Increase of Membrane Potential by Ginsenosides in Prostate Cancer and Glioma cells

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Abstract : Ginseng has an anti-cancer effect in several cancer models. As a mechanism study of ginsenoside-induced growth inhibition in cancer cells, we measured change of membrane potential in prostate cancer and glioma cells by ginsenosides, active constituents of ginseng. Membrane potential was estimated by measuring fluorescence change of DiBAC-loaded cells. Among 11 ginsenosides tested, ginsenosides Rb₂, Rg₃, and Rh₂ increased significantly and robustly the membrane potential in a concentration-dependent manner in prostate cancer and glioma cells. Ginsenosides Rc, Ro, and Rb₁ slightly increased membrane potential. The ginsenoside-induced membrane potential increase was not affected by treatment with pertussis toxin or U73122. The ginsenoside-induced membrane potential increase was not diminished in Na⁺-free or HCO₃⁻-free media. Furthermore, the ginsenoside-induced increase of membrane potential was not changed by EIPA (5-(*N*-ethyl-*N*-isopropyl)-amiloride), SITS (4-acetoamido-4'-isothiocyanostilbene-2,2'-disulfonic acid), and ome-prazole. In summary, ginsenosides Rb₂, Rg₃, and Rh₂ increased membrane potential in prostate cancer and glioma cells in a GPCR-independent and Na⁺ independent manner.

Key words: Prostate, glioma, ginseng, ginsenoside, membrane potential

INTRODUCTION

Prostate carcinoma is the most common malignancy and age-related cause of cancer death worldwide. Furthermore, its prevalence has progressively increased in recent decades.¹⁾ Localized prostate carcinoma is potentially curable by a radical operation or definitive radiotherapy. However, metastatic prostate carcinoma can only be effectively controlled by hormone manipulation, since the prostate is an androgen-dependent organ. Although many therapeutic protocols have been proposed, none currently available has proved to be dramatically effective.^{2,3)} An extensive search of other potential managements is now under way, however, the negative aspect of the newly developed managements is drug resistance or toxic side effects.^{4,5)}

Gliomas represent about half of all brain tumors, and among them, glioblastoma multiformes is thought to be Ginseng, the root of *Panax ginseng* C.A. Meyer, is a medicinal plant consumed worldwide and has been reported to have various biological effects including anticarcinogenesis. ⁸⁻¹⁰⁾ Ginsenosides, ginseng saponins, have been suggested to be the major effective ingredients in ginseng. ⁸⁻¹⁰⁾ Among them, ginsenoside Rh₂ with a dammarane skeleton is shown to have some biological effects on cell differentiation, such as stimulation of melanogenesis in melanoma cells, ¹¹⁾ glucocorticoid-like action in growth suppressive effect on various cancer cells, ¹²⁾ and modulation of protein kinase C activity in HL-60 cells. ¹³⁾ Ginsenoside Rg₃ has inhibitory effects on Ca²⁺ channel in sensory neurons and ventricular myocytes, cancer cells growth and platelet aggregation. ¹⁴⁻¹⁶⁾

Modulation of membrane potential plays important roles in neuronal cells and smooth muscle cells. To clarify anticancer effects of ginseng, we investigated the effect of ginsenosides on membrane potential of prostate cancer

the most malignant and common intracranial tumor.⁶⁾ Although generally not metastatic, glioblastoma cells exhibit highly migratory and invasive behavior.⁷⁾

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and glioma cells. Among 11 ginsenosides (Rb₁, Rb₂, Rc, Rd, Re, Rf, Rg₁, Rg₂, Rg₃, Rh₂, and Ro) tested, ginsenoside Rb₂, Rc, Rg₃, and Rh₂ increased the membrane potential in a concentration-dependent manner.

MATERIALS AND METHODS

Agents

Ginsenosides (Rb₁, Rb₂, Rc, Rd, Re, Rf, Rg₁, Rg₂, Rg₃, Rh₂, and Ro) were purified in Ko's laboratory, the Korea Ginseng and Tobacco Research Institute, and the purities were more than 99.9%. They were dissolved in absolute methanol and stored at - 20°C. DiBAC₄(3) was acquired from Biotium (Hayway, CA, USA). All other materials were purchased from Sigma-Aldrich (St. Louis, MO, USA).

Prostate cancer and glioma cell culture

LNCaP and PC3 cells were purchased from Korean Cell Line Bank (Seoul, Korea) and grown in monolayers at 37° C in a 5% $CO_2/95\%$ air atmosphere in growth medium which consisted of: 90% Minimum Eagles' Medium (MEM, Gibco Laboratories, Grand Island, NY), 10% fetal bovine serum, 2 mM glutamine, 1 mM sodium pyruvate, 100 units/ml penicillin, and 50 µg/ml streptomycin. Rat C6 glioma cells were maintained in high glucose DMEM containing 10% (v/v) fetal bovine serum, 100 units/ml penicillin, 50 µg/ml streptomycin, 2 mM of glutamine, and 1 mM of sodium pyruvate at 37° C in a humidified 5% CO_2 incubator.

Measurement of membrane potential

The cells were sedimented, resuspended with a Hepesbuffered medium consisting of 20 mM of Hepes (pH 7.4), 103 mM NaCl, 4.8 mM KCl, 1.2 mM KH₂PO₄, 1.2 mM MgSO₄, 0.5 mM CaCl₂, 25 mM NaHCO₃, 15 mM glucose and 0.1% bovine serum albumin (fatty acid free), and then incubated for 30 min with 5 μ M of DiBAC₄(3). Fluorescence emission at 530 nm wavelength from excitation wavelength (488 nm) were measured every 0.1 sec by F4500 fluorescence spectrophotometer (Hitachi, Japan). Membrane potential was estimated by measuring fluorescence change of DiBAC-loaded cells.¹⁷⁾

Data presentation

Representative traces for membrane potential were chosen out of 3 separate experiments and shown in Fig 1-6.

RESULTS

Effects of ginsenosides on membrane potential in prostate cancer and glioma cells

Among the ginsenosides tested (Rb₁, Rb₂, Rc, Rd, Re, Rf, Rg₁, Rg₂, Rg₃, Rh₂, and Ro), ginsenosides Rb₂, Rh₂, Rc, and Rg₃ induced significant increase of membrane potential in androgen dependent LNCaP prostate cancer cells (Fig. 1). Ginsenosides Ro and Rb₁ slightly increased membrane potential in the prostate cancer cells (Fig. 1). However, ginsenosides Rd, Re, Rf, Rg₁, and Rg₂ did not increase membrane potential (Fig. 1), suggesting ginsenoside type-specific action of the increase of membrane

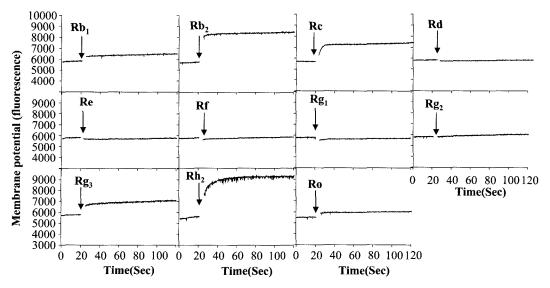


Fig. 1. Effects of ginsenosides on membrane potential in LNCaP prostate cells. Representative traces of membrane potential with 200 μM of each ginsenoside in DiBAC-loaded LNCaP cells were shown. Each ginsenoside was added at the arrow (30 sec).

potential. In androgen independent PC3 prostate cancer cells, we observed similar specificity of ginsenosides on membrane potential increase (data not shown). In C6 glioma cells, membrane potential was increased by ginsenoside Rb₂ and Rh₂ most prominently. Ginsenoside Rg₃, Rc, Rb₁, and Ro also induced slight to modest increase of membrane potential (Fig. 2). However, ginsenosides Rd, Re, Rf, Rg₁, and Rg₂ did not increase membrane potential (Fig. 2). In C6 glioma cells, increases of membrane potential by ginsenosides were stronger than those observed in

prostate cancer cells. Especially, ginsenoside Rb₂-induced increase of membrane potential was above the limit of the instrument and recorded as the highest number on scale (Fig. 2). Therefore, we choose C6 cells and continued further studies.

Ginsenosides Rb₂, Rh₂, and Rg₃ increased membrane potential and the increase sustained in C6 cells (Fig. 2). As shown in Fig 3, the ginsenosides-induced increases of membrane potential were concentration-dependent. Above 3 µM, increase of membrane potential was observed by

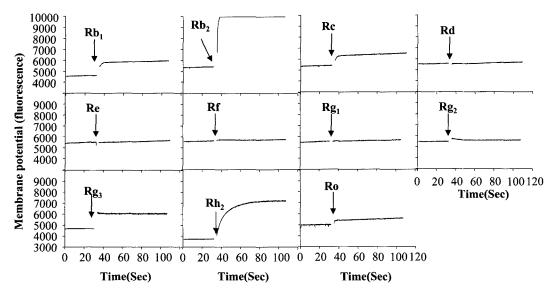


Fig. 2. Effects of ginsenosides on membrane potential in C6 glioma cells. Representative traces of membrane potential with 200 μM of each ginsenoside in DiBAC-loaded C6 glioma cells were shown. Each ginsenoside was added at the arrow (30 sec).

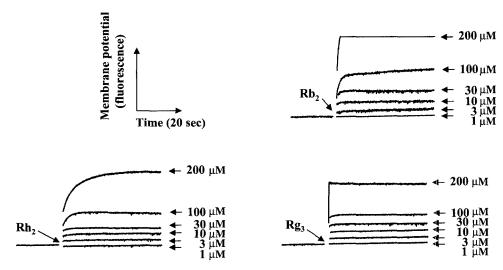


Fig. 3. Concentration-dependence of ginsenosides Rb₂, Rh₂, and Rg₃ on increase of membrane potential in C6 glioma cells. Representative trace of membrane potential with indicated concentration of each ginsenoside in DiBAC-loaded C6 glioma cells was shown. Each ginsenoside was added at the arrow (30 sec).

three ginsenosides and above $100 \,\mu\text{M}$, robust increases were observed (Fig. 3).

Involvement of G proteins and phospholipase C in ginsenoside-induced membrane potential increase

Pertussis toxin has been used to elucidate involvement of $G_{i/o}$ -type G proteins. Since G-protein-coupled receptor (GPCR) for ginsenoside action has been suggested, we treated C6 glioma cells with pertusis toxin (100 ng/ml, 24 hr). However, ginsenoside-induced increase of membrane potential was not ameliorated, suggesting no involvement of GPCR coupling to $G_{i/o}$ -type G proteins (Fig 4-A). We used U73122, a specific inhibitor of phospholipase C. Treatment of U73122 did not inhibit ginsenosides Rh_2 -induced membrane potential increase,

however, ginsenoside Rg_3 -induced increase of membrane potential was significantly blunted and ginsenoside Rb_2 -induced one was slightly (Fig 4-B). However, because increase of intracellular Ca^{2+} concentration was not observed by three ginsenosides, it is hard to conclude that ginsenosides activate U73122-sensitive phospholiase C and consequently increase the membrane potential. It is possible that U73122 may have another action site and action on the site influences the ginsenoside Rg_3 -induced increase of membrane potential.

Effect of Na⁺-free and HCO₃⁻-free media on ginsenoside-induced membrane potential increase

Intracellular pH is tightly regulated by pH regulator proteins such as Na⁺/H⁺ exchanger (NHE), bicarbonate

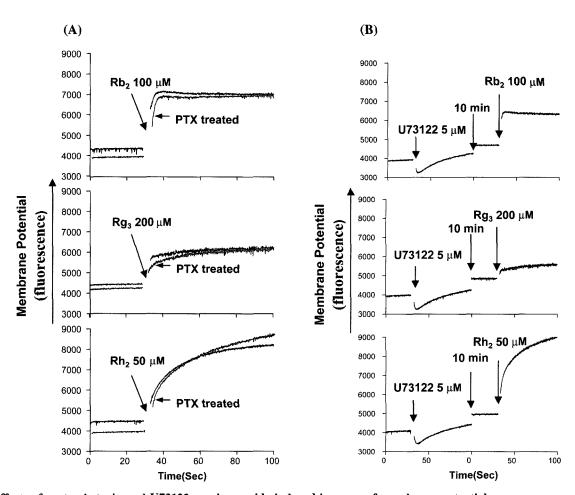


Fig. 4. Effects of pertussis toxin and U73122 on ginsenoside-induced increase of membrane potential.

A. Representative traces of membrane potential with 100 μ M of ginsenoside Rb₂, 200 μ M of ginsenoside Rg₃, or 50 μ M of ginsenoside Rh₂ in DiBAC-loaded C6 glioma cells treated with pertusiss toxin (100 ng/ml, 24 hr, dotted lines) or without pertussis toxin treatment (lines) were shown. B. Representative traces of membrane potential with 100 μ M of ginsenoside Rb₂, 200 μ M of ginsenoside Rg₃, or 50 μ M of ginsenoside Rh₂ in DiBAC-loaded C6 glioma cells. The cells were treated with U73122 (5 μ M, first arrow), after 10 min (second arrow) cells were treated with indicated ginsenosides (third arrow).

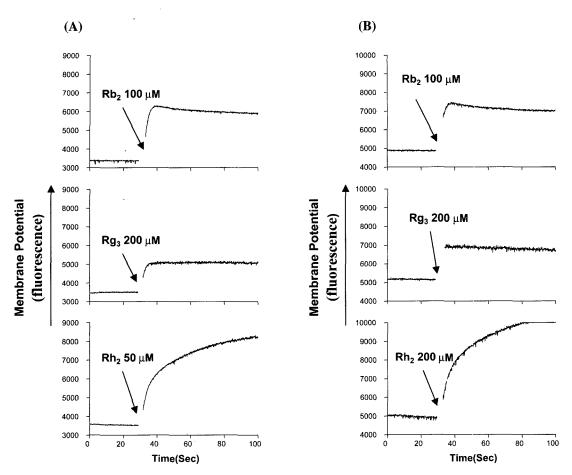


Fig. 5. Effects of Na⁺-free media and HCO₃⁻-free media on ginsenoside-induced increases of membrane potential.

Representative traces of membrane potential with 100 μM of ginsenoside Rb₂, 200 μM of ginsenoside Rg₃, or 50 μM of ginsenoside Rh₂ in DiBAC-loaded C6 glioma cells in Na⁺-free media (A) or in HCO₃⁻- free media (B) were shown.

transporter (BCT), and proton pump.²⁰⁾ Because such a regulation of ions could affect membrane potential, involvement of such regulators was studied. The effect of ginsenosides on membrane potential was measured in Na⁺-free or HCO₃⁻-free medium. Depletion of Na⁺ in the extracellular medium did not abrogate ginsenosideinduced increase of membrane potential, and also the membrane potential response in HCO3 -free medium was intact (Fig 5). To further confirm that NHE was not involved in ginsenoside-induced pH increase, EIPA (5-(Nethyl-N-isopropyl)-amiloride), a specific inhibitor of NHE, was used. As shown in Fig 6-A, EIPA treatment decrease the membrane potential dramatically by itself and such decrease of membrane potential lasted for 30 min. After 30 min of the EIPA treatment, ginsenosides were tested. Ginsenoside-induced increases of membrane potential were observed, however, the increments were lesser than non-treated groups. It is not possible to say that NHE is a component of ginsenoside-induced increase of membrane

potential, because the resting membrane potential was decreased by the treatment of EIPA. It is sure that EIPA-sensitive NHE is a regulator of membrane potential in C6 cells. Application of SITS (4-acetoamido-4'-isothiocy-anostilbene-2,2'-disulfonic acid), a specific inhibitor of BCT, or omeprazol, a specific inhibitor of proton pump, did not influence the ginsenoside activity significantly (Fig 6-B and C), excluding the possibility of involvement of BCT and proton pump in the response and supporting the result obtained in HCO₃-free medium. Therefore, the above results excluded the involvement of NHE, BCT, or proton pump in ginsenoside-induced increase of membrane potential.

DISCUSSION

Recently, Liu *et al.* reported that gingenoside Rg₃ inhibited proliferation of androgen dependent LNCaP cells.²¹⁾ Previously, we characterized effects of both ginsenosides

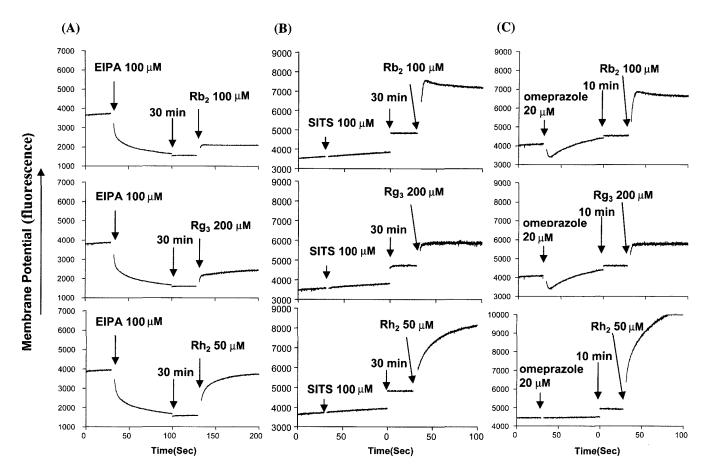


Fig. 6. Effects of EIPA, SITS, and omeprazole on ginsenoside-induced increase of membrane potential.

Representative traces of membrane potential with 100 μM of ginsenoside Rb₂, 200 μM of ginsenoside Rg₃, or 50 μM of ginsenoside Rh₂ in DiBAC-loaded C6 glioma cells. The cells were treated with EIPA (A, 100 μM, first arrow), SITS (B, 100 μM), or omeprazole (C, 20 μM), after 30 min (second arrow) cells were treated with indicated ginsenosides (third arrow).

Rg₃ and Rh₂ on androgen independent PC3 prostate cells.²²⁾ Inhibitory effects of ginsenosides Rg₃ and Rh₂ on the cell growth have previously been observed in other types of cancers. In particular, ginsenoside Rh₂ has been shown to be an apoptotic agent in rat glioma,²³⁾ human melanoma,²⁴⁾ ovarian cancer,²⁵⁾ breast cancer,²⁶⁾ hepatoma,²⁷⁾ and neuroblastoma.²³⁾ Also, ginsenoside Rh₂ has been reported to induce differentiation of melanoma,¹¹⁾ teratocarcinoma,¹²⁾ and leukemia.¹³⁾

Previously, we also reported cell detachment by ginsenoside Rg_3 and Rh_2 . There was differential cell detachment between ginsenoside Rg_3 and Rh_2 : Ginsenoside Rg_3 -induced detachment was greater than ginsenoside Rh_2 -induced one after 1 h of treatment, however, the detachment 24 h later was greater in ginsenoside Rh_2 -treated group than ginsenoside Rg_3 -treated group. Implication of the different action modes was further supported by differential modulations of MAP kinases by the

two ginsenosides.²²⁾ In androgen dependent LNCaP cells, inhibition of ERKs by ginsenosides Rg₃ and Rh₂ was an important trigger for cell detachment and growth inhibition, and modulation of p38 MAP kinase was additive in the case of the ginsenoside Rh₂. In androgen independent PC3 cells, modulation of p38 MAP kinase was important for the effects of both ginsenosides, and activation of JNK was additive in the case of ginsenosides Rg₃.²²⁾

As a mechanism study of ginsenoside-induced growth inhibition in cancer cells, we measured membrane potential in prostate cancer and glioma cells in the present study. Major findings of this communication are four folds. First, three ginsenosides, ginsenosides Rb_2 , Rh_2 and Rg_3 , were found to increase significantly and robustly membrane potential of androgen dependent and independent prostate cancer cells and C6 glioma cells. Second, the increase of membrane potential may not be related with $G_{1/0}$ type G proteins. Third, Na^+ ion or HCO_3^- ion is

not involved with the ginsenoside-induced increase of membrane potential. Last, NHE, BCT, and proton pump might not be related with the ginsenoside-induced increase of membrane potential. We could not observe any effect of ginsenosides on intracellular Ca²⁺ concentration in prostate cancer cells, excluding a possibility of that ginsenoside-induced Ca²⁺ increase causes increase of membrane potential.²²⁾

In the previous study, we showed increases of membrane potential by LPA, LPC, and SPC in C6 glioma cells. Although the precise mechanism for the increases was not elucidated, we found involvement of Na⁺ influx in the process and independence of suramin-sensitive GPCRs and pertussis toxin-sensitive G proteins.¹⁷⁾ However, in our study, ginsenoside-induced increase of membrane potential was not much changed by treatment of PTX, U73122, EIPA, SITS, or omeprazole. Further study is necessary to elucidate action mechanism of ginsenosides on membrane potential and to find relationship between anti-cancer effect of ginsenosides and increase of membrane potential by ginsenosides.

FOOTNOTE

Abbreviations used are: HBM hepes-buffered medium, EIPA 5-(*N*-ethyl-*N*-isopropyl)-amiloride, DiBAC₄(3) bis-(1,3-dibarbituric acid)-trimethine oxanol, SITS 4-acetoa-mido-4'-isothiocyanostilbene-2,2'-disulfonic acid, PTX pertussis toxin, GPCR G-protein-coupled receptor, NHE Na⁺/H⁺ exchanger, BCT bicarbonate transporter, LPA 1-oleoyl-*sn*-2-lysophosphatidic acid, LPC 1-palmitoyl-*sn*-2-lysophosphatidylcholine, SPC sphingosylphosphorylcholine.

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