

Isolation of *Serratia fonticola* from pirarucu *Arapaima gigas*

Casiano H. Choresca Jr¹, Ji Hyung Kim², Dennis K. Gomez³, Hwan Jang⁴,
Seong Joon Joh⁴, Se Chang Park^{2,3,*}

¹College of Veterinary Medicine, Seoul National University, Seoul 151-742, Korea

²Brain Korea 21 Program for Veterinary Science, College of Veterinary Medicine,
Seoul National University, Seoul 151-742, Korea

³KRF Zoonotic Disease Priority Research Institute, Seoul National University, Seoul, 151-742, Korea

⁴National Veterinary Research and Quarantine Service,
Anyang 430-824, Korea

(Accepted: February 13, 2008)

Abstract : The pirarucu, *Arapaima gigas* (body weight = 18.3 kg and total length = 102 cm) which had been reared in one of the private commercial aquaria for exhibition was found dead and submitted for diagnostic examination. A pure bacterial culture was isolated from the kidney, which was enlarged, and contained fluids. Result of the bacterial identification yielded *Serratia fonticola*. This paper describes the first isolation of *S. fonticola* from pirarucu.

Keywords : *Arapaima gigas*, fish, pirarucu, *Serratia fonticola*

Introduction

The pirarucu, *Arapaima gigas* is a gigantic tropical freshwater fish native to South America and Africa and undoubtedly one of the most important food fish species of the Amazon ichthyofauna [3, 8]. Starting as early as the late 18th century, a commercial fishery was developed for *Arapaima* to satisfy demand of bacalhau, a salted and dried cod for which *Arapaima* was found to be an excellent freshwater substitute [4]. The fish often grows up to 5 m in length and weigh as much as 200 kg [8]. In nature, pirarucus prey on small fish and other items such as mollusks, crustaceans and insects [3]. Recently, the use of pelleted feeds has been developed for intensive culture of this species [3]. The lack of knowledge about controlled breeding and general health management still hampers the pirarucu culture and needs to be overcome despite of all the advances achieved [3]. Due to its enormous commercial importance, *Arapaima* became increasingly scarce and commercially extinct near larger Amazonian cities [4]. In 1975, *Arapaima* became one of a few fish species listed by the Convention on International Trade

in Endangered species (CITE) II convention [4].

In this paper, five pirarucus (about 2 years old) were imported from Amazon River, South America. They were reared and confined in a 30 m³ aquarium tank equipped with heating and filtration systems and fed with frozen baitfish. They were displayed in one of the private commercial aquaria for indoor exhibition in Seoul, Korea for almost 3 years. In January 2007, four of the five pirarucus in the present aquarium showed abnormal swimming behavior and were later transferred to the other aquarium without any treatment. In the new aquarium, the four fishes showed lethargy, loss of appetite and died after 5 days. One of the pirarucus that were found dead was submitted for diagnostic examination. Fish showed no external body signs of disease, opercula and gills appeared to be normal, however, internally, kidney was enlarged and showed the presence of fluids. A pure bacterial culture was isolated from the enlarged kidney. The result of the bacterial identification yielded *Serratia fonticola*. This paper describes the first isolation of *S. fonticola* from pirarucu, *Arapaima gigas*.

*Corresponding author: Se Chang Park
College of Veterinary Medicine, Seoul National University, Seoul 151-742, Korea
[Tel: +82-2-880-1282, Fax: +82-2-880-1213, E-mail: parksec@snu.ac.kr]

Materials and Methods

Fish

Pirarucu (weighing 18.3 kg, total length 102 cm) that was found dead in one of the commercial aquaria was submitted for diagnostic examination.

Isolation and Identification of bacteria

Swabs from the gills, liver, spleen and kidney were streaked onto brain heart infusion agar (BHIA) (Becton, Dickinson and Company, USA), and incubated at 25°C for two days. Dense culture growth of bacteria was recovered from the kidney of the fish directly streaked onto BHIA. An isolated bacterium was restreaked again on fresh media to obtain the pure culture.

Gram staining and motility test were performed. An API 20E profile (bioMérieux, France) and Vitek System 2 (bioMérieux, France) were used for the purpose and to further characterize the isolate. The result of the bacterial identification was compared to reference strain from Bergey's Manual of Determinative Bacteriology [7].

Results

Result of the identification revealed that the isolated bacterium was Gram negative, rod in shape, and motile. Table 1 shows the result of bacterial identification as revealed by API 20E in comparison with the phenotypic characteristics from Bergey's Manual of Determinative Bacteriology.

The isolate was positive for β -galactosidase, arginine dihydrolase, lysine decarboxylase, ornithine decarboxylase, and utilization of citrate and fermentative. Negative for the production of H₂S, urease, tryptophane deaminase, production of indole, Voges-Proskauer reaction, gelatinase and utilization of sucrose.

The isolate was also positive for the utilization of glucose, mannitol, inositol, sorbitol, rhamnose, melibiose, amygladin and nitrite. Based on these features and with reference strain from the Bergey's Manual of Determinative Bacteriology, it only differed in arginine dihydrolase in which the isolate was positive while negative in Bergey's reference. However, we further characterized the isolated strain using Vitek 2 and the result showed 91% probability indicating that the isolate is closely related to *Serratia fonticola*.

Table 1. The characteristics of isolated strain as revealed by API 20E profile in comparison with the reference of *Serratia fonticola* strain from Bergey's Manual of Determinative Bacteriology

Characteristics	API 20E	Bergey's Manual ^a
β D Galactosidase (ONPG)	+	+
A-Dihydrolase	+	-
L-Decarboxylase	+	+
O-Decarboxylase	+	+
Citrate	+	+
H ₂ S	-	-
Urease	-	-
Tryptophane DeAminase	-	ns
Indole production	-	-
VP	-	-
Gelatinase	-	-
Glucose	+	+
Mannitol	+	+
Inositol	+	+
Sorbitol	+	+
Rhamnose	+	+
Sucrose	-	-
Melibiose	+	+
Amygladin	+	ns
Arabinose	+	+
Oxidase	-	-
NO ₂	+	+
O/F	F	F

+: positive, -: negative, ns: not stated, F: fermentative.

^aBergey's Manual of Determinative Bacteriology.

Discussion

In 1979, *S. fonticola* is an enterobacterium that was described as a new species of *Serratia* and was first isolated from water and soil [6]. It has been related to contaminants in the respiratory tract, and has been isolated in the droppings of European wild birds [11]. This bacterium was recognized to be a human pathogen. It has been described as a contaminant in cases involving wound sustained in a traffic accident [2], an abscess in an open fracture [13], a diarrheic process in an immunodepressed patient [14], and a wound from a bear bite [9]. It was reported that *S. fonticola* has an extensive distribution in aquatic environments [5] but no report yet in fish. So far, there were only a few reports on the isolation of other pathogenic *Serratia* species from fish [1, 10, 12, 15].

Many kinds of animals, including tropical fish, are usually kept in controlled condition like aquarium for exhibition but often suffer from stress or disease since their ecology and physiology are not always sufficiently understood [8]. The origin and mode of infection of the *S. fonticola* isolated from the pirarucu in the present study is not known. It is possible that the bacteria were transmitted to the fish via water, since poor water quality and filter system condition of the aquarium may also affect the development of the disease. Another possibility is transmission via aquarium tools and materials used inside the aquarium contaminated by fish carriers. In addition, it is also possible coming from baitfish contaminated with bacteria that were used as feeds. Stress was also speculated to be one of the reasons that caused the death of the fish. It may be due to inappropriate handling techniques of the animals and no acclimatization happened during the transfer of the fish to another aquarium by staff. Other physical conditions such as changes in temperature, dissolved oxygen and pH could also affect the animal condition, considering that these fish were held in an aquarium. Based on these speculations, it could possibly lead to an immunocompromised state of the fish paving the way for the entry of opportunistic bacteria such as *S. fonticola* that could cause bacterial infection leading to the mortality of the fish. Therefore, the maintenance of a biologically and chemically balanced environment and the appropriate regulation of physical conditions in the aquarium are required to avoid stress or disease. To our knowledge, this report was the first case of isolation of *S. fonticola* in fish. However, its direct association to the fish mortality is not certain.

Acknowledgments

This study was financially supported by a Korea Research Foundation Grant (KRF-2006-005-J02903) and the Research Institute for Veterinary Science, Seoul National University.

References

1. **Austin B, Stobie M.** Recovery of *Serratia plymuthica* and presumptive *Pseudomonas pseudoalcaligenes* from skin lesions in rainbow trout, *Oncorhynchus mykiss* (Walbaum), otherwise infected with enteric redmouth. *J Fish Dis* 1992, **15**, 541-543.
2. **Bollet C, Gannier M, Sainty JM, Orhesser P, De Micco P.** *Serratia fonticola* isolated from a leg abscess. *J Clin Microbiol* 1991, **29**, 834-835.
3. **de Andrade JIA, Ono EA, de Menezes GC, Brasil EM, Roubach R, Urbinati EC, Tavares-Dias M, Marcon JL, Affonso EG.** Influence of diets supplemented with vitamins C and E on pirarucu (*Arapaima gigas*) blood parameters. *Comp Biochem Physiol A Mol Integr Physiol* 2007, **146**, 576-580.
4. **Farias IP, Hrbek T, Brinkmann H, Sampaio I, Meyer A.** Characterization and isolation of DNA microsatellite primers for *Arapaima gigas*, an economically important but severely over-exploited fish species of the Amazon basin. *Mol Ecol Notes* 2003, **3**, 128-130.
5. **Farmer JJ III, Davis BR, Hickman-Brenner FW, McWhorter A, Huntley-Carter GP, Asbury MA, Riddle C, Wathen-Grady HG, Elias C, Fanning GR, Steigerwalt AG, O'Hara CM, Morris GK, Smith PB, Brenner DJ.** Biochemical identification of new species and biogroups of Enterobacteriaceae isolated from clinical specimens. *J Clin Microbiol* 1985, **21**, 46-76.
6. **Gavini F, Ferragut C, Izard D, Trinel PA, Leclerc H, Lefebvre B, Mossel DA.** *Serratia fonticola*, a new species from water. *Int J Syst Bacteriol* 1979, **29**, 92-101.
7. **Holt JG, Krieg NR, Sneath PHA, Staley JT, Williams ST.** *Bergey's Manual of Determinative Bacteriology*, 9th ed. pp. 216-245, The Williams & Wilkins Co., Baltimore, 1994.
8. **Kodama H, Nakanishi Y, Yamamoto F, Mikami T, Izawa H.** *Salmonella arizonae* isolated from a pirarucu, *Arapaima gigas* Cuvier, with septicemia. *J Fish Dis* 1987, **10**, 509-512.
9. **Kunimoto D, Rennie R, Citron DM, Goldstein EJC.** Bacteriology of a bear bite wound to a human: Case Report. *J Clin Microbiol* 2004, **42**, 3374-3376.
10. **McIntosh D, Austin B.** Recovery of an extremely proteolytic form of *Serratia liquefaciens* as a pathogen of Atlantic salmon, *Salmo salar*, in Scotland. *J Fish Biol* 1990, **36**, 765-772.
11. **Müller HE, Steigerwalt AG, Brenner DJ.** Isolation of *Serratia fonticola* from birds. *Zentralbl Bakteriell Mikrobiol Hyg* 1986, **261**, 212-218.
12. **Nieto TP, López LR, Santos Y, Núñez S, Toranzo AE.** Isolation of *Serratia plymuthica* as an opportunistic pathogen in rainbow trout, *Salmo gairdneri* Richardson.

- J Fish Dis 1990, **13**, 175-177.
13. **Pfyffer GE.** *Serratia fonticola* as an infectious agent. Eur J Clin Microbiol Infect Dis 1992, **11**,199-200.
14. **Soler C, Samson T, Hernandez E, Herve V, de Revel T.** *Serratia fonticola*, an opportunistic germ: observations of an immunodepressed patient. Med Mal Infect 2000, **30**, 599-600.
15. **Vigneulle M, Laurencin FB.** *Serratia liquefaciens*: a case report in turbot (*Scophthalmus maximus*) cultured in floating cages in France. Aquaculture 1995, **132**, 121-124.