

Differential Functions of Ras for Malignant Phenotypic Conversion

Aree Moon

College of Pharmacy, Duksung Women's University, Seoul 132-714, Korea

(Received January 10, 2006)

Among the effector molecules connected with the group of cell surface receptors, Ras proteins have essential roles in transducing extracellular signals to diverse intracellular events, by controlling the activities of multiple signaling pathways. For over 20 years since the discovery of Ras proteins, an enormous amount of knowledge has been accumulated as to how the proteins function in overlapping or distinct fashions. The signaling networks they regulate are very complex due to their multiple functions and cross-talks. Much attention has been paid to the pathological role of Ras in tumorigenesis. In particular, human tumors very frequently express Ras proteins constitutively activated by point mutations. Up to date, three members of the Ras family have been identified, namely H-Ras, K-Ras (A and B), and N-Ras. Although these Ras isoforms function in similar ways, many evidences also support the distinct molecular function of each Ras protein. This review summarizes differential functions of Ras and highlights the current view of the distinct signaling network regulated by each Ras for its contribution to the malignant phenotypic conversion of breast epithelial cells. Four issues are addressed in this review: (1) Ras proteins, (2) membrane localization of Ras, (3) effector molecules downstream of Ras, (4) Ras signaling in invasion. In spite of the accumulation of information on the differential functions of Ras, much more remains to be elucidated to understand the Ras-mediated molecular events of malignant phenotypic conversion of cells in a greater detail.

Key words: Ras, Tumorigenesis, Signaling, Invasion, Migration

INTRODUCTION

Normal cell proliferation in multicellular organism is tightly controlled to ensure that it occurs only when it is required. Break-down of normal growth regulation often leads to cancer. The Ras proteins are some of the first proteins identified that can regulate cell growth. Aberrant Ras function has been well acknowledged to be strongly associated with human cancer development. Biochemical and molecular approaches have been used to inhibit Rasmediated oncogenic activities. A major drawback of these approaches is in vivo toxicity. Since Ras proteins are central molecules responsible for cell survival, proliferation and other cellular processes, inhibition of general Ras activities can be detrimental not only to cancer cells but also to normal cells. Therefore, it is critical to identify Ras downstream signaling molecules that are required for malignant cancer cell behavior but less critical for normal cell functions. The functions and signaling pathways of

Ras proteins and therapeutic approaches to target Ras pathways are now understood in great detail as summarized in several excellent reviews (Magee and Marshall, 1999; Shields *et al.*, 2000; Downward, 2003; Hancock, 2003). The present review is aimed to highlight the current understanding of the differential functions of Ras proteins in the malignant phenotypic conversion of epithelial cells.

The following five themes are addressed in this review: (1) Ras proteins. It is generally assumed that there are three human ras genes that code for Ras proteins and there have been indications for different molecular functions of these proteins. The activation of different Ras isoforms can have distinct biochemical consequences for cells, rationalizing the mutation of specific Ras isoforms in different human tumors. (2) Membrane localization of Ras proteins. Since a considerable body of evidence has suggested that functional differences among the Ras isoforms could be due to variations in plasma membrane microlocalization, it is important to identify a link between interaction of Ras with the plasma membrane and the functional differences in Ras signaling. (3) Ras downstream effector molecules. Ras proteins exert biological

Correspondence to: Aree Moon, College of Pharmacy, Duksung Women's University, Seoul 132-714, Korea Tel: 82-2-901-8394 Fax: 82-2-901-8386

E-mail: armoon@duksung.ac.kr

activities by stimulating a multitude of downstream signaling cascades. This review summarizes the role of signaling molecules in Ras-induced cellular responses and the differential regulation of signaling pathways by Ras isoforms. (4) Ras signaling in invasion. It is well appreciated that the activated Ras proteins contribute significantly to several aspects of the malignant phenotypes including invasiveness, migration and angiogenesis. This review covers current understanding of tumor invasion with a major focus on the contribution of distinct signaling network exerted by different Ras isoforms to invasion and migration of breast epithelial cells.

(1) Ras proteins

The Ras proteins are guanine nucleotide-binding proteins that play an essential role in transducing extracellular signals to diverse cellular responses, including cell proliferation and differentiation (Boguski and McCormick, 1993; Campbell *et al.*, 1998). One of the most frequent defects in human cancer is the uncontrolled activation of the Ras-signaling pathways (Barbacid, 1987). Human tumors frequently express Ras proteins that have been activated by point mutations — about 20% of all tumors have undergone an activating mutation in one of the *ras* genes (Barbacid, 1987; Bos, 1989).

The Ras family consists of three identified members: Harvey-Ras (H-Ras), Kirsten-Ras (K-RasA and K-RasB), and N-Ras, proteins of 188-189 amino acids with a molecular weight of 21 kDa. Activating point mutation has been most frequently found in K-Ras (about 85% of total), then N-Ras (about 15%), then H-Ras (less than 1%) (Downward, 2003). While the N-terminal 85 amino acids

are identical and the middle 80 amino acids contain 85-90% homology between Ras proteins, the C-terminal sequence, so-called the hypervariable region (HVR), is highly divergent (Barbacid, 1987; Boguski and McCormick, 1993). Homology of HVR among the four Ras proteins was shown to be 4-15% (Shield *et al.*, 2000; Prior and Hancock, 2001). The sequence divergence between the Ras proteins is shown in Fig. 1.

An important question on Ras biology is whether the ubiquitously expressed, almost identical Ras isoforms have distinct functions. There are differences in the signal transduction pathways induced by Ras proteins, supporting unique functions of Ras family members (Carbone et al., 1991; Umanoff et al., 1995; Johnson et al., 1997; Koera et al., 1997; Yan et al., 1998). Although these Ras proteins share common signaling pathways leading to similar cellular responses, cell-type specific differences in the transforming potential between Ras proteins were also reported. One member of the ras family appears to be the preferred "target gene" in particular tumors. The single point mutation at amino acid residue 12 (glycine to aspartate) of H-Ras is more often found in mammary carcinoma, whereas the same mutation of N-Ras is detected in teratocarcinoma and leukemia (Franks and Teich, 1997). While H-Ras is more transforming than N- or K-Ras in murine fibroblasts, N-Ras is more transforming in human haemopoietic cells (Maher et al., 1995). Differences in the transforming activitites of Ras proteins appear to be due to unique sequences between amino acids 84 and 143 in Ras proteins (Maher et al., 1995). H-Ras, but not N-Ras, induces invasive and migrative phenotypes in human breast epithelial cells while both H-

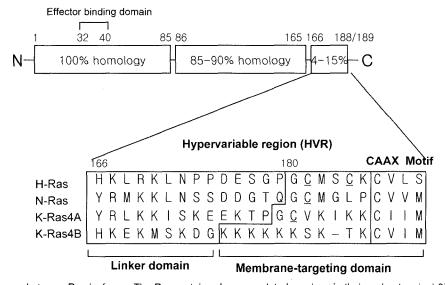


Fig. 1. Sequence divergence between Ras isoforms. The Ras proteins show complete homology in their amino-terminal 85 residues. The greatest sequence divergence is seen in the carboxy-terminal residues of 165-188/189, a region called hypervariable region (HVR). Cysteine palmitoylation sites are underlined.

Ras and N-Ras induce transformed phenotypes such as focus-forming morphological change and anchorage-independent growth (Moon et al., 2000; Kim et al., 2003).

(2) Membrane localization of Ras proteins

Since dynamic microdomain localization has implications for understanding how signaling complexes are assembled and disassembled in response to particular stimuli, it is important to identify a link between interaction of Ras with the plasma membrane (Magee and Marshall, 1999) and the functional differences in Ras signaling. The differences appear to result, at least in part, from differences in the mechanisms of membrane attachment of the three Ras isoforms (Yan *et al.*, 1998; Booden *et al.*, 2000).

The biological activity of Ras requires a proper localization to the inner surface of the plasma membrane (Willumsen et al., 1984). Several studies, however, demonstrate that Ras is also activated on and transmits signals from the endoplasmic reticulum and Golgi apparatus, suggesting that the plasma membrane may not be the exclusive platform from which Ras regulates signaling (Chiu et al., 2002; Bivona et al., 2003). Accumulating evidence has revealed differences in the way that Ras proteins are routed to the plasma membrane (reviewed in Hancock, 2003). Such differences seem reasonable since the HVR contains the signals responsible for the correct localization of Ras (Fig. 1). These signals include the Cterminal CAAX box (in which A = aliphatic amino acid and X = serine or methionine) (Casey et al., 1989; Gutierrez et al., 1989) and palmitoylation of two cysteine residues (Cys¹⁸¹ and Cys¹⁸⁴) in H-Ras, one cysteine (Cys¹⁸¹) in N-Ras, and a polybasic sequence of multiple lysines (Lys¹⁷⁵⁻¹⁸⁰) in K-Ras (Hancock et al., 1990; Kato et al., 1992). These moieties and possibly the entire HVR sequence confer membrane-anchoring capacity on Ras and are also involved in the membrane trafficking of Ras proteins (Choy et al., 1999).

Given that correct post-translational modification of Ras proteins is required for their biological activities, several strategies have been developed to inhibit the membrane anchoring of Ras proteins. The enzymes involved in this processing have become potential targets for therapeutic intervention (Seabra *et al.*, 1998; Cox and Der, 1997; Hancock *et al.*, 1989; Hancock *et al.*, 1990), the most common being the design of compounds that mimic the CAAX motif and compete for binding to farnesyltransferase (Cox and Der, 1997; Oliff, 1999; Downward, 2003). These farnesyltransferase inhibitors (FTI) as tumor therapeutic drugs, however, turned out to be selective for H-Ras since K-Ras and, to a lesser extent, N-Ras can also be modified by geranylgeranyltransferase.

The interactions of Ras proteins with plasma membrane can differ from one isoform to another due to their different

membrane anchoring moieties. Several recent studies (Niv et al., 2002; Prior et al., 2003; Roy et al., 1999; Parton and Hancock, 2004) revealed that differently anchored Ras proteins display different interactions with lipid rafts (Simons and Toomre, 2000), which are cholesterol/ sphingolipid-enriched microdomains that dynamically organize specific membrane proteins. H-Ras (wild type) but not K-Ras is significantly concentrated in cholesteroldependent rafts (Niv et al., 2002). Differential activation of Raf-1 and phosphatidylinositol 3-kinase (PI3K) by K-Ras and H-Ras (Yan et al., 1998) may be explained by the differences in the microdomain localization of these Ras proteins (Yan et al., 1998; Prior et al., 2001; Matallanas et al., 2003). Mutation of H-Ras C terminus changed effector pathway utilization (Booden et al., 2000), suggesting a role of the lipidated C-terminai in the biological functions of Ras proteins. These studies demonstrate that the localization of Ras proteins to different microdomains of the plasma membrane may be critical for signaling specificity. It has been demonstrated that H-Ras and N-Ras differentially regulate Rac1 activity which plays a key role in invasion and migration while they do not vary in their abilities to activate Raf-1 and PI3K in MCF10A cells (Shin et al., 2005). Detailed comparison of microlocalization of H-Ras and N-Ras needs to be performed to investigate whether differential microlocalization in plasma membrane accounts for the distinct regulation of signaling pathway and invasive phenotype by the highly homologous H-Ras and N-Ras proteins.

(3) Ras downstream effector molecules

Ras proteins exert their biological effects by activating several downstream effector molecules including Raf, PI3K and Ral (Marshall, 1996; Campbell et al., 1998). The first mammalian effector of Ras is the serine/threonine kinase Raf (Leevers et al., 1994). GTP-Ras binds to Raf and this interaction causes Raf relocation to the plasma membrane which seems to be critical for its activation (Marais et al., 1995). Ras-stimulated Raf activates the downstream kinase MAPK/ERK (MEK), which in turn phosphorylates extracellular signal-regulated kinases (ERKs) (Kyriakis et al., 1992; Alessi et al., 1994). ERKs can be transported into the nucleus following activation, stimulating nuclear transcription factors such as Ets family Elk1, Fos and Jun. The Raf-MEK-ERK pathway can mediate Ras-induced cell survival and proliferation by promoting cell-cycle progression (Yordy et al., 2000; Pruitt and Der, 2001; Vaudry et al., 2002).

In addition to the Raf-MEK-ERK pathway, the biological effects of Ras proteins are exerted through the activation of PI3K (Downward, 1998). Ras can directly interact with the catalytic subunit of PI3K and activate the molecule (Rodriguez-Viciana et al., 1994; Pacold et al., 2000). PI3K

controls the activity of a large number of downstream molecules by phosphorylating phosphatidylinositol 4, 5-bisphosphate to produce phosphatidylinositol 3, 4, 5-triphosphate which is a second messenger that binds to a number of proteins. Much attention has focused on the activation of Akt (also called protein kinase B) which has a strong anti-apoptotic function (Romashkova and Makarov, 1999) and seems to be an important mediator of Rasgenerated survival signal (Downward, 1998; Datta *et al.*, 1999; Khwaja *et al.*, 1997; Khwaja, 1999). Ras also activates three GDP-GTP exchange factors (RalGDS, RGL and RGL2/Rlf) to stimulate Ral (Feig *et al.*, 1996; Wolthuis and Bos, 1999).

The small GTP-binding protein Rac has been shown to be critical for the mitogenic and oncogenic effects of Ras by promoting actin cytoskeletal reorganization leading to membrane ruffling, lamellipodia formation, cell migration and invasion (Hancock *et al.*, 1990; Ridley *et al.*, 1992; Qiu *et al.*, 1995; Joneson *et al.*, 1996; Etienne-Manneville and Hall, 2002). Rac can be responsible for Ras-induced changes in the actin cytoskeleton associated with developing invasive carcinoma of mammary epithelial cells by modulating motility and invasion (Nobes *et al.*, 1995). Rac activation, which can occur through PI3K-dependent and PI3K-independent pathways, is important in Ras-induced transformation and invasion (Lambert *et al.*, 2002; Malliri *et al.*, 2002).

The signaling order between Rac and PI3K has been controversial in different cell systems. PI3K acts upstream of Rac1 in pathways for membrane ruffling, chemotaxis (Reif *et al.*, 1996; Welch *et al.*, 2003) and inducing malignant phenotype of signet-ring cell carcinoma (Xu *et al.*, 2003). Rac1 has been shown to act upstream of PI3K to promote cellular motility and invasiveness by disrupting the normal polarization of mammary epithelial cells (Keely *et al.*, 1997; Sachdev *et al.*, 2002). In H-Ras-activated MCF10A cell system, PI3K pathway was dependent on Rac activity while Rac activity was not affected by PI3K inhibition, suggesting that Rac1 may lie upstream of PI3K (Shin *et al.*, 2005).

The reason why the Ras proteins need many down-stream effector molecules has been speculated (Shields et al., 2000). Ras exerts a diverse spectrum of cellular responses depending on different cell systems. Utilization of distinct sets of multiple signaling pathways is required for the complex nature of the transformed phenotype exerted by oncogenic Ras, such as uncontrolled proliferation, loss of anchorage-dependent growth, invasion, metastasis and angiogenesis.

Despite the fact that the amino acid sequence corresponding to the effector binding loop, which spans residues 32-40, is identical among Ras proteins (Fig. 1), recent studies have demonstrated that the three Ras

isoforms can differentially activate the effector molecules. There are differences in the signal transduction pathways induced by Ras proteins, suggesting unique functions of different Ras family members at the molecular level (Carbone et al., 1991; Voice et al., 1999). K-Ras activates Rac more effectively than H-Ras (Walsh and Bar-Sagi, 2001) and is a more potent activator of membranerecruited Raf-1 than H-Ras (Yan et al., 1998). H-Ras activates PI3K more potently than K-Ras (Yan et al., 1998). Enhanced motility induced by H-Ras (Kim et al., 2003) suggested that H-Ras might be a more effective activator of the Rac pathway compared to N-Ras in MCF10A cells. Consistently, marked activation of Rac-MKK3/6-p38 pathway was exerted by H-Ras, but not by N-Ras, in MCF10A cells while the Raf-MEK-ERKs and PI3K-Akt pathways were activated by both H-Ras and N-Ras (Shin et al., 2005).

(4) Ras signaling in invasion

Elevated levels of the Ras protein have been found in 60-70% of human primary breast carcinomas (Clair *et al.*, 1987), although Ras mutations are infrequent in human breast cancer. Ras expression has been suggested as a marker for tumor aggressiveness of breast cancer, including the degrees of invasion to fat tissues, infiltration into lymphatic vessels, and tumor recurrence (Clair *et al.*, 1987; Watson *et al.*, 1991; Clark and Der, 1995). The activated Ras proteins contribute to malignant phenotypes including invasiveness and angiogenesis (Shields *et al.*, 2000; Downward, 2003).

Recent reports investigated the role of Ras and Ras-dependent signaling pathways in cell invasion and migration. The Ras-Pl3K-Akt pathway, which induces invasion and metastasis, downregulates RhoB, a suppressor of transformation, invasion and metastasis (Jiang *et al.*, 2004). The Rac-MKK3/6-p38 pathway, activated by H-Ras, but not by N-Ras, is critical to the H-Ras-induced invasive and migrative phenotypes in MCF10A breast epithelial cells (Kim *et al.*, 2003; Shin *et al.*, 2005). Anti-migratory and anti-invasive effect of somatostatin involves Rac, Pl3K, and ERKs pathways in human neuroblastoma cells (Pola *et al.*, 2003). The Ras-MEKK1 pathway mediates lysophosphatidic acid-induced ovarian cancer cell migration (Bian *et al.*, 2004).

An essential part of the metastatic process includes degradation of the basement membrane and the stromal extracellular matrix, which allows breast cells to migrate into neighboring tissues. Members of matrix metalloproteinase (MMP) family, especially, MMP-2 (72 kDa type IV collagenase, gelatinase A) and MMP-9 (92 kDa type IV collagenase, gelatinase B), have been shown to be deeply involved in tumor invasion and metastasis formation (Ura et al., 1989; Stetler-Stevenson, 1990; Liotta et al., 1991;

Tryggvason, 1993). In rat and human embryonic fibroblasts, H-Ras mediated transformation and invasiveness were shown to be associated with enhanced expression of MMP-9 mRNA and protein (Bernhard et al., 1990, 1994). In human breast epithelial cells, however, H-Ras-induced invasive phenotype is associated more closely with the expression of MMP-2 rather than MMP-9 (Moon et al., 2000), demonstrating that Ras-mediated cellular responses differ between epithelial cells and fibroblasts (Oldham et al., 1996). A direct correlation between high level of MMP-2 expression and an increased invasive capacity of tumor cell lines has been demonstrated both in vitro and in vivo (Sato et al., 1992; Stetler-Stevenson et al., 1996; Bodey et al., 2001). Mounting evidence demonstrates a role for MMP-2 in the invasion of breast cancer cells and risk for a relapse in breast cancer patients (Moon et al., 2000; Talvensaari-Mattila et al., 2001; Nakopoulou et al., 2003).

Ras signaling pathways responsible for regulating upregulation of MMP-2 and MMP-9 causing invasive phenotype have been elucidated. The role of ERKs pathway in regulation of MMP-9 and invasion was demonstrated (Simon et al., 1996; Gum et al., 1997). The p38 pathway was also shown to regulate MMP-9 expression and in vitro invasion (Simon et al., 1998; Simon et al., 2001). Recent reports show that induction of MMP-9 expression is mediated by Ras-ERK and PI3K-Akt pathways (Chung et al., 2004; Moon et al., 2004). Although the signaling pathways regulating MMP-2 expression have been relatively poorly elucidated thus far, a recent study showed that insulin-like growth factor-I up-regulated MMP-2 expression via PI3K-Akt signaling while concomitantly transmitting a negative regulatory signal via the Raf-ERK pathway in lung carcinoma cells (Zhang et al., 2004). Activation of Rac, PI3K, ERKs and p38 is crucial for H-Ras-induced up-regulation of MMP-2 and MMP-9 which play key roles in invasion and migration in MCF10A cells (Kim *et al.*, 2003; Shin *et al.*, 2005). The MKK6-p38 pathway alone was able to induce MMP-2 expression as well as invasive/migrative phenotype in MCF10A cells (Shin *et al.*, 2005). Activation of p38 pathway did not induce MMP-9 expression, suggesting that MMP-2 up-regulation by MKK3-p38 signaling plays a key role in invasion of breast epithelial cells. Based on the knowledge obtained in the MCF10A human breast epithelial cell system, a working model was proposed for the differential regulation of signaling pathways by H-Ras and N-Ras leading to proliferation, anchorage-independent growth, invasion and migration (Fig. 2).

Although MMP-2 and MMP-9 share structural and catalytic similarities, previous studies suggest that transcription of MMP-2 and MMP-9 may be independently regulated due to distinct arrays of cis-acting elements in the promoter. While the regulation of MMP-9 gene expression has been extensively studied (Sato, 1993; Sato and Seiki, 1993), the molecular basis of the regulating mechanisms for MMP-2 expression has not been well identified at the transcriptional level. AP-1 was demonstrated to play a crucial role in tumorigenesis especially in breast cancer (Li et al., 1997, 1998, 2000; Ludes-Meyers et al., 2001). Although MMP-2 was considered to be an AP-1-unresponsive gene in many cell types (Brown et al., 1990; Tryggvason et al., 1990; Qin et al., 1999), recent data indicate that a functional AP-1 site mediates MMP-2 transcription in cardiac cells and breast cancer cells (Bergman et al., 2003; Bachmeier et al., 2005). The putative binding elements for p53, AP-1, Ets-1, C/EBP, CREB, PEA3, Sp1, and AP-2 have been found in the region of MMP-2 promoter (Bian and Sun, 1997; Qin et

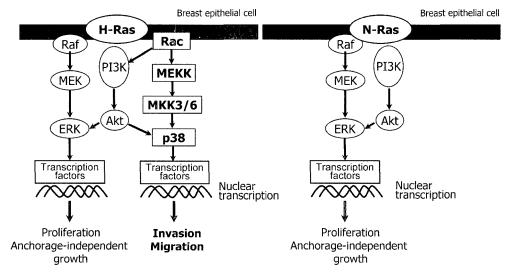


Fig. 2. Signaling pathways activated by H-Ras and N-Ras leading to proliferation and malignant phenotypic conversion in breast epithelial cell system

al., 1999; Bergman et al., 2003). Given that MMP-2 has been suggested as a key factor responsible for H-Rasinduced invasion and migration of breast epithelial cells, further studies need to be performed to characterize which elements are functionally active in the transcriptional activation of MMP-2 gene leading to the malignant phenotypic conversion of breast epithelial cells.

CONCLUSIONS

For over 20 years since Ras was discovered, an enormous amount of knowledge has been accumulated as to how the Ras proteins function. The signaling network they regulate is very complex with multiple functions and cross-talks. The review summarizes some of the current understanding of Ras signaling with a major focus in its contribution to the invasive phenotypic conversion of epithelial cells. Four issues have been addressed in this review. First, Ras family consists of three members: H-Ras, K-Ras (A and B), and N-Ras. The greatest sequence divergence is seen in the C-terminal 25 residues, HVR, which is therefore potentially responsible for the distinct function of each isoform of Ras. Secondly, it is important to identify a differential membrane localization of Ras isofoms since a proper localization to the plama membrane is essential for the biological activity. HVR contains sequences for membrane anchoring of Ras including CAAX motif and lipidated residues. Detailed comparison of microlocalization of Ras isoforms needs to be performed. Thirdly, Ras proteins transduce extracellular signals to diverse cellular functions by stimulating multiple signaling networks. Utilization of the distinct sets of multiple signal transduction pathways regulated by Ras proteins accounts for the differential functions of H-Ras, K-Ras, and N-Ras in cell systems. Lastly, this review summarized recent studies on the differential role of Ras proteins in regulating signal molecules leading to invasion and migration with an emphasis on the Ras-induced up-regulation of MMP-2 and MMP-9. Although much has been known on the differential functions of Ras proteins, much more remains to be elucidated to provide implications on detailed understanding of molecular events for malignant phenotypic conversion of breast cells induced by Ras proteins.

ACKNOWLEDGEMENTS

This work was financially supported by the Ministry of Education and Human Resources Development (MOE), the Ministry of Commerce, Industry and Energy (MOCIE) and the Ministry of Labor (MOLAB) through the fostering project of the Lab of Excellency. The author thanks Ms. Hae-Young Yong and Ms. In-Young Kim for helping prepare the manuscript.

REFERENCES

- Alessi, D. R., Saito, Y., Campbell, D. G., Cohen, P., Sithanandam, G., Rapp, U., Ashworth, A., Marshall, C. J., and Cowley, S., Identification of the sites in MAP kinase kinase-1 phosphorylated by p74raf-1. *EMBO J.*, 13, 1610-1619 (1994).
- Bachmeier, B. E., Albini, A., Vene, R., Benelli, R., Noonan, D., Weigert, C., Weiler, C., Lichtinghagen, R., Jochum, M., and Nerlich, A. G., Cell density-dependent regulation of matrix metalloproteinase and TIMP expression in differently tumorigenic breast cancer cell lines. *Exp. Cell Res.*, 305, 83-98 (2005).
- Barbacid, M., Ras genes. *Annu. Rev. Biochem.*, 56, 779-827 (1987).
- Bergman, M. R., Cheng, S., Honbo, N., Piacentini, L., Karliner, J. S., and Lovett, D. H., A functional activating protein 1 (AP-1) site regulates matrix metalloproteinase 2 (MMP-2) transcription by cardiac cells through interactions with JunB-Fra1 and JunB-FosB heterodimers. *Biochem. J.*, 369, 485-496 (2003).
- Bernhard, E. J., Gruber, S. B., and Muschel, R. J., Direct evidence linking expression of matrix metalloproteinase 9 (92 kDa gelatinase/collagenase) to the metastatic phenotype in transformed rat embryo cells. *Proc. Natl. Acad. Sci. U.S.A.*, 91, 4293-4297 (1994).
- Bernhard, E. J., Muschel, R. J., Hughes, E. N., and M. R., 92,000 gelatinase release correlates with the metastatic phenotype in transformed rat embryo cells. *Cancer Res.*, 50, 3872-3877 (1990).
- Bian, D., Su, S., Mahanivong, C., Cheng, R. K., Han, Q., Pan, Z. K., Sun, P., and Huang, S., Lysophosphatidic Acid stimulates ovarian cancer cell migration *via* a Ras-MEK kinase 1 pathway. *Cancer Res.*, 64, 4209-4217 (2004).
- Bian, J. and Sun, Y., Transcriptional activation by p53 of the human type IV collagenase (gelatinase A or matrix metalloproteinase 2) promoter. *Mol. Cell Biol.*, 17, 6330-6338 (1997).
- Bivona, T. G., Perez De Castro, I., Ahearn, I. M., Grana, T. M., Chiu, V. K., Lockyer, P. J., Cullen, P. J., Pellicer, A., Cox, A. D., and Philips, M. R., Phospholipase Cgamma activates Ras on the Golgi apparatus by means of Ras-GRP1. *Nature*, 7, 694-698 (2003).
- Bodey, B., Bodey, B. Jr., Groger, A. M., Siegel, S. E., and Kaiser, H. E., Invasion and metastasis: the expression and significance of matrix metalloproteinases in carcinomas of the lung. *In Vivo*, 15, 175-180 (2001).
- Boguski, M. S. and McCormick, F., Proteins regulating Ras and its relatives. *Nature*, 663, 643-654 (1993).
- Booden, M. A., Sakaguchi, D. S., and Buss, J. E., Mutation of Ha-Ras C terminus changes effector pathway utilization. *J. Biol. Chem.*, 275, 23559-23568 (2000).
- Bos, J. L., Ras oncogenes in human cancer: a review. Cancer

- Res., 49, 4682-4689 (1989).
- Brown, P. D., Levy, A. T., Margulies, I. M., and Liotta, L. A., and Stetler-Stevenson, W. G., Independent expression and cellular processing of Mr 72,000 type IV collagenase and interstitial collagenase in human tumorigenic cell lines. *Cancer Res.*, 50, 6184-6191 (1990).
- Campbell, S. L., Khosravi-Far, R., Rossman, K. L., Clark, G. J., and Der, C. J., Increasing complexity of Ras signaling. *Oncogene*, 17, 1395–1413 (1998).
- Carbone, A., Gusella, G. L., Radzioch, D., and Varesio, L., Human Harvey-ras is biochemically different from Kirsten- or N-ras. *Oncogene*, 6, 731-737 (1991).
- Casey, P. J, Solski, P. A., Der, C. J., and Buss, J. E., p21ras is modified by a farnesyl isoprenoid. *Proc. Natl. Acad. Sci. U.S.A.*, 86, 8323-8327 (1989).
- Chiu, V. K., Bivona, T., Hach, A., Sajous, J. B., Silletti, J., Wiener, H., Johnson, R. L. II, Cox, A. D., and Philips, M. R., Ras signalling on the endoplasmic reticulum and the Golgi. *Nat. Cell Biol.*, 4, 343-350 (2002).
- Choy, E., V. K., Chiu, J., Silletti, M., Feoktistov, T., Morimoto, D., Michaelson, I. E., Ivanov, and M. R. Philips., Endomembrane trafficking of Ras: the CAAX motif targets proteins to the ER and Golgi. *Cell*, 98, 69-80 (1999)
- Chung, T. W., Lee, Y. C., and Kim, C. H., Hepatitis B viral HBx induces matrix metalloproteinase-9 gene expression through activation of ERK and PI-3K/AKT pathways: involvement of invasive potential. FASEB J., 18, 1123-1125 (2004).
- Clair, T., Miller, W., and Cho-Chung, Y., Prognostic significance of the expression of the ras protein with a molecular weight of 21, 000 by human breast cancer. *Cancer Res.*, 49, 5290-5293 (1987).
- Clark, G. J. and Der, C. J., Aberrant function of the Ras signal transduction pathway in human breast cancer. *Breast Cancer Res. Treat.*, 35, 133-144 (1995).
- Cox, A. D. and Der, C. J., Fanesyltransferase inhibitors and cancer treatment: targeting simply Ras? *Biochem. Biophys. Acta*, 1333, F51-F71 (1997).
- Datta, S. R., Brunet, A., and Greenberg, M. E., Cellular survival: aplay in three Akts. *Genes Dev.*, 13, 2905-2927 (1999).
- Downward, J., Mechainsms and consequences of activation of protein kinase B/Akt. *Curr. Opin. Cell Biol.*, 10, 262-267 (1998).
- Downward, J., Targeting ras signaling pathways in cancer therapy. *Nature*, 3, 11-22 (2003).
- Etienne-Manneville, S. and Hall, A., Rho GTPases in cell biology. *Nature*, 420, 629-635 (2002).
- Feig, L. A., Urano, T., and Canto, S., Evidence for a Ras/Ral signaling cascade. *Trends Biochem. Sci.*, 21, 438-441 (1996).
- Franks, L. M. and Teich, N. M., Cellular and Molecular Biology of Cancer. Oxford University Press, (1997).
- Gum, R., Wang, H., Lengyel, E., Juarez, J., and Boyd, D., Regulation of 92 kDa type IV collagenase expression by the

- jun aminoterminal kinase- and the extracellular signal-regulated kinase-dependent signaling cascades. *Oncogene*, 14, 1481-1493 (1997).
- Gutierrez, L., Magee, A. I., Marshall, C. J., and Hancock, J. F., Post-translational processing of p21ras is two-step and involves carboxyl-methylation and carboxy-terminal proteolysis. *EMBO J.*, 8, 1093-1098 (1989).
- Hancock, J. F., Magee, A. I., Childs, J. E., and Marshall, C. J., All Ras proteins are polyisoprenylated but only some are palmitoylated. *Cell*, 57, 1167-1177 (1989).
- Hancock, J. F., Peterson, H., and Marshall, C. J., A polybasic domain or palmitoylation is required in addition to the CAAX motif to localize p21ras to the plasma membrane. *Cell*, 63, 133-139 (1990).
- Hancock, J. F., Ras proteins: different signals from different locations. *Nat. Rev. Mol. Cell Biol.*, 4, 373-384 (2003).
- Jiang, K., Sun, J., Cheng, J., Djeu, J. Y., Wei, S., and Sebti, S., Akt mediates Ras downregulation of RhoB, a suppressor of transformation, invasion, and metastasis. *Mol. Cell Biol.*, 24, 5565-5576 (2004).
- Johnson, L., Greenbaum, D., Cichowski, K., Mercer, K., Murphy, E., Schmitt, E., Bronson, R. T., Umanoff, H., Edelmann, W., Kucherlapati, R., and Jacks, T., K-ras is an essential gene in the mouse with partial functional overlap with N-ras. *Genes Dev.*, 11, 2468-2481 (1997).
- Joneson, T., White, M., Wigler, M., and Bar-Sagi, D., Stimulation of membrane ruffling and MAP kinase activation by distinct effectors of Ras. *Science*, 271, 810-812 (1996).
- Kato, K., Cox, A. D., Hisaka, M. M., Graham, S. M., Buss, J. E., and Der, C. J., Isoprenoid addition to Ras protein is the critical modification for its membrane association and transforming activity. *Proc. Natl. Acad. Sci. U.S.A.*, 89, 6403– 6407 (1992).
- Keely, P. J., Westwick, J. K., Whitehead, I. P., Der, C. J., and Parise, L. V., Cdc42 and Rac1 induce integrin-mediated cell motility and invasiveness through PI(3)K. *Nature*, 390, 632-636 (1997).
- Khwaja, A., Akt is more than just a Bad kinase. *Nature*, 401, 33-34 (1999).
- Khwaja, A., Rodriguez-Viciana, P., Wennstrom, S., Warne, P. H. and Downward, J., Matrix adhesion and Ras transformation both activate a phosphoinositide 3-OH kinase and protein kinase B/Akt cellular survival pathway. *EMBO J.*, 16, 2783-2793 (1997).
- Kim, M. S., Lee, E. J., Choi kim, H. R., and Moon, A., p38 kinase is a key signaling molecule for H-ras-induced cell motility and invasive phenotype in human breast epithelial cell. *Cancer Res.*, 63, 5454-5461 (2003).
- Koera, K., Nakamura, K., Nakao, K., Miyoshi, J., Toyoshima, K., Hatta, T., Otani, H., Aiba, A., and Katsuki, M., K-ras is essential for the development of the mouse embryo. *Oncogene*, 15, 1151-1159 (1997).
- Kyriakis, J. M., App, H., Zhang, X. F., Banerjee, P., Brautigan, D.

L., Rapp, U. R., and Avruch, J., Raf-1 activates MAP kinase-kinase. *Nature*, 358, 417-421 (1992).

- Lambert, J. M., Lambert, Q. T., Reuther, G. W., Malliri, A., Siderovski, D. P., Sondek, J., Collard, J. D., and Der, C. J., Tiam1 mediates Ras activation of Rac by a PI(3)Kindependent mechnism. *Nature*, 417, 625-821 (2002).
- Leevers, S. J., Paterson, H. F., and Marshall, C. J., Requirement for Ras in Raf activation is overcome by targeting Raf to the plasma membrane. *Nature*, 369, 411-414 (1994).
- Li, J. J., Cao, Y., Young, M. R., and Colburn, N. H., Induced expression of dominant-negative c-jun downregulates NFkappaB and AP-1 target genes and suppresses tumor phenotype in human keratinocytes. *Mol. Carcinog.*, 29, 159-169 (2000).
- Li, J. J., Rhim, J. S., Schlegel, R., Vousden, K. H., and Colburn, N. H., Expression of dominant negative Jun inhibits elevated AP-1 and NF-kappaB transactivation and suppresses anchorage independent growth of HPV immortalized human keratinocytes. *Oncogene*, 16, 2711-2721 (1998).
- Li, J. J., Westergaard, C., Ghosh, P., and Colburn, N. H., Inhibitors of both nuclear factor-kappaB and activator protein-1 activation block the neoplastic transformation response. *Cancer Res.*, 57, 3569-3576 (1997).
- Liotta, L. A., Steeg, P. S., and Stetler-Stevenson, W. G., Cancer metastasis and angiogenesis: An imbalance of positive and negative regulation. *Cell*, 64, 327-336 (1991).
- Ludes-Meyers, J. H., Liu, Y., Munoz-Medellin, D., Hilsenbeck, S. G., and Brown, P. H., AP-1 blockade inhibits the growth of normal and malignant breast cells. *Oncogene*, 20, 2771-2780 (2001).
- Magee, T. and Marshall, C., New insights into the interaction of Ras with the plasma membrane. *Cell*, 98, 9-12 (1999).
- Maher, J., Baker, D. A., Manning, M., Dibb, N. J., and Roberts, I. A. G., Evidence for cell-specific differences in transformation by N-, H- and K-ras. Oncogene, 11, 1639-1647 (1995).
- Malliri, A., Van der Kammen, R. A., Clark, K., Van der Valk, M., Michiels, F., and Collar, J. G., Mice deficient in the Rac activator Tiam1 are resistant to Ras-induced skin tumors. *Nature*, 417, 867-871 (2002).
- Marais, R., Light, Y., Paterson, H. F., and Marshall, C. J., Ras recruits Raf-1 to the plasma membrane for activation by tyrosine phosphorylation. *EMBO J.*, 14, 3136-3145 (1995).
- Marshall, C. J., Ras effectors. *Curr. Opin. Cell Biol.*, 8, 197-204 (1996).
- Matallanas, D., Arozarena, I., Berciano, M. T., Aaronson, D. S., Pellicer, A., Lafarga, M., and Crespo, P., Differences on the inhibitory specificities of H-Ras, K-Ras, and N-Ras (N17) dominant negative mutants are related to their membrane microlocalization. *J. Biol. Chem.*, 278, 4572-4581 (2003).
- Moon, A., Kim, M. S., Kim, T. G., Kim, S. H., Kim, H. E., Chen, Y. Q., and Choi Kim, H. R., H-ras, but not N-ras, induces an invasive phenotype in human breast epithelial cells: a role for MMP-2 in the H-ras-induced invasive phenotype. *Int. J.*

- Cancer, 85, 176-181 (2000).
- Moon, S. K., Cha, B. Y., and Kim, C. H., ERK1/2 mediates TNF-alpha-induced matrix metalloproteinase-9 expression in human vascular smooth muscle cells *via* the regulation of NF-kappaB and AP-1: Involvement of the ras dependent pathway. *J. Cell Physiol.*, 198, 417-427 (2004).
- Nakopoulou, L., Tsirmpa, I., Alexandrou, P., Louvrou, A., Ampela, C., Markaki, S., and Davaris, P. S., MMP-2 protein in invasive breast cancer and the impact of MMP-2/TIMP-2 phenotype on overall survival. *Breast Cancer Res. Treat.*, 77, 145-155 (2003).
- Niv, H., Gutman, O., Kloog, Y., and Henis, Y. I., Activated K-Ras and H-Ras display different interactions with saturable nonraft sites at the surface of live cells. *J. Cell Biol.*, 157, 865-872 (2002).
- Nobes, C. D. and Hall, A., Rho, Rac, and Cdc42 GTPases regulate the assembly of multimolecular focal complexs associated with actin stress fibers, lamlipodia, and filopodia. *Cell*, 81, 53-62 (1995).
- Oldham, S. M., Clark, G. J., Gangarosa, L. M., Coffey, R. J., and Der, C. J., Activation of the Raf-1/MAP kinase cascade is not sufficient for Ras transformation of RIE-1 epithelial cells. *Proc. Natl. Acad. Sci. U.S.A.*, 93, 6924-6928 (1996).
- Oliff, A., Rarnesyltransferse inhibitors: targeting the molecular basis of cancer. *Biochim. Biophys. Acta*, 1423, C19-C30 (1999).
- Pacold, M. E., Suire, S., Perisic, O., Lara-Gonzlez, W., Davis, C. T., Walker, E. H., Hawkins. P. T., Stephens, L., Eccleston, J. R., and Williams, R. L., Crystal strucure and functional analysis of Ras binding to its effector phosphoinositide 3-kinaseã. *Cell*, 103, 931-943 (2000).
- Parton, R. G. and Hancock, J. F., Lipid rafts and plasma membrane microorganization: insights from Ras. *Trends Cell Biol.*, 14, 141-147 (2004).
- Pola, S., Cattaneo, M. G., and Vicentini, L. M., Anti-migratory and anti-invasive effect of somatostatin in human neuroblastoma cells: involvement of Rac and MAP kinase activity. *J. Biol. Chem.*, 278, 40601-40606 (2003).
- Prior, I. A., Harding, A., Yan, J., Sluimer, J., Parton, R. G., and Hancock, J. F., GTP-dependent segregation of H-ras from lipid rafts is required for biological activity. *Nat. Cell Biol.*, 3, 368-375 (2001).
- Prior, I. A. and Hancock, J. F., Compartmentalization of ras proteins. *J. Cell Sci.* 114, 1603-1608 (2001).
- Prior, I. A., Muncke, C., Parton, R. G., and Hancock, J. F., Direct visualization of Ras proteins in spatially distinct cell surface microdomains. *J. Cell Biol.*, 160, 165-170 (2003).
- Pruitt, K. and Der, C. J., Ras and Rho regulation of the cell cycle and oncogenesis. *Cancer Lett.*, 171, 1-10 (2001).
- Qin, H., Sun, Y., and Benveniste, E. N., The transcription factors Sp1, Sp3, and AP-2 are required for constitutive matrix metalloproteinase-2 gene expression in astroglioma cells. *J. Biol. Chem.*, 274, 29130-29137 (1999).

- Qui, R., McCormick, F., and Symons, M., An essential role for Rac in Ras transformation. *Nature*, 374, 457-459 (1995).
- Reif, K., Nobes, C. D., Thomas, G., Hall, A., and Cantrell, D. A., Phosphatidylinositol 3-kinase signals activate a selective subset of Rac/Rho-dependent effector pathways. *Curr. Biol.* 6, 1445-1455 (1996).
- Ridley, A. J., Paterson, H. F., Johnston, C. L., Diekmann, D., and Hall, A., The small GTP-binding protein rac regulates growth factor-induced membrane ruffling. *Cell*, 70, 401-410 (1992).
- Rodriguez-Viciana, P., Warne, P. H., Dhand, R., Vanhaesebroeck, B., Gout, I., Fry, M. J., Waterfield, M. D., and Downward, J., Phosphatidylinositol-3-OH kinase as a direct target of Ras. *Nature*, 370, 527-532 (1994).
- Romashkova, J. A. and Makarov, S. S., NF-kappaB is target of AKT in anti-apoptotic PDGF signaling. *Nature*, 401, 86-90 (1999).
- Roy, S., Luetterforst, R., Harding, A., Apolloni, A., Etheridge, M., Stang, E., Rolls, B., Hancock, J. F., and Parton, R. G., Dominant-negative caveolin inhibits H-Ras function by disrupting cholesterol-rich plasma membrane domains. *Nat. Cell Biol.*, 1, 98–105 (1999).
- Sachdev, P., Zeng, L., and Wang, L. H., Distinct role of phosphatidylinositol 3-kinase and Rho family GTPases in Vav3-induced cell transformation, cell motility, and morphological changes. *J. Biol. Chem.*, 277, 17638-17648 (2002).
- Sato, H., Kida, Y., Mai, M., Endo, Y., Sasaki, T., Tanaka, J., and Seiki, M., Mutation spectra of smoky coal combustion emissions in Salmonella reflect the TP53 and KRAS mutations in lung tumors from smoky coal-exposed individuals. *Oncogene*, 7, 77-83 (1992).
- Sato, H., Kita, M., and Seiki, M., v-Src activates the expression of 92-kDa type IV collagenase gene through the AP-1 site and the GT box homologous to retinoblastoma control elements. A mechanism regulating gene expression independent of that by inflammatory cytokines. *J. Biol. Chem.*, 268, 23460-23468 (1993).
- Sato, H. and Seiki, M., Regulatory mechanism of 92 kDa type IV collagenase gene expression which is associated with invasiveness of tumor cells. *Oncogene*, 8, 395-405 (1993).
- Seabra, M. C., Membrane association and targeting or prenylated Ras-like GTPases. *Cell signal.*, 10, 167-172 (1998).
- Shields, J. M., Pruitt, K., McFall, A., Shaub, A., and Der, C. J., Understanding Ras: 'it ain't over 'til it's over'. *Trends Cell Biol.*, 10, 147-154 (2000).
- Shin, I. C., Kim, S. H., Song, H., Choi Kim, H. R., and Moon, A., H-Ras-specific activation of Rac-MKK3/6-p38 pathway. *J. Biol. Chem.*, 15, 14675-14683 (2005).
- Simon, C., Goepfert, H., and Boyd, D., Inhibition of the p38 mitogen-activated protein kinase by SB 203580 blocks PMA-induced Mr 92,000 type IV collagenase secretion and *in vitro*

- invasion. Cancer Res., 58, 1135-1139 (1998).
- Simon, C., Juarez, J., Nicolson, G. L., and Boyd, D., Effect of PD 098059, a specific inhibitor of mitogen-activated protein kinase kinase, on urokinase expression and *in vitro* invasion. *Cancer Res.*, *56*, 5369-5374 (1996).
- Simon, C., Simon, M., Vucelic, G., Hicks, M. J., Plinkert, P. K., Koitschev, A., and Zenner, H. P., The p38 SAPK pathway regulates the expression of the MMP-9 collagenase via AP-1-dependent promoter activation. *Exp. Cell Res.*, 271, 344-355 (2001).
- Simons, K. and Toomre, D., Lipid rafts and signal transduction. *Nat. Rev. Mol. Cell Biol.*, 1, 31–39 (2000).
- Steller-Stevenson, W. G., Type-IV collagenases in tumor invasion and metastasis. *Cancer Metast. Rev.*, 9, 289-303 (1990).
- Stetler-Stevenson, W. G., Hewitt, R., and Corcoran, M., Matrix metalloproteinases and tumor invasion: from correlation and causality to the clinic. *Semin. Cancer Biol.*, 7, 147-154 (1996).
- Talvensaari-Mattila, A., Paakko, P., Blanco-Sequeiros, G., and Turpeenniemi-Hujanen, T., Matrix metalloproteinase-2 (MMP-2) is associated with the risk for a relapse in postmenopausal patients with node-positive breast carcinoma treated with antiestrogen adjuvant therapy. Breast Cancer Res. Treat., 65, 55-61 (2001).
- Tryggvason, K., Huhtala, P., Tuuttila, A., Chow, L., Keski-Oja, J., and Lohi, J., Structure and expression of type IV collagenase genes. *Cell. Differ. Dev.*, 32, 307-312 (1990).
- Tryggvason, K., Type-IV collagenase in invasive tumors. *Breast Cancer Res. Treat.*, 24, 209-218 (1993).
- Umanoff, H., Edelmann, W., Pellicer, A., and Kucherlapati, R., The murine N-ras gene is not essential for growth and development. *Proc. Natl. Acad. Sci. U.S.A.*, 92, 1709-1713 (1995).
- Ura, H., Bonfil, R. D., Reich, R., Reddel, R., Pfeifer, A., Harris, C. C., and Klein-Szanto, A. J., Expression of type IV collagenase and procollagen genes and its correlation with the tumorigenic, invasive, and metastatic abilities of oncogene-transformed human bronchial epithelial cells. *Cancer Res.* 49, 4615-4621 (1989).
- Vaudry, D., Stork, P. J., Lazarovici, P., and Eiden, L. E., Signaling pathways for PC12 cell differentiation: making the right connections. Science, 296, 1648-1649 (2002).
- Voice, J. K., Klemke, R. L., Le, A., and Jackson, J. H., Four Human Ras Homologs Differ in Their Abilities to Activate Raf-1, Induce Transformation, and Stimulate Cell Motility. *J. Biol. Chem.*, 274, 17164-17170 (1999).
- Walsh, A. B. and Bar-Sagi, D., Differential activation of the Rac pathway by Ha-Ras and K-Ras. *J. Biol. Chem.*, 276, 15609-152001 (2001).
- Watson, D. M., Elton, R. A., Jack, W. J., Dixon, J. M., Chetty, U., and Miller, W. R., The H-ras oncogene product p21 and prognosis in human breast cancer. *Breast Cancer Res.*

- Treat., 17, 161-169 (1991).
- Welch, H. C., Coadwell, W. J., Stephens, L. R., and Hawkins, P. T., Phosphoinositide 3-kinase-dependent activation of Rac. FEBS Lett., 546, 93-97 (2003).
- Willumsen, B. M, Christensen, A, Hubbert, N. L, Papageorge, A. G, and Lowy, D. R., The p21 ras C-terminus is required for transformation and membrane association. *Nature*, 310, 583-586 (1984a).
- Willumsen, B. M., Norris, K, Papageorge, A. G, Hubbert, N. L, and Lowy, D. R., Harvey murine sarcoma virus p21 ras protein: biological and biochemical significance of the cysteine nearest the carboxy terminus. *EMBO J.*, 3, 2581-2585 (1984b).
- Wolthuis, R. M. and Bos, J. L., Ras caught in another affaif: the RHOad less traveled gets congested. *Oncogene*, 17, 1415-1438 (1999).

- Xu, Q., Karouji, Y., Kobayashi, M., Ihara, S., Konishi, H., and Fukui, Y., The PI 3-kinase-Rac-p38 MAP kinase pathway is involved in the formation of signet-ring cell carcinoma. *Oncogene*, 22, 5537-5544 (2003).
- Yan, J., Roy, S., Apolloni, A., Lane, A., and Hancock, J. F., Ras isoforms vary in their ability to activate Raf-1 and phosphoinositide 3-kinase. *J. Biol. Chem.*, 273, 24052-24056 (1998).
- Yordy, J. S. and Muise-Helmericks, R. C., Signal transduction and the Ets family of transcription factors. *Oncogene*, 19, 6503-6513 (2000).
- Zhang, D., Bar-Eli, M., Meloche, S., and Brodt, P., Dual regulation of MMP-2 expression by the type 1 insulin-like growth factor receptor: the phosphatidylinositol 3-kinase/Akt and Raf/ERK pathways transmit opposing signals. *J. Biol. Chem.*, 279, 19683-19690 (2004).