

Effect of Herb Distillate on Hepatic Xanthine Oxidase Activity and Serum Lipid Profiles in Carbon Tetrachloride-Administered Rats

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Abstract

In order to evaluate the hepatoprotective effect of an herb distillate, ie., a mixture of 28 traditional Korean herbs, germanium, tormarine and Gijangsoo (Gijang water), CCl4 was intraperitoneally administered to rats before or after supplementation of the diluted herb distillate (HD) for 2 weeks. Then hepatic xanthine oxidase activity and serum lipid profiles were determined. The experimental groups had higher feed intake than the normal control (NC), but had lower weight gain. Water intake and the amount of feces were not significantly different, but urine was excreted in lower amounts in all the experimental groups compared to the NC. Liver weights in the HD-supplemented groups were lower than that of the distilled water-supplemented groups (DW-groups) after CCl4-administration. Serum ALT activities in all the experimental groups were higher than that of the NC-group. However, the increasing activity of serum ALT in the HD-supplemented groups (HD-groups) was lower than that of the DW-groups. Total serum and LDL-cholesterol levels were higher in all the CCl4-administered groups than in the NC-groups, and serum HDL-cholesterol levels were lower in all the experimental groups compared with the NC-groups. Meanwhile, the increasing rate of total serum and LDL-cholesterol levels and the decreasing rate of HDL-cholesterol in the HD-groups were lower than that of the DW-groups. But, levels of serum TG were similar among all the experimental groups. The activities of hepatic xanthine oxidase (XOD) type O of the CCl4-administered rats showed a significant increase in and an increasing rate of XOD in the HD-groups, which was lower than that of the DW-groups. On the other hand, GST activities in all the experimental groups were significantly decreased, and the decreasing rate was lower in the HD-groups than in the DW-groups. The hepatic contents of GSH and LPO in all the rats were not changed by CCl₄ administration. These results suggest that the decreased liver damage in the HD-supplemented groups was due to the inhibition of XOD-type O activity by constituents of HD, as well as by a prevention/inhibition of serum lipid profile changes in CCl4-treated rats. However, further detailed studies are needed to support this hypothesis.

Key words: herb distillate, carbon tetrachloride, xanthine oxidase activity, hepatic lipid profiles

INTRODUCTION

It is well-known that water serves as the universal solvent in which a variety of solutes are dissolved. Water in the body is related to nutrient absorption, transport, and metabolism, as well as excretion of waste products, maintenance of osmolarity, pH, BP and temperature control. Water intake is strongly influenced by habit or health. Healthy people have a remarkable ability to maintain the tonicity of their body fluid. Water balance is regulated by hypothalamic ADH and the kidneys by the osmolarity of body fluid. But, diseases, such as diabetes mellitus, and renal disease alter water balance (1). Water quality is critical for maintaining human health. However, with worldwide industrial development leading to water pollution, water quality has become too low

and insufficient for maintaining health.

Germanium, tournaline, and Gijangsoo (made from yellow earth: loess) have been known to irradiate in the far-infrared portion of the light spectrum, which activates anti-oxidant activity in the herbal mixture by affecting heat transport (2). Also, bioactive water made by treatment of far-infrared light produced from ceramic stone has a protective effect on alcohol-induced hepatic injury in pigs (3). It is also well-known that water is converted from higher to relatively lower molecular mass clusters by ceramics, jade, yellow soil or granite porphyry (mackban-stone), which emit far infrared rays (4-6). However, low molecular mass water clusters have only been studied a little in relation to their effect on human health.

The vapor of specific medicines during distillation

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results in what is referred to as herb distillate. Medicines contain various volatiles which have been researched for use in promoting health, treatment of disease, and for beauty care products (7-9). In the present study, we examined the detoxification effect of an herb distillate made from 28 traditional Korean herbs, as well as ceramic mixtures of germanium, tourmaline, and Gijangsoo, which is traditionally used in Korean medicine for preventing hepatic damage and cancer (4-6), on CCL₄induced toxicity in rats. CCl4 was intraperitoneally injected into rats before or after supplementation of a diluted herb distillate for 2 weeks. Body weight, feed intake, water intake, organ weight, serum lipid profiles, content of hepatic glutathione, lipid peroxide content, and xanthine oxidase, glutathione S-transferase and serum alanine aminotransferase activity were determined to investigate liver damage in the CCl4-administered rats.

MATERIALS AND METHODS

Materials

Various herbs (Table 1) were purchased from a commercial firm in Daegu, Korea and stored at 4°C until used. Tourmaline, germanium (38 microliters per liter) was obtained from the Uncheon mine in Gyeongsansi, Korea. Gijangsoo (Gijang water) was obtained from the Haksan mine, Gyeongsansi, Korea.

Preparation of the herb distillate

One hundred g of herbs were added to 4 L of distilled water and distilled at 120°C for 5 hours, as shown in Table 1. The diluted herb distillate (HD) was prepared by adding herb distillate to a final concentration of 2% into ozone-treated (0.5 microliters per liter) water.

Animals, experimental plots and preparation of diets

Male Sprague-Dawley rats with a mean mass of 195 ±5 g were purchased from Hyochang Science (Daegu, Korea). The experimental animals were divided into five groups (10 rats/group), as shown in Fig. 1. The following notafious were indicated: Normal (designated NC), distilled water (DW) supplement after CCl₄ injection (CCl₄-DW), DW supplement for 2 weeks and CCl₄ injection before 2 days of sacrifice (DW-CCl₄), diluted herb distillate (HD) supplement after CCl₄ injection (CCl₄-HD), and HD supplement for 2 weeks and CCl₄ injection before 2 days of sacrifice (HD-CCl₄). All animals were fed a prepared basal diet according to the AIN-76A diet (Teklad, Indiana, USA), as shown in Table 2. The ratio of carbohydrate, protein and lipid content in the basal diet was adjusted to 60:20:15.

Rats were individually housed in stainless steel cages with wire bottoms in a room maintained at $20\pm2^{\circ}\mathrm{C}$ and

Table 1. Materials for preparation of herb distillate

No Ko	rean name	e Scientific name	Ratio (%)
1 Dan	ggui	Angelica gigas	2.2
2 Gan	ncho	Glycyrrhizae glabra	2.2
3 Sasa	am	Codonopis lanceolata	2.2
4 Sam	yak	Disocorea japonica	2.2
5 Hwa	angkee	Astragalus membranaceus	2.2
6 Suk	jihwayang	; Rehmanniae radix	1.0
7 Has	uo	Pleuropterus multflorus	2.5
8 Gun	gang	Zingiber officinale	2.5
9 Yuk	gae	Cinnamomum loureirii	2.5
10 Hw	acho	Zanthoxylum piperitum	2.5
11 Dan		Salvia miltiorrhiza	2.5
12 Bon		Impatiens balsamina	3.0
13 Hon	ıgwha	Carthamus tinctorius	3.0
14 Dok	hwal	Aralia contientalis	3.0
15 Mal	_	Ephedra sinica	3.0
16 Gal		Pueraria thunbergiana radix	4.5
17 Wo	osul	Achyranthes japonica radi x	3.0
18 Mol	chwa	Chaenomeles sinensis	3.0
19 Hae	_	Pinusdensiflora neeles	3.0
20 Jern	_	Boehmera niver radi x	3.0
21 Suk		Artemisia princeps var. orientalis	4.0
22 You	ıgsullan	Agave americana	2.0
23 Hwa	anggum	Scutellaria baicalensis	2.5
	angyeun	Coptis chinensis	2.5
25 Hwa	angback	Phellodendron amurense	2.5
26 Heta	gae	Hovenia dulcis	1.5
27 Ork		Corn powder	12.0
	manium	Germanium	5.0
29 Tou	rmaline	Tourmaline	7.0
30 Gija	ngsoo	Gijang water made from yellow earth	8.0

Experimental groups

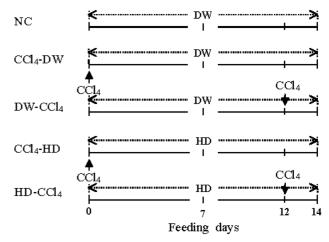


Fig. 1. Experimental design for control and treatment groups. NC, normal control; DW, distilled water; HD, herb distillate.

 $60\pm5\%$ relative humidity. The room was exposed to alternative 12-hours of light and dark. Rats were fed an animal diet (Purina Co., Seoul, Korea) during an initial 1-week acclimation period. All rats were allowed to eat their respective diets and drink their water freely.

Table 2. Compositions of basic diets (g/kg diet)

Ingredients	Content
Corn starch	150
Casein	200
Corn oil	50
Sucrose	500
Cellulose	50
AIN mineral mixture ¹⁾	35
AIN vitamin mixture ²⁾	10
DL-Methionine	3
Choline bitartrate	2
Total	1,000

AIN mineral mixture (g/kg): calcium lactate 620.0, sodium chloride 74.0, potassium phosphate di-basic 220.0, potassium sulfate 52.0, magnesium oxide 23.0, manganous carbonate 3.3, ferric citrate 6.0, zinc carbonate 1.0, cupric carbonate 0.2, potassium iodate 0.01, sodium selenite 0.01, chromium potassium sulfate 0.5, finely powdered to make 1,000 g.

⁴AIN vitamin mixture (mg/kg): thiamin-HCl, 600; riboflavin, 600; pyridoxine-HCl, 700; nicotinic acid, 3,000; D-calcium pantothenate, 1,600; folic acid, 200; D-biotin, 20; vitamin B12, 2.5; vitamin A, 400,000 IU; vitamin D3, 100,000 IU; vitamin E, 7,500 IU; vitamin K 75, finely powdered to make 1,000 g.

Induction of hepatic damage

Fifty percent CCl₄ (Sigma Chem. Co., St. Louis, MO, USA) mixed with olive oil (1:1; v/v) was administered intraperitoneally at 0.1 mL of 50% the CCl₄ per 100 g body weight once daily for 2 days to CCl₄-DW and CCl₄-HD groups before supplementation of HD or DW. The DW-CCl₄ and HD-CCl₄ groups were administered the CCl₄ solution 1 and 2 days before sacrifice; The NC group was administered olive oil.

Weight gain, feed and water intake, and feed efficiency ratio

Body weight, feed and water intake were measured daily. The feed efficiency ratio (FER) was calculated by dividing the weight gains into the feed intake weekly.

Preparation of analytical samples

After the treatment of the experimental groups for 2 weeks, rats were fasted for 24 hours and anesthetized with ethylether; and blood was collected from the abdominal aorta. The collected blood was centrifuged at 3,000 rpm ($\times g$) for 10 minutes at room temperature and the serum separated was kept at -70°C. The internal organs, such as the liver were exhaustively perfused with cold physiological saline solution though the portal vein and quickly removed. The liver was homogenized with 4 volumes of 0.25 M sucrose, centrifuged at $1,000 \times g$ for 10 minutes and the supernatant was recentrifuged at $10,000 \times g$ for 20 minutes. The pellet was resuspended

with 0.25 M sucrose for use as a mitochondrial fraction and the supernatant was used as a post-mitochondrial fraction (PMF). The collected urine and feces were measured everyday.

Analysis of blood and urine sugar, and serum lipids

The contents of triglyceride, total cholesterol and HDL-cholesterol in the serum were measured using kit reagents from AM 157S-K, Asan Pharm Co., Seoul, Korea; AM 202-K, Asan Pharm Co., Seoul, Korea, and AM 203-K, Asan Pharm Co., Seoul, Korea; respectively. The content of LDL-cholesterol was calculated using the method of Friedewald et al. (10).

Measurement of hepatic xanthine oxidase (XOD), glutathione S-transferase (GST) activities and serum alanine aminotransferase (ALT) activities

Xanthine oxidase (11) and glutathione S-transferase (12) activities in the liver were determined and expressed as uric acid nmole per minute per mg of PMF protein and thioether nmole per minute per mg of PMF protein, respectively. Alanine aminotransferase (ALT) activity in the serum was determined by using a kit reagent (Asan Pharm. Co., Seoul, Korea) and expressed as a Karmen unit.

Determination of glutathione, lipid peroxide and protein content

The hepatic glutathione content and lipid peroxide content were determined by the method of Ellman (13) and Satho (14), respectively. The protein content was determined by the Lowry (15) method with bovine serum albumin as a standard.

Statistical analysis

All experiments were carried out with 10 replicates and the means ± standard errors are reported. The means of the main effects were separated by Duncan's multiple range test using the SPSS (Statistical Package for Social Sciences, SPSS Inc., Chicago, IL, USA).

RESULTS AND DISCUSSION

Weight gain, feed intake, feed efficiency ratio and water intake

The results of body weight, weight gain, feed intake, feed efficiency ratio, water intake, excreted amounts of feces and urine are shown in Table 3. There was no significantly different body weight (mean 221 g) among the experimental groups. Weight gains of the CCl₄-DW and CCl₄-HD groups compared with the NC group were 42.7% and 47.8%, but those of the DW-CCl₄ and HD-CCl₄ groups were 53.9% and 48.5%, respectively.

The feed intake in all the experimental groups were

Table 3. Weight gain, feed intakes, FER, water intakes, amounts of feces and urine on rats supplemented with diluted herb distillate for 2 weeks after or before carbon tetrachloride administration

Measurements	$NC^{1)}$	CCl_4 - $DW^{2)}$	DW-CCl ₄ ³⁾	CCl ₄ -HD ⁴⁾	HD-CCl ₄ ⁵⁾
Initial body weight (g)	$207.9 \pm 6.23^{67)}$	224.3 ± 8.21^{a}	$219.0 \pm 8.45^{\circ}$	$226.5 \pm 5.37^{\circ}$	229.4±4.63 ^a
Final body weight (g)	$266.5 \pm 8.50^{\circ}$	249.3 ± 7.00^{b}	$250.6 \pm 4.90^{\mathrm{b}}$	254.5 ± 5.00^{ab}	257.8 ± 6.20^{ab}
Weight gain (g/week)	$29.3 \pm 3.80^{\circ}$	$12.5 \pm 8.90^{\rm b}$	15.8 ± 6.60^{b}	$14.0 \pm 4.10^{\circ}$	$14.2 \pm 4.50^{\circ}$
Feed intakes (g/week)	$195.3 \pm 8.00^{\circ}$	$210.6 \pm 4.50^{\mathrm{a}}$	$210.0 \pm 4.60^{\text{a}}$	$203.8 \pm 6.00^{\text{ab}}$	$203.1 \pm 9.70^{\mathrm{ab}}$
FER ⁶⁾	0.15 ± 0.02	0.06 ± 0.02^{b}	$0.08 \pm 0.03^{\circ}$	0.07 ± 0.02^{b}	0.04 ± 0.01^{b}
Water intakes (mL/week)	$215.3 \pm 12.3^{\circ}$	$253.5 \pm 14.4^{\circ}$	229.8 ± 27.0^{ab}	259.5 ± 15.9 ,*	$255.8 \pm 14.9^{\circ}$
Feces amounts (g/day)	$2.4 \pm 0.03^{\circ}$	$2.1 \pm 0.05^{\circ}$	$1.9 \pm 0.04^{\circ}$	$2.1 \pm 0.04^{\circ}$	$1.9 \pm 0.06^{\circ}$
Urine amounts (mL/day)	8.7 ± 0.14^{a}	8.0 ± 0.12^{c}	7.8 ± 0.10^{a}	$8.3\pm0.13^{\circ}$	$8.0 \pm 0.08^{\circ}$

¹⁻⁵⁾See Fig. 1. ⁶⁾FER (feed efficiency ratio): weight gain/feed intakes.

significantly higher than that of the NC group, but the feed efficiency ratios were significantly lower than that of the NC group, showing no significant difference between each experimental group. Water intake in all the groups were similar, but the amount of feces in the DW-CCl₄ and HD-CCl₄ groups were significantly lower compared with the NC, CCl₄-DW and CCl₄-HD groups. The amount of urine in all the other groups were significantly lower than that of the NC group.

Organ weights

Table 4 shows the weight of organs and their relative % of total body weight. The liver and kidney weight per body weight in all the experimental groups increased to a greater extent compared with the NC group. Also, the CCl₄-HD and HD-CCl₄ groups were lower than those of the CCl₄-DW and DW-CCl₄ groups. While the weight of the spleen, heart and testicles to the body weight of the CCl₄-DW and DW-CCl₄ groups were similar to that of the NC group, the CCl₄-HD and HD-CCl₄ groups showed a lower weight of organs than the NC and CCl₄-DW groups.

The kidney weights to body weight (%) in the CCl₄-HD and HD-CCl₄ groups were lower than those of the CCl₄-DW and DW-CCl₄ groups. Unlike the kidney weights, the weight of the spleen, heart and testicles to

body weight (%) of the CCl₄-DW and DW-CCl₄ groups were similar to that of the NC group, and the weight (%) of the CCl₄-HD and HD-CCl₄ groups were lower than those of the NC and CCl₄-DW groups.

The liver weight to body weight (%) in the CCl₄-HD and HD-CCl₄ groups was lower than those of the CCl₄-DW and DW-CCl₄ groups, which is likely to suggest that the supplemented HD caused a better-damaged liver by CCl₄. Also, the finding that the liver weight and serum ALT activity in the HD-supplemented groups was lower compared to the distilled water-supplemented groups means that HD had a protective effect on liver damage.

Serum lipid content

The levels of triglyceride (TG), total cholesterol, HDL-cholesterol and LDL-cholesterol in serum are shown in Table 5. The level of serum triglyceride (TG) in all the experimental groups was lower compared with the NC group (Table 5). The HD-supplemented groups were similar in total cholesterol level to the NC group, but lower than the supplemented distilled water groups. On the other hand, HDL-cholesterol content in all the experimental groups was significantly decreased compared to that of the NC group. The decreasing degree was the lowest in the CCl₄-HD group and highest in

Table 4. Weight of organs and serum alt activity on rats supplemented with diluted herb distillate for 2 weeks before or after carbon tetrachloride administration

Internal organs	$NC^{1)}$	$CCl_4\text{-}DW^{2)}$	DW-CCl ₄ ³⁾	$CCl_4\text{-HD}^{4)}$	HD-CCl ₄ ⁵⁾
Body wt (g)	$266.5 \pm 8.50^{a7)}$	249.3 ± 7.00^{b}	$250.6 \pm 4.90^{\mathrm{ab}}$	$254.5 \pm 5.00^{\mathrm{ab}}$	$258.0 \pm 7.20^{\mathrm{ab}}$
Liver wt (%)	$3.51 \pm 0.18^{\circ}$	3.77 ± 0.17^{a}	3.76 ± 0.28^{a}	3.66 ± 0.20^{a}	$3.64 \pm 0.21^{\circ}$
Kidney wt (%)	$1.21\!\pm\!0.10^{\rm a}$	$1.32 \pm 0.12^{\rm a}$	$1.35 \pm 0.10^{\rm a}$	1.29 ± 0.08^a	1.26 ± 0.08^{a}
Spleen wt (%)	0.34 ± 0.04^a	0.35 ± 0.06^{a}	0.34 ± 0.05^a	0.29 ± 0.08^a	0.28 ± 0.06^{a}
Heart wt (%)	0.64 ± 0.05^{a}	0.62 ± 0.02^{a}	0.60 ± 0.04^{a}	0.59 ± 0.04^{a}	$0.54 \pm 0.06^{\circ}$
Testicle wt (%)	1.31 ± 0.14^{a}	1.25 ± 0.12^{a}	$1.28\!\pm\!0.08^a$	$1.22 \pm 0.10^{\rm a}$	1.24 ± 0.12^{a}
Serum ALT ⁶⁾	$20.2\!\pm\!2.6^{\rm d}$	65.5 ± 9.6^{b}	$86.7 \pm 10.2^{\circ}$	$38.7 \pm 6.5^{\circ}$	$69.8\!\pm\!10.5^{ ext{ab}}$

¹⁻⁵⁾See Fig. 1. ⁶⁾Karmen unit/mL of serum.

 $^{^{7}}$ Values are means \pm SE of 10 rats, different superscripts within a row (a \sim c) indicate significant difference (p<0.05).

 $^{^{7)}}$ Values are means \pm SE of 10 rats, different superscripts within a row (a \sim b) indicate significant difference (p<0.05).

Table 5. Lipid profiles and atherogenic index in serum on rats supplemented with diluted herb distillate for 2 weeks after or before carbone tetrachloride administration (mg/dL)

Groups ¹⁾	Triglyceride	Total cholesterol	HDL-cholesterol	LDL-cholesterol ²⁾	Atherogenic index ³⁾
NC	105.4 ± 14.4^{a4}	$104.9 \pm 9.5^{\mathrm{b}}$	62.3 ± 2.3^{a}	$21.5 \pm 8.2^{\circ}$	0.68 ± 0.10^{e}
CCl_4 -DW	86.3 ± 19.2^{ab}	$114.1\pm12.1^{\text{ab}}$	30.5 ± 2.9^{d}	66.3 ± 9.2^{ab}	2.74 ± 0.08^{a}
$DW-CCI_4$	79.6 ± 14.0^{ab}	$127.6 \pm 11.1^{\circ}$	$44.0 \pm 2.5^{\circ}$	67.7 ± 10.4^{a}	$1.90\pm0.14^{\circ}$
CCl_4 -HD	80.7 ± 18.6^{ab}	$104.8 \pm 9.5^{\circ}$	$49.8\pm1.5^{\circ}$	$38.9 \pm 6.0^{\circ}$	$1.10 \pm 0.10^{\text{d}}$
$\mathrm{HD}\text{-}\mathrm{CCl}_4$	71.8 ± 13.8^{b}	$105.2\pm14.7^{ m ab}$	$42.3 \pm 2.8^{\circ}$	$48.5 \pm 10.2^{\mathrm{ab}}$	$1.49 \pm 0.12^{\circ}$

 $[\]overline{}^{1)}$ See Fig. 1.

the CCl₄-DW group. Also, LDL-cholesterol levels in all the experimental groups were significantly increased compared with that of the NC group. But the CCl₄-HD group was lower than the CCl₄-DW group. The atherogenic index, showing the same tendency to the LDL-cholesterol, was lowest in the CCl₄-HD group.

Serum TG is transported by very low density lipoprotein (VLDL), derived from the liver, to the extrahepatic tissues (16,17). It is well-known that the level of VLDL in serum is a parameter on the release of TG from the liver (18-20). In abnormal conditions such as liver damage, TG can not be released from the liver and accumulate in the liver due to the inhibition of VLDL apolipoprotein synthesis (18,20-22). Under these conditions, levels of serum TG were decreased and hepatic TG was increased (23,24). On the basis of the above results, in concert with the data in the present paper, we assume that the cause of the decreased serum TG level in the CCl₄-treated rats may be related to the inhibition of VLDL apolipoprotein synthesis in the hepatic tissue, and that HD may not restore the decreased serum TG level in the CCl₄-treated rats.

It is widely accepted that the liver mainly scavenged serum LDL derived from VLDL (25) and that the level of serum LDL is increased in hepatic-damaged tissue (20,24,26,27). In addition, activities of hepatic LCAT and hepatic HDL apolipoprotein synthesis are inhibited in the damaged liver (22,24,26,27). In general, herb dis-

tillate contains biologically active small molecules such as sesquiterpenoids, triterpennoids and acetylene derivatives (28). It has been shown that decursin in danggui modulates drug-metabolizing enzyme activities (29,30), and salvianic acid in dansam exhibits a protective effect on carbon tetrachloride-induced liver damage in rats (31). Therefore, those active components, which are presumably present in the herb distillate used in this study, may contribute to preventing CCL₄-induced liver damage. These results suggested that the herb distillate might be able to regulate levels of serum LDL- and HDL-cholesterol under conditions where liver damage

Contents of glutathione and lipid peroxide, and enzyme activities

The levels of hepatic glutathione and lipid peroxide, and enzyme activities are shown in Table 6. The content of glutathione was unchanged in all the experimental groups by CCl₄ administration, but the content of hepatic LPO was higher compared with the NC group and the content of LPO by supplementing HD decreased more greatly compared with the distilled water-supplemented groups. Total XOD activity was significantly changed in only the CCl₄-HD group. However, the activity of XOD type O was higher in all the experimental groups than the NC group. The HD-supplemented groups was lower than the distilled water-supplemented groups. On the other hand, the GST activity of CCl₄-DW, DW-CCl₄,

Table 6. Contents of hepatic glutathione (GSH) and lipid peroxide (LPO), and activities of xanthine oxidase (XOD) and glutathione s-transferase (GST) on rats supplemented with diluted herb distillate for 2 weeks after or before carbon tetrachloride administration

Groups ¹⁾	GSH	LPO	XOD (uri	GST (CDG ²⁾ nmol/		
Groups	(Jmol/g)	(MDA nmol/g)	Total	O-type	O/T (%)	mg protein/min)
NC	4.80 ± 0.26^{a3}	$14.01 \pm 0.34^{\circ}$	4.14 ± 0.08^{d}	1.03 ± 0.02^{d}	24.88 ± 2.81 a	$370.8 \pm 32.3^{\circ}$
CCl_4 -DW	5.09 ± 0.24^{a}	$22.15 \pm 3.26^{\mathrm{b}}$	$4.31 \pm 0.04^{\rm c}$	$1.24\pm0.02^{\mathrm{b}}$	$28.77 \pm 1.60^{\mathrm{a}}$	$298.5 \pm 41.0^{\mathrm{ab}}$
$DW-CCl_4$	5.28 ± 0.23^{a}	$35.66 \pm 4.91^{\circ}$	$5.15\pm0.07^{ ext{a}}$	$1.35 \pm 0.03^{\circ}$	26.21 ± 2.89^{a}	$247.8 \pm 36.6^{\circ}$
CCl ₄ -HD	5.27 ± 0.22^{a}	$17.41 \pm 2.26^{\text{bc}}$	$3.86 \pm 0.05^{\circ}$	$1.08\pm0.02^{\circ}$	27.98 ± 2.29^{a}	309.6 ± 45.5^{80}
HD-CCl ₄	5.10 ± 0.23^{a}	$27.17 \pm 4.06^{ ext{ab}}$	4.45 ± 0.08^{b}	$1.20\pm0.02^{\circ}$	26.97 ± 4.21^{a}	$278.2 \pm 29.0^{\rm b}$

¹⁾See Fig. 1. ²⁾CDG: conjugated dinitrophenol glutathione.

 $^{^{20}}$ LDL-cholesterol = Total cholesterol - HDL-cholesterol - (TG/5).

 $^{^{3)}}$ Atherogenic index = (Total cholesterol – HDL-cholesterol) / HDL-cholesterol.

⁴⁾Values are means ± SE of 10 rats, different superscripts within a row (a~e) indicate significant difference (p<0.05).

³⁾Values are means ± SE of 10 rats, different superscripts within a column (a~d) indicate significant difference (p<0.05).

CCl₄-HD, and HD-CCl₄ compared with the NC group were decreased by 19.5, 33.2, 16.5, and 25%, respectively.

XOD is classified into two forms: dehydrogenase (type D) using NAD as an electron acceptor, and oxidase (type-O) using O2 as an electron acceptor (11). The XOD in rat liver tissue is mainly an NAD⁺-dependent dehydrogenase under normal conditions (11,32). Under pathological conditions, XOD type-D can be converted to XOD type-O either reversibly by sulfhydryl oxidation (33) or irreversibly by proteolytic cleavage (34). The XOD type-O uses molecular oxygen as an electron acceptor and consequently generates the superoxide anion (35), which participates in the generation of other reactive oxygen species (ROS), including hydrogen peroxide, hydroxyl radical and singlet oxygen. Excessive formation of ROS in the body results in diseases such as cancer, metabolic diseases, and inflammation. However, the ROS scavenging system, such as superoxide dismutase, catalase, glutathione peroxidase, GST and reduced glutathione (36-41), protects cells from the destructive effect of ROS. Nevertheless, an imbalance between ROS generating and scavenging systems induces an interaction between ROS and important cell components such as DNA, RNA, protein, and lipids, resulting in tissue damage (42-44).

In this study, the increase of lipid peroxide as a parameter indicating cell membrane damage (45) by CCl₄ in all the experimental groups resulted from excessively generated oxygen free radical by the increased XOD type-O. On the other hand, the decrease of lipid peroxide in the HD groups probably resulted from the inhibition of the D- to O-type conversion by antioxidative components of the herb distillate (28,31), as well as the prevention or inhibition of liver damage by scavenging oxygen free radicals. However, further detailed studies on the components in herb distillate are needed to confirm these findings.

REFERENCES

- Marieb EN. 1998. Human Anantomy & Physiology. 4 ed. Benjamin/Cummings Science Publishing, California, USA. p 1006-1007.
- Park JH, Jin JH, Kim HJ, Park HR, Lee SC. 2005. Effect
 of far-infrared irradiation on the antioxidant activity of
 extracts from rice hulls. J Korean Soc Food Sci Nutr 34:
 131-134.
- 3. Lee CW, Jeong WI, Noh DH, Jeong DH, Do SH, Kim YK, Kwon OD, Kim TH, Jeong KS. 2005. Protective effects of bio-active ceramic water on alcohol-induced hepatic injury in pigs. *J Vet Med Sci* 67: 403-409.
- 4. Kim EJ, Lee KI, Park KY. 2004. The growth inhibition against gastric cancer cell in germanium or soybean

- sprouts cultured with germanium. Korean J Soc Food Cookery Sci 20: 287-291.
- Yoo BH, Park CM, Oh TJ, Han SH, Kang HH, Chang IS. 2002. Investigation of jewelry powders radiating far-infrared rays and the biological effects on human skin. J Cosmet Sci 53: 175-184.
- Kang JY, Kang SC, Park S. 2000. Effect of filtrate of loess suspension on growth and quality soybean sprouts. J Korean Soc Agric Chem Biotechnol 43: 266-270.
- Kim SD, Ku YS, Lee IZ, Kim ID, Youn KS. 2001. Gerneral components and sensory evaluation of hot water extract from Liriopis tuber. J Korean Soc Food Sci Nutr 30: 20-24.
- Miao TJ, Xiong RC. 1984. The use of Chinese herbs folium chinensis decoction plus eucalyptus distillate for preoperative preparation of intestinal tract. J Tradit Chin Med 4: 149-152.
- Kim SH, Hwang SY, Park OS, Kim MK, Chung YJ. 2005. Effect of Pinus densiflora extract on blood glucose level, OGTT and biochemical parameters in streptozotocin induced diabetic rats. J Korean Soc Food Sci Mutr 34: 973-979.
- Friedewald WT, Levy RI, Fredrickson DS. 1972. Estimation of the concentration of the low-density lipoprotein cholesterol in plasma without use of the preparative ultracentrifuge. Clin Chem 18: 499-502.
- 11. Stirpe F, Della Corte E. 1969. The regulation of rat liver xanthine oxidase. *J Biol Chem* 244: 3855-3863.
- Habig WH, Pabist MJ, Jakoby WB. 1974. Glutathione S-transferase. The first enzymatic step in mercapturic acid formation. J Biol Chem 249: 7130-7139.
- Ellman GL. 1959. Tissue sulfhydryl group. Arch Biochem Biophys 82: 70-77.
- Satho K. 1978. Serum lipid peroxide in cerebrovascular disorders determined by a new colorimetric method. Clin Chim Acta 90: 37-43.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RL. 1951.
 Protein measurement by folin phenol reagent. J Biol Chem 193: 265-275.
- Sandhofer F. 1994. Physiology and pathophysiology of the metabolism of lipoproteins. Wien Med Wochenschr 144: 286-290.
- Corvilain B. 1997. Lipoprotein metabolism. Rev Med Brux 18: 3-9.
- Robbinson DS. 1962. The development in the rat of fatty livers associated with reduced plasma-lipoprotein synthesis. Biochim Biophys Acta 62: 163-165.
- Yokota F, Igarashi Y, Suzue R. 1982. Effects of ethionine feeding on fatty liver and plasma lipoprotein fractions in rats. J Nutr 112: 405-409.
- Wakasugi JW, Katami K, Ikeda T, Tomikawa M. 1985.
 Action of malotilate on reduced serum cholesterol level in rats with carbon tetrachloride-induced liver damage. Japan J Pharmacol 38: 391-401.
- Maldonado V, Chan L, Melendez J, Rincon AR, Zhu HJ, Panduro A. 1994. Regulation of apo B mRNA expression in liver and intestine during liver regeneration induced by CCl₄. Biochim Biophys Acta 1211: 1-6.
- Boll M, Weber LW, Becker E, Stampfl A. 2001. Hepatocyte damage induced by carbon tetrachloride: inhibited lipoprotein secretion and changed lipoprotein composition. Z Naturforsch [C] 56: 283-290.
- Barisione G, Fontana L, Cottalasso D, Domenicotti C, Pronzato MA, Nanni G. 1993. Changes in lipogly-

- coprotein metabolism in toxic fatty liver. Minerva Gastroenterol Dietol 39: 101-112.
- 24. Honma T, Suda M. 1997. Changes in plasma lipoproteins as toxicity markers for carbon tetrachloride, chloroform, and dichloromethane. *Ind Health* 35: 519-531.
- Spector AA. 1984. Plasma lipid transport. Clin Physiol Biochem 2: 123-134.
- Honma T. 1990. Effect of trichloroethylene, 1,1,1-trichloroethane and carbon tetrachloride on plasma lipoproteins of rats. Ind Health 28: 159-174.
- Gergely J, Kulcsar A, Harsfalvi J. 1995. Changes in fat metabolism in acute carbon tetrachloride intoxication of rats. Acta Pharm Hung 65: 3-4.
- 28. Yamada H. 1991. Natural products of commercial potential as medicines. Curr Opin Biotech 2: 203-207.
- 29. Shin KH, Han JM, Lee IR. 1996. Effect of the constituents of Angelicae ginantis radix on hepatic drug metabolizing enzymes. Kor J Pharmacogn 27: 323-327.
- Kim MR, El-Aty AM, Kim IS, Shim JH. 2006. Determination of volatile flavor components in danggui cultivars by solvent-free injection and hydrodistillation followed by gas chromatographic-mass spectrophometric analysis. J Chromatogr A 1116: 259-264.
- 31. Wang CY, Ma FL, Liu JT, Tian JW, Fu FH. 2007. Protective effect of salvianic acid A on acute liver injury induced by carbon tetrachloride in rats. *Biol Pharm Bull* 30: 44-47.
- Yoon CG, Huh K. 1989. Effect of carbon tetrachloride intoxication on the type conversion of xanthine dehydrogenase into xanthine oxidase in rats. Arch Pharm Res 10: 36-41.
- Nishino T, Nishino T. 1997. The conversion from the dehydrogenase type to the oxidase type of rat liver xanthine dehydrogenase by modification of cysteine residues with fluorodinitrobenzene. *J Biol Chem* 272: 29859-29864.

- Nishino T, Tamura I. 1991. The mechanism of conversion of xanthine dehydrogenase to oxidase and the role of the enzyme in reperfusion injury. Adv Exp Med Biol 309: 327-337.
- 35. Fridovich I. 1970. Quantitative aspects of the production of superoxide anion radical by milk xanthine oxidase. *J Biol Chem* 245: 4053-4057.
- 36. Aebi H. 1974. Catalase. In *Methods of Enzymatic Analysis*. Bergmeyer HU, ed. Academic Press, New York. Vol 2, p 673-674.
- Chow CK, Tappel AL. 1974. Respose of glutathione peroxidase to dietary selenium in rats. J Nutr 104: 444-451.
- 38. Fried R. 1975. Enzymatic and non-enzymatic assay of superoxide dismutase. *Biochimie* 57: 657-660.
- Jacoby JB. 1978. The glutathione S-transferases: a group of multifunctional detoxification proteins. Adv Enzymol Relat Areas Mol Biol 46: 383-414.
- McCord JM. 1974. Free radical and inflammation: protection of snovial fluid by superoxide dismutase. Science 185: 529-531.
- Hayes JD, McLellan LI. 1999. Glutathione and glutathione-dependent enzymes represent a co-ordinately regulated defence against oxidative stress. Free Radic Res 31: 273-300.
- 42. Freeman BA, Crapo JD. 1982. Biology of disease: free radicals and tissue injury. Lab Invest 47: 412-426.
- Lebovitz BE, Siegel BV. 1980. Aspects of free radical reactions in biological systems: aging. J Gerontol 35: 45-56.
- Frank L. 1991. Developmental aspects of experimental pulmonary oxygen toxicity. Free Radic Biol Med 11: 463-494.
- 45. Plaa GL, Witschin H. 1976. Chemicals, drugs and lipid peroxidation. Am Rev Toxicol Pharmacol 16: 125-141.

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