

The Effects of Hyaluronic Acid-Carboxymethylcellulose Membrane (GUARDIX-MB[®]) Barriers on Prevention of Post-operation Peritoneal Adhesions in Dogs

Sang-Mook Lee, Hwan-Soo Jang*, Jae-Sung Bae**, Jung-Eun Kim and Kwang-Ho Jang¹

College of Veterinary Medicine, Kyungpook National University, Daegu 702-701, Korea Department of *Pharmacology and **Physiology, School of Medicine, Kyungpook National University, Daegu 700-721, Korea

(Accepted: December 15, 2008)

Abstract : The aim of this study was to determine the effectiveness of hyaluronic acid-carboxymethylcellulose membrane (GUARDIX-MB[®]) barriers on prevention against post-operative peritoneal adhesions. In this study, fourteen mongrel dogs were divided into two experimental groups: 0.1 % hyaluronic acid (0.1HA) group and hyaluronic acid-carboxymethylcellulose membrane (HA-CMC) group. In order to induce adhesions, the anti-mesenteric serosa of the ileum was exteriorized and then abraded in a standard manner by scraping with a scalpel blade to create homogenous petechial hemorrhagic surface over a 1 × 1 cm area. Solution of 0.1HA were simply coated over the abraded tissues, 1.5 × 1.5 cm HA-CMC membrane was placed over the abraded tissues, allowed to spread across the intra-abdominal organs before closure of the abdomen. On day 1 before and day 1, 4, 7, 14, and 21 after operation, venous blood specimens were collected for measurement of fibrinogen and total WBC. The adhesions were blindly assessed 3 weeks later by using a computerized tensiometer. The fibrinogen and total WBC values of two groups showed no statistical significances. The mean tensile strength (gram force, gf) of formed adhesions day 21 after surgery was 88.1 ± 55.70 gf in the 0.1 % HA group and 24.8 ± 22.69 gf in the HA-CMC group. The tensile strength values of adhesion separation HA-CMC membrane group was significantly lower than the 0.1HA group (p<0.05). Therefore, we suggest that HA-CMC membrane reduce peritoneal adhesions may be applicable to preventing post-operative intraperitoneal adhesions in dogs.

Key Words: hyaluronic acid-carboxymethylcellulose membrane, peritoneal adhesions, tensile strength, dogs.

Introduction

Postoperative adhesions is an important and so far unsolved surgical problem (38,43). Adhesions were present in more than 94 % of patients who underwent major abdominal operations (4,5). Intra-abdominal adhesions are common cause of mechanical bowel obstructions (3,11,25), female infertility (6,17), and then they may be related to chronic abdominal pain (22,35). Adhesions are fibrinous or fibrous bands that form abnormal unions between two or more surfaces that are normally covered with the serosa (23).

Adhesion formation (10,20,21,30,31) begins with a fibrin matrix; cellular elements become prominent in the matrix at 1 to 3 days. Vascular granulation tissue containing macrophages, fibroblasts, and giant cells gradually replaces this matrix (19). After 4 days, most of the fibrin disappears, macrophages become the predominant leukocyte, and a larger number of fibroblasts and associated collagen are present. At day 5, small vascular channels containing endothelial cells are seen, and within the adhesion while collagen deposition

¹Corresponding author. E-mail : khojang@knu.ac.kr and organization advance (15). During the second week, the relatively few cells present are predominantly fibroblasts. Mesothelium (13) often covers well-defined adhesions, which contain blood vessels and connective tissue fibers, including elastin (30).

To prevent or reduce the complication of intraperitoneal adhesions numerous studies have been done on animals (9). Various adjuvants have been tried to prevent postoperative adhesions, these are as follow: anti-inflammatory agents (NSAIDs, corticosteroids), antibiotics, rubricate (fluid) agents (normal saline, chlorohexidine, sodium carboxymethylcellose) (40), hyaluronic acid, dextran 70, and antioxidative agents (vitamin E). In the past decade, significant progress has been made with the development of adhesion barriers. These include Gore-Tex[®] membrane; oxidized, regenerated cellulose (Interceed[®]) (2,18); and hyaluronic acid-carboxymethy-cellulose (HA-CMC) membrane (Seprafilm[®]; Genzyme, Somerville, NJ). Hyaluronic acid (HA) has been shown to prevent adhesion formation in both experimental models and human subjects (1).

HA-CMC membrane (GUARDIX-MB[®]) (7,26,39,41) is applied directly to specific sites of surgical trauma to provide a physical barrier that separates traumatized tissue from other tissues during normal healing. Composed of sodium hyaluronate and carboxymethylcellulose, HA-CMC membrane is slowly resorbed into the body, maintaining a barrier effect while the body's normal tissue repair takes place.

Accordingly the aim of this study was to evaluate the effectiveness of HA-CMC membrane barriers by comparing with HA capacity of which to prevent the formation of postoperative adhesions in a noninfectious environment is well known.

Materials and Methods

1. Experimental animals

Fourteen healthy adult mongrel dogs (weighing 4.9 ± 1.4 kg) were used in the study. They were vaccinated with DHPPL and dewormed with febantel (Drontal[®] Plus, Bayer Korea Ltd, Korea). Experiments were started after an initial adaption period for about ten days. The dogs were housed in each cage, and the food and water were fed *ad libitum*. All dogs were divided into two groups containing seven animals, respectively: 0.1 % hyaluronic acid-treated group (0.1HA), and hyaluronic acid-carboxymethylcellulose membrane (HA-CMC) group.

2. Preparation of materials

The HA-CMC membrane was kindly provided by BIO-RANE. Co., Ltd. The anti-adhesion membrane was prepared by lyophilizing HA-CMC solution and cross-linking properly with 1-ethyl-3- (3-dimethylaminopropyl) carbodiimide (EDAC).

HA was used as a 0.1 % solution. 0.1 % HA solution was prepared by adding 22.5 ml of sterile water at 1 % HA (Sodium hyaluronate 25 mg/ 2.5 ml, HYAL[®], Shinpoong Pharm Co, Korea) and then mixing with Vortex Genie 2, and filtering with syringe filter (0.45 Micron, 25 mm). All materials were stored at 4 °C until used.

3. Surgical Procedures

Feed was withheld from experimental dogs 12 hours prior to surgery. Anesthesia for surgery was similar for all groups. Each dog was administered atropine sulfate (Atropine[®], Dai Han Pharm Co, Korea, 0.05 mg/kg) subcuticularly. Anesthesia was induced and maintained with intramuscularly tiletmine/zolazepam (Zoletil[®], Virbac Co, Korea, 5 mg/kg). Dogs were positioned in dorsal recumbency and the entire abdomen was clipped. The abdomen of each dog was prepared with povidone-iodine solution and 70 % alcohol for aseptic surgery. Sterile surgical instruments and supplies were used in all surgical procedures. The surgical site was draped in a common manner. A 5 cm ventral midline incision was made through the skin, subcutaneous tissue, and peritoneum.

0.1HA Group: After the abdomen was opened, the ileum and ileocecal junction were examined and then exteriorized. Total five distinct surgical lesions were made in order to induce adhesions. The antimesenteric defects of the ileum approximately 5 cm from the ileocecal junction were created by light scraping about 1×1 cm areas with a sterile No. 10 scalpel blade to promote petechial bleeding, and allowed to air dry for 10 minutes. The total abrasions were 5 places at intervals of 5 cm. In this group, solution of 0.1HA were simply coated over the abraded tissues (3 ml/abraded area). And then the small bowel was returned to its original location. Before closure of the abdomen, liquid product was allowed to spread across the intraperitoneal organs (3 ml/kg) through a catheter using a syringe. The catheter was removed, and abdominal wall was closed using a simple interrupted pattern with 2-0 polyglactin 910 (Vicryl[®], Johnson&Johnson Medical Korea Ltd, Korea) and the skin was closed by a simple interrupted pattern with 3-0 nylon (BLUE NYLON[®], Ailee Co, Ltd).

HA-CMC Group: Antimesenteric defects to induce adhesions were created at five locations using a similar method described 0.1HA Group. In this group, 1.5×1.5 cm HA-CMC membrane was placed over the abraded tissues. The abdominal cavity was closed using with same method used in 0.1HA group.

One surgeon carried out all the operations and was blind to the randomization while he was performing the abrasions.

4. Postoperative Evaluations

The experimental animals were monitored daily for signs of abdominal pain, drainage, feed consumption or defecation. Enrofloxacin (Baytril, Bayer Korea Ltd, Korea, 5 mg/ kg) was administered subcuticularly to reduce the risk of postoperative infection for 3 days.

Fibrinogen and WBC values : On day -1 (day 1 before operation), 1, 4, 7, 14, and 21 after operation, venous blood specimens were collected from all experimental animals for measurement of fibrinogen and total WBC.

Adhesion Evaluation : Three weeks later, all the animals were sacrificed and the intra-abdominal cavity was inspected through ventral midline incision. And then a postmortem examination was conducted immediately. Adhesions were identified, and the adhesion site was excised to test. The tensile strength of the adhesion site was evaluated with tensiometer (H500DM[®], Hounsfield Co, UK) (Fig 1). Both adhesive tissue ends were secured tightly in a clamp so as not to slip from the clamp during tensile test. The clamp was advanced at the rate of 0.7 cm/min. During the operation of tensiometer, breaking strength of specimens was measured. Tests were performed at the room temperature of 20 °C, and specimens were moistened with a very fine mist of normal saline while clamped.

5. Statistical analysis

Values are expressed as mean \pm SD. Analysis of differences between treated groups was performed using analysis of variance followed by Student's t test. Differences in the separation strength of two groups at P<0.05 were considered statistically significant.



Fig 1. The tensiometer used in the measurement of the force required to separate adhesions.

Results

The second laparotomy was performed in all experimental groups at 3 weeks after surgery. All animals had insignificant postoperative signs during the experimental periods.

1. Laboratory Values

Fibrinogen: The fibrinogen concentration was normal on day -1 in all dogs, On day 1, values of fibrinogen was increased higher than those on day -1. The concentration of fibrinogen of 0.1HA group was decreased gradually from day 7, and recovered to normal ranges on day 14. The concentration of fibrinogen in HA-CMC group was decreased gradually from day 4, and recovered to normal ranges on day 21 (Fig 2). There was no statistical significance between the two groups.

WBC: The total WBC was normal on day -1 in all dogs, On day 1, they were increased higher than those on day -1. Total WBC numbers decreased gradually from day 4, and recovered from normal ranges on day 14 (Fig 3). There was no significant difference between the two groups.

2. Tensile Strength

All of the animals in 0.1HA group were admitted for occurrences of adhesions. Adhesions were occurred among five animals in HA-CMC group except two animals (Table 1).

The mean adhesion separation strength (gram force, gf) was 88.1 \pm 55.70 gf in 0.1HA group, and 24.8 \pm 22.69 gf in HA-CMC group. Values of adhesion formation in HA-CMC group was significantly reduced, compared with those in 0.1HA group (P<0.05) (Table 2).

Adhesions were identified in total sites; serosa to serosa (18 of 70, 25.7%), serosa to mesentery (27 of 70, 38.6%), serosa to omentum (5 of 70, 7.1%), serosa to parietal perito-



Fig 2. Changes of fibrinogen concentration of blood serum over time in dogs intraperitoneal adhesion was induced. 0.1HA : 0.1 % hyaluronic acid-treated group HA-CMC : hyaluronic acid-carboxymethylcellulose membrane-treated group. Mean \pm SD.



Fig 3. Changes of total WBC count in dogs intraperitoneal adhesion was induced. Mean \pm SD.

 Table 1. Adhesion incidences in each dog of groups on day 21

 after operation (no. of adhesion sites

Crown	No. of animal							Total
Gloup -	1	2	3	4	5	6	7	- 10141
0.1HA	3	3	6	3	4	7	3	29/35
HA-CMC	4	7	0	2	7	3	0	23/35

neum (2 of 70, 2.9 %). Total number of adhesions was 29 of 35 in 0.1HA group, and 23 of 35 in HA-CMC group (Table 3).

The mean strength (gram force, gf) of the adhesion separation in each sites ; serosa to serosa (121.8 \pm 107.65 gf), serosa to mesentery (61.0 \pm 31.64 gf), serosa to omentum (16.5 \pm 2.12 gf), serosa to parietal peritoneum (306 gf) in 0.1HA group, and serosa to serosa (56.3 \pm 39.57 gf), serosa to mesentery (35.8 \pm 19.76 gf), serosa to omentum (7.3 \pm 4.04 gf), serosa to parietal peritoneum (56 gf) in HA-CMC group (Fig 4).

Gross appearance of intraperitoneal adhesion in 0.1HA group was dense and broad, but that in HA-CMC group was

Group —	No. of animal							
	1	2	3	4	5	6	7	(mean \pm SD)
0.1HA	60.6 ± 14.97	201.3 ± 153.08	102.1 ± 101.95	327.6 ± 14.50	454.5 ± 50.73	80.4 ± 75.67	90 ± 16.70	88.1 ± 55.70
HA-CMC	26.2 ± 22.85	732.7 ± 13.11	0	224.5 ± 2.12	67.1 ± 38.42	23.3 ± 28.57	0	$24.8 \pm 22.69^*$

Table 2. The mean strength values of the adhesion separation in each dog of groups

* p<0.05 compare with the 0.1HA

 Table 3. Postoperative locations of adhesions in dogs 21 after operation (no. of adhesion sites)

	Location							
-	Total	S-S	S-M	S-O	S-PP			
0.1HA	29	10	16	2	1			
HA-CMC	23	8	11	3	1			

Locations of adhesions : S-S = serosa-serosa; S-M = serosa-mesentery; S-O = serosa-omentum; S-PP = serosa-parietal peritoneum.

filmy and thin (Fig 5).

Discussion

Injury or inflammation of a serosal surface generally initiates adhesion formation processes (8). Destroyed peritoneum fails to absorb fibrinogen-rich fluid exudate. In addition activation of inflammatory cells (42), secretion of inflammatory cytokines, and activation of the complementcoagulation cascades creates thrombin, which converts fibrinogen to fibrin in the exudate (24). Fibrin deposition not degraded within the first days of injury results in the progression of a fibrinous adhesion into a fibrous form with the movement and propagation of fibroblasts into the fibrin matrix. If degradation occurs, mesothelium (13) repairs its defect without adhesion formation (10,16,20,21,30,31). Small bowel is the most frequently observed location of adhesion formation because it occupies most of the abdomen and in located in the middle of the peritoneal cavity(14). For these reasons, it is very important to identify an anti-adhesive



Fig 4. The strength values of the adhesion separation in each sites.

product to prevent adhesions, especially between small bowel segments and between small bowel and other organs. Therefore, we abraded illeum to induce adhesion in our experiment.

Numerous agents have been used to reduce postoperative adhesions. No agent has yet proved effective against the formation of intra-abdominal abscesses and adhesions in bacterial peritonitis in patients. A promising new concept may be the application of intra-abdominal agents (12) containing the polysaccharide hyaluronan (36), which hae been shown to prevent adhesions in experimental and clinical studies (32). HA, a naturally occurring glycosaminoglycan, forms a highly viscous solution coating the serosal surfaces. Many studies showed HA decreases inflammation (34), interferes with



Fig 5. Gross appearance of adhesiolysis in dogs intraperitoneal adhesions was induced. 0.1HA (left) group and HA-CMC (right) group.

fibrin formation, prevents adhesion (29), and stimulates fibrinolytic and TNF- α (27) response. Preventing the formation of postoperative adhesions among these reactions (33) is believed viscous polysaccharides solutions reduce serosal trauma by coating the peritoneal surfaces which become damaged during abdominal surgery.

Application of a mechanical barrier between the injured peritoneum is a method to prevent adhesion formation. HA-CMC membrane is such a barrier (28). It is a biologically resorbable membrane, which composed of HA absorbed CMC film. The ideal barrier should be non-reactive, absorbable and easy to use, and it should remain in the lesion site during critical stages of healing. The safety and efficacy of this membrane in preventing postoperative adhesion has been shown in clinical and experimental studies (39). Therefore, we supposed that the HA-CMC membrane functions as a physical barrier by separating the serosal surfaces may be beneficial in the treatment of intraperitoneal adhesions related to inflammation and fibrin.

Bleeding is likely to cause a massive extravasation of fibrinogen into the peritoneal cavity. Adhesion formation begins with fibrin deposition within an inflammatory exudate formed over an injured serosal layer (37). Our result showed that the fibrinogen concentration of HA-CMC treated group were higher than those of 0.1HA treated group on day 1 and 4, but were decreased gradually from day 4, and recovered to normal ranges on day 21. We have no idea about the reason the fibrinogen concentration of HA-CMC treated group was higher. Further studies is warranted on the relation of HA-CMC membrane and fibriongen concentration.

The value of total WBC was normal on day -1 in all dogs. On day 1 after operation, total WBC were increased higher than those on day -1. Total WBC concentraton decreased gradually from day 4, and recovered to normal ranges on day 14. In this study, we suppose that total WBC count increase is resulted from inflammation after operation and the change of total WBC count could be caused by the inflammatory reaction rather than change in the adhesion formation.

In present study, we performed by using a HA-CMC membrane and 0.1 % HA solution. All of the animals in 0.1HA group and five animals in HA-CMC group were admitted for occurrences of adhesions. Values of adhesion formation in HA-CMC group was statistically significant reductions in compared with those in 0.1HA group (P<0.05). In this results, we can consider that HA-CMC membrane reduced the incidence and severity of adhesion because of the physical properties of the membrane; Reduce fibrin deposits by limiting the inflammatory response, facilitate the degradation of fibrin with fibrinolytic stimulators and separate surfaces during the time that the fibrin remains sticky. The results are in agreement with similar studies (39). Almost all studies related to HA-CMC membrane were about the physical or barrier effects of the material. HA-CMC membrane is a temporary proven to reduce the incidence, extent, and severity of adhesions in patients undergoing abdominal or pelvic laparotomy. It was designed to prevent adhesions by separating traumatized tissue surfaces (44). When applied to traumatized tissues, HA-CMC membrane hydrates and becomes a gel and remains in place during the critical 7-day healing period. One limitation of HA-CMC membrane is its handing characteristics. It is somewhat brittle and sometimes difficult to apply. Theoretically, dislocation is possible after application, and this may interfere with the membrane's antiadhesions effect.

Therefore we suggest that the HA-CMC membrane was effective on preventing the formation of postoperative small intestinal adhesions in the dog. Further studies about histological analysis are needed to find inflammatory reaction and fibrotic response in adhesive sites.

References

- Abraham N. Morse. Robert A. Hammer. Jeffrey L. Cornella. Joseph C. Loftus. Validation of a Mouse Adhesion Reduction Model Using Seprafilm[®]. Journal of Gynecologic Surgery 2005; 21: 147-153.
- Arnold PB, Green CW, Foresman PA, Rodeheaver GT. Evaluation of resorbable barriers for preventing surgical adhesions. Fertil Steril 2000; 73: 157-161.
- Ara C, Kirimlioglu H, Karabulut AB, Coban S, Hascalik S, Celik O, Yilmaz S, Kirimlioglu V. Protective effect of melatonin against oxidative stress on adhesion formation in the rat cecum and uterine horn model. Life Sci. 2005;77:1341-1350.
- Bae JS, Jang KH, Kwon YS, Jang HS, Kim JE, Park SI, Lim JH, Li WX and Lee SJ. The effect of hyaluronic acid and vitamin E combination on preventing postoperative intraperitoneal adhesion formation in dogs. Korean J Vet Clin 2003; 20: 42-48.
- Becker JM, Stucchi AF. Intra-abdominal adhesion prevention: are we getting any closer? Ann Surg 2004; 240: 202-204.
- Berkkanoglu M, Zhang L, Ulukus M, Cakmak H, Kayisli UA, Kursun S, Arici A. Inhibition of chemokines prevents intraperitoneal adhesions in mice. Hum Reprod 2005; 20: 3047-3052.
- Bristow RE, Montz FJ. Prevention of adhesion formation after radical oophorectomy using a sodium hyaluronatecarboxymethylcellulose (HA-CMC) barrier. Gynecol Oncol 2005; 99: 301-308.
- Cheong JT, Lee KK, Jang KH. Prevention of abdominal Re-adhesions by sodium carboxymethylcelluloes in dogs with abdominal adhesions. Korean J Vet Clin Med 1997; 14: 161-167.
- Cohen Z, Senagore AJ, Dayton MT, Koruda MJ, Beck DE, Wolff BG, Fleshner PR, Thirlby RC, Ludwig KA, Larach SW, Weiss EG, Bauer JJ, Holmdahl L. Prevention of postoperative abdominal adhesions by a novel, glycerol/ sodium hyaluronate/carboxymethylcellulose-based bioresorbable membrane: a prospective, randomized, evaluator-blinded multicenter study. Dis Colon Rectum 2005; 48: 1130-1139.
- Cornum R, Bell J, Gresham V, Brinkley W, Beall D, MacPhee M. Intraoperative use of the absorbable fibrin adhesive bandage: long term effects. J Urol 1999; 162: 1817-1820.

- 11. Fazio VW, Cohen Z, Fleshman JW, van Goor H, Bauer JJ, Wolff BG, Corman M, Beart RW Jr, Wexner SD, Becker JM, Monson JR, Kaufman HS, Beck DE, Bailey HR, Ludwig KA, Stamos MJ, Darzi A, Bleday R, Dorazio R, Madoff RD, Smith LE, Gearhart S, Lillemoe K, Gohl J. Reduction in adhesive small-bowel obstruction by Seprafilm adhesion barrier after intestinal resection. Dis Colon Rectum 2006; 49: 1-11.
- Ferland R, Mulani D, Campbell PK. Evaluation of a sprayable polyethylene glycol adhesion barrier in a porcine efficacy model. Hum Reprod 2001; 16: 2718-2723.
- Frederik J. T. van Oosterom, Jan J. B. van Lanschot, Johannes Oosting, Huug Obertop Hyaluronic acid/carboxymethylcellulose membrane surrounding an intraperitoneal or subcutaneous jejunojejunostomy in rats. Eur J Surg 2000; 166: 654-658.
- 14. Han TS, Lee JM, Yun YM, Kang 쇼, Shin TK, Kang YH, Kim NJ, Kim HS, Lee KK, Cheong JT. Sodium carboxymethylcellulose and hyaluronic acid on prevention of intra-abdominal adhesion in rats. Korean J Vet Clin Med 2003; 20: 323-327.
- Hellebrekers BW, Emeis JJ, Kooistra T, Trimbos JB, Moore NR, Zwinderman KH, Trimbos-Kemper TC. A role for the fibrinolytic system in postsurgical adhesion formation. Fertil Steril 2005; 83: 122-129.
- Jang HY, Yoon HY, Kim JY, Han HJ, Lee BR, Won HJ, Jeong SW. Evaluation of sodium carboxymethylcellulose for prevention of adhesion in intestinal anastomosis in dogs. Korean J Vet Res 2005; 45: 417-421
- Jang KH, Kwon YS, Kim JE, Kwon EJ, Oh TH, Lee KW, Jang IH. Effect of carboxymethyl chitosan on postoperative intraperitoneal adhesion formation in the rat. Korean J Vet Res 2000; 40: 635-643.
- Jang YK, Kim HH, Lee HC, Yeon SC, Lee HJ. Reduction of postoperative adhesions by a synthetic solution of sodium carboxymethylcellulose and dextran or interceed following intestinal anastomosis in dogs. Korean J Vet Clin Med 2004; 21: 349-354.
- Jeremy Thompson. Pathogenesis and Prevention of Adhesion Formation. Digestive Surgery 1998; 15: 153-157.
- Jung DK, Cheong JT, Lee KK, Kim HS, Choi NJ, Yeon SC, Seo KM. The effect of sodium carboxymethylcellulose on prevention of post-operative pleural and pericardial adhesions in dogs. Korean J Vet Clin Med 2000; 17: 368-375.
- Kang YH, Cheong JT, Yeon SC. Prevention of uterine adhesion by sodium carboxymethylcellulose in dogs. Korean J Vet Clin Med 2000; 17: 381-387.
- 22. Kayaoglu HA, Ozkan N, Hazinedaroglu SM, Ersoy OF, Koseoglu RD. An assessment of the effects of two types of bioresorbable barriers to prevent postoperative intraabdominal adhesions in rats. Surg Today 2005; 35: 946-950.
- Klein ES, Asculai SS, Ben-Ari GY. Effects of hyaluronic acid on fibroblast behavior in peritoneal injury. J Surg Res 1996; 61: 473-476.
- Kwon YS, Jang KH. Effects of carboxymethyl chitosan fabric and low molecular weight heparin on reducing adhesin formation in the rat. Korea Vet Res 2003; 43: 703-708.
- 25. LeBlanc KA, Stout RW, Kearney MT, Paulson DB. Comparison of adhesion formation associated with Pro-Tack (US

Surgical) versus a new mesh fixation device, Salute (ONUX Medical). Surg Endosc 2003; 17: 1409-1417.

- Lee YW, Lee YM. Evaluation on effectiveness for preventing pest surgical adhesion of sodium hyaluronatesodium carboxymethylcelluose (HA/CMC) membrane in rat cecum/peritonium model. Membrane journal 2005; 15: 213-223.
- Mirastschijski U, Johannesson K, Jeppsson B, Agren MS. Effect of a matrix metalloproteinase activity and TNF-alpha converting enzyme inhibitor on intra-abdominal adhesions. Eur Surg Res 2005; 37: 68-75.
- Miyamoto K, Masuda K, Inoue N, Okuma M, Muehleman C, An HS. Anti-adhesion properties of a thrombin-based hemostatic gelatin in a canine laminectomy model: a biomechanical, biochemical, and histologic study. Spine 2006; 31: 91-97.
- 29. Oh DS, Manning MM, Emmanuel J, Broyles SE, Stone HH. Repair of full-thickness defects in alimentary tract wall with patches of expanded polytetrafluoroethylene. Ann Surg 2002; 235: 708-711.
- Ozel H, Avsar FM, Topaloglu S, Sahin M. Induction and assessment methods used in experimental adhesion studies. Wound Repair Regen 2005; 13: 358-364.
- Ozmen MM, Ozalp N, Zulfikaroglu B, Abbasoglu L, Kacar A, Seckin S, Koc M. Histoacryl blue versus sutured left colonic anastomosis: experimental study. ANZ J Surg 2004; 74: 1107-1110.
- Reijnen MM, Bleichrodt RP, van Goor H. Pathophysiology of intra-abdominal adhesion and abscess formation, and the effect of hyaluronan. Br J Surg 200; 90: 533-541.
- Reijnen MM, Skrabut EM, Postma VA, Burns JW, van Goor H. Polyanionic polysaccharides reduce intra-abdominal adhesion and abscess formation in a rat peritonitis model. J Surg Res 2001; 101: 248-253.
- Sawada T, Hasegawa K, Tsukada K, Kawakami S. Adhesion preventive effect of hyaluronic acid after intraperitoneal surgery in mice. Hum Reprod 1999; 14: 1470-1472.
- Schafer M, Krahenb hl L, Buchler MW. Comparison of adhesion formation in open and laparoscopic surgery. Dig Surg 1998; 15: 148-152.
- Seeger JM, Kaelin LD, Staples EM, Yaacobi Y, Bailey JC, Normann S, Burns JW, Goldberg EP. Prevention of postoperative pericardial adhesions using tissue-protective solutions. J Surg Res 1997; 68: 63-66.
- Sulaiman H, Dawson L, Laurent GJ, Bellingan GJ, Herrick SE. Role of plasminogen activators in peritoneal adhesion formation. Biochem Soc Trans 2002; 30: 126-131.
- Tarhan OR, Barut I, Sutcu R, Akdeniz Y, Akturk O. Pentoxifylline, a methyl xanthine derivative, reduces peritoneal adhesions and increases peritoneal fibrinolysis in rats. Tohoku J Exp Med 2006; 209: 249-255.
- Tarhan OR, Eroglu A, Cetin R, Y Nce A, Bulbul M, Altuntas YR. Effects of seprafilm on peritoneal fibrinolytic system. ANZ J Surg 2005; 75: 690-692.
- 40. Tingstedt B, Nehez L, Axelsson J, Lindman B, Andersson R. Increasing anastomosis safety and preventing abdominal adhesion formation by the use of polypeptides in the rat. Int J Colorectal Dis 2006; 21: 566-572.
- 41. Vrijland WW, Tseng LN, Eijkman HJ, Hop WC, Jakimowicz JJ, Leguit P, Sta, Swank DJ, Haverlag R, Bonjer

HJ, Jeekel H. Fewer intraperitoneal adhesions with use of hyaluronic acid-carboxymethylcellulose membrane: a randomized clinical trial. Ann Surg 2002; 235: 193-199.

- Vural B, Canturk NZ, Esen N, Solakoglu S, Canturk Z, Kirkali G, Sokmensuer C. The role of neutrophils in the formation of peritoneal adhesions. Hum Reprod 1999; 14: 49-54.
- 43. Wang XC, Gui CQ, Zheng QS. Combined therapy of

allantoin, metronidazole, dexamethasone on the prevention of intra-abdominal adhesion in dogs and its quantitative analysis. World J Gastroenterol 2003; 9: 568-571.

44. Yamaner S, Kalayci M, Barbaros U, Balik E, Bulut T. Does hyaluronic acid-carboxymethylcellulose (HA-CMC) membrane interfere with the healing of intestinal suture lines and abdominal incisions? Surg Innov 2005; 12: 37-41.