# Effects of cranberry powder on serum lipid profiles and biomarkers of oxidative stress in rats fed an atherogenic diet

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#### **Abstract**

This study investigated that the antioxidative effect of freeze-dried cranberry powder against protein and lipid oxidation and ameliorative effect of serum lipid profile in rat fed atherogenic diet. Six weeks old male Sprague-Dawley rats were divided into the following four groups: normal diet group with 5% corn oil (control), atherogenic diet group with 5% corn oil, 10% lard, 1% cholesterol, and 0.5% sodium cholate (HFC), atherogenic plus 2% cranberry powder diet group (HFC+C2), and atherogenic plus 5% cranberry powder diet group (HFC+C5), and respective diet and water were fed daily for 6 weeks. After the experimental period, the serum lipid profile, such as total cholesterol, HDL-cholesterol, LDL-cholesterol, and triglyceride, ferric reducing ability of plasma (FRAP), plasma phenolics content, superoxide dismutase (SOD) activity, serum protein carbonyl and thiobarbituric acid reactive substances (TBARS) levels were examined. Total phenolic compound and total flavonoid levels in freeze-dried cranberry powder were 9.94 mg/g and 8.12 mg/g, respectively. Serum total cholesterol and LDL-cholesterol levels were not significantly different for cranberry powder treatment, but serum HDL-cholesterol level was significantly increased in HFC+C5 group compared with HFC group. Plasma FRAP value tended to be increased by cranberry powder treatment though there was no significant difference. Plasma total phenol concentrations and SOD activities were not significantly different among all groups. Serum protein carbonyl and TBARS levels were significantly decreased in HFC+C5 group compared with HFC group. Overall results suggested that freeze-dried cranberry powder might have the serum lipid improving effect, as well as antioxidative effect demonstrated by its protective effect against protein and lipid oxidation.

Key Words: Antherogenic diet, protein carbonyl, TBARS, FRAP, cranberry powder

## Introduction

In recent years, there has been increasing interest in health and disease prevention in relation to oxidative stress. Enhanced oxidative stress was shown to cause general damage on cells by promoting the oxidation of protein and DNA and lipid peroxidation (Cadenas & Davies, 2000). Several studies have found that biomarkers of oxidative stress were improved by various plant food materials such as tea (Sriram *et al.*, 2008), cocoa (Ramiro-Puig *et al.*, 2007), red wine (Das *et al.*, 2007), and grape juice (Dani *et al.*, 2008), which are known to be high in phenolic compounds. In addition, polyphenolic constituents of wines and grapes juices have been reported to act as free radical scavengers and antimicrobial agents (Sanchez-Moreno *et al.*, 1999; Shahidi & Wanasundara, 1992).

Cranberry has been shown to prevent urinary tract infections, inhibit peptic ulcer-associated bacterium, *Helicobacter pylori*, and induce apoptosis of carcinoma cell (Gotteland *et al.*, 2008; Kontiokari *et al.*, 2001; Krestry *et al.*, 2008). In addition, cranberry was known to have antioxidant properties as shown

in plasma antioxidant status, protection of RBC against hemolysis and a cellular antioxidant activity (Villarreal *et al.*, 2007; Wolfe & Liu, 2007).

Cranberry is rich in phenolic phytochemicals such as phenolic acids, flavonoids and ellagic acid. These phytochemicals act as antioxidants, and showed health benefits including reduction of oxidative damage that can cause cancer, heart disease, and other degenerative diseases (Çelik *et al.*, 2008; Vattem *et al.*, 2005). Chu and Liu (2005) reported that phytochemicals in cranberries could inhibit LDL oxidation and induce expression of LDL receptors. Mckay and Blumberg (2007) also reported that cranberry reduced the risk of cardiovascular disease (CVD) by inhibition of LDL oxidation and platelet aggregation, and reducing blood pressure via other anti-thrombotic and anti-inflammatory mechanisms.

Typical forms of cranberry found in markets are dried and ground whole fruits or dried juice. Recently, freeze-dried cranberry powder and 100% cranberry juice are available and introduced their health benefit on prevention of urinary tract infections (UTIs). Piljac-Žegarac *et al.* (2008) reported that total

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phenolics level in cranberry juice was 1546.9 mg/L and Chen et al. (2001) reported that cranberry juice had 400 mg/L of total phenolics. Vinson et al. (2008) reported that the amount of total phenols in cranberry foods was higher in dried or frozen products than in 100% cranberry juices.

However, most studies used cranberry extract or juice for testing its *in vivo* antioxidant function. In addition, it is limited that investigators studied freeze-dried cranberry powder and its antioxidant and cardiovascular protective effects. Therefore, we employed freeze-dried cranberry powder and investigated its possible health benefits using animal model. In the present paper, we report the antioxidative effect against protein and lipid oxidation and serum lipid profile ameliorative effect of freeze-dried cranberry powder in rats fed atherogenic diet.

#### Materials and Methods

#### Animals and diet

Six-week-old, 30 male Sprague-Dawley rats, weighing 160-190 g, were obtained from Animals Co. (Daehan, Korea) and maintained under a specific condition with a temperature-controlled (23 ± 2°C) and humidity-controlled (50 ± 10%). After a 7-day acclimation period, animals were divided into the following four groups on the basis of comparable mean body weight: 5% corn oil normal diet group (Control, n=7), atherogenic diet group containing 5% corn oil, 10% lard, 1% cholesterol, and 0.5% sodium cholate (HFC, n=7), atherogenic plus 2% cranberry powder diet group (HFC + C2, n=8), and atherogenic plus 5% cranberry powder diet (HFC + C5, n=8). Freeze-dried cranberry powder was purchased from Mastertaste Co. (USA) and all diets were the modified AIN-76 (American Institute of Nutrition, 1977) (Table 1). The experimental

Table 1. Composition of control and experimental diet (%)

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Grou	Control	HFC	HFC + C2	HFC + C5
Casein	20	20	20	20
DL-methionine	0.3	0.3	0.3	0.3
Corn starch	29.0	17.5	15.5	12.5
Sucrose	40	40	40	40
Cellulose	1.0	1.0	1.0	1.0
Corn oil	5	5	5	5
Lard	•	10	10	10
AIN-mineral Mix <sup>1)</sup>	3.5	3.5	3.5	3.5
AIN-vitamin Mix <sup>2)</sup>	1.0	1.0	1.0	1.0
Choline chloride	0.2	0.2	0.2	0.2
Cranberry powder	-	-	2.0	5.0
Cholesterol	-	1.0	1.0	1.0
Sodium cholate	-	0.5	0.5	0.5

Control: normal diet control group,

HFC: atherogenic diet control group,

HFC + C2: atherogenic plus 2% cranberry power diet group,

HFC + C5: atherogenic plus 5% cranberry power diet group.

diets and water were fed ad libitum for 6 weeks from the beginning of the experiment. Food intake was recorded daily and the body weight was measured weekly. The end of the experimental period, the animals anesthetized with ethyl ether after 12 hours fasting and blood from the saphenons vein and the liver samples were collected. Blood samples were collected in vacuum tube (Becton-Dickinson, Meylan, France) without anticoagulant for lipid profiles and biochemical analysis, and with heparin for erythrocyte lysate preparation. All blood samples were immediately centrifuged (3,000 rpm, 20 min, 4°C) for the separation of serum, plasma, and erythrocyte. Erythrocyte samples were lysed in 4 times with ice cold HPLC-grade water and centrifuged (10,000 g, 15 min, 4°C), and the supernatant (erythrocyte lysate) was collected for erythrocyte superoxide dismutase (SOD) activity analysis. All samples were stored at -80°C and thawed only once, just before analysis.

#### Lipid profiles

Serum total cholesterol, HDL-cholesterol, and triglyceride were detected by enzymatic assay using automatic biochemical analyzing system (Vitalab Selectra E, Vital Scientific N.V., the Netherlands). LDL-cholesterol level was calculated by a formula of Friendewald *et al.* (1972).

## Ferric reducing ability of plasma (FRAP)

Plasma antioxidant status was evaluated using ferric reducing ability of plasma (FRAP) assay (Benzie & Strain, 1996). At low pH, the ferric-tripyridyltriazine (Fe(III)-TPTZ) complex is reduced to ferrous tripyridyltriazine (Fe(II)-TPTZ), which is color marker, depends on the ability of sample. Two hundred seventy µl of pre-warmed (at 37°C) freshly prepared FRAP reagent (25 ml of 300 mM sodium acetate buffer pH 3.6, 2.5 ml of 10 mM 2,4,6-tri(2-pyridil)-s-triazine (TPTZ) in 40 mM HCl, 2.5 ml of 20 mM ferric chloride solution) were mixed with 30 µl of plasma diluents. After 15 min incubation at 37°C, absorbance was read at 550 nm. FRAP values are measured by detection of the absorbance change of test sample as the reducing power of antioxidants present in the plasma comparing with that of trolox standard.

# Total phenolics and total flavonoid assay

The total soluble phenolic compounds were determined in cranberry powder and plasma with the Folin-Ciocalteu reagent according to the method of Singleton *et al.* (1965) using gallic acid as standard. Mixture of 50 µl sample (plasma or 100 mg freeze-dried cranberry powder in 1 ml of distilled water) and 865 µl distilled water was incubated with 75 µl Folin-Ciocalteu reagent at room temperature for 5 min. Two hundred twenty five µl of 20% Na<sub>2</sub>CO<sub>3</sub> and 285 µl of distilled water were added to the reaction mixture which was incubated at room temperature for 2 hours, and the absorbance was read at 760 nm. The total

<sup>1)</sup> Composition of AIN-76 salt mixture

<sup>2)</sup> Composition of AIN-76 vitamin mixture

phenolic content is expressed as gallic acid equivalent (GAE) in mM of plasma and milligrams per gram of dried sample.

The total flavonoid level in freeze-dried cranberry powder was measured by spectropotometric method using catechin as standard (Jia et al., 1999). Five hundred µl of diluted sample with distilled water and 75 µl of 5% NaNO<sub>2</sub> solution were mixed and incubated at room temperature for 5 min. This mixture was added 150 µl of 10% AlCl<sub>3</sub> and allowed to stand for a further 5 min before 0.5 ml of 1 M NaOH was added. The solution was measured the absorbance at 510 nm. The results were expressed as catechin equivalent in milligrams per gram of freeze-dried cranberry powder.

Plasam and erythrocyte lysate superoxide dismutase (SOD) activity

Superoxide dismutase (SOD) activities of plasma and erythrocyte lysate were measured by microplate assay method based on the detection of superoxide radicals generated by xanthine oxidase and hypoxanthine using the commercial kit (Cayman chemical superoxide dismutase assay kit, Cayman Chemical Co., USA).

Serum protein carbonyl and thiobarbituric acid reactive substances levels

Serum protein carbonyl levels were determined by enzyme linked immunosorbent assay (ELISA) based on the detection of protein hydrazones form from the reaction of dinitrophenylhydrazine (DNP) with protein carbonyl using the Biocell PC test kit (BioCell Corp., Ltd., New Zealand).

Serum thiobarbituric acid reactive substances (TBARS), which is the important marker of lipid peroxidation, was determined by spectrophotometric method using the OXItek TBARS kit (ZeptoMetrix Co., USA).

## Statistical analysis

Statistical programs available in SAS program (version 9.1, SAS Inc, Cary, NC, USA) were utilized for data analysis. The significance of difference among the groups was assessed using one-way analysis of variance and Duncan's multiple range tests. Values are presented as the mean ± standard deviation, and a significance test on all results was conducted at level of p<0.05 unless otherwise stated.

#### Results

Total phenolics and total flavonoids in cranberry powder

Total phenolic compound and total flavonoid levels in freeze-dried cranberry powder were 9.94 mg/g and 8.12 mg/g respectively (Table 2).

Table 2. Total phenolic compound and total flavonoid levels in cranberry powder

	Content
Total phenolic compound (mg gallic acid equivalents/g)	9.94 ± 0.13
Total flavonoid (mg catechin equivalents/g)	8.12 ± 0.25

Values are mean ± SD.

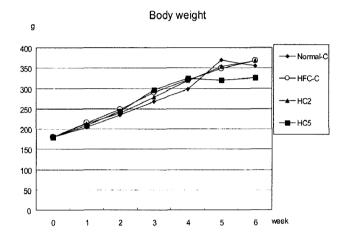


Fig. 1. Change of body weights in rat. Values are mean,

#### Weights and food intake

Weight gains, mean of food intake, and food efficiency ratio (FER) are shown in Table 3 and body weight change are shown in Fig. 1. Body weights were gradually increased in all groups, and the weight gains and mean of food intake were decreased in HFC+C5 group (146.2 g, 13.5 g/day) compared with control (182.5 g, 15.5 g/day) or HFC groups (194.5 g, 15.4 g/day), but FER was not significant difference between HFC+C5 (0.25 g/100 g body weight) and control (0.26 g/100 g body weight) or HFC group (0.31 g/100 g body weight).

The final body weight also was decreased in HFC + C5 group compared with control or HFC groups (Table 3). The liver weight and relative liver weight were significantly increased in all atherogenic diet groups than in normal diet group, but the liver weight was decreased in HFC + C5 group (17.8 g) compared with the HFC group (23.0 g) (Table 3).

# Lipid profiles

The results about the effect of cranberry powder on serum lipid profiles are shown in Table 4. Average of serum triglyceride level in control group was significantly higher than that in all atherogenic diet groups. Serum total cholesterol and LDL-cholesterol levels were significantly increased by atherogenic diets, but there was no significant difference for cranberry powder treatment. Serum HDL-cholesterol level and HDL/LDL ratio were significantly decreased in all atherogenic diet groups compared with normal diet group. However serum HDL-

Table 3. Effects of cranberry powder on food intake, food intake efficiency ratio (FER), weight gains, final body weight, liver weight, and relative liver weight in diet-induced hyperlipidemia rat

Group	) (n)	Food intake (g/day)	FER <sup>2)</sup> (g/100 g body weight)	Weight gains <sup>3)</sup> (g)	Final body weight (g)	Liver weight (g)	Relative liver weight (g/100 g body weight)
Control	(7)	15.5 ± 0.6°	0.26 ± 0.03 <sup>ab</sup>	182.5 ± 16.3 <sup>a</sup>	362.5 ± 19.6 <sup>a</sup>	12.4 ± 1.6°	$3.42 \pm 0.36^{b}$
HFC	(7)	$15.4 \pm 0.9^{a}$	$0.31 \pm 0.04^{ab}$	194.5 ± 34.4°	374.5 ± 34.5 <sup>a</sup>	$23.0 \pm 3.6^{a}$	$6.13 \pm 0.42^{a}$
HFC + C2	(8)	14.7 ± 0.5 <sup>b</sup>	$0.31 \pm 0.06^{a}$	192.9 ± 38.7°	$372.9 \pm 40.0^{a}$	21.3 ± 3.9°	5.66 ± 0.53°
HFC + C5	(8)	$13.5 \pm 0.5^{\circ}$	$0.25 \pm 0.06^{b}$	146.2 ± 28.5 <sup>b</sup>	$326.3 \pm 24.8^{b}$	17.8 ± 2.5 <sup>b</sup>	5.48 ± 0.86°

n: number of animals,

Control: normal diet control group,

HFC: atherogenic diet control group,

HFC + C2: atherogenic plus 2% cranberry power diet group,

HFC + C5: atherogenic plus 5% cranberry power diet group.

1) Values are mean ± SD,

Means with the different superscripts are significantly different at p<0.05 by Duncan's multiple range test.

2) FER: food intake efficiency ratio

<sup>3)</sup> Total body weight gains for 6 weeks

Table 4. Effects of cranberry powder on serum lipid profiles in diet-induced hyperlipidemia rat1)

Group	p (n)	Triglyceride (mg/dl)	Total cholesterol (mg/dl)	HDL-cholesterol (mg/dl)	LDL-cholesterol (mg/dl)	HDL/LDL ratio
Control	(7)	112.5 ± 49.2 <sup>a</sup>	200.5 ± 20.8 <sup>b</sup>	62.9 ± 3.3°	113.3 ± 14.1 <sup>b</sup>	0.535 ± 0.040°
HFC	(7)	51.9 ± 15.6 <sup>b</sup>	456.1 ± 82.5°	38.7 ± 8.3°	407.0 ± 81.8°	0.098 ± 0.023 <sup>b</sup>
HFC + C2	(8)	71.3 ± 35.2 <sup>b</sup>	468.8 ± 170.3 <sup>a</sup>	36.6 ± 6.1°	417.9 ± 171.2°	$0.098 \pm 0.036^{b}$
HFC + C5	(7)	$70.0 \pm 29.2^{b}$	504.1 ± 99.1 <sup>a</sup>	49.7 ± 12.6 <sup>b</sup>	436.3 ± 95.5°	$0.106 \pm 0.046^{b}$

n: number of animals.

Control: normal diet control group,

HFC: atherogenic diet control group,

HFC + C2: atherogenic plus 2% cranberry power diet group,

HFC + C5: atherogenic plus 5% cranberry power diet group.

 $^{1)}$  Values are mean excluding one of the data over 2SD  $\pm$  SD,

Means with the different superscripts are significantly different at p<0.05 by Duncan's multiple range test.

Table 5. Effects of cranberry powder on plasma FRAP and total phenolic levels in dief-induced hyperlipidemia rat<sup>1)</sup>

Group	(n)	FRAP <sup>2)</sup> (μΜ)	Total phenolic level (mM gallic acid equivalents)
Control	(7)	172.9 ± 54.2 <sup>NS</sup>	6.08 ± 0.20 <sup>NS</sup>
HFC	(6)	180.6 ± 59.0	6.27 ± 0.17
HFC + C2	(8)	$228.8 \pm 82.5$	$6.16 \pm 0.39$
HFC + C5	(8)	217.4 ± 64.2	$6.06 \pm 0.30$

n: number of animals,

Control: normal diet control group,

HFC: atherogenic diet control group,

HFC + C2: atherogenic plus 2% cranberry power diet group,

HFC + C5: atherogenic plus 5% cranberry power diet group.

1) Values are mean ± SD,

Means with the different superscripts are significantly different at p<0.05 by Duncan's multiple range test

<sup>2)</sup> FRAP: the ferric reducing ability of plasma

cholesterol was significantly increased in HFC + C5 group (49.7 mg/dl) compared with HFC group (38.7 mg/dl) (Table 4).

Plasma FRAP, total phenolics levels and SOD activity

Plasma FRAP and total phenolics levels were detected to examine the effects of cranberry powder on antioxidant status. Average of plasma FRAP value shown tended to be increased by cranberry powder treatment, but there was no significant difference (control; 172.9  $\mu$ M, HFC; 180.6  $\mu$ M, HFC + C2; 228.8  $\mu$ M, HFC + C5; 217.4  $\mu$ M). Plasma total phenolic content was

Table 6. Effects of cranberry powder on plasma and erythrocyte lysate SOD activity in diet-induced hyperlipidemia rat<sup>1)</sup>

Group (n)		SOD <sup>2)</sup> activity in plasma (U/ml)	SOD activity in erythrocyte (U/ml)	
Control	(7)	23.1 ± 9.8 <sup>NS</sup>	653.6 ± 338.1 <sup>NS</sup>	
HFC	(7)	27.0 ± 12.3	657.8 ± 218.1	
HFC + C2	(8)	$29.0 \pm 10.9$	578.7 ± 273.4	
HFC + C5	(8)	26.2 ± 8.1	422.7 ± 205.4	

n: number of animals,

Control: normal diet control group,

HFC: atherogenic diet control group,

HFC + C2: atherogenic plus 2% cranberry power diet group,

HFC + C5: atherogenic plus 5% cranberry power diet group.

1) Values are mean ± SD,

Means with the different superscripts are significantly different at p<0.05 by Duncan's multiple range test.

2) SOD: superoxide dismutase

not significantly different for atherogenic diet or cranberry powder treatment (Table 5).

Plasma SOD activity was also not significantly different by atherogenic diet or cranberry powder treatment, but erythrocyte SOD activity showed a tendency to be decreased by cranberry powder treatment although there was not significant (Table 6).

## Serum protein carbonyls and TBARS levels

Serum protein carbonyl and TBARS levels were measured to investigate the antioxidant effects of cranberry powder against

**Table 7.** Effects of cranberry powder on serum protein carbonyl and TBARS levels in diet-induced hyperlipidemia rat<sup>1)</sup>

Group (n)		Protein carbonyl (nmol/mg)	TBARS <sup>2)</sup> (nmol/ml)	
Control	(7)	0.58 ± 0.11 <sup>a</sup>	14.7 ± 3.3 <sup>ab</sup>	
HFC	(7)	$0.51 \pm 0.05^{ab}$	15.9 ± 3.8 <sup>a</sup>	
HFC + C2	(8)	$0.47 \pm 0.04^{b}$	15.6 ± 6.0 <sup>a</sup>	
HFC + C5	(8)	$0.24 \pm 0.05^{\circ}$	$10.7 \pm 3.6^{b}$	

n: number of animals,

Control: normal diet control group,

HFC: atherogenic diet control group,

HFC + C2: atherogenic plus 2% cranberry power diet group,

HFC + C5: atherogenic plus 5% cranberry power diet group.

<sup>17</sup> Values are mean ± SD,

Means with the different superscripts are significantly different at p<0.05 by Duncan's multiple range test

<sup>2)</sup> TBARS: thiobarbituric acid reactive substances

oxidative damage such as protein oxidation and lipid peroxidation. Mean serum protein carbonyl level was found to be significantly decreased in HFC + C2 (0.47 mmol/mg) and HFC + C5 groups (0.24 nmol/mg) as compared to control group (0.58 nmol/mg) and significantly decreased in HFC + C5 group as compared to HFC group (0.51 nmol/mg) (Table 8). Serum TBARS level was also significantly decreased in HFC + C5 group (10.7 nmol/ml) compared with HFC group (15.9 nmol/ml) (Table 7).

#### Discussion

This study examined the serum lipid profile ameliorative and antioxidative effects of freeze-dried cranberry powder in rats fed atherogenic diet. Cranberry is known to be a good source of antioxidants, which have health benefits such as anti-adhension activity, antiviral, and anticancer properties (Bomser *et al.*, 1996; Howell *et al.*, 2005).

In this study, we used freeze-dried cranberry powder, and its total phenolics and total flavonoid levels were 9.94 mg/g and 8.12 mg/g, respectively. Sun et al. (2002) reported that cranberry had the highest total phenolic content, followed by apple, red grape, strawberry, pineapple, banana, peach, lemon, orange, pear, and grapefruit. Celik et al. (2008) reported that total phenolics contents of cranberry fruits were different among each maturation stage and dark red cranberry had 4.745 mg gallic acid equivalents/g of total phenolics. Vinson et al. (2005) also reported that the average total phenols were 663 mg/100 g for fresh cranberries and 870 mg/100 g for dried cranberries, which was a similar level with our study. According to the study of Vinson et al. (2008), the amount of total phenols in cranberry foods on fresh weight basis was higher in dried or frozen products than in juice or sauce forms. Piljac-Žegarac et al. (2008) reported that total phenolics level in cranberry juice was 1546.9 mg/L. Chen et al. (2001) reported that cranberry juice had 400 mg/L of total phenolics including flavonoids, and the major forms of phenolics and flavonoids were benzoic acid, quercetin and

myricetin. In a study of Bailey *et al.* (2007), concentrated cranberry extract with 30% phenolics was used to investigate its effects on the recurrent urinary tract infections (UTIs) in women, since the concentrated active phenolics might be more effective in preventing UTIs. However, the optimum level and form of administration was not suggested. In the current study, we used just freeze-dried powder because its total phenolics level was higher than fresh or juice type of cranberry and it was a natural food form without excess processes such as concentration and extraction.

Mckay and Blumberg (2007) suggested that polyphenols in cranberry might reduce the cardiovascular disease risk factor such as LDL oxidation, platelet aggregation, and high blood pressure. Ghazala et al. (2008) also reported that plasma LDL-cholesterol and LDL oxidation were reduced by cranberry extract fortification. However, Ruel et al. (2006) reported that total and LDL-cholesterol were not changed but HDL-cholesterol was increased by cranberry juice consumption in men. In the current study, we also observed similar results that serum total cholesterol and LDL-cholesterol levels were not significantly different in rats with and without cranberry powder treatment, but serum HDL-cholesterol level was significantly increased by 5% cranberry powder fortification. Deyhim et al. (2007) also reported that drinking cranberry juice increased plasma antioxidant capacity but did not change the cholesterol concentrations in liver and plasma of rat and it assumed that cranberry juice might increase antioxidant status without affecting cholesterol homeostasis.

In case of *in vitro* study, cranberry was known that it has abundant phytochemicals and the highest cellular antioxidant activity among tested food materials including apple, red grape and green grape (Wolfe & Liu, 2007). Villarreal *et al.* (2007) also reported that plasma antioxidant status was increased and red blood cell was protected against hemolysis by drinking cranberry juice in rats. In this study, plasma total antioxidant capacity measure by FRAP tended to be increased by cranberry powder treatment, although there was no significant difference. In several short-term intervention studies for cranberry juice, it was shown that plasma antioxidant capacity was increased by cranberry juice consumption (Ruel *et al.*, 2005; Zhang & Zue, 2004).

Pedersen *et al.* (2000) reported that total phenolics level in plasma was slightly increased four hours following 500 ml cranberry juice consumption for one time. However, according to the study of Ruel *et al.* (2005), dietary and possibly active compounds from cranberry juice were rapidly degraded in plasma after consumption. This metabolic degradation might explain in part the findings observed in a study by Duthie *et al.* (2006) and the current study. Duthie *et al.* (2006) showed that total phenols in fasting plasma were not different between cranberry juice consumption and placebo groups in their 2 week study. When rats were fed atherogenic diet fortified with cranberry powder, we also failed to observe the significant changes in total phenolics content of fasting plasma.

In this study, plasma and erythrocyte SOD activities were not significantly different by atherogenic diet or cranberry powder treatment. Similarly, Duthie et al. (2006) found that plasma SOD activities were not different between cranberry juice consumption and placebo groups. However, there was a conflict finding observed in a study by Devhim et al. (2007), which reported that cranberry juice increased plasma SOD activity. For biomarkers of protein and lipid oxidation, mean serum protein carbonyl and TBARS levels were significantly decreased in 5% cranberry powder treated group compared with atherogenic diet fed group in this study. Serum protein carbonyl formation has been proposed to be an early marker for protein oxidation (Reznick & Packer, 1994) and serum TBARS is regarded as a marker of lipid peroxidation. In a human intervention study with the dried cranberry juice, serum protein oxidation level was significantly decreased by a 1200 mg of dried cranberry juice consumption (Valentova et al., 2007). Deyhim et al. (2007) reported that cranberry juice increased plasma antioxidant capacity and reduced malondialdehyde concentrations. Vattem et al. (2005) also reported that cranberry phenolics decreased MDA formation in oxidatively stressed porcine muscle and it suggested that exogenously treated phenolic phytochemicals could be reducing the oxidative stress. Including the results from the current study, several trials have shown beneficial effect of cranberry consumption on protein and lipid oxidation. These consensus outcomes may explain the antioxidative effect of cranberry powder and juice against oxidative damage.

Overall results of the present study showed that 5% of the freeze-dried cranberry powder treatment increased HDL-cholesterol and reduced protein carbonyl and TBARS levels in rat fed atherogenic diet. It assumed that cranberry powder might have the plasma lipid improving effect and antioxidant effect against oxidative damage. However, since the current study was done in animal model, the findings from this study cannot be applied to the human diet directly. Therefore, further studies are needed to evaluate health benefits and effective dose of cranberry powder in humans. In addition, it is suggested that the mechanisms of antioxidant action and lipid improving effect of cranberry powder and its components should be elucidated for better understanding the health benefits of this plant food with high phenolics.

#### Literature cited

- American Institute of Nutrition (1977). Report of the American Institute of Nutrition Ad Hoc Committee on Standards for Nutritional Studies. *J Nutr* 107:1340-1348.
- Bailey DT, Dalton C, Daugherty FJ & Tempesta MS (2007). Can a concentrated cranberry extract prevent recurrent urinary tract infections in women? A pilot study. *Phytomedicine* 14:237-241.
- Benzie IF & Strain JJ (1996). The Ferric Reducing Ability of Plasma (FRAP) as a Measure of "Antioxidant Power": The FRAP Assay. Anal Biochem 239:70-76.

- Bomser J, Madhavi DL, Singletary K & Smith MA (1996). In vitro anticancer activity of fruit extracts from Vaccinium species. Planta Med 62:212-216.
- Cadenas E & Davies KJ (2000). Mitochondrial free radical generation, oxidative stress, and aging. Free Radic Biol Med 29:222-230.
- Çelik H, Özgen M, Serçe S & Kaya C (2008). Phytochemical accumulation and antioxidant capacity at four maturity stages of cranberry fruit. Scientia Horticulturae 117:345-348.
- Chen H, Zuo Y & Deng Y (2001). Separation and determination of flavonoids and other phenolic compounds in cranberry juice by high-performance liquid chromatography. *J Chromatogr A* 913: 387-395
- Chu YF & Liu RH (2005). Cranberries inhibit LDL oxidation and induce LDL receptor expression in hepatocytes. *Life Sci* 77:1892-1901
- Dani C, Pasquali MA, Oliveira MR, Umezu FM, Salvador M, Henriques JA & Moreira JC (2008). Protective effects of purple grape juice on carbon tetrachloride-induced oxidative stress in brains of adults Wistar rats. *J Med Food* 11:55-61.
- Das S, Santani DD & Dhalla NS (2007). Experimental evidence for the cardioprotective effects of red wine. Exp Clin Cardol 12:5-10.
- Deyhim F, Patil BS, Villarreal A, Lopez E, Garcia K, Rios R, Garcia C, Gonzales C & Mandadi K (2007). Cranberry juice increases antioxidant status without affecting cholesterol homeostasis in orchidectomized rats. *J Med Food* 10:49-53.
- Duthie SJ, Jenkinson AM, Crozier A, Mullen W, Pirie L, Kyle J, Yap LS, Christen P & Duthie GG (2006). The effects of cranberry juice consumption on antioxidant status and biomarkers relating to heart disease and cancer in healthy human volunteers. *Eur J Nutr* 45:113-122.
- Friedewald WT, Levy RI & Fredrickson DS (1972). Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clin Chem* 18:499-502.
- Ghazala S, Aldoori WH, Davis PJ & Alkanani T (2008). Fortification of atherogenic diet with cranberry extract and spinach: plasma LDL oxidation and other atherosclerosis risk factors in rats. *Atheroscler Suppl* 9:11.
- Gotteland M, Andrews M, Toledo M, Muñoz L, Caceres P, Anziani A, Wittig E, Speisky H & Salazar G (2008). Modulation of *Helicobacter* pylori colonization with cranberry juice and *Lactobacillus johnsonii* La1 in children. Nutrition 24:421-426.
- Howell AB, Reed JD, Krueger CG, Winterbottom R, Cunningham DG & Leahy M. (2005). A-type cranberry proanthocyanidins and uropathogenic bacterial anti-adhesion activity. *Phytochemistry* 66: 2281-2291.
- Jia Z, Tang M & Wu J (1999). The determination of flavonoid contents in mulberry and their scavenging effects on superoxides radicals. Food Chem 64:555-559.
- Kontiokari T, Sundqvist K, Nuutinen M, Pokka T, Koskela M & Uhari M (2001). Randomised trial of cranberry-lingonberry juice and *Lactobacillus GG* drink for the prevention of urinary tract infections in women. *BMJ* 322:1571-1575.
- Kresty LA, Howell AB & Baird M (2008). Cranberry proanthocyanidins induce apoptosis and inhibit acid-induced proliferation of human esophageal adenocarcinoma cells. J Agric Food Chem 56:676-680.
- Mckay DL & Blumberg JB (2007). Cranberries (Vaccinium macrocarpon) and cardiovascular disease risk factor. *Nutr Rev* 65:490-502.
- Pedersen CB, Kyle J, Jenkinson AM, Gardner PT, McPhail DB & Duthie GG (2000). Effects of blueberry and cranberry juice

- consumption on the plasma antioxidant capacity of healthy female volunteers. Eur J Clin Nutr 54:405-408.
- Piljac-Žegarac J, Valek L, Martinez S & Belščak A (2008). Fluctuations in the phenolic content and antioxidant capacity of dark fruit juices in refrigerated storage. *Food Chem* (Epub ahead of print).
- Ramiro-Puig E, Urpí-Sardà M, Pérez-Cano FJ, Franch A, Castellote C, Andrés-Lacueva C, Izquierdo-Pulido M & Castell M (2007). Cocoa-enriched diet enhances antioxidant enzyme activity and modulates lymphocyte composition in thymus from young rats. J Agric Food Chem 55:6431-6438.
- Reznick AZ & Packer L (1994). Oxidative damage to proteins: spectrophotometric method for carbonyl assay. *Methods Enzymol* 233:357-363.
- Ruel G, Pomerleau S, Couture P, Lamarche B & Couillard C (2005). Change in plasma antioxidant capacity and oxidized low-density lipoprotein levels in men after short-term cranberry juice consumption. *Metab clin exp* 54:856-861.
- Ruel G, Pomerleau S, Couture P, Lemieux S, Lamarche B & Couillard C (2006). Favourable impact of low-calorie cranberry juice consumption on plasma HDL-cholesterol concentrations in men. Br J Nutr 96:357-364.
- Sanchez-Moreno C, Larrauri JA & Saura-Calixto F (1999). Free radical scavenging capacity and inhibition of lipid oxidation of wines, grapes juices and related polyphenolic constituents. Food Res Intern 32:407-412.
- Shahidi F & Wanasundara PKJPD (1992). Phenolic antioxidants. Crit Rev Food Sci Nutr 32:67-103.
- Singleton VL & Rossi JA (1965). Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *Am J Enol Vitic* 16:144-158.
- Sriram N, Kalayarasan S & Sudhandiran G (2008). Enhancement of antioxidant defense system by epigallocatechin-3-gallate during

- bleomycin induced experimental pulmonary fibrosis. *Biol Pharm Bull* 31:1306-1311.
- Sun J, Chu YF, Wu X & Liu RH (2002). Antioxidant and antiproliferative activities of common fruits. J Agric Food Chem 50:7449-7454.
- Valentova K, Stejskal D, Bednar P, Vostalova J, Cíhalík C, Vecerova R, Koukalova D, Kolar M, Reichenbach R, Sknouril L, Ulrichova J & Simanek V (2007). Biosafety, antioxidant status, and metabolites in urine after consumption of dried cranberry juice in healthy women: a pilot double-blind placebo-controlled trial. J Agric Food Chem 55:3217-3224.
- Vattern DA, Ghaedian R & Shetty K (2005). Enhancing health benefits of berries through phenolic antioxidant enrichment: focus on cranberry. Asia Pac J Clin Nutr 14:120-130. Review.
- Vattern DA, Randhir R & Shetty K (2005). Cranberry phenolicsmediated antioxidant enzyme response in oxidatively stressed porcine muscle. *Process Biochem* 40:2225-2238.
- Villarreal A, Stoecker BJ, Garcia C, Garcia K, Rios R, Gonzales C, Mandadi K, Faraji B, Patil BS & Deyhim F (2007). Cranberry juice improved antioxidant status without affecting bone quality in orchidectomized male rats. *Phytomedicine* 14:815-820.
- Vinson JA, Bose P, Proch J, A Kharrat H & Samman N (2008). Cranberries and cranberry products: powerful in vitro, ex vivo, and in vivo sources of antioxidants. J Agric Food Chem 56:5884-5891.
- Vinson JA, Zubik L, Bose P, Samman N & Proch J (2005). Dried fruits: excellent in vitro and in vivo antioxidants. J Am Coll Nutr 20:44-50.
- Wolfe KL & Liu RH (2007). Cellular antioxidant activity (CAA) assay for assessing antioxidants, foods, and dietary supplements. J Agric Food Chem 55:8896-8907.
- Zhang K & Zuo Y (2004). GC-MS determination of flavonoids and phenolic and benzoic acids in human plasma after consumption of cranberry juice. J Agric Food Chem 52:222-227.