

Identification of *Aspergillus nidulans* from cooked eggs produced by permitted factory

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Abstract

Fungus generally doesn't produce toxic or harmful substances so it has low chances to cause food poisoning. However it leads to change appearance, odor and characteristics of the contaminated foods and result in sanitary risk problems. Therefore the contamination of fungi should be prevented since they are not proper for human consumption. Green fungi with white outline raised from the air cell of cooked eggs which were collected by Gyeonggi Livestock Veterinary Service in August, 2006. The results came out after the cultivation using Sabouraud's Dextrose Agar(SDA). The conidium appeared white and monospore, the shape of colony was round and oval. Conidiophore was brown and granulated and wrinkles and formed. It was confirmed as *Aspergillus nidulans* based on the dying using Lactophenol cotton blue, the observation of septum and vesicle from the grown spores, and rDNA sequencing

Key words : Fungi, spore, Lactophenol cotton blue, rDNA sequencing, *Aspergillus nidulans*

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Introduction

Generally the mold very well grow in everywhere with proper moisture, temperature and nutrition. When condition is not good for themselves, they overcome it by forming spore. Mold spores are scattered in everywhere such as air,

soil, utensils, clothes, food, and they germinate in moist and warm circumstance. According to the report¹⁾ we live with the mold every day even though there were slight seasonal differences in the density of the mold spore. In April, for instance, the density of indoor and outdoor were 17.1sp/m³ and 46.0sp/m³,

respectively. In July, that of indoor was 287.4sp/m³ and outdoor 475.8 sp/m³. In October, the former was 474.7 sp/m³ and the latter 1105.9sp/m³

The unsatisfied cases of mold contamination in eggs were not known in Korea because the individual standard of it is not set up in Food Hygiene Act or Livestock Processing Act. In addition because mold contamination case is classified as foreign material case, there is no case which reported just mold pollution in eggs.

This study would like to report the results of identification *Aspergillus nidulans* from cooked eggs produced by legal factory.

Methods and Results

Cultivation and macroscopic finding

The eggs collected from Gyonggi province in August, 2006, had thick green brimmed whitish fluffy colony mold in airsack (Fig 1). The mold was examined by common lab protocol²⁾, then cultivation and microscopic observation was performed as usual. The mold was inoculated in Sabouraud Dextrose (SD) agar and observed. It formed fluffy colony of 2 cm diameter (Fig 2) 2 days after inoculation, and bright green center (Fig 3) was shown 3 days after culture. Following a week, it formed granular wrinkle, and the center of that became thick green with white brim (Fig 4).

cultivation samples (Fig 5) were prepared. The mold was inoculated in the end of SD agar and it was covered with cover glass and cultivated with it. The inoculated glass was stained with a drop of lactophenol cotton blue (BD, USA) and it was investigated with microscope (× 40). The mold was suspected of genus *Aspergillus* because ascus was formed in conidiospore and septum (Fig 6).



Fig 1. *Aspergillus nidulans* on cooked egg



Fig 2. *Aspergillus nidulans* on SDA 2 days after culture

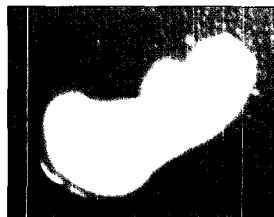


Fig 3. *Aspergillus nidulans* on SDA 3 days after culture

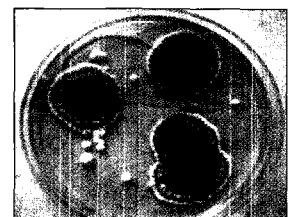


Fig 4. *Aspergillus nidulans* on SDA 7 days after culture

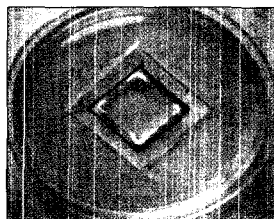


Fig. 5. Slide culture

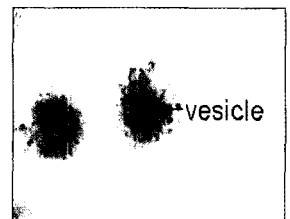


Fig 6. Slide culture of *Aspergillus nidulans* mounted in lactophenol cotton blue

Microscopic examination

For microscopic examination the slide

Analysis of rDNA sequencing

The additive analysis for identification

was conducted by the Korean Microbe Preservation Center. The mold was identified as *A. nidulans* because of very high homology (99%) after ITS-5.8S rDNA sequencing (Fig 7, 8, 9).

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>06-105
GGCTGCTCGGGGCGCCAACTCCACCGGTGAATACTAACACTGTTGCTTCGGGGGGAACCCCTCGGGGGGAGCGCGCCG
GGCACTACTGAACCTTCATGCTCGAGAGTATGCACTCTGAGTCTGAATATAAAATCAGTCAAACCTTCAACAATGGATCTCTTG
GTTCCGGCATCGATGAAGAAGCAGCGAAGTGCATAGTAATGTGAATTCGAGAAATTCAGTGAATCATCGAGTCTTTGACGCA
CATTTGCGCCCCCTGGCATTCCGGGGGCATGCGCTGTCCGAGGGTCAATGCTGCCATCAAGCCCGGCTTGTGTGTTGGGTGCTCG
TCCCCCGGGGGGACGCGCCGAAAGCAGCGCGCCACCGTGTCCGCTCCGAGGGTATGGGGCTTTGTACCCGCTCGACTA
GGCCCGCGCGGGGCGCCAGCGGAGTCTCCAGCAATTTTCTTCAGGT
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Fig 7. ITS-5.8S rDNA sequencing result (sample: 06-105)

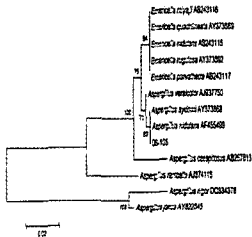


Fig. 8. Dendrogram 1 (sample: 06-105)



Fig. 9. Dendrogram 2 (sample: 06-105)

and others. It doesn't cause disease, but sometimes the disease could be developed if the immunity is weakened^{3,4}. For instance *Aspergillus nidulans* is the etiologic agent of cutaneous aspergillosis⁵, maxillary sinus disease^{6,7}, osteomyelitis⁸, pulmonary disease^{9,10}, guttural pouch mycosis in a horse^{11,12}, and a cerebral abscess¹³.

As the resistance mechanism of eggs against contamination of the mold, the surface could prevent the infiltration of microbes by the cuticula layer which is originated from the protein mucus secreted from the oviduct. Air sac is located in the blunt end of egg which has a lot of pores, thus through these microbes can penetrate into the egg. The shell can protect the egg from microbes because it is composed of keratin and reticular system¹⁴. So, in order to protect this system in the course of egg heating procedure, we have to treat the egg with special care.

In conclusion, the mold of the cooked egg collected from 2006 August Gyeonggi province represented the similarity with *A. nidulans* by the rDNA sequencing.

References

1. Kim CH, Lee KY, Kim WK, et al. 2001. The seasonal differences in the density of the indoor and outdoor mold spore in Seoul area. *The Korean Academy of Asthma, Allergy and Clinical Immunology*. 2001 Spring Conference Abstract : 432
2. Quinn PJ, Carter ME, Markey BK, et al. 2000. *Clinical veterinary microbiology*

Discussion

Aspergillus belongs to the Ascomycota and has the common feature of holding diploid ascospore in the ascus and has the characteristics of having ascus in reproducing process. Today there are about 1,000 species in the world. Because of having diploid ascospore, some can get syngensis process and generally others are forming 8 spores. Also, hypha is independent in each cell and feature is determined by the characteristics of septum.

Aspergillus is frequently found from nature like indoor air and easily discovered even from the food by-product or soil. This mold can corrupt the foods

- logy. Mosby International Limited : 367-380.
3. Jeung KJ. 2005. *Picture book to the microbiology III*. Institute of microbiology Seoul National University : 190-191.
 4. Won BW, Nam SY, Choi HS, et al. 1999. *Biological science*. Hyungseul Publish Co, Seoul : 565-582
 5. Lucas GM, Tucker P, Merz WG. 1999. Primary cutaneous *Aspergillus nidulans* infection associated with a Hickman catheter in a patient with neutropenia. *Clin Infect Dis* 29 : 1594-1596.
 6. Horre R, Schumacher G, Marklein G, et al. 2002. Case Report. Maxillary sinus infection due to *Emericella nidulans*. *Mycoses* 45 : 402-405.
 7. Mitchell RG, Chaplin AJ, Mackenzie DW. 1987. *Emericella nidulans* in a maxillary sinus fungal mass. *J Med Vet Mycol* 25 : 339-341.
 8. Dotis J, Panagopoulou P, Filioti J, et al. 2003. Femoral osteomyelitis due to *Aspergillus nidulans* in a patient with chronic granulomatous disease. *Infection* 31 : 121-124.
 9. Mizuki M, Chikuba K, Tanaka K. 1994. A case of chronic necrotizing pulmonary aspergillosis due to *Aspergillus nidulans*. *Mycopathologia* 128 : 75-9.
 10. Rosen-Wolff A, Koch A, Friedrich W, et al. 2004. Successful elimination of an invasive *Aspergillus nidulans* lung infection by voriconazole after failure of a combination of caspofungin and liposomal amphotericin B in a boy with chronic granulomatous disease. *Pediatr Inf Dis J* 23 : 584-586.
 11. Cabanes FJ, Monreal L, Majo N, et al. 2002. Guttural pouch mycosis by *Emericella nidulans* in a horse. *Rev Iberoam Micol* 19 : 208-211.
 12. Guillot J, Collobert C, Gueho E, et al. 1997. *Emericella nidulans* as an agent of guttural pouch mycosis in a horse. *J Med Vet Mycol* 35 : 433-435.
 13. Morris A, Schell WA, McDonagh D, et al. 1995. Pneumonia due to *Fonsecaea pedrosoi* and cerebral abscesses due to *Emericella nidulans* in a bone marrow transplant recipient. *Clin Infect Dis* 21 : 1346-1348.
 14. Kang HJ, Kim BH, Kim SJ, et al. 1993. *Veterinary Public Health*. Manundang, Seoul : 644-656.