

Chemical and Antimicrobial Properties of Essential Oils from Three Coniferous Trees *Abies koreana*, *Cryptomeria japonica*, and *Torreya nucifera*

Hyun Jeong Oh^{1,*}, Hyo Min Ahn¹, Kyoung Ha So¹, Sang Suk Kim², Pil Yong Yun³, Gyeong Lyong Jeon^{1,4} and Key Zung Riu⁴

¹Bio-Agr. Co., Jeju Bio-Industry Development Center R105, Jeju 690-121, Korea

²SkincareCosmetic, Inc., Jeju Bio-Industry Development Center R103, Jeju 690-121, Korea

³Jeju Hi-Tech Industry Development Institute, Jeju 690-121, Korea

⁴Faculty of Biotechnology, Cheju National University, Jeju 690-756, Korea

Received August 7, 2007; Accepted September 11, 2007

Three coniferous essential oils were extracted from *Abies koreana*, *Cryptomeria japonica*, and *Torreya nucifera* by hydrodistillation. The chemical composition of each oil was analyzed by GC-MS, and their antimicrobial activities were tested against two bacteria and one yeast strains. Forty-seven compounds were identified from *A. koreana* oil, 39 from *C. japonica*, and 59 from *T. nucifera*. Main components of the essential oils were limonene (23.5%), bornyl acetate (17.9%), α -pinene (11.1%), and camphene (10.2%) in *A. koreana*, kaurene (26.3%), γ -eudesmol (19.0%), elemol (6.9%), and sabinene (5.1%) in *C. japonica*, limonene (13.5%), δ -cadinene (10.5%), α -bisabolol (10.2%), and α -copaene (7.7%) in *T. nucifera*. Among the three coniferous trees tested, the essential oils of *A. koreana* exhibited higher and broader antimicrobial activity against the tested organisms than those of *C. japonica* and *T. nucifera*.

Key words: *Abies koreana*, antimicrobial, coniferous, gas chromatography-mass spectrometry, volatile compound

Essential oils are the concentrates of the volatile compounds, which comprise terpenoids, hydrocarbons, alcohols, and aldehydes from the herbal plants. They have been regarded as useful antibacterial, antifungal, antiviral, antioxidizing, and anticancer agents [Burt, 2004; Edris, 2007]. They were also used in the production of pharmaceuticals, food flavorings, cosmetics, and fragrances by the industries.

Investigations on the compositions of the essential oils of firs and coniferous trees have been performed recently, including *Abies koreana* Wils., *Abies alba*, *A. nephrolepsis*, softwood leaves, and *Pinus* species [Baran *et al.*, 2007; Duquesnoy *et al.*, 2007; Hong *et al.*, 2004; Kurose *et al.*, 2007; Su *et al.*, 2006; Yang *et al.*, 2002]. However, little information is available on the antimicrobial and antifungal activities of the essential oils extracted from the coniferous trees.

In our search for the available and useful essential oil sources, among the coniferous trees, *A. koreana*, *Cryptomeria japonica*, and *Torreya nucifera*, native to the Jeju Island, were selected. *A. koreana* (Korean fir) is a member of the family Pinaceae with cone shape and is one of the fragrant, alpine conifer trees, endemic to the high mountains in Southern Korea including Mt. Dukyu, Mt. Chiri, and Mt. Halla [Lee, 1996]. *A. koreana* has been reported to have antibacterial, cytotoxic, and memory-enhancing effects [Kim *et al.*, 2001; Kim *et al.*, 2006; Jeong *et al.*, 2007]. The Japanese cedars, *C. japonica* (Taxodiaceae), were artificially planted in Jeju. It has been reported that various parts of *C. japonica* have antibacterial, antifungal, and antitermiticidal activities [Cha *et al.*, 2007]. The Japanese torreyia (*Torreya nucifera*), an evergreen tree indigenous to Japan, is a member of the family Taxaceae. Although studies have been performed on the composition of the essential oils of the coniferous trees, few have been done on the Japanese torreyia seed oil used as edible oil, hair oil, light, and medicine [Endo *et al.*, 2006].

In this study, the antimicrobial activities of the essential

*Corresponding author

Phone: 82-64-721-0399; Fax: 82-64-726-0399

E-mail: bbjeon0@kornet.net

Abbreviations: MICs, minimum inhibitory concentrations

oils isolated from three coniferous trees were investigated using the disc diffusion method and the minimal inhibitory concentration (MIC) assay. The chemical compositions of the essential oils isolated from the coniferous trees were also investigated.

Materials and Methods

Plant materials and preparation of essential oils.

The needles of *A. koreana*, *C. japonica*, and *T. nucifera* were collected in June, 2007. This research was carried out using the samples of hydrodistilled oils. The needles of trees were kept in a deep-freezer before use. The needles of trees were distilled for 6 h using simultaneous steam distillation apparatus to obtain the essential oil. Anhydrous sodium sulphate was used to absorb the little water contained in the essential oil.

GC/MS analysis. Analyses were carried out by GC/MS using an Agilent Technologies 6890 N equipped with a 5973 network mass selective detector, split/splitless injector, autosampler, and column HP-5MS (30 m × 0.25 mm i.d., 0.25 µm film thickness). The oven temperature was programmed from 60 to 200°C at 2°C/min. The carrier gas was helium at a flow rate of 1 mL/min. The GC-MS was carried out on a 6890 N mass spectrometer operating in the EI mode at 70 eV. The oils were analyzed individually by injecting 1 mL of 1 : 5 (v/v) solution of the essential oil in dichloromethane. The identification of the chemical constituents was based on the comparisons of their retention times and mass spectra with those obtained from the authentic sample Wiley (ver.7.0) libraries spectra, and literatures [Adams, 1995].

Antimicrobial activity assay. The bacterial strains, *Escherchia coli* ATCC 25922 and *Staphylococcus epidermidis* KCTC 3958, and the yeast strain, *Candida albicans* KCTC 7965, were used in this study. The bacterial strains were cultured overnight at 37°C in LB agar and Cornebacterium agar. The yeast was cultured at 30°C overnight in yeast malt agar.

Antimicrobial activities of the essential oil against bacteria and yeast strains were determined by the paper disc assay and the broth dilution method. The disc diffusion method was employed for the determination of antimicrobial activities of the essential oils. Briefly, a suspension of the tested microorganism (0.1 mL of 10⁸ cells per mL) was spread on to the solid media plates. Filter paper discs (8 mm in diameter) were impregnated with 40 µL of the oil and placed on the inoculated plates. These plates were incubated at 37°C for 24 h for bacteria and at 30°C for yeast. The diameters of the inhibition zones were measured in millimeters. All tests were performed in triplicates.

The minimum inhibitory concentrations (MICs) of the essential oils were determined for the using the broth dilution method. The antibacterial activities were determined after incubation at 37°C for 24 h (bacteria) or at 30°C for 24 h (candida) using the dilution techniques. MICs were determined as the lowest concentration of visible growth in the broth. Turbidity of the broth indicates the microorganism growth (OD₆₀₀).

Results and Discussion

Chemical composition of the essential oils. The yield of essential oils from *A. koreana*, *C. japonica*, and *T. nucifera* were 0.53 ± 0.10, 0.84 ± 0.06, and 0.37 ± 0.04% (v/w), respectively. The chemical components analyzed by GC-MS are presented in Table 1.

Forty-seven compounds were identified from the essential oil of *A. koreana*, of which 11 are monoterpene hydrocarbons (55.7%), 8 oxygenated monoterpenes (21.3%), 20 sesquiterpene hydrocarbons (18.2%), 6 oxygenated sesquiterpenes (2.1%), and 2 others (2.2%). The most abundant compound was limonene (23.5%), followed by bornyl acetate (17.9%), α-pinene (11.1%), camphene (10.2%), β-himachalene (4.2%), β-myrcene (3.4%), γ-selinene (3.1%), γ-gurjunene (2.4%), β-eudesmene (2.3%), β-pinene (2.1%), and other minor constituents.

The essential oil of *C. japonica* consisted of 39 compounds including 13 monoterpene hydrocarbons (17.8%), 6 oxygenated monoterpenes (5.9%), 9 sesquiterpene hydrocarbons (3.3%), 8 oxygenated sesquiterpenes (41.8%), 2 diterpenes (26.6%), and 1 of 2-hexanal (0.3%). The most abundant compound was kaurene (26.3%), followed by γ-eudesmol (19.0%), α-eudesmol (7.9%), elemol (6.9%), β-eudesmol (6.0%), sabinene (5.1%), 4-terpineol (4.6%), α-pinene (3.0%), γ-terpinene (2.2%), and other minor constituents.

Fifty-nine compounds were identified from the essential oil of *T. nucifera* oil, more than those of *A. koreana* and *C. japonica*. Among the compounds, 13 monoterpene hydrocarbons (32.1%), 6 oxygenated monoterpenes (1.0%), 23 sesquiterpene hydrocarbons (34.6%), 9 oxygenated sesquiterpenes (15.3%), 7 diterpenes (3.0%), and 1 of 2-hexanal (0.2%). Both monoterpene and sesquiterpene hydrocarbons were more abundant than the oxygenated derivatives, constituting 32.1 and 34.6% of the total volatile compounds, respectively. The most abundant compound was limonene (13.5%), followed by δ-cadinene (10.5%), α-Bisabolol (10.2%), α-copaene (7.7%), δ-3-carene (7.1%), α-pinene (5.3%), β-farnesene (2.7%), caryophyllene (2.6%), α-terpinolene (2.3%), α-humulene (2.2%), and other minor constituents.

The major components and their percentages were

Table 1. Chemical composition of the essential oils of the three valuable conifers in Jeju Island

Components ^a	Formula	Area (%)		
		<i>A. koreana</i>	<i>C. japonica</i>	<i>T. nucifera</i>
Monoterpenes				
Tricyclene	C ₁₀ H ₁₆	2.30	0.10	tr ^b
α-Thujene	C ₁₀ H ₁₆	- ^c	0.57	-
α-Pinene	C ₁₀ H ₁₆	11.11	2.96	5.34
α-Fenchene	C ₁₀ H ₁₆	-	-	0.68
Camphene	C ₁₀ H ₁₆	10.16	0.61	-
β-Pinene	C ₁₀ H ₁₆	2.07	-	0.67
Sabinene	C ₁₀ H ₁₆	-	5.06	0.22
β-Myrcene	C ₁₀ H ₁₆	3.41	1.26	1.24
1-Phellandrene	C ₁₀ H ₁₆	0.06	0.13	tr
δ-3-Carene	C ₁₀ H ₁₆	1.74	0.62	7.08
α-Terpinene	C ₁₀ H ₁₆	tr	1.42	0.09
dl-Limonene	C ₁₀ H ₁₆	23.49	1.89	13.52
γ-Terpinene	C ₁₀ H ₁₆	0.10	2.15	0.20
α-Terpinolene	C ₁₀ H ₁₆	1.27	0.93	2.33
α-Ocimene	C ₁₀ H ₁₆	-	0.10	0.74
		55.71	17.8	32.11
Oxygenated Monoterpenes				
Linalool	C ₁₀ H ₁₈ O	0.21	0.06	0.10
p-Menth-2-en-1-ol	C ₁₀ H ₁₈ O	-	0.10	-
Camphor	C ₁₀ H ₁₆ O	-	-	0.06
α-Camphenal	C ₁₀ H ₁₆ O	0.07	-	-
L-Borneol	C ₁₀ H ₁₈ O	1.27	-	-
4-Terpineol	C ₁₀ H ₁₈ O	0.09	4.63	0.29
α-Terpineol	C ₁₀ H ₁₈ O	0.41	0.29	0.22
β-Citronellol	C ₁₀ H ₂₀ O	-	-	0.25
Fenchyl acetate	C ₁₂ H ₂₀ O ₂	0.97	-	-
Bornyl acetate	C ₁₂ H ₂₀ O ₂	17.87	0.71	0.08
Sabinyl acetate	C ₁₂ H ₁₈ O ₂	-	0.06	-
Geranyl acetate	C ₁₂ H ₂₀ O ₂	0.42	-	-
		21.31	5.85	1.00
Sesquiterpenes				
δ-Elemene	C ₁₅ H ₂₄	-	tr	0.17
α-Longipinene	C ₁₅ H ₂₄	-	-	0.48
α-Ylangene	C ₁₅ H ₂₄	-	-	0.06
Isodene	C ₁₅ H ₂₄	-	-	0.06
α-Copaene	C ₁₅ H ₂₄	-	-	0.06
(-)-Alloaromadendrene	C ₁₅ H ₂₄	-	-	tr
β-Elemene	C ₁₅ H ₂₄	0.25	0.17	0.12
α-Gurjunene	C ₁₅ H ₂₄	0.13	-	0.14
Junipene	C ₁₅ H ₂₄	0.19	-	-
α-Chamigrene	C ₁₅ H ₂₄	0.13	-	-
Caryophyllene	C ₁₅ H ₂₄	1.50	0.07	2.64
β-Cubebene	C ₁₅ H ₂₄	-	-	1.18
α-Guaiene	C ₁₅ H ₂₄	0.12	-	-
α-Selinene	C ₁₅ H ₂₄	0.06	-	-
γ-Elemene	C ₁₅ H ₂₄	-	-	0.08
β-Farnesene	C ₁₅ H ₂₄	0.15	0.11	2.66

Table 1. (Continued)

Components ^a	Formula	Area (%)		
		<i>A. koreana</i>	<i>C. japonica</i>	<i>T. nucifera</i>
(+)-Aromadendrene	C ₁₅ H ₂₄	-	-	0.06
α-Amorphene	C ₁₅ H ₂₄	-	-	3.70
Germacrene-D	C ₁₅ H ₂₄	-	0.30	1.60
β-Eudesmene	C ₁₅ H ₂₄	2.31	-	-
γ-Selinene	C ₁₅ H ₂₄	3.09	-	-
α-Muurolene	C ₁₅ H ₂₄	-	0.28	0.06
cis-α-Bisabolene	C ₁₅ H ₂₄	-	-	0.28
β-Bisabolene	C ₁₅ H ₂₄	0.98	-	2.00
γ-Gurjunene	C ₁₅ H ₂₄	2.41	-	-
γ-Cadinene	C ₁₅ H ₂₄	-	0.21	-
7-epi-α-Selinene	C ₁₅ H ₂₄	1.08	-	-
δ-Cadinene	C ₁₅ H ₂₄	0.19	1.78	10.52
α-Cadinene	C ₁₅ H ₂₄	-	-	0.05
Germacrene B	C ₁₅ H ₂₄	0.28	-	0.90
trans-γ-Bisabolene	C ₁₅ H ₂₄	0.52	-	-
Cis-α-Bisabolene	C ₁₅ H ₂₄	0.08	-	-
Junipene	C ₁₅ H ₂₄	0.36	-	-
α-Copaene	C ₁₅ H ₂₄	-	-	7.71
Ledene	C ₁₅ H ₂₄	0.18	-	-
Thujopsene	C ₁₅ H ₂₄	-	-	0.07
β-Himachalene	C ₁₅ H ₂₄	4.18	-	-
Viridiflorene	C ₁₅ H ₂₄	-	0.42	-
		18.19	3.34	34.62
Oxygenated Sesquiterpenes				
α-Humulene	C ₁₅ H ₂₄ O	0.66	0.05	2.24
Elemol	C ₁₅ H ₂₆ O	-	6.87	-
Palustrol	C ₁₅ H ₂₆ O	-	-	0.22
α-Bisabolol	C ₁₅ H ₂₆ O	0.67	-	10.20
Junipercamphor	C ₁₅ H ₂₆ O	0.07	0.40	-
Farnesol	C ₁₅ H ₂₆ O	-	-	0.18
tau-Cadinol	C ₁₅ H ₂₆ O	-	-	1.67
β-Eudesmol	C ₁₅ H ₂₆ O	-	5.98	-
α-Eudesmol	C ₁₅ H ₂₆ O	-	7.86	-
Nerolidol	C ₁₅ H ₂₆ O	0.26	0.06	0.35
Caryophyllene oxide	C ₁₅ H ₂₄ O	0.09	-	0.17
Ledol	C ₁₅ H ₂₆ O	-	-	0.05
α-Eudesmol	C ₁₅ H ₂₆ O	-	1.59	-
γ-Eudesmol	C ₁₅ H ₂₆ O	0.32	19.02	-
Hexahydrofarnesyl acetone	C ₁₈ H ₃₆ O	-	-	0.21
		2.07	41.83	15.29
Diterpenes				
Isopimaradiene	C ₂₀ H ₃₂	-	0.32	0.13
Kaurene	C ₂₀ H ₃₂	-	26.31	0.12
Phytol	C ₂₀ H ₄₀ O	-	-	0.59
Dehydroabietal	C ₂₀ H ₂₈ O	-	-	0.38
Ferruginol	C ₂₀ H ₃₀ O	-	-	1.24
Methyl dehydroabietate	C ₂₁ H ₃₀ O ₂	-	-	0.29
4-Epidehydroabietol	C ₂₀ H ₃₀ O	-	-	0.24
		0.00	26.63	2.99

Table 1. (Continued)

Components ^a	Formula	Area (%)		
		<i>A. koreana</i>	<i>C. japonica</i>	<i>T. nucifera</i>
Others				
2-Hexenal	C ₆ H ₁₀ O	0.34	0.26	0.23
Santene	C ₉ H ₁₄	1.84	-	-
		2.18	0.26	0.23
Total		99.46	95.71	86.24
Grouped components				
Monoterpene hydrocarbons (%)		55.71	17.80	32.11
Oxygenated monoterpenes (%)		21.31	5.85	1.00
Sesquiterpenes hydrocarbons (%)		18.19	3.34	34.62
Oxygenated sesquiterpenes (%)		2.07	41.83	15.29
Diterpenes (%)		0.00	26.63	2.99
Others (%)		2.18	0.26	0.23
Total identified (%)		99.46	95.71	86.24

a: Components were analyzed on the Hp-5 column

b: Trace (<0.05%)

c: Not detected

Table 2. Antimicrobial activity of the coniferous essential oils by the diffusion method

Microorganisms	Inhibition zones (mm) ^a		
	<i>Abies koreana</i>	<i>Cryptomeria japonica</i>	<i>Torreya nucifera</i>
<i>Escherichia coli</i>	14.5 ± 1.32	- ^b	-
<i>Staphylococcus epidermidis</i>	27.30 ± 1.53	21.17 ± 1.04	11.67 ± 0.28
<i>Candida albicans</i>	34.0 ± 2.83	11.5 ± 2.12	-

a: Average value (triplicate) ± standard deviation.

b: Not detected.

Table 3. Determination of MIC (Minimum Inhibitory Concentration) of the essential oils

Microorganisms	MIC (mg/mL)		
	<i>Abies koreana</i>	<i>Cryptomeria japonica</i>	<i>Torreya nucifera</i>
<i>Escherichia coli</i>	2.05 ± 0.51	>26.16	>25.95
<i>Staphylococcus epidermidis</i>	3.81 ± 0.50	18.60 ± 2.01	>20.76 ± 2.44
<i>Candida albicans</i>	2.05 ± 0.51	7.56 ± 1.00	>25.95

limonene (23.5%) and bornyl acetate (17.9%) in *A. koreana*, kaulene (26.3%) and γ -eudesmol (19.0%) in *C. japonica*; and limonene (13.5%) and δ -cadinene (10.5%) in *T. nucifera*.

A. koreana is distributed in the alpine regions of southern Korean peninsula and Jeju Island. Our results on the main components of the essential oils of *A. koreana* grown in Mt. Halla were limonene, bornyl acetate, α -pinene, and camphene. These data were very similar to those reported for the needles of *A. koreana*, originated from Mt Halla in Jeju [Baran *et al.*, 2007], the major components being bornyl acetate, camphene, α -pinene, and limonene. On the other hand, the main constituents of

the essential oils of *A. koreana* grown in Mt. Dukyu were borneol, α -pinene, β -pinene, and bornyl acetate, and camphene and limonene were contained in trace amounts [Jeong *et al.*, 2007]. These changes might have arisen from the environmental differences, including climatic, seasonal, geographical, and genetic differences, among the mountainous regions (Mt. Dukyu, Mt. Chiri, and Mt. Halla) [Perry *et al.*, 1999].

Antimicrobial activities of the essential oils. The antimicrobial activities of the essential oils from *A. koreana*, *C. japonica*, and *T. nucifera* were evaluated by the disk diffusion method (Table. 2), and the MICs were determined against the two bacteria (*E. coli* and *S.*

epidermidis) and one yeast (*C. albicans*) strains (Table. 3). The results showed that the essential oil of *A. koreana* has stronger and broader spectrum of antimicrobial activities than those of *C. japonica* and *T. nucifera*. The yeast *C. albicans*, with the strongest inhibition zone (34 mm), was the most susceptible strain tested against the oil of *A. koreana*. The oil of *A. koreana* was effective against all strains tested in the study, at a range of 2.05-3.81 mg/mL, whereas the essential oil of *C. japonica* showed activities against *S. epidermidis* and *C. albicans*. All tested oils inhibited the growth of *S. epidermidis*. The *A. koreana* oil had the lowest MIC value of 3.81 mg/mL. It is of interest to note that the antifungal effect on the growth inhibition of *C. albicans* was observed in all essential oils except from that of *T. nucifera*, suggesting that the composition of each essential oil may vary in its effectiveness against *C. albicans*. Interestingly, these results indicate that the essential oil from *A. koreana* has both antibacterial and antifungal effects. *A. koreana* oil exhibited stronger activity than did *C. japonica* and *T. nucifera*.

The differences in the antimicrobial activity could be due to the differences in the chemical compositions of the oils. The content of monoterpene hydrocarbons (especially α -pinene) in *A. koreana* was higher than those of *C. japonica* and *T. nucifera*. In addition, α -pinene and β -pinene have been reported to have significant antibacterial activities [Couladis *et al.*, 2003; Chalchat *et al.*, 2000]. It is possible that these essential oils identified from the coniferous trees can be used antibacterial or antifungal agents in foods or in other products.

Acknowledgments. This study was supported by Jeju Hi-Tech Industry Development Institute.

References

- Adams RP (1995) Identification of essential oil components by gas chromatography/mass spectroscopy. Allured Publishing: Carol Stream, IL.
- Bagci E, and Digrak M (1996) Antimicrobial activity of essential oils of some *Abies* (Fir) species from Turkey. *Flavour Fragr J* **11**(4), 251-256.
- Baran S, Reuss SH, Konig WA, and Kalemba D (2007) Composition of the essential oil of *Abies koreana* Wils. *Flavour Fragr J* **22**, 78-83.
- Burt S (2004) Essential oils: their antibacterial properties and potential applications in foods- a review. *Int J Food Microbiol* **94**, 223-253.
- Cha JD, Jeong MR, Jeong SI, Moon SE, Kil BS, Yun SI, Lee KY, and Song YH (2007) Chemical composition and antimicrobial activity of the essential oil of *Cryptomeria japonica*. *Phytother Res* **21**, 295-299.
- Chalchat JC, Chiro F, Garry R, Lacosete J, and Santos V (2000) Photochemical hydroperoxidation of terpenes: Antimicrobial activity of α -pinene, β -pinene, and limonene hydroperoxides. *J Essent Oil Res* **12**, 125-126.
- Couladis M, Chinou IB, Tzakou O, and Petrakis PV (2003) Composition and antimicrobial activity of the essential oil of *Hypericum rumeliacum* subsp. *apollinis* (Boiss. & Heldr.). *Phytother Res* **17**, 152-154.
- Duquesnoy E, Casrola V, and Casanova J (2007) Composition and chemical variability of the twig oil of *Abies alba* Miller from Corsica. *Flavour Fragr J* **22**, 293-299.
- Edris AE (2007) Pharmaceutical and therapeutic potentials of essential oils and their individual volatile constituents: A review. *Phytother Res* **21**, 308-323.
- Endo Y, Osada Y, Kimura F, and Fujimoto K (2006) Effects of Japanese torreyia (*Torreya nucifera*) seed oil in lipid metabolism in rats. *Nutr* **22**, 553-558.
- Hong E J, Na KJ, Choi IG, Choi KC, and Jeung EB (2004) Antibacterial and antifungal effects of essential oils from coniferous trees. *Biol Pharm Bull* **27**(6), 863-866.
- Jeong SI, Lim JP, and Jeon H (2007) Chemical composition and antibacterial activities of the essential oil from *Abies koreana*. *Phytother Res* July 18 (in press).
- Kim HJ, Le QK, Lee MH, Kim TS, Lee HK, Kim YH, Bae K, and Lee IS (2001) A cytotoxic secocycloartenoid from *Abies koreana*. *Arch Pharm Res* **24**, 527-531.
- Kim KH, Bu YM, Jeong SG, Lim JP, Kwon YG, Cha DS, Jeon SR, Eun JS, and Jeon H (2006) Memory-enhancing effect of a supercritical carbon dioxide fluid extract of the needles of *Abies koreana* on scopolamine-induced amnesia in mice. *Biosci Biotechnol Biochem* **70**, 1821-1826.
- Kurose K, Okamura D, and Yatagai M (2007) Composition of the essential oils from the leaves of nine *Pinus* species and the cones of three of *Pinus* species. *Flavour Fragr J* **22**, 10-20.
- Lee TB (1996) "Illustrated Flora of Korea" (in Korean), Hyangmoon-Sa, Seoul, p. 50.
- Perry NB, Anderson RE, and Brennan NJ (1999) Essential oils from dalmatian sage (*Salvia officinalis* L.): variations among individuals, plant parts, seasons and sites. *J Agric Food Chem* **47**, 2048-2054.
- Su YC, Ho CL, and Wang EIC (2006) Analysis of leaf essential oils from the indigenous five conifers of Taiwan. *Flavour Fragr J* **21**, 447-452.
- Yang JK, Kang BK, Kim TH, Hong SC, Seo WT, and Choi MS (2002) Efficient extraction methods and analysis of essential oil from softwood leaves. *Kor J Biotechnol Bioeng* **17**, 357-364.