

Cytotoxic Activities of Green and Brown Seaweeds Collected from Jeju Island against Four Tumor Cell Lines

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Abstract

Methanolic and aqueous extracts from 37 seaweed species (10 green and 27 brown seaweeds) collected from Jeju Island coast were prepared at high (70°C) and room (20°C) temperatures and examined for cytotoxic activity against 4 tumor cell lines: U937 (human monoblastoid leukemia cell line), HL60 (human promyelocytic leukemia cell line), HeLa (woman cervical carcinoma cell line) and CT26 (mouse colon carcinoma line). Both MeOH extracts of *Desmarestia tabacoides* and *Dictyota dichotoma* possessed strong cytotoxic activities against all the tumor cell lines tested, but the aqueous extract exhibited no activity. On the other hand *Ecklonia cava* showed strong cytotoxic activities for the 20°C aqueous extract against the three tumor cells except HeLa cell. *Sagassum coreanum* and *Sagassum siliquastrum* 20°C aqueous extracts also exhibited strong cytotoxic activities against U937, HL60, HeLa cells. Even though green seaweeds showed less activity than brown seaweeds, 20°C aqueous extracts of *Codium contractum* and *Codium fragile* exhibited strong cytotoxic activities against HL60 or CT26 cells, respectively.

Key words: cytotoxic activity, seaweeds extract, Jeju Island, tumor cells

INTRODUCTION

Cancer is a disease characterized by uncontrolled cell growth that presents over 100 distinct clinical pathologies (1). Cancer is the largest single cause of death in both men and women, claiming over 6 million lives each year worldwide. In the last few decades, basic cancer research has produced remarkable advances in understanding the biology and genetics of cancer (2). Recently, many anti-cancer drugs have been developed and applied by clinical doctors. Chemotherapeutic agents and radiation, which cause DNA mutation in actively dividing cells, were intended to selectively kill cancer cells while having limited effect on normal cells. Unfortunately, these cytotoxic agents, while effective in managing certain types of cancer, were limited in their utility due to their toxicity in normal dividing cell populations, resulting in adverse side effects. Therefore, the research into developing new and safe drugs has become a subject of great interest to the pharmaceutical industry (3).

Marine bioresources are known to be attractive as they sometimes yield new compounds showing several kinds of different bioactivities which are not possible in land plants. Screening of algal extracts for biologically active compounds began in the 1950s with simple antibiotic assays and soon expanded to include testing for products

with antiviral, antibacterial, antifungal, anti-mitotic or anti-tumorigenic activities (4-7). Studies on antitumor effects of algal species have been reported by a number of researchers (8-12). Seaweeds contain rich bioactive constituents such as minerals and trace elements identified as bodily requirements, plus they have other nutrients, many of which are known to offer protection against a variety of health complications. The composition of seaweeds is quite different from the land plants. This facilitates the isolation of certain important bioactive materials which are not obviously possible with land plants. Different aspects of biochemical studies have isolated different components of seaweeds which are antioxidants, anticancer, antiangiogenic, anticoagulant, antibacterial, antifungal, anti-inflammatory and immunomodulatory compounds (13-18).

The objective of the present study was to investigate the growth inhibition effects of the methanolic and aqueous extracts from 37 seaweed species on four tumor cell lines (U937, HL60, HeLa and CT26).

MATERIALS AND MHEHODS

Chemicals

RPMI-1640, fetal bovine serum (FBS) and phosphate buffer saline (PBS) were purchased from Gibco BRL Co.

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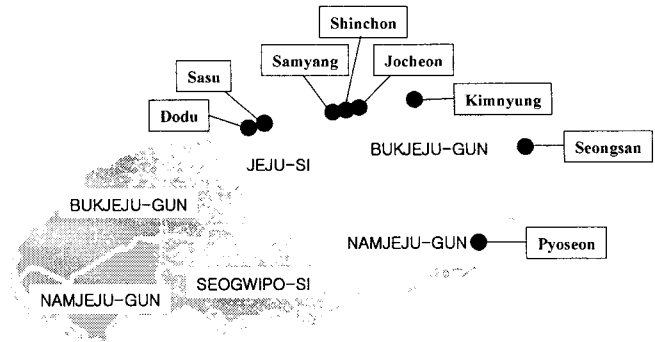
Table 1. Jeju green and brown seaweeds used in this study

Scientific name	Korea name	Collected space
Green algae		
<i>Monostroma nitidum</i>	참홀파래	Jocheon
<i>Enteromorpha compressa</i>	납작파래	Jocheon
<i>Enteromorpha intestinalis</i>	창자파래	Dodu
<i>Enteromorpha linza</i>	잎파래	Jocheon
<i>Enteromorpha</i> sp.	파래류	Dodu
<i>Ulva conglobata</i>	모란갈파래	Jocheon
<i>Ulva pertusa</i>	구멍갈파래	Jocheon
<i>Chaetomorpha linum</i>	실염주말	Samyang
<i>Codium contractum</i>	몽우리청각	Shinchon
<i>Codium fragile</i>	청각	Kimnyung
Brown algae		
<i>Papenfussiella kuromo</i>	연두털말	Sasu
<i>Ishige okamurai</i>	패	Jocheon
<i>Ishige sinicola</i>	넓패	Seongsan
<i>Leathesia difformis</i>	바위두룩	Seongsan
<i>Petrospongium rugosum</i>	바위주름	Jocheon
<i>Colpomenia sinuosa</i>	볼레기말	Seongsan
<i>Endarachne binghamiae</i>	미역쇠	Seongsan
<i>Hydroclathrus clathratus</i>	그물바구니	Seongsan
<i>Scytosiphon lomentaria</i>	잘룩이고리매	Jocheon
<i>Myelophycus simplex</i>	바위수염	Dodu
<i>Desmarestia tabacoides</i>	담배산말	Kimnyung
<i>Undaria pinnatifida</i>	말미역	Seongsan
<i>Ecklonia cava</i>	감태	Pyoseon
<i>Laminaria ochotensis</i>	다시마	Seongsan
<i>Dictyopteris prolifera</i>	가시뻬대그물말	Sasu
<i>Dictyota dichotoma</i>	참그물바탕말	Jocheon
<i>Pachydictyon</i> sp.	참가죽	Sasu
<i>Padina arborescens</i>	부켓살	Seongsan
<i>Myagropsis myagroides</i>	외톨개모자반	Seongsan
<i>Hizikia fusiformis</i>	돛	Jocheon
<i>Sargassum coreanum</i>	큰잎모자반	Jocheon
<i>Sargassum fulvellum</i>	모자반	Seongsan
<i>Sargassum horneri</i>	잇바디 팽생이모자반	Seongsan
<i>Sargassum piluliferum</i>	구슬모자반	Seongsan
<i>Sargassum siliquastrum</i>	파배기모자반	Seongsan
<i>Sargassum thunbergii</i>	지충이	Jocheon

(Gaithersburg, MD, USA). Dimethyl sulfoxide (DMSO) and 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium-bromide (MTT) were purchased from Sigma Co. (St. Louis, MO, USA).

Preparation of seaweed extracts

Seaweeds were collected along the Jeju Island coast (Table 1 and Fig. 1) of Korea during a period from February 2004 to March 2005. Of the Jeju coastal seaweeds, 27 species of brown and 10 species of green seaweeds were collected, and then salt, epiphytes and sand were removed using tap water. Finally the seaweeds were rinsed carefully with freshwater and stored in a medical freezer at -20°C . The frozen samples were lyophilized and homogenized with a grinder before extraction. The powdered samples were then extracted for 24

**Fig. 1.** Map of the sampling stations in the study area, Jeju Island, S. Korea.

h first with 80% MeOH under continuous shaking at 70°C and 20°C , and then aqueous extracts were prepared from the residue. The result was four unique extracts: 70 ME (methanolic extract at 70°C), 20 ME (methanolic extract at 20°C), 70 AE (aqueous extract at 70°C) and 20 AE (aqueous extract at 20°C). The methanolic extracts were first subjected to evaporation and then dissolved in DMSO and used for experiments adjusting the final concentration of DMSO in culture medium to $<0.01\%$. Respective water extracts were freeze dried and a known amount of the powder was again dissolved in water.

Cell culture

U937 (human monoblastoid leukemia cell line), HL60 (human promyelocytic leukemia cell line), HeLa (woman cervical carcinoma cell line) and CT26 (mouse colon carcinoma line) were grown in RPMI 1640 medium supplemented with 10% (v/v) heat inactivated fetal bovine serum (FBS), penicillin (100 U/mL) and streptomycin (100 $\mu\text{g}/\text{mL}$). Cultures were maintained at 37°C in 5% CO_2 incubator.

Cell growth inhibition assay

The cytotoxicity of methanolic and aqueous extracts from marine algae was determined by a colorimetric MTT assay. Suspension cells (U937 and HL60 cells) were seeded at a concentration of 2×10^4 cells/mL together with the extracts (100 $\mu\text{g}/\text{mL}$) and incubated up to 72 h before MTT treatment. Attach cells (HeLa and CT26 cells) were seeded in a 96-well plate at a concentration of 2×10^4 cells/mL. Sixteen hours after plating, the cells were treated with the extract samples (100 $\mu\text{g}/\text{mL}$). The cells were then incubated for an additional 72 h at 37°C . MTT stock solution (50 μL ; 2 mg/mL in PBS) was then added to each well for a total reaction volume of 250 μL . After incubating for 4 h, the plate was centrifuged at 2,000 rpm for 5 min and the supernatants were aspirated. The formazan crystals in each well were dissolved in 150 μL of DMSO. The amount of purple formazan was

determined by measuring the absorbance at 540 nm.

Cell growth inhibition calculation

The percentage of cancer cell growth inhibition was calculated according to the following equation.

$$\% \text{ growth inhibition} = 1 - (\text{CS}/\text{CC}) \times 100$$

Where CS is the cells treated with seaweed extracts; CC is the untreated control cells grown with the same amount of distilled water or DMSO added as used as the solvent in the sample treatment.

Statistical analysis

The overall growth inhibition activities of methanolic and aqueous extracts on tumor cell lines are expressed

as the means \pm SD. These data were analyzed for significance using Student's *t*-test. $p < 0.05$ was considered to be a significant difference.

RESULTS AND DISCUSSION

Although many anticancer agents have been developed and used, their side effects and resistance to anticancer drugs are serious problems to be overcome in the treatment of cancer. There is a need, therefore, to develop of safer and better therapeutic drugs from natural bioresources. Recently, there has been increasing interest in the cancer therapeutic potential of natural plants, suggesting that many plants have anticancer activities that

Table 2. Growth inhibition activity of methanolic and aqueous extracts (100 $\mu\text{g}/\text{mL}$) from Jeju seaweeds on U937 cells

Scientific name	U937 cell growth inhibition rate (%)			
	20 ME ¹⁾	70 ME	20 AE	70 AE
Green seaweeds				
<i>Monostroma nitidum</i>	17.98 \pm 12.08	21.12 \pm 6.00	16.89 \pm 11.57	10.26 \pm 4.35
<i>Enteromorpha compressa</i>	32.33 \pm 7.05	60.31 \pm 3.52	18.84 \pm 1.80	28.73 \pm 6.46
<i>Enteromorpha intestinalis</i>	44.03 \pm 5.34	48.58 \pm 0.37	29.55 \pm 2.61	24.74 \pm 0.47
<i>Enteromorpha linza</i>	25.68 \pm 15.12	42.82 \pm 5.41	45.03 \pm 1.55	26.65 \pm 7.05
<i>Enteromorpha sp.</i>	39.22 \pm 3.36	60.90 \pm 6.82	30.35 \pm 6.29	26.39 \pm 11.01
<i>Ulva conglobata</i>	24.53 \pm 5.63	51.78 \pm 3.41	30.80 \pm 6.25	14.05 \pm 4.94
<i>Ulva pertusa</i>	23.99 \pm 7.37	35.23 \pm 4.92	30.01 \pm 0.28	13.79 \pm 21.15
<i>Chaetomorpha linum</i>	27.40 \pm 7.29	27.99 \pm 14.82	31.21 \pm 10.63	14.51 \pm 3.54
<i>Codium contractum</i>	47.80 \pm 6.47	52.36 \pm 1.70	62.08 \pm 0.98	36.35 \pm 7.00
<i>Codium fragile</i>	44.32 \pm 0.25	51.80 \pm 16.71	45.03 \pm 1.55	26.65 \pm 7.05
Brown seaweeds				
<i>Papenfussiella kuromo</i>	39.94 \pm 4.15	34.96 \pm 0.07	28.76 \pm 11.12	14.45 \pm 5.32
<i>Ishige okamurai</i>	34.47 \pm 5.00	20.05 \pm 5.24	43.24 \pm 2.94	36.36 \pm 5.48
<i>Ishige sinicola</i>	2.95 \pm 1.07	15.01 \pm 7.13	29.02 \pm 0.82	26.76 \pm 2.86
<i>Leathesia difformis</i>	1.74 \pm 5.24	26.73 \pm 7.71	34.89 \pm 14.46	48.90 \pm 7.23
<i>Petrospongiium rugosum</i>	8.46 \pm 1.80	15.83 \pm 1.93	11.54 \pm 2.52	15.21 \pm 9.14
<i>Colpomenia sinuosa</i>	16.72 \pm 15.05	25.00 \pm 17.57	6.66 \pm 0.47	26.19 \pm 6.25
<i>Endarachne binghamiae</i>	35.40 \pm 2.21	44.50 \pm 2.13	25.55 \pm 2.13	33.93 \pm 1.21
<i>Hydroclathrus clathratus</i>	48.11 \pm 2.82	63.53 \pm 5.78	38.19 \pm 10.23	24.01 \pm 6.16
<i>Scytosiphon lomentaria</i>	25.56 \pm 4.98	18.60 \pm 11.06	20.06 \pm 1.88	19.36 \pm 13.16
<i>Myelophycus simplex</i>	26.14 \pm 12.57	23.35 \pm 13.19	22.20 \pm 8.83	26.07 \pm 11.53
<i>Desmarestia tabacoides</i>	86.37 \pm 5.34	79.43 \pm 5.43	* ²⁾	*
<i>Undaria pinnatifida</i>	44.61 \pm 5.90	59.15 \pm 0.41	22.60 \pm 2.37	20.75 \pm 13.79
<i>Ecklonia cava</i>	37.66 \pm 9.83	33.31 \pm 1.39	74.39 \pm 1.72	33.87 \pm 3.76
<i>Laminaria ochotensis</i>	33.78 \pm 5.82	12.80 \pm 2.04	26.42 \pm 3.19	41.94 \pm 5.26
<i>Dictyopteris prolifera</i>	51.83 \pm 13.27	14.36 \pm 12.16	23.48 \pm 11.75	9.17 \pm 0.47
<i>Dictyota dichotoma</i>	84.33 \pm 0.52	76.47 \pm 1.26	37.14 \pm 7.00	20.45 \pm 11.75
<i>Pachydictyon sp.</i>	69.18 \pm 1.93	42.40 \pm 5.11	18.21 \pm 7.46	22.30 \pm 1.68
<i>Padina arborescens</i>	34.65 \pm 7.37	20.44 \pm 10.31	19.19 \pm 3.27	21.62 \pm 5.89
<i>Myagropsis myagroides</i>	20.55 \pm 12.60	25.68 \pm 12.75	48.75 \pm 10.73	9.37 \pm 4.10
<i>Hizikia fusiformis</i>	47.45 \pm 5.65	45.13 \pm 1.39	29.77 \pm 1.06	25.61 \pm 7.28
<i>Sargassum coreanum</i>	20.02 \pm 8.45	21.49 \pm 1.93	80.80 \pm 3.45	10.63 \pm 4.35
<i>Sargassum fulvellum</i>	11.53 \pm 3.56	17.77 \pm 3.15	46.83 \pm 0.19	26.03 \pm 5.02
<i>Sargassum horneri</i>	29.61 \pm 8.08	31.08 \pm 2.00	58.27 \pm 10.92	19.85 \pm 7.00
<i>Sargassum piluliferum</i>	31.29 \pm 11.47	26.99 \pm 0.52	27.51 \pm 5.69	36.02 \pm 6.34
<i>Sargassum siliquastrum</i>	15.79 \pm 7.49	24.27 \pm 4.45	72.03 \pm 3.92	18.19 \pm 6.13
<i>Sargassum thunbergii</i>	23.60 \pm 11.12	25.20 \pm 4.67	32.20 \pm 6.29	29.71 \pm 4.90

¹⁾20 ME: methanolic extract at 20°C, 70 ME: methanolic extract at 70°C, 20 AE: aqueous extract at 20°C, 70 AE: aqueous extract at 70°C.

²⁾Not determined.

could be therapeutically useful. The objective of the present study was to investigate the growth inhibition effects of the methanolic and aqueous extracts of 37 marine alga species on four tumor cell lines (U937, HL60, HeLa and CT26 cell).

The U937 cell growth inhibitory effects of seaweed extracts are shown in Table 2. Ten brown seaweeds and five green seaweeds inhibited the growth of U937 cells by more than 50%. Of those extracts, *Desmarestia tabacoides* 20 ME, *Dictyota dichotoma* 20 ME, *Sargassum coreanum* 20 AE, *Ecklonia cava* 20 AE and *Sargassum siliquastrum* 20 AE exhibited growth inhibitions of 86.37%, 84.33%, 80.80%, 74.39% and 72.03%, respec-

tively, on U937 cells.

The HL60 cell growth inhibitory effects of the seaweed extracts are shown in Table 3. Growth inhibition activity of eleven brown seaweeds and seven green seaweeds on HL60 cells exceeded 50%. Excellent growth inhibitory activities in this assay was exhibited by *D. tabacoides* 20 ME (89.27%), *D. dichotoma* 20 ME (85.85%), *S. siliquastrum* 20 AE (75.78%), *S. coreanum* 20 AE (73.94%) and *E. cava* 20 AE (72.24%).

The growth inhibitory effects of seaweed extracts against HeLa and CT26 cells are shown in Table 4 and 5. Growth inhibition activity of six brown seaweeds and two green seaweeds on HeLa cells exceeded 50% and

Table 3. Growth inhibition activity of methanolic and aqueous extracts (100 µg/mL) of Jeju seaweeds on HL60 cells

Scientific name	HL60 cell growth inhibition rate (%)			
	20 ME ¹⁾	70 ME	20 AE	70 AE
Green seaweeds				
<i>Monostroma nitidum</i>	10.93 ± 4.77	31.38 ± 4.71	46.49 ± 0.83	37.27 ± 0.79
<i>Enteromorpha compressa</i>	20.73 ± 4.13	63.97 ± 1.23	42.30 ± 7.99	33.01 ± 6.54
<i>Enteromorpha intestinalis</i>	33.17 ± 9.43	35.16 ± 4.89	35.53 ± 8.78	45.06 ± 0.75
<i>Enteromorpha linza</i>	26.91 ± 2.99	45.65 ± 12.02	58.35 ± 0.22	40.31 ± 3.34
<i>Enteromorpha</i> sp.	25.76 ± 8.21	64.23 ± 2.76	51.52 ± 2.59	49.91 ± 0.04
<i>Ulva conglobata</i>	4.88 ± 2.37	16.67 ± 3.27	51.83 ± 4.61	43.14 ± 5.49
<i>Ulva pertusa</i>	24.23 ± 7.01	27.52 ± 6.27	45.06 ± 4.88	35.28 ± 10.54
<i>Chaetomorpha linum</i>	16.50 ± 4.79	5.98 ± 2.51	60.90 ± 4.35	57.24 ± 0.57
<i>Codium contractum</i>	43.61 ± 0.90	49.92 ± 13.57	71.80 ± 10.54	53.51 ± 0.66
<i>Codium fragile</i>	68.94 ± 0.09	33.21 ± 1.09	67.76 ± 1.23	59.72 ± 1.45
Brown seaweeds				
<i>Papenfussiella kuromo</i>	26.46 ± 3.05	32.76 ± 0.46	27.33 ± 3.60	37.45 ± 2.02
<i>Ishige okamurai</i>	36.10 ± 14.81	19.09 ± 5.41	46.30 ± 3.38	53.17 ± 2.28
<i>Ishige sinicola</i>	17.11 ± 4.62	18.92 ± 2.99	30.50 ± 3.34	49.01 ± 8.34
<i>Leathesia difformis</i>	23.72 ± 2.18	31.63 ± 5.52	50.86 ± 0.72	49.19 ± 5.53
<i>Petrospongium rugosum</i>	25.26 ± 2.56	24.51 ± 8.80	14.38 ± 3.25	28.54 ± 5.93
<i>Colpomenia sinuosa</i>	22.89 ± 3.16	21.79 ± 1.03	34.19 ± 2.33	47.20 ± 2.20
<i>Endarachne binghamiae</i>	20.33 ± 0.33	27.07 ± 2.75	44.22 ± 0.00	40.78 ± 1.36
<i>Hydroclathrus clathratus</i>	43.41 ± 2.99	0.85 ± 1.09	48.94 ± 6.50	50.25 ± 1.49
<i>Scytosiphon lomentaria</i>	4.19 ± 0.57	17.98 ± 3.56	29.75 ± 2.11	52.76 ± 6.10
<i>Myelophycus simplex</i>	14.02 ± 3.18	23.45 ± 2.85	47.11 ± 3.65	46.40 ± 4.48
<i>Desmarestia tabacoides</i>	89.27 ± 1.38	74.29 ± 7.94	* ³⁾	*
<i>Undaria pinnatifida</i>	30.80 ± 8.11	49.25 ± 0.62	42.08 ± 5.23	35.43 ± 0.40
<i>Ecklonia cava</i>	33.65 ± 7.12	21.79 ± 3.68	72.24 ± 1.23	33.98 ± 8.78
<i>Laminaria ochotensis</i>	20.80 ± 3.27	13.92 ± 3.20	37.14 ± 1.84	38.11 ± 0.22
<i>Dictyopteris prolifera</i>	46.34 ± 0.92	35.73 ± 5.23	38.01 ± 10.69	48.54 ± 0.83
<i>Dictyota dichotoma</i>	85.85 ± 3.33	76.06 ± 10.06	52.95 ± 0.13	42.20 ± 5.50
<i>Pachydictyon</i> sp.	45.61 ± 15.86	47.68 ± 4.31	43.01 ± 5.49	34.53 ± 1.49
<i>Padina arborescens</i>	- ²⁾	1.34 ± 4.79	36.37 ± 15.50	45.03 ± 0.18
<i>Myagropsis myagroides</i>	13.82 ± 3.96	24.76 ± 0.76	25.28 ± 10.54	44.16 ± 4.13
<i>Hizikia fusiformis</i>	63.94 ± 13.52	39.72 ± 4.31	39.19 ± 2.20	52.20 ± 1.54
<i>Sargassum coreanum</i>	27.52 ± 12.25	26.42 ± 8.16	73.94 ± 5.40	39.81 ± 1.76
<i>Sargassum fulvellum</i>	22.03 ± 0.00	29.07 ± 3.97	46.68 ± 0.48	38.42 ± 5.05
<i>Sargassum horneri</i>	28.78 ± 0.23	30.53 ± 5.00	38.48 ± 2.15	48.54 ± 3.03
<i>Sargassum piluliferum</i>	27.31 ± 4.98	24.11 ± 10.52	37.22 ± 10.64	58.20 ± 9.31
<i>Sargassum siliquastrum</i>	9.88 ± 6.50	29.07 ± 7.42	75.78 ± 2.20	40.62 ± 1.32
<i>Sargassum thunbergii</i>	18.75 ± 7.40	8.22 ± 3.99	21.83 ± 1.71	36.43 ± 10.23

¹⁾Samples are the same as in Table 2.

²⁾Not detected. ³⁾Not determined.

Table 4. Growth inhibition activity of methanolic and aqueous extracts (100 µg/mL) of Jeju seaweeds on HeLa cells

Scientific name	HeLa cell growth inhibition rate (%)			
	20 ME ¹⁾	70 ME	20 AE	70 AE
Green seaweeds				
<i>Monostroma nitidum</i>	- ²⁾	-	18.28 ± 4.54	17.20 ± 3.14
<i>Enteromorpha compressa</i>	40.79 ± 1.52	72.00 ± 4.89	35.45 ± 1.66	20.37 ± 0.23
<i>Enteromorpha intestinalis</i>	33.52 ± 1.18	34.19 ± 6.48	24.81 ± 0.64	25.44 ± 2.96
<i>Enteromorpha linza</i>	24.67 ± 2.24	23.52 ± 2.37	8.57 ± 4.92	12.76 ± 3.74
<i>Enteromorpha</i> sp.	43.33 ± 1.07	49.74 ± 0.31	11.90 ± 1.48	11.38 ± 9.53
<i>Ulva conglobata</i>	16.56 ± 1.99	14.76 ± 5.05	24.89 ± 2.91	19.20 ± 4.76
<i>Ulva pertusa</i>	22.92 ± 1.74	31.10 ± 5.34	21.29 ± 4.12	7.59 ± 2.68
<i>Chaetomorpha linum</i>	20.65 ± 0.90	12.60 ± 2.84	32.96 ± 5.15	34.14 ± 3.42
<i>Codium contractum</i>	30.54 ± 0.84	51.81 ± 4.61	19.59 ± 4.65	23.38 ± 4.49
<i>Codium fragile</i>	47.14 ± 3.20	33.72 ± 3.09	21.26 ± 1.64	25.81 ± 1.87
Brown seaweeds				
<i>Papenfussiella kuromo</i>	27.00 ± 4.80	20.75 ± 0.56	8.60 ± 1.07	28.61 ± 2.17
<i>Ishige okamurai</i>	5.12 ± 1.56	18.15 ± 1.97	15.63 ± 0.59	19.20 ± 0.64
<i>Ishige sinicola</i>	15.65 ± 1.46	17.12 ± 5.00	4.19 ± 1.00	15.98 ± 1.73
<i>Leathesia difformis</i>	9.53 ± 1.90	25.59 ± 4.17	24.13 ± 4.53	28.32 ± 6.75
<i>Petrospongium rugosum</i>	6.39 ± 1.18	21.72 ± 2.43	30.05 ± 3.10	16.81 ± 6.20
<i>Colpomenia sinuosa</i>	9.82 ± 12.15	24.58 ± 12.09	3.38 ± 1.09	28.68 ± 0.23
<i>Endarachne binghamiae</i>	23.19 ± 1.74	32.05 ± 2.53	2.90 ± 1.10	2.84 ± 0.91
<i>Hydroclathrus clathratus</i>	34.63 ± 0.25	32.16 ± 3.74	1.43 ± 1.09	28.68 ± 0.23
<i>Scytosiphon lomentaria</i>	18.51 ± 3.03	16.56 ± 4.44	-	7.09 ± 0.00
<i>Myelophycus simplex</i>	16.56 ± 4.10	19.94 ± 1.68	25.81 ± 1.32	11.79 ± 1.64
<i>Desmarestia tabacoides</i>	90.93 ± 1.62	71.76 ± 9.12	* ³⁾	*
<i>Undaria pinnatifida</i>	22.32 ± 7.30	53.14 ± 8.20	5.35 ± 2.55	1.68 ± 1.01
<i>Ecklonia cava</i>	48.57 ± 8.03	25.33 ± 2.18	38.24 ± 0.23	-
<i>Laminaria ochotensis</i>	6.04 ± 3.37	6.20 ± 2.58	-	6.77 ± 2.11
<i>Dictyopteris prolifera</i>	35.77 ± 0.12	21.89 ± 6.04	8.14 ± 1.07	28.61 ± 2.17
<i>Dictyota dichotoma</i>	81.37 ± 1.12	72.25 ± 2.62	6.84 ± 1.26	22.04 ± 5.92
<i>Pachydictyon</i> sp.	63.74 ± 0.64	35.95 ± 6.11	3.70 ± 1.31	18.08 ± 5.04
<i>Padina arborescens</i>	11.32 ± 1.85	14.02 ± 0.06	7.35 ± 4.56	10.60 ± 1.78
<i>Myagropsis myagroides</i>	14.23 ± 2.68	-	14.98 ± 2.77	20.70 ± 0.42
<i>Hizikia fusiformis</i>	25.02 ± 8.20	26.61 ± 6.85	6.98 ± 3.84	16.91 ± 6.52
<i>Sargassum coreanum</i>	56.12 ± 4.36	48.72 ± 9.22	78.74 ± 12.12	16.74 ± 2.87
<i>Sargassum fulvellum</i>	20.79 ± 0.75	8.46 ± 3.61	19.00 ± 5.69	14.52 ± 3.70
<i>Sargassum horneri</i>	17.05 ± 9.78	10.00 ± 1.93	24.26 ± 3.51	18.61 ± 1.16
<i>Sargassum piluliferum</i>	6.20 ± 2.74	11.23 ± 1.81	13.87 ± 4.72	22.17 ± 0.55
<i>Sargassum siliquastrum</i>	12.56 ± 0.44	13.57 ± 1.37	84.02 ± 1.20	20.34 ± 8.23
<i>Sargassum thunbergii</i>	17.51 ± 4.77	13.42 ± 2.92	-	12.76 ± 3.74

¹⁾Samples are the same as in Table 2.

²⁾Not detected. ³⁾Not determined.

twenty brown seaweeds and nine green seaweeds species on inhibited the growth of CT26 cells by over 50%. Especially, the *D. tabacoides* 20 ME, *S. siliquastrum* 20 AE, *D. dichotoma* 20 ME and *S. coreanum* 20 AE showed highly potent inhibitory effects with 90.93%, 84.02%, 81.37% and 78.74% growth inhibition on HeLa, respectively. Among all the seaweed extracts, tested *D. tabacoides* 20 ME (87.23%), *E. cava* 20 AE (85.72%), *D. dichotoma* 20 ME (84.45%) and *Codium fragile* 20 AE (73.41%) exhibited the greatest growth inhibition on CT26 cells.

The growth inhibitory potential of the four tumor cells were over 80% in the 20 ME of *D. tabacoides* and *D. dichotoma* among brown seaweeds. The growth inhib-

itory potential of U937, HL60 and HeLa cell were over 70% in the 20 AE of *S. siliquastrum* and *S. coreanum* among brown seaweeds. The growth inhibitory potential of U937, HL60 and CT26 cell were over 70% in the 20 AE of *E. cava*.

Taking into consideration all of the results regarding the tumor cell growth inhibitory activities of the algal extracts, we have tried to analyze significant differences between the extraction solvents (methanol and water) and between the extracting temperatures (20 and 70°C). As shown in Table 6, there was no significant difference between extracts at either temperature. However a significant difference between the extraction solvents was observed in HL60, CT26 and HeLa cell. Aqueous ex-

Table 5. Growth inhibition activity of methanolic and aqueous extracts (100 µg/mL) of Jeju seaweeds on CT26 cells

Scientific name	CT26 cell growth inhibition rate (%)			
	20 ME ¹⁾	70 ME	20 AE	70 AE
Green seaweeds				
<i>Monostroma nitidum</i>	29.93 ± 10.49	52.74 ± 6.20	46.14 ± 13.01	51.12 ± 8.94
<i>Enteromorpha compressa</i>	47.67 ± 4.84	66.85 ± 1.74	46.42 ± 4.66	47.17 ± 2.78
<i>Enteromorpha intestinalis</i>	38.63 ± 6.58	59.45 ± 0.58	32.87 ± 5.16	35.11 ± 0.40
<i>Enteromorpha linza</i>	44.59 ± 12.49	54.86 ± 1.07	54.63 ± 1.99	25.35 ± 5.46
<i>Enteromorpha</i> sp.	48.01 ± 5.33	53.29 ± 9.59	69.83 ± 1.80	58.50 ± 2.68
<i>Ulva conglobata</i>	27.33 ± 11.78	50.14 ± 3.68	53.37 ± 2.78	32.44 ± 1.99
<i>Ulva pertusa</i>	40.34 ± 8.33	42.26 ± 9.20	40.73 ± 3.38	36.94 ± 5.96
<i>Chaetomorpha linum</i>	31.85 ± 4.17	25.67 ± 0.89	61.50 ± 2.13	52.32 ± 3.48
<i>Codium contractum</i>	32.33 ± 6.20	49.73 ± 1.16	65.61 ± 0.74	62.78 ± 0.79
<i>Codium fragile</i>	42.05 ± 0.19	35.27 ± 0.68	73.41 ± 6.21	69.36 ± 4.90
Brown seaweeds				
<i>Papenfussiella kuromo</i>	37.60 ± 8.04	53.22 ± 3.39	55.55 ± 6.65	53.44 ± 0.30
<i>Ishige okamurai</i>	30.89 ± 4.36	37.05 ± 5.33	50.46 ± 3.68	61.04 ± 0.82
<i>Ishige sinicola</i>	34.59 ± 3.39	37.88 ± 4.55	50.29 ± 0.98	46.65 ± 3.02
<i>Leathesia difformis</i>	44.59 ± 4.36	47.19 ± 2.62	58.50 ± 3.67	60.32 ± 2.28
<i>Petrospongium rugosum</i>	42.12 ± 1.07	35.41 ± 7.85	48.74 ± 1.99	46.00 ± 2.28
<i>Colpomenia sinuosa</i>	47.40 ± 3.49	35.55 ± 11.72	50.98 ± 7.15	65.03 ± 2.78
<i>Endarachne binghamiae</i>	27.40 ± 2.42	31.23 ± 6.01	39.13 ± 11.04	46.59 ± 0.65
<i>Hydroclathrus clathratus</i>	50.55 ± 4.07	53.63 ± 6.10	65.45 ± 3.38	64.12 ± 2.09
<i>Scytosiphon lomentaria</i>	19.11 ± 6.88	21.37 ± 0.19	53.29 ± 4.25	72.60 ± 1.14
<i>Myelophycus simplex</i>	27.33 ± 9.20	29.59 ± 4.65	24.80 ± 6.46	39.19 ± 1.80
<i>Desmarestia tabacoides</i>	87.23 ± 3.23	77.32 ± 4.98	* ²⁾	*
<i>Undaria pinnatifida</i>	44.45 ± 1.26	59.45 ± 0.39	28.73 ± 3.84	27.63 ± 5.40
<i>Ecklonia cava</i>	51.44 ± 0.87	35.21 ± 2.71	85.72 ± 0.90	41.04 ± 10.13
<i>Laminaria ochotensis</i>	37.19 ± 1.84	66.92 ± 4.17	45.20 ± 8.67	58.38 ± 8.99
<i>Dictyopteris prolifera</i>	65.21 ± 2.52	57.74 ± 7.65	29.78 ± 9.73	52.32 ± 3.28
<i>Dictyota dichotoma</i>	84.45 ± 2.03	50.48 ± 5.91	62.08 ± 1.59	55.76 ± 0.40
<i>Pachydictyon</i> sp.	48.08 ± 9.02	45.14 ± 8.21	50.77 ± 5.26	30.41 ± 7.85
<i>Padina arborescens</i>	20.62 ± 0.29	23.63 ± 4.75	50.40 ± 5.89	50.35 ± 4.50
<i>Myagropsis myagroides</i>	27.81 ± 6.35	40.89 ± 9.59	53.37 ± 3.58	50.42 ± 1.79
<i>Hizikia fusiformis</i>	67.95 ± 2.13	41.03 ± 1.07	52.25 ± 7.15	41.15 ± 6.75
<i>Sargassum coreanum</i>	38.97 ± 15.80	48.90 ± 14.92	7.87 ± 0.20	14.96 ± 3.67
<i>Sargassum fulvellum</i>	25.75 ± 2.97	24.99 ± 9.78	46.77 ± 3.58	20.22 ± 9.53
<i>Sargassum horneri</i>	35.82 ± 8.56	33.15 ± 1.74	50.63 ± 0.30	23.10 ± 4.67
<i>Sargassum piluliferum</i>	41.51 ± 6.59	33.01 ± 1.74	51.05 ± 11.02	51.47 ± 0.10
<i>Sargassum siliquastrum</i>	36.85 ± 6.38	34.59 ± 2.23	27.74 ± 7.85	11.94 ± 4.57
<i>Sargassum thunbergii</i>	41.99 ± 1.65	62.12 ± 6.49	35.90 ± 6.29	62.60 ± 1.39

¹⁾Samples are the same as in Table 2.

²⁾Not determined.

Table 6. The overall growth inhibition activity of methanolic and aqueous extracts on tumor cell lines

Sample ¹⁾	U937	HL60	CT26	HeLa
Methanolic extract	33.42 ± 2.02 ^{NS2)}	29.05 ± 2.08 ^{a3)}	42.03 ± 1.55 ^a	25.69 ± 2.12 ^b
Aqueous extract	29.38 ± 1.80	44.72 ± 1.41 ^b	47.68 ± 1.83 ^b	18.03 ± 1.75 ^a
Extracts at 20°C	33.43 ± 2.06	36.71 ± 2.22 ^a	44.75 ± 1.76 ^a	21.88 ± 2.24 ^a
Extracts at 70°C	29.37 ± 1.74	36.94 ± 1.8 ^a	44.96 ± 1.6 ^a	21.84 ± 1.72 ^a

¹⁾The mean of tumor cell growth inhibitory activity values of total methanolic extracts at 20 and 70°C, total aqueous extracts at 20 and 70°C, total methanolic and aqueous extracts at 20°C and total methanolic and aqueous extracts at 70°C.

²⁾Not significant.

³⁾Means with different letters within a column are significantly different (p<0.05).

tracts were superior to methanolic extracts for inhibiting cell growth of HL60 and CT26, and the opposite was true for HeLa.

The activities reported in methanolic extracts can be

attributed to the pigments and phlorotannis of the brown seaweeds (19,20). The predominant pigments of brown seaweeds have been reported to be fucoxanthine and chlorophyll (21,22). The polyphenolic compounds of the

brown seaweeds are referred to as phlorotannins and those are readily soluble in polar solvents like methanol and water (19). This suggests that the tumor cell growth inhibitory activities of the aqueous extracts of brown seaweeds can be attributed to the phlorotannins. Previous studies have reported that fucoxanthin inhibits the growths of human neuroblastoma GOTO cells, human leukemia cells, and prostate cancer cells (21-24). Several phlorotannins of brown seaweeds have been identified for their potential bioactivities such as antioxidant, antibacterial and antihyaluronase (25-36). Therefore these results suggest that further intensive studies on the anticancer activities of those extracts are needed. The aqueous extract of *Codium contractum* exhibited the highest tumor cell growth inhibition activity (71.80%) of the green seaweed extracts. This activity can not be due to the chlorophylls, because they are hydrophobic and relatively insoluble in water. Thus, it might be suggested that it can be due to some other water soluble constituents of the seaweed such as protein or low molecular weight polysaccharide. Seaweeds contain many different kinds of compounds which could possess potent anticancer activity. As has been reported previously, other seaweed compounds such as β -carotin and lutein as well as chlorophyll related compounds have documented high anticancer activities (37). Those compounds quench radical species and thereby reduce cancer cell formation *in vitro* and *in vivo* (38). Hence, it is clear that the similar kinds of compounds may be associated with the high anti-proliferative effect of the present study.

In conclusion, methanolic and aqueous extracts from 37 marine algal species collected from Jeju Island were evaluated for their cytotoxic activities against four tumor cells (U937, HL60, CT26 and HeLa). Most seaweed species tested in this study showed potential anticancer activities. Of the seaweed species *D. tabacoides*, *D. dichotoma*, *S. siliquastrum* and *S. coreanum* showed excellent cytotoxic activities. However, further studies are essential to purify anticancer active compounds to elucidate relationships between structure and activity which might help with future drug design. Therefore, seaweeds present in Jeju Island are possible candidates for future anticancer drug discovery.

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