

## Antibacterial Activity of Sophoraflavanone G Isolated from the Roots of *Sophora flavescens*

CHA, JEONG-DAN<sup>1</sup>, MI-RAN JEONG<sup>2</sup>, SEUNG-IL JEONG<sup>3</sup>, AND KYUNG-YEOL LEE<sup>1\*</sup>

<sup>1</sup>Institute of Oral Bioscience and Department of Oral Microbiology, School of Dentistry, Chonbuk National University, Jeonju 361-763, Korea

<sup>2</sup>Research Center of Bioactive Materials, Chonbuk National University, Jeonju 361-763, Korea

<sup>3</sup>Jeonju Biomaterials Institute, Jeonju 361-763, Korea

Received: December 3, 2006

Accepted: January 15, 2007

**Abstract** This study investigated the antibacterial activities of sophoraflavanone G from *Sophora flavescens* in combination with two antimicrobial agents against oral bacteria. The combined effect of sophoraflavanone G and the antimicrobial agents was evaluated using the checkerboard method to obtain a fractional inhibitory concentration (FIC) index. The sophoraflavanone G+ampicillin (AM) combination was found to have a synergistic effect against *S. mutans*, *S. sanguinis*, *S. sobrinus*, *S. gordonii*, *A. actinomycetemcomitans*, *F. nucleatum*, *P. intermedia*, and *P. gingivalis*, whereas the sophoraflavanone G+gentamicin (GM) combination had a synergistic effect against *S. sanguinis*, *S. criceti*, *S. anginosus*, *A. actinomycetemcomitans*, *F. nucleatum*, *P. intermedia*, and *P. gingivalis*. Neither combination exhibited any antagonistic interactions (FIC index > 4). In particular, the MICs/MBCs for all the bacteria were reduced to one-half ~ one-sixteenth as a result of the drug combinations. A synergistic interaction was also confirmed by time-kill studies for nine bacteria where the checkerboard suggested synergy. Thus, a strong bactericidal effect was exerted through the drug combinations, plus *in vitro* data suggested that sophoraflavanone G combined with other antibiotics may be microbiologically beneficial rather than antagonistic.

**Keywords:** *Sophora flavescens*, sophoraflavanone G, antibacterial activity, checkerboard method, time-kill method, synergic effect

Dental plaque is a film of microorganisms on the tooth surface that plays an important part in the development of caries and periodontal diseases [9, 26, 31]. The further accumulation of plaque around the gingival margin and subgingival region can lead to a shift in its microbial

composition from streptococcus-dominated to a larger number of *Actinomyces* spp. and increased number of capnophilic and obligatory anaerobic bacteria, such as *Porphyromonas gingivalis* [23, 24, 31, 37]. Several studies have already reported on the antibacterial effects of teeth-cleaning chewing sticks on cariogenic bacteria and periodontal pathogens, particularly bacteroides species, and their inhibitory action on dental plaque formation [1, 32, 38]. Frequently used antibacterial chemicals include povidone iodine products, chlorhexidine, and cetylpyridinium chloride, plus natural antibacterial substances have also attracted attention [2, 3, 13, 21, 25, 28, 29].

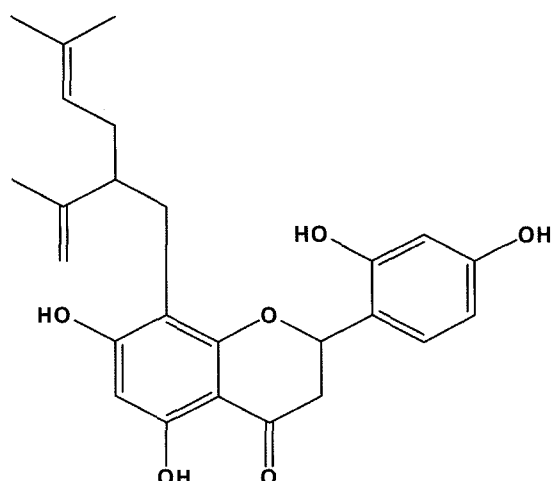
Sophorae Radix, the dried roots of *Sophora flavescens* AITON (Leguminosae), is an Oriental traditional medicine that has well-known antibacterial, antiviral, antiprotozoal, antiinflammatory, and antipyretic effects and is used as an insecticide and for the treatment of skin and mucosal ulcers, sores, diarrhea, gastrointestinal hemorrhages, arrhythmia, and eczema [6, 11, 16, 17, 19, 35, 39–41]. In addition to flavonoids with the regular prenyl side chains, *S. flavescens* also produces diverse flavanones with lavandulyl chains, irregular monoterpenoid groups, such as kurarinone and sophoraflavanone G [10, 12, 27, 30]. Recent pharmaceutical studies have shown that the lavandulyl side chain is essential for the antitumor activity and phospholipase-C $\gamma$ 1-inhibition activity of the flavonoids isolated from this plant [7, 22]. Sophoraflavanone G, one of the main lavandulylated flavanones isolated from the dried roots of *S. flavescens*, is known to possess antimalarial, antimicrobial, antiviral, and antioxidant activities, and to inhibit the production of nitric oxide and prostaglandin E $_2$  in lipopolysaccharide-treated RAW cells [4, 8, 15, 17, 34].

The roots of *S. flavescens* used in the present study were originally collected in October 2001 from Jinan, Jeonbuk Province, Korea. The authenticity of the plant was confirmed by Y. S. Ju, College of Oriental Medicine, Woosuk University, and

\*Corresponding author

Phone: 82-63-270-4023; Fax: 82-63-270-4037;

E-mail: kyleecnu@chonbuk.ac.kr



**Fig. 1.** Structure of 5,7,2',4'-tetrahydroxy-8-lavandulylflavanone (sophoraflavanone G) isolated from *Sophora flavescens*.

a voucher specimen (JS01-3) was deposited in the Herbarium of the Department of Bio and Medicinal Chemistry, College of Natural Science, Mokwon University. Sophoraflavanone G was isolated from the roots of *S. flavescens* based on the method described previously [27]. The sophoraflavanone

G isolated from the dried roots of *S. flavescens* was identified based on a comparison with the spectral data in reported literature [12, 27] (Fig. 1). Sophoraflavanone G:  $C_{25}H_{28}O_6$  ( $M_R$ : 424); pale yellow needle;  $[\alpha]_D^{25} -49^\circ$  ( $c$  1.0 in MeOH); UV (MeOH)  $\lambda_{max}$ =340, 293 nm.

The oral bacterial strains used in this study were *Streptococcus mutans* (ATCC 25175), *Streptococcus sanguinis* (ATCC 10556), *Streptococcus sobrinus* (ATCC 27607), *Streptococcus rattii* (KCTC: Korean collection for type cultures 3294), *Streptococcus criceti* (KCTC 3292), *Streptococcus anginosus* (ATCC 31412), *Streptococcus gordonii* (ATCC 10558), *Actinobacillus actinomycetemcomitans* (ATCC 43717), *Fusobacterium nucleatum* (ATCC 10953), *Prevotella intermedia* (ATCC 25611), and *Porphyromonas gingivalis* (ATCC 33277). A Brain-Heart Infusion broth supplemented with 1% yeast extract (Difco Laboratories, Detroit, MI, U.S.A.) was used for all the bacterial strains, except *P. intermedia* and *P. gingivalis*. The minimum inhibitory concentrations (MICs) were determined as the lowest test sample concentration that resulted in a complete inhibition of visible growth in the broth [14, 20, 33], whereas the minimum bactericidal concentrations (MBCs) were determined as the lowest concentration of sophoraflavanone G that killed 99.9% of the test bacteria when plating on the

**Table 1.** Checkerboard assay of sophoraflavanone G and ampicillin against oral bacteria.

Strains	Agent	MIC/MBC ( $\mu\text{g/ml}$ )		FIC <sup>b</sup> ( $\mu\text{g/ml}$ )	FICI <sup>2</sup>	Outcome
		Alone	Combination <sup>a</sup>			
<i>S. mutans</i> ATCC 25175	Sophoraflavanone G	3.2/3.2	0.4 /0.4	0.125	0.375	Synergistic
	Ampicillin	0.25/0.5	0.0625/0.125	0.25		
<i>S. sanguinis</i> ATCC 10556	Sophoraflavanone G	3.2/3.2	0.8/1.6	0.25	0.5	Synergistic
	Ampicillin	1/2	0.25/0.5	0.25		
<i>S. sobrinus</i> ATCC 27607	Sophoraflavanone G	3.2/3.2	0.8/1.6	0.25	0.31	Synergistic
	Ampicillin	0.25/0.5	0.0156/0.0312	0.063		
<i>S. rattii</i> KCTC 3294	Sophoraflavanone G	1.6/3.2	0.8/0.8	0.5	1	Additive
	Ampicillin	1/2	0.5/0.5	0.5		
<i>S. criceti</i> KCTC 3292	Sophoraflavanone G	1.6/3.2	0.4/0.8	0.25	0.75	Additive
	Ampicillin	0.5/1	0.25/0.25	0.5		
<i>S. anginosus</i> ATCC 31412	Sophoraflavanone G	3.2/6.4	0.8/0.8	0.25	0.75	Additive
	Ampicillin	1/1	0.5/0.5	0.5		
<i>S. gordonii</i> ATCC 10558	Sophoraflavanone G	0.8/0.8	0.1/0.2	0.125	0.375	Synergistic
	Ampicillin	1/2	0.025/0.5	0.25		
<i>A. actinomycetemcomitans</i> ATCC 43717	Sophoraflavanone G	3.2/3.2	0.2/0.4	0.063	0.31	Synergistic
	Ampicillin	32/64	8/16	0.25		
<i>F. nucleatum</i> ATCC 51190	Sophoraflavanone G	6.4/12.8	0.4/0.8	0.063	0.5	Synergistic
	Ampicillin	4/4	1/1	0.25		
<i>P. intermedia</i> ATCC 49049	Sophoraflavanone G	3.2/6.4	1.6/1.6	0.25	0.5	Synergistic
	Ampicillin	4/8	1/2	0.25		
<i>P. gingivalis</i> ATCC 33277	Sophoraflavanone G	0.2 /0.8	0.1/0.1	0.25	0.31	Synergistic
	Ampicillin	0.25/0.5	0.0156/0.0312	0.063		

<sup>a</sup>The checkerboard test was performed as previously described [5]. The MICs and MBCs of sophoraflavanone G with ampicillin against oral bacteria are indicated.

<sup>b</sup>The interaction was defined as synergistic if the FIC index was less than or equal to 0.5, additive if the FIC index was greater than 0.5 and less than or equal to 1.0, indifferent if the FIC index was greater than 1.0 and less than or equal to 2.0, and antagonistic if the FIC index was greater than 2.0 [5, 18].

**Table 2.** Checkerboard assay of sophoraflavanone G and gentamicin against oral bacteria.

Strains	Agent	MIC/MBC ( $\mu\text{g/ml}$ )		FIC <sup>b</sup> ( $\mu\text{g/ml}$ )	FICI <sup>2</sup>	Outcome
		Alone	Combination <sup>a</sup>			
<i>S. mutans</i> ATCC 25175	Sophoraflavanone G	3.2/3.2	0.8/0.8	0.25	0.75	Additive
	Gentamicin	8/16	4/4	0.5		
<i>S. sanguinis</i> ATCC 10556	Sophoraflavanone G	3.2/6.4	0.4/0.4	0.125	0.375	Synergistic
	Gentamicin	64/64	16/16	0.25		
<i>S. sobrinus</i> ATCC 27607	Sophoraflavanone G	3.2/3.2	1.6/3.2	0.5	0.75	Additive
	Gentamicin	4/8	1/2	0.25		
<i>S. ratti</i> KCTC 3294	Sophoraflavanone G	1.6/3.2	0.8/0.8	0.5	1	Additive
	Gentamicin	16/32	8/8	0.5		
<i>S. criceti</i> KCTC 3292	Sophoraflavanone G	1.6/3.2	0.4/0.4	0.25	0.281	Synergistic
	Gentamicin	8/16	0.25/0.25	0.031		
<i>S. anginosus</i> ATCC 31412	Sophoraflavanone G	3.2/6.4	0.8/0.8	0.25	0.281	Synergistic
	Gentamicin	32/32	1/2	0.031		
<i>S. gordonii</i> ATCC 10558	Sophoraflavanone G	0.8/0.8	0.1/0.1	0.125	0.63	Additive
	Gentamicin	32/32	16/16	0.5		
<i>A. actinomycetemcomitans</i> ATCC 43717	Sophoraflavanone G	3.2/3.2	0.8/0.8	0.25	0.5	Synergistic
	Gentamicin	4/8	1/1	0.25		
<i>F. nucleatum</i> ATCC 51190	Sophoraflavanone G	6.4/12.8	1.6/1.6	0.25	0.5	Synergistic
	Gentamicin	2/4	0.5/0.5	0.25		
<i>P. intermedia</i> ATCC 49049	Sophoraflavanone G	3.2/6.4	0.4/0.8	0.125	0.375	Synergistic
	Gentamicin	16/32	4/4	0.25		
<i>P. gingivalis</i> ATCC 33277	Sophoraflavanone G	0.2/0.8	0.05/0.1	0.25	0.31	Synergistic
	Gentamicin	256/512	16/32	0.063		

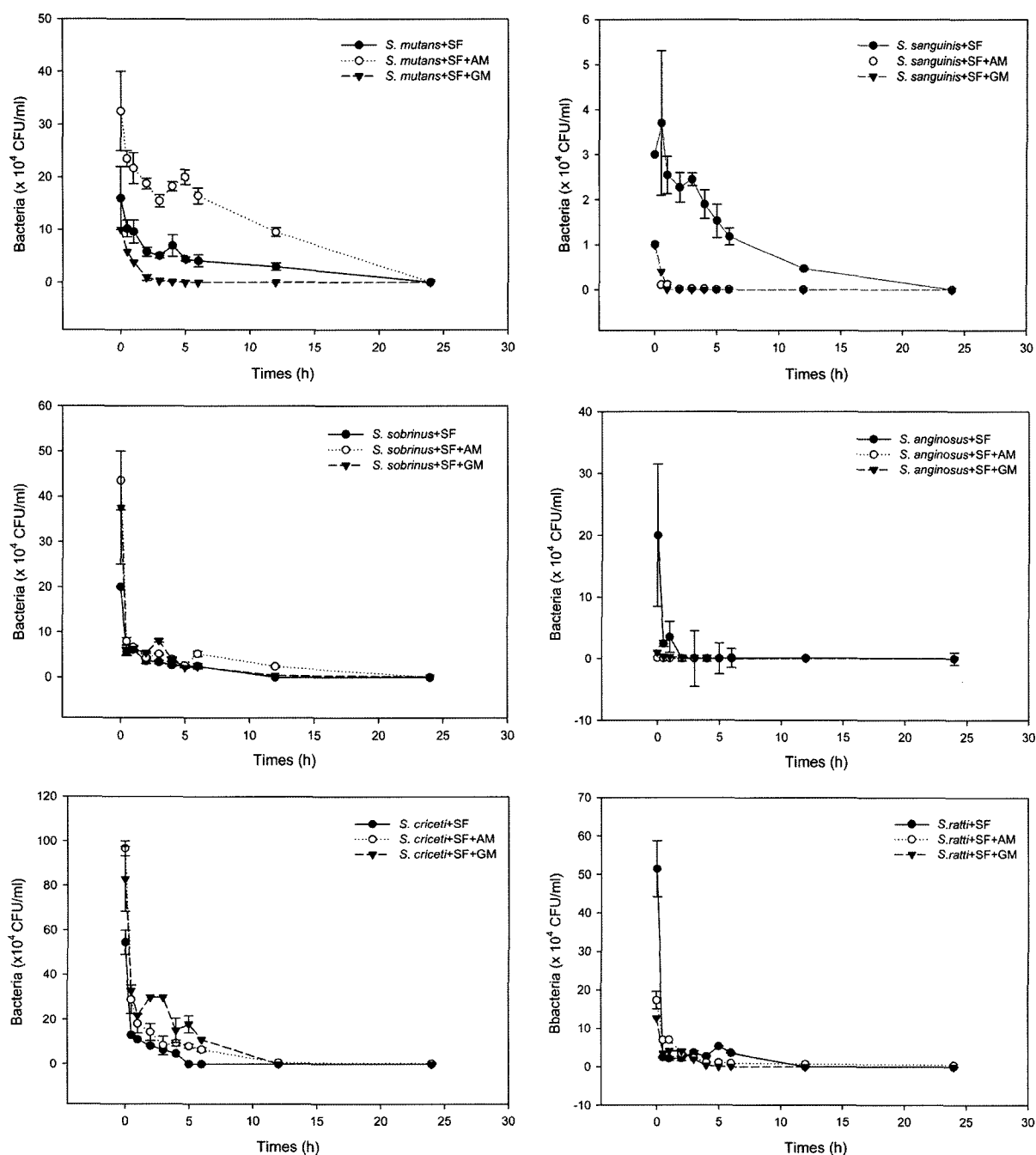
<sup>a</sup>The checkerboard test was performed as previously described [5]. The MICs and MBCs of sophoraflavanone G with gentamicin against oral bacteria are indicated.

<sup>b</sup>The interaction was defined as synergistic if the FIC index was less than or equal to 0.5, additive if the FIC index was greater than 0.5 and less than or equal to 1.0, indifferent if the FIC index was greater than 1.0 and less than or equal to 2.0, and antagonistic if the FIC index was greater than 2.0 [5, 18].

appropriate agar plate. In addition, the antibacterial effects of combining sophoraflavanone G with certain antibiotics were assessed using a checkerboard test, as previously described [5, 14]. The antimicrobial combinations assayed included sophoraflavanone G from *S. flavescens* plus ampicillin or gentamicin. As such, the fractional inhibitory concentration index (FICI) was determined as the sum of the FICs of each drug, which in turn was defined as the MIC of each drug when used in combination, divided by the MIC of the drug when used alone. The interaction was then defined as synergistic if the FIC index was less than or equal to 0.5, additive if the FIC index was greater than 0.5 and less than or equal to 1.0, indifferent if the FIC index was greater than 1.0 and less than or equal to 2.0, and antagonistic if the FIC index was greater than 2.0 [5, 18]. The MIC/MBC of sophoraflavanone G was found to be either 0.4/0.8 or 1.6/3.2  $\mu\text{g/ml}$ , the MIC/MBC of ampicillin either 0.25/0.5 or 32/64  $\mu\text{g/ml}$ , and the MIC/MBC of gentamicin either 0.5/1 or 256/512  $\mu\text{g/ml}$  (Tables 1 and 2). When combined with sophoraflavanone G, the MIC/MBC of ampicillin was reduced  $\geq 8$ -fold for *S. mutans*, *S. sanguinis*, *S. sobrinus*, *S. gordonii*, *A. actinomycetemcomitans*, *F. nucleatum*, *P. intermedia*, and *P. gingivalis*, reflecting a synergistic effect, as defined by

the  $\text{FICI} \leq 0.5$ . The addition of sophoraflavanone G led to a reduced single dilution for *S. ratti*, *S. criceti*, and *S. anginosus*, as defined by the  $\text{FICI} \leq 0.5-1$  (Table 1). The combination of gentamicin and sophoraflavanone G resulted in a decrease in the MIC/MBC for all the bacteria, where the MIC/MBC of 2-256/4-512  $\mu\text{g/ml}$  for gentamicin became 0.25-32/0.25-32  $\mu\text{g/ml}$ . The FICI classified the combination of sophoraflavanone G and gentamicin as additive for *S. mutans*, *S. sobrinus*, *S. ratti*, and *S. gordonii* and synergistic for *S. sanguinis*, *S. criceti*, *S. anginosus*, *A. actinomycetemcomitans*, *F. nucleatum*, *P. intermedia*, and *P. gingivalis* (Table 2).

Many attempts have already been made to eliminate *S. mutans* from the oral flora [9, 23]. For example, antibiotics, such as ampicillin, chlorhexidine, erythromycin, penicillin, tetracycline, and vancomycin, have been shown to be very effective in preventing dental caries [1, 2, 25, 29, 38], whereas various flavonoid derivatives from *S. flavescens*, such as quercetin, sophoraflavanone G, and kaempferol, have been found to exhibit antimicrobial and antimalaria activity [4, 7, 17]. In addition, phytoalexins, defensive compounds produced by plants against microbial infections, have been purified from *Sophora exigua* (Leguminosae) and their growth inhibitory effects on oral cariogenic bacteria determined *in vitro* [36].

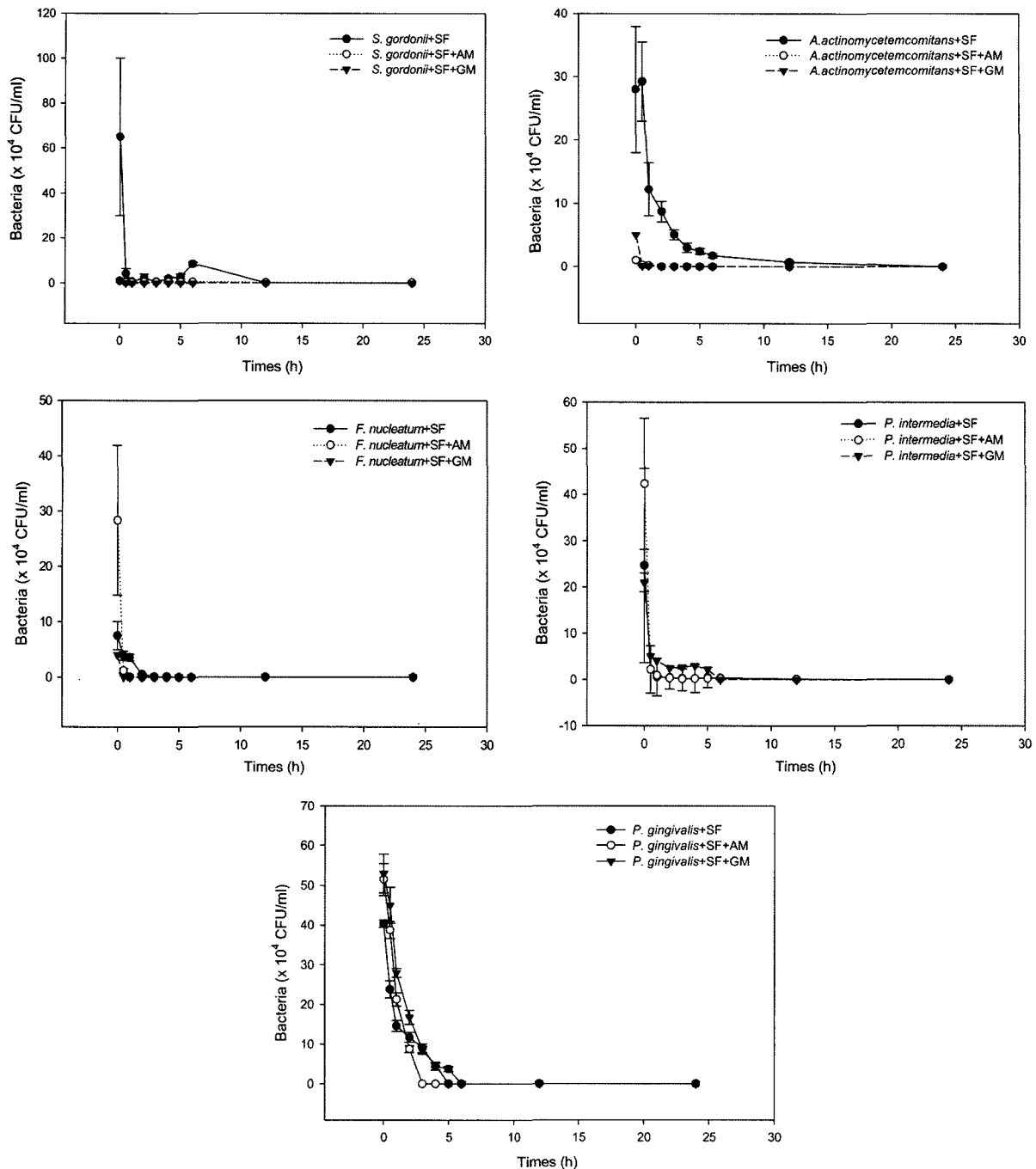


**Fig. 2.** Time-kill curves for MIC of sophoraflavanone G alone and when combined with MIC of ampicillin and gentamicin against *S. mutans*, *S. sanguinis*, *S. sobrinus*, *S. anginosus*, *S. criceti*, and *S. ratti*.

Bacteria were incubated with sophoraflavanone G (●), sophoraflavanone G+ampicillin (○), and sophoraflavanone G+gentamicin (▼) over time. Data points are the mean values±SEM of six experiments. CFU, colony-forming units.

The bactericidal activities of the drugs investigated in this study as regards oral bacteria were also evaluated using time-kill curves. As such, tubes containing sophoraflavanone G and oral bacteria were incubated at 37°C in an anaerobic chamber and viable counts performed 0, 0.5, 1, 2, 3, 4, 5, 6, 12, and 24 h after the addition of the antimicrobial agents. Agar plates were incubated for up to 48 h in an

anaerobic chamber at 37°C. The colony counts were performed in duplicate, and the means taken. Cultures of all the bacteria, with a cell density of  $10^5$ – $10^6$  CFU/ml, were exposed to the MIC of sophoraflavanone G alone and with ampicillin or gentamicin several times. When exposed to sophoraflavanone G alone, the rate of killing CFU/ml increased after 1 h, whereas the combination of



**Fig. 3.** Time-kill curves for MIC of sophoraflavanone G alone and when combined with MIC of ampicillin and gentamicin against *S. gordonii*, *A. actinomycetemcomitans*, *F. nucleatum*, *P. intermedia*, and *P. gingivalis*.

Bacteria were incubated with sophoraflavanone G (●), sophoraflavanone G+ampicillin (○), and sophoraflavanone G+gentamicin (▼) over time. Data points are the mean values±SEM of six experiments. CFU, colony-forming units.

sophoraflavanone G and ampicillin or gentamicin produced a more rapid rate of killing after 30 min (Figs. 2 and 3). Furthermore, the combination of sophoraflavanone G and ampicillin or gentamicin killed all the bacteria within 5 h, except for *S. mutans*. Thus, the drug combinations exhibited a strong bactericidal effect. The antibacterial agents

currently used to prevent dental caries include xylitol, tea extracts, essential oils, and antibiotics. Xylitol, a natural sweetener derived from xylose, is presently widely used in chewing gum, toothpaste, and mouthwash [1, 2, 25, 28, 29].

Accordingly, the present findings suggest that sophoraflavanone G fulfills the conditions required for

novel cariogenic bacteria and periodontal pathogens as a bactericide species drug, and may be useful for the treatment of oral bacteria infections. However, for medicinal purposes, the safety and toxicity of this compound still need to be addressed. The difference in susceptibility may also allow the formulation of products that will selectively kill or inhibit certain organisms, while having a minimal effect on the commensal microorganisms.

## Acknowledgments

The authors would like to thank Prof. Y. S. Ju, College of Oriental Medicine, Woosuk University, for confirming the plant authenticity. This paper was supported by research funds of Chonbuk National University in 2005.

## REFERENCES

- Al-Lafi, T. and H. Ababneh. 1995. The effect of the extract of the miswak (chewing sticks) used in Jordan and the Middle East on oral bacteria. *Int. Dent. J.* **45**: 218–222.
- Botelho, M. G. 2005. The antimicrobial activity of a dentin conditioner combined with antibacterial agents. *Oper. Dent.* **30**: 75–82.
- Cha, J. D., M. R. Jeong, S. C. Jeong, S. E. Moon, J. Y. Kim, B. S. Kil, and Y. H. Song. 2005. Chemical composition and antimicrobial activity of the essential oils of *Artemisia scoparia* and *A. capillaries*. *Planta Med.* **2**: 186–190.
- Chen, L., X. Cheng, W. Shi, Q. Lu, V. L. Go, D. Heber, and L. Ma. 2000. Inhibition of growth of *Streptococcus mutans*, methicillin-resistant *Staphylococcus aureus*, and vancomycin-resistant enterococci by kurarinone, a bioactive flavonoid isolated from *Sophora flavescens*. *J. Clin. Microbiol.* **43**: 3574–3575.
- Climo, M. W., R. L. Patron, and G. L. Archer. 1999. Combinations of vancomycin and beta-lactams are synergistic against staphylococci with reduced susceptibilities to vancomycin. *Antimicrob. Agents Chemother.* **43**: 1747–1753.
- Dai, S., M. Y. Chan, S. S. Lee, and C. W. Ogle. 1986. The antiarrhythmic effects of *Sophora flavescens* Ait. in rats and mice. *Am. J. Chin. Med.* **14**: 119–123.
- DeNaeyer, A., W. Vander Berghe, V. Pockock, S. Milligan, G. Haegeman, and D. De Keukeleire. 2004. Estrogenic and anticarcinogenic properties of kurarinone, a lavandulyl flavanone from the roots of *Sophora flavescens*. *J. Nat. Prod.* **67**: 1829–1832.
- Ding, P. L., Z. X. Liao, H. Huang, P. Zhou, and D. F. Chen. 2006. (+)-12alpha-Hydroxysophocarpine, a new quinolizidine alkaloid and related anti-HBV alkaloids from *Sophora flavescens*. *Bioorg. Med. Chem. Lett.* **16**: 1231–1235.
- Hamada, S. and H. D. Slade. 1980. Biology, immunology, and cariogenicity of *Streptococcus mutans*. *Microbiol. Rev.* **44**: 331–384.
- Hirobumi, Y., Z. Ping, and I. Kenichiro. 2002. Origin of two isoprenoid units in a lavandulyl moiety of sophoraflavanone G from *Sophora flavescens* cultured cells. *Phytochemistry* **60**: 263–267.
- Hsiang, C. Y., C. L. Hsieh, S. L. Wu, I. L. Lai, and T. Y. Ho. 2001. Inhibitory effect of anti-pyretic and anti-inflammatory herbs on herpes simplex virus replication. *Am. J. Chin. Med.* **29**: 459–467.
- Iinuma, M., T. Tanaka, M. Mizuno, Y. Shirataki, I. Yokoe, M. Komatsu, and F. A. Lang. 1990. Two flavanones in *Sophora leachiano* and some related structures. *Phytochemistry* **29**: 2667–2669.
- Jeon, S. M., H. J. Kim, K. B. Lee, J. W. Kim, and M. N. Kim. 2001. Antibacterial effect of the surface-modified biomedical polyurethane against *Staphylococcus aureus* and *Staphylococcus epidermidis*. *J. Microbiol. Biotechnol.* **11**: 259–265.
- Jung, H. J., K. S. Choi, and D. G. Lee. 2005. Synergistic killing effect of synthetic peptide P20 and cefotaxime on methicillin-resistant nosocomial isolates of *Staphylococcus aureus*. *J. Microbiol. Biotechnol.* **15**: 1039–1046.
- Jung, H. J., S. S. Kang, J. J. Woo, and J. S. Choi. 2005. A new lavandulylated flavonoid with free radical and ONOO-scavenging activities from *Sophora flavescens*. *Arch. Pharm. Res.* **28**: 1333–1336.
- Kang, T. H., S. J. Jeong, W. G. Ko, N. Y. Kim, B. H. Lee, M. Inagaki, T. Miyamoto, R. Higuchi, and Y. C. Kim. 2000. Cytotoxic lavandulyl flavanones from *Sophora flavescens*. *J. Nat. Prod.* **63**: 680–681.
- Kim, Y. C., H. S. Kim, Y. Wataya, D. H. Sohn, T. H. Kang, M. S. Kim, Y. M. Kim, G. M. Lee, J. D. Chang, and H. Park. 2004. Antimalarial activity of lavandulyl flavanones isolated from the roots of *Sophora flavescens*. *Biol. Pharm. Bull.* **27**: 748–750.
- Kitahara, T., Y. Aoyama, Y. Hirakata, S. Kamihira, S. Kohno, N. Ichikawa, M. Nakashima, H. Sasaki, and S. Higuchi. 2006. *In vitro* activity of lauric acid or myristylamine in combination with six antimicrobial agents against methicillin-resistant *Staphylococcus aureus* (MRSA). *Int. J. Antimicrob. Agents* **27**: 51–57.
- Kuroyanagi, M., T. Arakawa, Y. Hirayama, and T. Hayashi. 1999. Antibacterial and antiandrogen flavonoids from *Sophora flavescens*. *J. Nat. Prod.* **62**: 1595–1599.
- Lee, M. J., D. H. Bae, D. H. Lee, K. H. Jang, D. H. Oh, and S. D. Ha. 2006. Reduction of *Bacillus cereus* in cooked rice treated with sanitizers and disinfectants. *J. Microbiol. Biotechnol.* **16**: 639–642.
- Lee, S. H. and C. J. Kim. 1999. Antibacterial activity of antimycotic miconazole against methicillin-resistant *Staphylococcus aureus*. *J. Microbiol. Biotechnol.* **9**: 572–575.
- Lee, S. W., H. S. Lee, J. Y. Nam, O. E. Kwon, J. A. Baek, J. S. Chang, M. C. Rho, and Y. K. Kim. 2005. Kurarinone isolated from *Sophora flavescens* Ait inhibited MCP-1-induced chemotaxis. *J. Ethnopharmacol.* **97**: 515–519.
- Loesche, W. J. 1986. Role of *Streptococcus mutans* in human dental decay. *Microbiol. Rev.* **50**: 353–380.

24. Mayrand, D. and S. C. Holt. 1988. Biology of a saccharolytic black-pigmented *Bacteroides* species. *Microbiol. Rev.* **52**: 134–152.
25. Roldan, S., E. G. Winkel, D. Herrera, M. Sanz and A. J. Van Winkelhoff. 2003. The effects of a new mouthrinse containing chlorhexidine, cetylpyridinium chloride and zinc lactate on the microflora of oral halitosis patients: A dual-centre, double-blind placebo-controlled study. *J. Clin. Periodontol.* **30**: 427–434.
26. Rosan, B. and R. J. Lamont. 2000. Dental plaque formation. *Microbes Infect.* **2**: 1599–1607.
27. Ryu, S. Y., S. K. Kim, Z. No, and J. W. Ahn. 1996. A novel flavonoid from *Sophora flavescens*. *Planta Med.* **62**: 361–363.
28. Saeki, Y., Y. Ito, M. Shibata, Y. Sato, K. Okuda, and I. Takazoe. 1989. Antimicrobial action of natural substances on oral bacteria. *Bull. Tokyo Dent. Coll.* **30**: 129–135.
29. Shiraiishi, T. and Y. Nakagawa. 2002. Evaluation of the bactericidal activity of povidone-iodine and commercially available gargle preparations. *Dermatology* **204**: 37–41.
30. Shirataki, Y., I. Yokoe, M. Noguchi, T. Tomimori, and M. Komatsu. 1988. Studies on the constituents of *Sophora* species XXII. Constituents of the root of *Sophora moorcroftii* Benth. Ex Baker(1). *Chem. Pharm. Bull.* **36**: 2220–2225.
31. Slots, J. and M. A. Listgarten. 1988. *Bacteroides gingivalis*, *Bacteroides intermedius* and *Actinobacillus actinomycetemcomitans* in human periodontal diseases. *J. Clin. Periodontol.* **15**: 85–93.
32. Sote, E. O. and M. Wilson. 1995. *In-vitro* antibacterial effects of extracts of Nigerian tooth-cleaning sticks on periodontopathic bacteria. *Afr. Dent. J.* **9**: 15–19.
33. Sung, W. S., H. J. Jung, I. S. Lee, H. S. Kim, and D. G. Lee. 2006. Antimicrobial effect of furaneol against human pathogenic bacteria and fungi. *J. Microbiol. Biotechnol.* **16**: 349–354.
34. Tashiro, M., F. Suzuki, Y. Shirataki, Y. Yokote, K. Akahane, N. Motohashi, M. Ishihara, Y. Jiang, and H. Sakagami. 2002. Effects of prenylflavanones from *Sophora* species on growth and activation of mouse macrophage-like cell line. *Anticancer Res.* **22**: 53–58.
35. Tsai, J. C., S. Tsai, and W. C. Chang. 2004. Effect of ethanol extracts of three Chinese medicinal plants with anti-diarrheal properties on ion transport of the rat intestinal epithelia. *J. Pharmacol. Sci.* **94**: 60–66.
36. Tsuchiya, H., M. Sato, M. Iinuma, J. Yokoyama, M. Ohyama, T. Tanaka, I. Takase, and I. Namikawa. 1994. Inhibition of the growth of cariogenic bacteria *in vitro* by plant flavanones. *Experientia* **50**: 846–849.
37. van Winkelhoff, A. J., T. J. M. van Steenberghe, and J. DeGraaff. 1988. The role of black-pigmented *Bacteroides* in human oral infections. *J. Clin. Periodontol.* **15**: 145–155.
38. Wolinsky, L. E. and E. O. Sote. 1983. Inhibiting effect of aqueous extracts of eight Nigerian chewing sticks on bacterial properties favouring plaque formation. *Caries Res.* **17**: 253–257.
39. Xiang, Q., M. Q. Tan, and Y. H. Huang. 2002. Anti-leukemia effect of *Sophora flavescens* combined with the low molecular weight natural tumor suppressor of the human fetal liver and its mechanism. *Hunan Yi Ke Da Xue Xue Bao* **27**: 108–110.
40. Youn, H. J., J. Lakritz, G. E. Rottinghaus, H. S. Seo, D. Y. Kim, M. H. Cho, and A. E. Marsh. 2004. Anti-protozoal efficacy of high performance liquid chromatography fractions of *Torilis japonica* and *Sophora flavescens* extracts on *Neospora caninum* and *Toxoplasma gondii*. *Vet. Parasitol.* **125**: 409–414.
41. Zhang, Y., H. Zhu, G. Ye, C. Huang, Y. Yang, R. Chen, Y. Yu, and X. Cui. 2006. Antiviral effects of sophoridine against coxsackievirus B3 and its pharmacokinetics in rats. *Life Sci.* **78**: 1998–2005.