. 4.4 mmol). DPPA (950 μ L. 4.4 mmol) consecutively. After stirred at rt for 2 h. the reaction mixture was treated with H₂O (1 mL), then evaporated *in vacuo* and purified by silica gel column chromatography (n-hexane/EtOAc 2 : 1) to afford the product 12 (470 mg. 70%) as a colorless liquid. ¹H-NMR (400 MHz, CDCl₃) δ 6.81 (1H. d, J= 8 Hz), 6.75-6.72 (2H. m), 4.71 (1H, b, NH), 3.94 (1H. m), 3.87 (3H. s. OMe), 3.86 (3H. s. OMe), 3.42-3.26 (2H. m), 2.86-2.67 (2H, m), 1.43 (9H. s). ¹³C-NMR (100 MHz, CDCl₃) δ 155.0, 148.9, 147.7, 129.5, 121.2, 112.1, 111.3, 79.6, 55.8, 55.7, 53.0, 51.2, 37.5, 28.2, EI-HRMS calcd for C₁₆H₂₄N₄O₄ 336.1798, found: 336.1800.

(S)-Ethyl-3-(azidomethyl)-6,7-dimethoxy-3,4-dihydroisoquinoline-1-carboxylate(13). To a solution of 12 (350 mg, 1.04 mmol) in anhydrous CH₂Cl₂ (10 mL) was added AlCl₃ (278 mg. 2.08 mmol). After 1 h. COCICOOEt (173 μL, 1.56 mmol) was added. The reaction mixture was stirred at rt for 30 min and solid NaHCO₃ (263 mg, 3.12 mmol) was slowly added. Being stirred at rt for 1 h, the reaction mixture was then treated with saturated NaHCO3 solution slowly, water and extracted with CH₂Cl₂ (20 mL ×3). The organic phases were washed with brine, dried over anhydrous MgSO4 and concentrated in vacuo. The residue was purified by silica gel column chromatography (n-hexane/EtOAc 1 : 1) to afford the oxalate amide product (213 mg, 61%) as a colorless liquid. H-NMR (400 MHz, CDCl₃) δ 6.83-6.70 (3H, m), 4.38-4.29 (3H, m), 3.88 (3H, s. OMe), 3.87 (3H, s. OMe), 3.52-3.38 (2H, m), 2.92-2.79 (2H, m), 1.40 (3H. t, J = 7.2 Hz). ¹³C-NMR (100 MHz, CDCl₃) δ 160.3, 156.1, 149.1, 148.0, 128.5, 121.2, 112.0, 111.3. 63.4, 55.8. 52.3. 50.5, 37.0, 13.9. EI-HRMS calcd for $C_{15}H_{20}N_4O_5$ 336.1434, found: 336.1429. A mixture of the oxalate amide (127 mg, 0.378 mmol) and P₂O₅ (751 mg, 2.65 mmo) in anhydrous CHCl₃ (10 mL) was refluxed at 80 °C for 12 h. Then, the reaction mixture was cooled by ice-water bath. slowly treated with NaOH 2 M solution until pH = 9 and extracted with CH₂Cl₂ (20 mL ×3). The combined organic layer was dried over anhydrous Na2SO4 and concentrated in vacuo. The resultant residue was purified by silica gel column chromatography (n-hexane/EtOAc 1 : 1) to yield the product 13 (77 mg, 64%) as a liquid. 'H-NMR (400 MHz, CDCl₃) δ 7.28 (1H, s), 6.73 (1H, s), 4.45 (2H, m), 3.93 (3H, s, OMe), 3.89 (3H, s, OMe), 3.75 (2H. m). 3.61 (1H. m). 2.79 (1H, dd, J = 5.6, 15.6 Hz). 2.70 (1H, m), 1.42 (3H, t. J = 7.2 Hz). ¹³C-NMR (100 MHz, CDCl₃) δ 64.7, 159.2, 151.9, 147.6,

130.5, 118.6, 110.4, 110.1, 62.0, 57.3, 56.0, 55.9, 54.9, 28.2, 14.1. EI-HRMS calcd for $C_{15}H_{18}N_4O_4$ 318.1328, found: 318.1329.

Cyclic lactam (11). Compound 13 (28 mg. 0.088 mmol) in MeOH (7 mL) was hydrogenated at rt for 6 h using Pd/C 10% (14 mg). After filtration through a Celite layer and evaporation in vacuum, the resultant residue was purified by silica gel column chromatography (EtOAc/MeOH 10:1) to give the lactam product (11.8 mg, 54%) as a colorless oil. ¹H-NMR (400 MHz, CDCl₃) δ 6.83 (1H, s), 6.60 (1H, s), 6.39 (1H, s, NH), 4.34 (1H, s), 3.86 (3H, s. OMe), 3.82 (3H, s. OMe), 3.80 (2H, m), 3.22 (2H, m), 2.77 (1H, bs, NH), 2.65 (1H, d, J = 17.2)Hz). ¹³C-NMR (100 MHz, CDCl₃) δ 172.1, 148.7, 147.4, 126.9, 124.6, 111.6, 109.8, 57.2, 56.0, 55.9, 48.3, 43.2, 33.4. EI-HRMS calcd for $C_{13}H_{16}N_2O_3$ 248.1161, found: 248.1161. To a solution of the above lactam (10 mg, 0.04 mmol) in anhydrous CH₂Cl₂ (5 mL) were added anhydrous pyridine (4 μL, 0.048 mmol) and CICOOMe (7 μL, 0.08 mmol) successively. After stirred overnight at rt, the reaction mixture was evaporated under reduced pressure and purified by silica gel column chromatography (EtOAc/MeOH 10:1) to yield the product 11 (11 mg. 99%) as a white solid. ¹H-NMR (400 MHz, CDCl₃) δ 6.86 (1H, s), 6.60 (1H, s), 6.50 (0.3H, s, NH), 6.36 (0.7H, s, NH), 5.42 (0.3H, s), 5.31 (0.7H, s), 4.98 (0.7H, bm), 4.85 (0.3H, bm), 3.88 (3H, s, OMe), 3.84 (3H, s, OMe), 3.74 (3H, s), 3.38 (1H, dd, J = 7.2, 16.4 Hz), 3.20 (1H, dd, J =1.2, 12 Hz), 2.70 (1H, d, J = 17.6 Hz). ¹³C-NMR (100 MHz, CDCl₃) & 169.8, 154.7, 149.0, 147.6, 124.7, 124.4, 111.4, 109.6, 56.5, 56.0, 55.9, 53.1, 47.1, 42.5, 32.6. EI-HRM\$ calcd for C₁₅H₁₈N₂O₅ 306.1216, found: 306.1217.

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