# Efficacy of Pesticides and Growth Hormones against Root Disease Complex of Mulberry (Morus alba L.) 

Vorkady Nishitha Naik* and Dinesh Dutta Sharma<br>Central Sericulture Research and Training Institute, Mysore - 570008, Karnataka, India

(Received 30 October 2007; Accepted 22 November 2007)


#### Abstract

During mulberry cultivation, root disease complex caused by the association of root knot nematode (Meloidogyne incognita) with root rot pathogens like Fusarium solani and Botryodiplodia theobromae poses serious loss in leaf production. Therefore, an attempt was made to assess the efficacy of eight pesticides (Metayalaxyl + Mancozeb, Thiophanate methyl, Mancozeb, Bitertanol, Phenomiphos, Phorate, Thionazin \& Carbofuran) and two growth hormones (Salicylic acid and Indole 3 acetic acid) at 0.1 and $0.2 \%$ concentrations under in vitro conditions against nematode (hatching of eggs and mortality of larvae) and root rot pathogens (poisoned food technique) for short listing the treatments to develop an IDM strategy. Results revealed that among the pesticides and growth hormones, Carbofuran followed by Salicylic acid were found to be effective at $0.2 \%$ concentration against both nematode and pathogenic fungi. Both the chemicals inhibited the hatching of nematode eggs by 83.5$\mathbf{7 8 . 9 \%}$ and $\mathbf{8 0 - 7 6 \%}$ larval mortality over the control and reduced the mycelial growth of both the pathogenic fungi to an extent of 75.5-77.8\%. Though Mancozeb inhibited both the pathogenic fungi strongly (77 $\mathbf{- 8 0 \%}$ ), it did not show any effectiveness against nematode. The rest of the chemicals were found either moderately or poorly effective in reducing the growth of pathogenic fungi, hatching of nematode eggs and enhancing the mortality of larvae. The two effective chemicals viz, Carbofuran and Salicylic acid, which rated as strong inhibitors against both nematode and pathogenic fungi, can be exploited in developing an IDM package as one of the component for better man-


[^0]agement of root disease complex in mulberry.

Key words: Pesticides, Mulberry, Root disease complex, Root knot nematode, Pathogenic fungi.

## Introduction

Mulberry (Morus alba L.) a sole food plant of silkworms (Bombyx mori L ) is affected by two major soilborne diseases viz., root knot (Meloidogyne incognita) and root rot (Fusarium solani and Botryodiplodia theobromae), which affect the established plantations resulting in severe leaf yield loss up to $20 \%$ besides altering the nutritional status of the leaf (Sharma et al., 2003; Sharma and Gupta, 2005). Recently, root disease complex [association of root knot nematode (M. incognita) with root rot pathogens like Fusarium solani and B. theobromae] is a major production constraint during mulberry cultivation resulting in leaf yield loss to an extent of $15 \%$ (Naik and Sharma, 2004; Naik, 2006). Reports on the management of this disease are negligible (Sharma et al., 2003; Sharma and Gupta, 2005).

Furthermore, to obtain good quality mulberry leaf, it is essential to manage the array of pathogens that infest mulberry crop and the interactions among them. Till today, chemicals are being used as the most common component especially for soilborne diseases for developing an Integrated Disease Management package (Nehra, 2005). Various nematicides/fungicides have been tested against root disease complex in certain agricultural/horticultural crops and successfully reduced the severity of disease and improving the plant growth characters (Verma, 1993; Poornima and Vadivelu, 1993; Singh and Goswami, 2001). Besides, growth hormones are not only known to produce the pathogenesis related (PR) proteins such as chitinase, B-1 and 3-glucanase, which induce systemic resistance in plants against various foliar and soilborne
pathogens but also inhibited the pathogens growth to a great extent (Yalpani et al., 1993; Klessig and Malamy, 1994; Dann et al., 1996; Nandi et al., 2003). Hence, the present study was under taken to find out the efficacy of eight pesticides and two growth hormones against pathogens causing root disease complex viz., nematode (hatching of eggs and mortality of larvae) and root rot pathogens like B. theobromae and F. solani (poisoned food technique) under in vitro conditions using as one of the component in developing an Integrated Disease Management strategy.

## Materials and methods

Four fungicides such as Dithomyl (Metayalaxyl+Mancozeb; $8+64=72 \% \mathrm{WP}$ ), Roko (Thiophanate methyl 70\% WP), Dithane M-45 (Mancozeb 75\% WP) and Baycor (Bitertanol $25 \%$ WP), four nematicides like Namacure (Phenomiphos 10 G ), Thimet (Phorate 10 G ), Nemaphos (Thionazin 10 G ) and Furadan (Carbofuran 3 G ) and two growth hormones (Salicylic acid and Indole 3 acetic acid) at 0.1 and $0.2 \%$ concentrations were tested under in vitro conditions against pathogens causing root disease complex viz., M. incognita (hatching of eggs and mortality of larvae) and pathogenic fungi ( $B$. theobromae and $F$. solani) by poisoned food technique to assess their efficacy. Different concentration of each fungicide/nematicide was prepared using the following formula.
$\mathrm{D}=\frac{\mathrm{A} \times \mathrm{B}}{\mathrm{C}}$
Where, A: Required concentration (\%) of the fungicide/ nematicide
B : Required quantity of solution
C: \% Active ingredient available in the formulation
D : Required quantity of the product
The concentrations of growth hormones (Salicylic acid and Indole-3-acetic acid) like 0.1 and $0.2 \%$ were obtained by suitable dilution of respective appropriate concentration of stock solutions.

## Testing against nematode

To study the effect of chemicals on ovicidal action, four uniform size matured egg masses ( 1500 eggs) were suspended in each glass cavity block containing 5 ml solution of 0.1 and $0.2 \%$ concentrations of different chemicals along with a control (distilled water) and kept for incubation at $28 \pm 2^{\circ} \mathrm{C}$ for 10 days. After incubation, the hatched larvae were counted every alternate day. The hatching percentage and percentage inhibition over the control were calculated and categorized in to following 5 grades
(Naik, 2006).

| Grade | Inhibition of patho- Degree of effective- <br> gens <br> ness |  |
| :---: | :---: | :---: |
| I | Nil | Non inhibitor |
| II | $00.1-025 \%$ | Poor |
| III | $25.1-050 \%$ | Moderate |
| IV | $50.1-075 \%$ | Good |
| V | $75.1-100 \%$ | Strong |

For studying the larvicidal action, several egg masses were kept in glass cavity blocks containing 5 ml of sterile distilled water. The cavity blocks were incubated at $28 \pm$ $2^{\circ} \mathrm{C}$ overnight for hatching. 100 freshly hatched larvae were added to 5 ml solution of 0.1 and $0.2 \%$ chemicals separately in three replications along with control (larvae in distilled water) for comparison. Observations were recorded on mortality of larvae after 48 and 72 hrs in terms of dead larvae to calculate the mortality percentage. The larval mortality was judged by transferring the immobilized larvae into distilled water for an hour to confirm their death. The degree of effectiveness of each chemical was categorized into $1-5$ grades.

## Testing against pathogenic fungi

The inhibitory effect of chemicals on mycelial growth of both the pathogenic fungi (B. theobromae and $F$. solani) was assayed by poisoned food technique (Sharvelle, 1961). For this, the calculated quantity of each chemical at 0.1 and $0.2 \%$ concentrations were mixed in 100 ml of lukewarm Potato Dextrose Agar medium separately and poured into the Petriplates under aseptic condition. A four mm mycelial disc from 7 days old culture of both the pathogenic fungi was placed at the center of the plate separately for each chemical and concentration. A control was also maintained without amendment of chemical. Three replicates were maintained for each treatment. The plates were incubated at $28 \pm 2^{\circ} \mathrm{C}$ for 7 days. After incubation, the effectiveness of each chemical was determined measuring the radial growth ( mm ) of the fungal pathogens in the treated plates and comparing with the growth in the control. Inhibition percent was calculated by the formula of Vincent (1947) and effectiveness of each chemical was categorized into 1-5 grades.
Inhibition (\%)=
$\underline{\text { Radial growth in control(mm)-Radial growth in treated }}$
Radial growth in control

## Results

The results on ovicidal (hatching of M. incognita eggs) and larvicidal (mortality of larvae) properties of chemicals

Table 1. Comparative efficacy of different chemicals on ovicidal action against M. incognita (hatching of eggs) as assessed under in vitro conditions

| $\begin{gathered} \text { Sl. } \\ \text { No. } \end{gathered}$ | Chemicals | Concen -tration (\%) | No of larvae hatched /1500 eggs on |  |  |  |  | Total larvae hatched | Average hatched larvae | Inhibition over control (\%) | Inhibition grade |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | Days after |  |  |  |  |  |  |  |  |
|  |  |  | 2 | 4 | 6 | 8 | 10 |  |  |  |  |
| 1 | Control (D.W.) | -- | 520 | 314 | 256 | 164 | 110 | 1364 | 272.8 | - | - |
| 2 | Dithomyl(Metayalaxyl +Mancozeb) | 0.1 | 275 | 220 | 199 | 176 | 143 | 1013 | 202.6 | 25.7 | III |
|  |  | 0.2 | 256 | 205 | 187 | 169 | 82 | 899 | 179.8 | 34.1 | III |
| 3 | Roko (Thiophanate methyl) | 0.1 | 262 | 228 | 190 | 169 | 133 | 982 | 196.4 | 28.0 | III |
|  |  | 0.2 | 255 | 207 | 173 | 138 | 103 | 876 | 175.2 | 35.8 | III |
| 4 | Dithane M-45 <br> (Mancozeb) | 0.1 | 242 | 209 | 190 | 164 | 137 | 942 | 188.4 | 30.9 | III |
|  |  | 0.2 | 222 | 199 | 173 | 156 | 100 | 850 | 170.0 | 37.7 | III |
| 5 | Baycor (Bitertanol) | 0.1 | 302 | 257 | 206 | 154 | 58 | 977 | 195.4 | 28.4 | III |
|  |  | 0.2 | 288 | 258 | 189 | 148 | 35 | 918 | 183.6 | 32.7 | III |
| 6 | Nemacure (Phenomiphos) | 0.1 | 198 | 162 | 120 | 98 | 85 | 663 | 132.6 | 51.4 | IV |
|  |  | 0.2 | 152 | 132 | 108 | 72 | 35 | 499 | 99.8 | 63.4 | IV |
| 7 | Thimet (Phorate) | 0.1 | 171 | 145 | 106 | 88 | 63 | 573 | 114.6 | 58.0 | IV |
|  |  | 0.2 | 140 | 110 | 89 | 50 | 34 | 423 | 84.6 | 69.0 | IV |
| 8 | Nemaphos (Thionazin) | 0.1 | 174 | 108 | 83 | 69 | 61 | 495 | 99.0 | 63.7 | IV |
|  |  | 0.2 | 147 | 100 | 76 | 54 | 31 | 408 | 81.6 | 70.1 | IV |
| 9 | Furadan (Carbofuran) | 0.1 | 185 | 152 | 65 | 0 | 0 | 402 | 80.4 | 70.5 | IV |
|  |  | 0.2 | 121 | 68 | 36 | 0 | 0 | 225 | 45.0 | 83.5 | V |
| 10 | Salicylic acid | 0.1 | 198 | 175 | 96 | 0 | 0 | 469 | 93.8 | 65.6 | IV |
|  |  | 0.2 | 127 | 89 | 72 | 0 | 0 | 288 | 57.6 | 78.9 | V |
| 11 | Indole 3 acetic acid | 0.1 | 196 | 160 | 126 | 109 | 88 | 679 | 135.8 | 50.2 | IV |
|  |  | 0.2 | 172 | 151 | 111 | 84 | 58 | 576 | 115.2 | 57.8 | IV |
|  |  | SE $\pm$ | CD at 5 \% |  |  |  |  |  |  |  |  |
| Days (A) |  | 0.45 |  |  |  |  |  |  |  |  |  |
| Treatments (B) |  |  |  |  |  |  |  |  |  |  |  |
| Concentration (C) |  |  |  |  |  |  |  |  |  |  |  |
| A $\times \mathrm{B} \times \mathrm{C}$$\mathrm{CV} \%$ |  |  |  |  |  |  |  |  |  |  |  |
|  |  | 2.56 |  |  |  |  |  |  |  |  |  |

against nematode presented in Tables $1 \& 2$ clearly indicate that the concentration of $0.2 \%$ was found to be more toxic than $0.1 \%$. Among all the chemicals, Carbofuran (nematicide) and Salicylic acid (growth hormone) were able to suppress the hatching of $M$. incognita eggs to a maximum extent of 83.5 and $78.9 \%$, respectively at $0.2 \%$ concentration over the control. In both the cases, the maximum hatching was noticed on the second day and minimum on the sixth day (Table 1). Further, these two chemicals also caused higher larval mortality upto 80.0 and $76.0 \%$, respectively (Table 2) at $0.2 \%$ concentration after 72 hrs of exposure. Though there was a significant difference between Carbofuran and Salicylic acid with respect to suppression of hatching and increase in larval mortality, they showed strong nematicidal activity and based on the extent of inhibition they were rated under
class V as 'strong inhibitors'. The rest of the chemicals exhibited moderate to good nematicidal properties in suppression of egg hatching and increasing larval mortality. They were rated as 'moderate' to 'good' inhibitors belonging to classes III and IV. However, among the 3 chemicals, Dithane M-45 was not found effective against nematode.

With regards to the pathogenic fungi ( $B$. theobromae and $F$. solani), among all the chemicals under study, Mancozeb, Carbofuran and Salicylic acid inflicted maximum inhibition of mycelial growth at $0.2 \%$ concentration (Table 3). They inhibited the mycelial growth of B. theobromae by $77.0,76.0$ and $75.9 \%$ while in the case of $F$. solani, the growth was inflicted to an extent of $80.0,77.8$ and $75.5 \%$, respectively over the control. The values of the above three chemicals did not show any significant

Table 2. Comparative efficacy of different chemicals on larvicidal action (larval mortality) against M. incognita as assessed under in vitro conditions

difference. Thus, these three chemicals proved their strong fungicidal activity and can be grouped under class V as 'strong inhibitors'. The remaining chemicals were effective only as 'moderate' to 'good' against both the pathogenic fungi under classes III and IV. However, among the above three chemicals, Mancozeb was not found effective against nematode.

## Discussion

In the present investigation, among the 10 chemicals screened, Mancozeb had the maximum and strong fungicidal efficacy against both the pathogenic fungi (B. theobromae and F. solani). However, it did not exhibit any strong activity against nematode, which clearly proves its fungicidal action only. Whereas, Carbofuran being a nem-
aticide/insecticide and Salicylic acid being a growth hormone, strongly inhibited the nematode and both the pathogenic fungi indicating that having anti nematode and anti fungal properties. Carbofuran prepared by treating by 2,3-dihydro-2, 2-dimethyl-7-benzofuranol with methyl isocynate, might be toxic to both nematode and pathogenic fungi for inhibiting them. Various workers have reported the effectiveness of Carbofuran against root knot nematode in mulberry (Govindaiah and Sharma, 1994; Sharma et al., 1998; 2001; Ramakrishnan and Senthilkumar, 2003) and is also used for the control of disease complex caused by the association of nematode and soilborne fungal pathogens in certain crops (Sitaramaiah and Kumari, 1997; Mahapatra and Swain, 1999; Singh and Goswami, 2001; Haseeb et al., 2005; Shreenivasa et al., 2005). The present findings are also in conformity with the results of various workers who reported that certain

Table 3. Effect of different chemicals on pathogenic fungi as assessed under in vitro conditions

phytohormones/ chemical elicitors were found to be effective in reducing the growth and incidence of many fungal and nematode diseases/pathogens in certain crops including mulberry (Reuveni et al., 1994; Subramanian and Kumar, 1997; Vidhyasekaran, 1998; Nandi et al., 2000; Sahu, 2003).

Thus, the above observations under in vitro studies suggest that chemicals viz., Carbofuran and Salicylic acid exhibited strong inhibitory properties against nematode and root rot fungi, can be utilized as one of component for further refinement to formulate an integrated technology to manage the root disease complex in mulberry.

## Acknowledgements

The authors are highly thankful Mrs. M. Rekha, ScientistC (Statistics), CSRTI, Mysore for rendering help in sta-
tistical analysis.

## References

Dann, E. K., P. Menwle, J. P. Metranx, and B. J. Deverall (1996) The effect of pathogen inoculation or chemical treatment on activities of PR proteins in leaves of Phaseolus vulgaris L. Physiological and Molecular Plant Pathology 49, 307-319.
Haseeb, A., A. Sharma and P. K. Shukla (2005) Studies on the management of root-knot nematode, Meloidogyne incognita - wilt fungus, Fusarium oxysporum disease complex of green gram, Vigna radiata cv. ML-1108. J. Zhejiang University Science 6B, 736-742.
Govindaiah and D. D. Sharma (1994) Root knot nematode, Meloidogyne incognita infesting mulberry-A Review. Indian J. Seric. 33, 110-113.

Klessig, D. F. and J. Malamy (1994) The salicylic acid signal
in plants. Plants Mol. Biol. 26, 1439-1458.
Mahapatra, S. N. and P. K. Swain (1999) Avoidable yield loss due to Meloidogyne incognita and Fusarium oxysporum in black gram. Indian J. Neamtol. 29, 78-111.
Naik, V. N. and D. D. Sharma (2004) Efficacy of plant extracts, oil cakes and bioagents against pathogens causing disease complex in mulberry (Morus Spp.); in Workshop on promotion of bio-pesticides and bio-fertilizers in agriculture. Babu, V. S. and Valentina, G. (eds.), pp. 125-132, National Institute of Rural Development, Hyderabad, India.
Naik, V. N. (2006) Interaction of Meloidogyne incognita with soilborne fungal pathogens and their management in mulberry (Morus spp.). Ph. D. Thesis, Mysore University, Mysore, India.
Nandi, B., N. C. Sukul and S. P. S. Babu (2000) Exogenous salicylic acid reduces Meloidogyne incognita infestation of tomato. Allelopathy J. 7, 285-288.
Nandi, B., N.C. Sukul, N. Banerjee, S. Sengupta, P. Das and S.P.S. Babu (2003) Induction of pathogenesis related protein by salicylic acid and resistance to root- knot nematode in tomato. Indian J. Nematol. 33, 111-115.
Nehra, S. (2005) Integrated management of plant parasitic nematodes; in Plant diseases biocontrol management. Nehra, S. (eds.), pp.367-399, Aavishkar Publishers, Distributors, Jaipur, India.
Poornima, K. and S. Vadivelu (1993) Comparative efficacy of nematicides, oil cakes and plant extracts in the management of Meloidogyne incognita, Pratylenchus delattrei and Rotylenchulus reniformis on brinjal. Indian J. Nematol. 23, 170173.

Ramakrishnan, S. and T. Senthilkumar (2003) Plant parasitic nematodes, a serious threat to mulberry-A review. Indian $J$. Seri. 42, 82-92.
Reuveni, R.; V. Agapov, M. Reuveni and M. Reviv (1994) Effects of foliar sprays of phosphates on powdery mildew (Sphaerotheca pannosa) of roses. J. Phytopathol. 142, 331337.

Sahu, S. (2003) Evaluation of chemical elicitors for the induction of systemic acquired resistance in mulberry against leaf spot (Cercospora moricola Cooke) disease. M.Sc. Thesis, University of Mysore, Mysore, India.
Sharma, D. D. (1998) Nematicidal effect of culture filtrates of biocontrol agents on hatching of Meloidogyne incognita
eggs in comparison with Rugby 10 G and neem oil cake. Int. J. Tropical Plant Pathology 16, 239-242.

Sharma, S. D., A. Mishra, R. N. Pandey and S. J. Patel (2001) Sensitivity of Trichoderma harzianum to fungicides. J. Mycol. Pl. Pathol. 31, 251-253.
Sharma, D. D.; V. N. Naik, N. B. Chowdary and V. R. Mala (2003) Soilborne diseases of mulberry and their management - A review. Int. J. Indust. Entomol. 7, 93-106.
Sharma, D.D. and V. P. Gupta (2005) Soilborne diseases of mulberry and their management; in A text Book on Mulberry Crop Protection. Govindaiah; V. P. Gupta, D. D. Sharma; S. Rajadurai and V. N. Naik (eds.), pp. 195-228, Central Silk Board, Bangalore, India.
Shreenivasa, K. R., K. Krishnappa, B. M. R. Reddy, N. G. Ravichandra, K. Karuna and V. Kantharaju (2005) Integrated management of nematode complex on banana. Indian $J$. Nematol. 35, 37-40.
Singh, S. and B. K. Goswami (2001) Studies on the management of disease complex caused by root knot nematode, Meloidogyne incognita and wilt fungus, Fusarium oxyspor$u m$ on chickpea by nematode and carbofuran. Indian J. Nematol. 31, 122-125.
Sitaramaiah, K. and K. K. Kumari (1997) Evaluation of nematicides for the control Phytophthora-Meloidogyne disease complex in Tobacco. Indian J. Nematol. 27, 36-40.
Sharvelle, E. C. (1961) The nature and use of modern fungicides. Burgess Publishing Co., Minn, USA, p. 308.
Subramanian, S. and C. V. S. Kumar (1997) Induction of systemic resistance in greengram to Meloidogyne incognita by salicylic acid and phosphates. Int. J. Tropical Plant Diseases 15, 219-222.
Verma, K. K. and D. C. Gupta (1993) Germplasm evaluation of some rabi pluses against root-knot nematode, Meloidogyne javanica. Indian J. Nematol. 23, 209-210.
Vidhyasekaran, P. (1998) Molecular biology of pathogenesis and induced systemic resistance, Indian Phytopath. 51, 111120.

Vincent, J. M. (1947) Distoration of fungal hyphae in presence of certain inhibitors. Nature 96, 596.
Yalapani, N., V. Shulaev and I. Raskin (1993) Endogenous salicylic acid levels correlate with accumulation of PR-proteins and virus resistance in tobacco. Phytopathology 83, 702-708.


[^0]:    *To whom the correspondence addressed
    Central Sericultural Research and Training Institute, Mysore570 008, India. Tel: 091-0821-2362406; Fax: 091-08212362757; E-mail: nishi_naik2002@yahoo.co.in

