

Synthesis of New Anthracycline Derivatives Containing Lactic or Stearic Acid Moiety

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Novel anthracycline analogues **2-9** as potential anticancer agents were synthesized from daunomycin (**1a**) and doxorubicin (**1b**). Compounds **2**, **6**, and **7** were prepared by the nucleophilic displacement type esterification of a 14-bromodaunomycin (**1c**) with a sodium lactate, and stearic acid, respectively. Compounds **3-5** and **7-9** were prepared by the reaction of either daunomycin (**1a**) or doxorubicin (**1b**) with L-lactic and stearic acids in the presence of EDCI/PP reagents.

Key Words : Daunomycin, Doxorubicin, Anthracycline derivatives, Lactic acid, Stearic acid

Introduction

The anthracycline antibiotics, daunomycin (**1a**) and doxorubicin (**1b**), are the member of a very important class of cytotoxic agent that has been used for many years in the treatment of many different types of cancer. Doxorubicin (**1b**), a well known anticancer drug, is considered as one of the most active single anticancer agents because of its broad antitumour spectrum.¹ Chemotherapy with doxorubicin, however, is largely limited by the cumulative dose-related cardiotoxicity, resulting in congestive heart failure. It is generally believed that the formation of free radicals plays a crucial role, and the involvement of radicals in the mechanism of doxorubicin-induced cardiotoxicity has been the subject of extensive reviewing^{2,3} (Figure 1).

As a result, numerous synthetic efforts have been devoted to overcome these disadvantages, culminating in the development of daunomycin (**1a**) or doxorubicin (**1b**) derivatives.⁴⁻⁷ Many examples of coupling daunomycin (**1a**) or doxorubicin (**1b**) with some amino acids⁸⁻¹³ as well as blending **1a** or **1b** with some amino acids⁴ are reported.

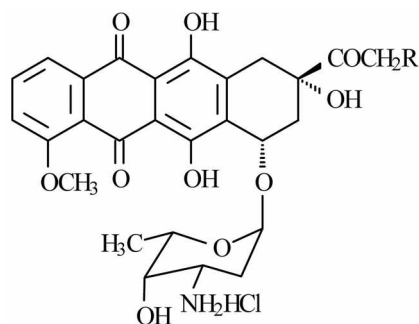
L-Lactic acid occurs in small quantities in the blood and muscle fluid of man and animals. The lactic acid concentration increases in muscle and blood after vigorous activity.

In muscle, pyruvic acid is reduced to lactic acid during exertion. Stearic acid is also a promising material having strong adsorption property in a cell wall.¹⁴ Therefore, anthracyclines containing a lactic or stearic acid residue are expected to be good potential delivery drugs, maybe due to diminishing cardiotoxicity and undesirable side effects. Herein, we describe the synthesis of new anthracycline derivatives *via* coupling of C₁₄-OH and C₃-NH₂ in DM (**1a**) and DX (**1b**) with two kinds of acid molecules, L-lactic and stearic acid.

Results and Discussion

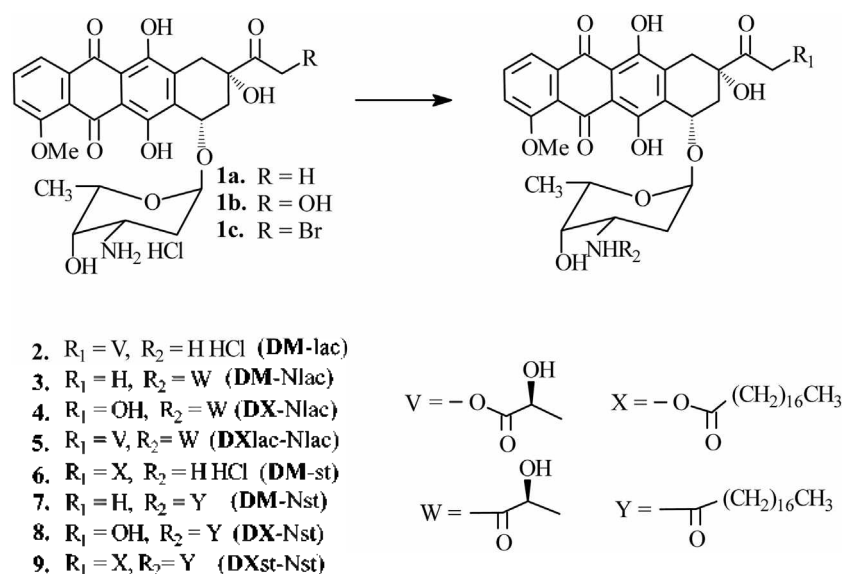
In the previous papers, we described the total synthesis of anthracyclinone derivatives through Michael type condensation¹⁵⁻¹⁸ or Friedel-Crafts acylation.^{19,20} We reported the successful preparation of a new aglycone containing an ester linkage at C-14 through a nucleophilic displacement esterification method.^{19,21} In more recent papers, we have also reported the successful synthesis of some new anthracycline analogues from commercially available anticancer agents, such as daunomycin (**1a**) and doxorubicin (**1b**).⁸⁻¹³ Several new anthracycline derivatives were synthesized using acylation methods (Scheme 1). The synthesis of 14-bromo DM (**1c**) from **1a** was accomplished by the known procedure.^{9,21-23} All compounds **2-9** were obtained through acylation of the C-14 hydroxyl group in the aglycone and/or the amino group at C-3' in the glycone with L-lactic acid and stearic acid.

DM-lac (**2**) and DM-st (**6**), potential prodrugs, were prepared by the reaction of 14-bromo DM (**1c**) with a sodium lactate or stearic acid. For the purpose of comparing the activity of **2** and **6**, carboamidation compounds, DM-Nlac (**3**), DX-Nlac (**4**), DM-Nst (**7**), and DX-Nst (**8**) were synthesized by amidation of the amino group at C-3' of sugar moiety in **1a** or **1b** with the corresponding acids. In addition, *N*-acylation compounds, DXlac-Nlac (**5**) and DXst-Nst (**9**), were prepared through esterification of the C₁₄-OH in DX (**1b**) with the corresponding acids followed by amidation of the amino group at the sugar moiety with the corresponding



1a. Daunomycin R = H
1b. Doxorubicin R = OH

Figure 1. Chemical structures of daunomycin and doxorubicin.



Scheme 1. Synthesis of new anthracycline analogues 2-9.

acids.

First, 14-bromo DM (**1c**) was synthesized in best yield when a minimum quantity of co-solvent (methanol/1,4-dioxane, v/v = 1 : 2) was used, because it diminished the formation of side product from dimethylketalization of ketone at C-13. The rate of bromination depended on the reaction temperature and time. The optimal conditions are bromination at 30 °C for 40 min.

DM-lac (**2**) was synthesized as follows: To a 14-bromo DM (**1c**) prepared by introducing Br atom at C-14 of daunomycin (**1a**) was added a solution of sodium lactate in acetone; the mixture was heated at reflux at for 4 hr. After removing the solvent under reduced pressure, the residue was dissolved in THF and treated with ethereal HCl followed by stirring at -20 °C for 2 hr and further stirring at room temperature for 3 hr to give DM-lac (**2**). DM-st (**6**) was synthesized from DM (**1c**) and stearic acid in triethylamine/acetone as described for the preparation of DM-lac (**2**).

Many attempts to prepare DM-Nlac (**3**) and DX-Nlac (**4**) through direct coupling of the amino group in daunomycin (**1a**) or doxorubicin (**1b**) with L-lactic acid using 1,3-dicyclohexylcarbodiimide (DCC)/4-(dimethylamino) pyridine (DMAP)²⁴ failed. Reactants (**1a**, **1b**) and DCU (dicyclohexylurea) were observed as main products. Eventually, DM-Nlac (**3**) was synthesized by coupling of the NH_2HCl in **1a** with L-lactic acid using 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDCI) in the presence of catalytic amounts of 4-pyrrolidinopyridine (PP).^{25,26} DX-Nlac (**4**) was synthesized from DX (**1b**) as described for the preparation of DM-Nlac (**3**). For the reaction of **1b**, however, the competition between the C-14 hydroxyl group and the amine group at C-3' was observed. Both products of DX-Nlac (**4**) and DXlac-Nlac (**5**) were formed and the product ratio depended on the amounts of L-lactic acid and EDCI used in the reaction. DXlac-Nlac (**5**) was prepared using 2.2 equivalent of the corresponding acid and EDCI under the

same conditions.

Synthesis of DM-Nst (**7**) and DX-Nst (**8**) was carried out as follows: stearic acid and EDCI (1.2 equiv) was dissolved in dry DMF and the mixture was stirred at 0 °C for 30 min; to the reaction mixture was added DM (**1a**) or DX (**1b**) and catalytic amounts of PP, and the mixture was then stirred at room temperature for 3 hr to give DM-Nst (**7**) and DX-Nst (**8**). DXst-Nst (**9**) was synthesized from **1b** as described for the preparation of **8** by increasing the amounts of stearic acid (2.2 equiv) and EDCI (2.2 equiv).

In conclusion, we synthesized the new anthracycline analogues including lactic or stearic acid as potential anti-cancer agents. Detailed biological tests of the title glycosides in vitro will be reported elsewhere in the future.

Experimental Section

All reactions were carried out under argon atmosphere in dried glassware. All solvents were carefully dried and distilled as reported.²⁷ Bulk grade hexane was distilled before use. Merck pre-coated silica gel plates (Art.5554) with fluorescent indicator were used as analytical TLC. Gravity column chromatography and flash column chromatography were carried out on silica gel (230-400 mesh from Merck). ¹H and ¹³C NMR spectra were recorded on a JEOL JNM EX-400 spectrometer. Chemical shifts were internally referenced to TMS for ¹H or to solvent signals for ¹³C. Infrared spectra were recorded on a Nicolet 5-DXB series FT-IR spectrophotometer. Mass spectra were obtained on a JEOL JMS HX-110/110A Tandem mass spectrometer (FAB⁺, ESI). UV-VIS absorption spectra were recorded on a Hitachi-556 spectrophotometer. Optical rotations were determined using the Rudolph AUTOPOL IV apparatus with a 0-100-1.5 polarimeter sample tube. Melting points were obtained on a Büchi 510 melting point apparatus and are uncorrected.

Daunomycin-14-lactate hydrochloride (2). A solution of 14-bromodaunomycin hydrochloride (**1c**, 0.30 g, 0.46 mmol) and sodium lactate (0.10 g, 0.93 mmol) in acetone (300 mL) was heated at reflux for 4 hr. Upon completion of the reaction the solvent was evaporated. The residue was dissolved in dry THF (150 mL), ethereal HCl was added, and the mixture was stirred at $-20\text{ }^{\circ}\text{C}$ for 2 hr and further stirred at room temperature for 3 hr. The organic solvent was concentrated by a rotary evaporator and the residue was purified by column chromatography on silica gel ($\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH}/\text{HCO}_2\text{H}/\text{H}_2\text{O} = 88 : 15 : 2 : 1$) to give daunomycin-14-lactate hydrochloride (**2**, 0.29 g, 97%) as a red powder: mp $120\text{--}122\text{ }^{\circ}\text{C}$; $[\alpha]_{\text{D}}^{20} +110.98^{\circ}$ (c 0.1, CH_3OH); IR (KBr) 3432, 2939, 1793, 1738, 1609, 1387, 1283, 1215, 1129, 987, 793 cm^{-1} ; ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 13.91 (s, 1H, PhOH), 13.24 (s, 1H, PhOH), 8.29 (s, 2H, NH_2), 7.78 (m, 2H, ArH), 7.55 (m, 1H, ArH), 5.60 (d, 1H, $J = 6.3$ Hz, C_4H), 5.50 (s, 1H, C_9OH), 5.28 (d, 1H, $J = 18.0$ Hz, C_{14}H), 5.11 (d, 1H, $J = 3.9$ Hz, $\text{C}_{7\text{eq}}\text{H}$), 5.28 (d, 1H, $J = 18.0$ Hz, C_{14}H), 4.84 (m, 1H, C_1H), 4.16 (q, 1H, $J = 6.3$ Hz, C_5H), 4.03 (q, 1H, $J = 6.8$ Hz, C_{16}H), 3.93 (s, 3H, C_4OCH_3), 3.63 (m, 1H, C_4OH), 3.32 (m, 1H, C_3H), 3.04 (d, 1H, $J = 17.5$ Hz, $\text{C}_{10\text{eq}}\text{H}$), 2.73 (d, 1H, $J = 17.5$ Hz, $\text{C}_{10\text{ax}}\text{H}$), 2.20 (d, 1H, $J = 14.6$ Hz, $\text{C}_{8\text{eq}}\text{H}$), 2.09 (dd, 1H, $J = 3.9, 14.6$ Hz, $\text{C}_{8\text{ax}}\text{H}$), 1.67–1.90 (m, 2H, C_2H), 1.19 (d, 3H, $J = 6.1$ Hz, C_5CH_3), 1.11 (d, 3H, $J = 6.8$ Hz, C_{17}CH_3); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 208.13, 186.14, 183.31, 180.57, 174.18, 165.05, 160.71, 136.16, 135.07, 134.54, 134.07, 119.74, 118.89, 110.58, 99.30, 75.02, 69.43, 66.70, 66.15, 65.98, 65.77, 56.57, 50.67, 46.52, 35.94, 31.82, 28.99, 28.29, 20.59, 16.70; UV (CH_3OH): λ_{max} (log ϵ) = 205 (0.19), 249 (0.05), 485 (0.03); Mass (FAB^+ , Na) m/z 639 ($\text{M-HCl} + \text{Na}$) $^+$.

Daunomycin-3'-N-lacticarboamide (3). The mixture of L-lactic acid (0.08 mL, 1.06 mmol) and EDCI (0.20 g, 1.06 mmol) in dry DMF (80 mL) was stirred on an ice bath for 30 min and allowed to warm to room temperature. To the stirred solution was added daunomycin hydrochloride (**1a**, 0.50 g, 0.88 mmol) and catalytic amounts of 4-pyrrolidinopyridine, and the mixture was stirred then for 12 hr. The reaction mixture was dissolved in CH_2Cl_2 (200 mL), washed with water (2×200 mL) and brine (2×200 mL), dried over MgSO_4 , and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel ($\text{CH}_2\text{Cl}_2/\text{Hexane}/\text{CH}_3\text{OH} = 8 : 1 : 1$) to give daunomycin-3'-N-lacticarboamide (**3**, 0.50 g, 94%) as a red powder: mp $127\text{--}129\text{ }^{\circ}\text{C}$; $[\alpha]_{\text{D}}^{20} +120.01^{\circ}$ (c 0.1, CH_3OH); IR (KBr) 3399, 2927, 2855, 1716, 1622, 1580, 1529, 1411, 1289, 1236, 1207, 1119, 985, 821, 760, 620 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 13.95 (s, 1H, PhOH), 13.21 (s, 1H, PhOH), 7.99 (d, 1H, $J = 7.8$ Hz, ArH), 7.76 (dd, 1H, $J = 7.8, 8.3$ Hz, ArH), 7.35 (d, 1H, $J = 8.3$ Hz, ArH), 6.90 (d, 1H, $J = 8.3$ Hz, NH), 5.49 (s, 1H, C_9OH), 5.16 (d, 1H, $J = 5.8$ Hz, C_4H), 4.90 (d, 1H, $J = 3.9$ Hz, $\text{C}_{7\text{eq}}\text{H}$), 4.48 (m, 1H, C_1H), 4.09–4.22 (m, 2H, C_5H , $\text{Nlac}\alpha\text{H}$), 4.03 (s, 3H, C_4OCH_3), 3.66 (m, 1H, C_4OH), 3.23 (m, 1H, C_3H), 3.19 (d, 1H, $J = 19.0$ Hz, $\text{C}_{10\text{eq}}\text{H}$), 2.86 (d, 1H, $J = 19.0$ Hz, $\text{C}_{10\text{ax}}\text{H}$), 2.41 (s,

3H, C_{14}CH_3), 2.29 (d, 1H, $J = 14.1$ Hz, $\text{C}_{8\text{eq}}\text{H}$), 2.04 (dd, 1H, $J = 3.9, 14.1$ Hz, $\text{C}_{8\text{ax}}\text{H}$), 1.85 (m, 2H, C_2H), 1.32 (d, $J = 6.8$ Hz, 3H, C_5CH_3), 1.27 (d, 1H, $J = 7.3$ Hz, $\text{Nlac}\beta\text{CH}_3$); ^{13}C NMR (100 MHz, CDCl_3) δ 212.05, 186.95, 186.53, 174.29, 160.93, 156.41, 155.74, 135.67, 135.42, 134.37, 134.03, 120.76, 119.75, 118.34, 111.38, 111.16, 100.59, 76.57, 69.92, 69.25, 68.40, 66.97, 56.58, 45.15, 35.03, 33.28, 29.67, 24.85, 21.01, 16.74; UV (CH_3OH): λ_{max} (log ϵ) = 235 (0.32), 252 (0.20), 482 (0.99); Mass (FAB^+ , Na) m/z 623 ($\text{M} + \text{Na}$) $^+$.

Doxorubicin-3'-N-lacticarboamide (4). The mixture of L-lactic acid (0.09 mL, 1.24 mmol) and EDCI (0.24 g, 1.24 mmol) in dry DMF (100 mL) was stirred on an ice bath for 30 min and allowed to warm to room temperature. To the stirred solution was added doxorubicin hydrochloride (**1b**, 0.60 g, 1.03 mmol) and catalytic amounts of 4-pyrrolidinopyridine, and the mixture was then stirred for 14 hr. The reaction mixture was dissolved in CH_2Cl_2 (200 mL), washed with water (2×200 mL) and brine (2×200 mL), dried over MgSO_4 , and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel ($\text{CH}_2\text{Cl}_2/\text{Hexane}/\text{CH}_3\text{OH} = 8 : 1 : 1$) to give doxorubicin-3'-N-lacticarboamide (**4**, 0.59 g, 92%) as a red powder: mp $116\text{--}117\text{ }^{\circ}\text{C}$; $[\alpha]_{\text{D}}^{20} +72.01^{\circ}$ (c 0.1, CH_3OH); IR (KBr) 3425, 2926, 2254, 2128, 1728, 1658, 1580, 1530, 1441, 1411, 1377, 1286, 1241, 1118, 1020, 992, 822, 764, 708, 624 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 13.95 (s, 1H, PhOH), 13.24 (s, 1H, PhOH), 8.00 (d, 1H, $J = 7.8$ Hz, ArH), 7.78 (dd, 1H, $J = 7.8, 8.3$ Hz, ArH), 7.40 (d, 1H, $J = 8.3$ Hz, ArH), 7.06 (d, 1H, $J = 8.7$ Hz, NH), 5.51 (d, 1H, $J = 5.8$ Hz, C_4H), 5.24 (s, 1H, C_9OH), 4.95 (d, 1H, $J = 3.9$ Hz, $\text{C}_{7\text{eq}}\text{H}$), 4.83 (m, 1H, C_1H), 4.77 (s, 2H, C_{14}H), 4.09–4.30 (m, 2H, C_5H , $\text{Nlac}\alpha\text{H}$), 4.07 (s, 3H, C_4OCH_3), 3.62 (m, 1H, C_4OH), 3.31 (m, 1H, C_3H), 3.23 (d, 1H, $J = 18.5$ Hz, $\text{C}_{10\text{eq}}\text{H}$), 2.98 (d, 1H, $J = 18.5$ Hz, $\text{C}_{10\text{ax}}\text{H}$), 2.37 (d, 1H, $J = 14.1$ Hz, $\text{C}_{8\text{eq}}\text{H}$), 2.15 (dd, 1H, $J = 3.9, 14.1$ Hz, $\text{C}_{8\text{ax}}\text{H}$), 1.90–2.04 (m, 2H, C_2H), 1.10–1.30 (m, 6H, C_5CH_3 , $\text{Nlac}\beta\text{CH}_3$); ^{13}C NMR (100 MHz, CDCl_3) δ 213.87, 186.91, 186.54, 174.58, 160.83, 156.15, 155.51, 135.57, 135.28, 133.81, 133.56, 120.69, 119.58, 118.30, 111.27, 111.08, 100.74, 76.26, 69.34, 68.25, 67.08, 65.28, 56.49, 44.73, 35.60, 33.57, 29.44, 29.29, 20.40, 16.82; UV (CH_3OH): λ_{max} (log ϵ) = 234 (0.39), 253 (0.24), 485 (0.11); Mass (FAB^+ , Na) m/z 639 ($\text{M} + \text{Na}$) $^+$.

Doxorubicin-14, 3'-N-dilactate (5). The mixture of L-lactic acid (0.08 mL, 1.14 mmol) and EDCI (0.22 g, 1.14 mmol) in dry DMF (50 mL) was stirred on an ice bath for 30 min and allowed to warm to room temperature. To the stirred solution was added doxorubicin hydrochloride (**1b**, 0.30 g, 0.46 mmol) and catalytic amounts of 4-pyrrolidinopyridine, and the mixture was stirred for 13 hr. The resulting mixture was diluted with CH_2Cl_2 (300 mL), washed with water (2×200 mL) and brine (2×200 mL), dried over MgSO_4 , and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel ($\text{CH}_2\text{Cl}_2/\text{Hexane}/\text{CH}_3\text{OH} = 8 : 2 : 1$) to give **5** (0.28 g, 88%) as a red powder: mp $103\text{--}105\text{ }^{\circ}\text{C}$; $[\alpha]_{\text{D}}^{20} +145.29^{\circ}$ (c 0.1,

CH₃OH); IR (KBr) 3425, 2940, 1720, 1630, 1577, 1530, 1418, 1280, 1215, 1124, 985, 782, 705 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 13.98 (s, 1H, PhOH), 13.28 (s, 1H, PhOH), 8.30 (d, 1H, *J* = 7.8 Hz, ArH), 7.80 (dd, 1H, *J* = 7.8, 8.3 Hz, ArH), 7.41 (d, 1H, *J* = 8.3 Hz, ArH), 7.00 (d, 1H, *J* = 8.3 Hz, sugarNH), 5.52 (d, 1H, *J* = 5.8 Hz, C₄H), 5.38 (d, 1H, *J* = 18.1 Hz, C₁₄H), 5.30 (d, 1H, *J* = 18.1 Hz, C₁₄H), 5.21 (s, 1H, C₉OH), 4.89 (d, 1H, *J* = 4.1 Hz, C_{7eq}H), 4.76 (m, 1H, C₁H), 4.30 (m, 1H, C₁₆H), 4.20 (q, 1H, *J* = 6.3 Hz, C₅H), 4.18 (m, 1H, NlacαH), 4.08 (s, 3H, C₄OCH₃), 3.67 (m, 1H, C₄OH), 3.45 (m, 1H, C₃H), 3.27 (d, 1H, *J* = 18.5 Hz, C_{10eq}H), 2.99 (d, 1H, *J* = 18.5 Hz, C_{10ax}H), 2.17 (d, 1H, *J* = 14.1 Hz, C_{8eq}H), 2.13 (dd, 1H, *J* = 4.1, 14.1 Hz, C_{8ax}H), 1.75 (m, 2H, C₂H), 1.25 (m, 9H, C₅CH₃, C₁₈CH₃, Nlac CH₃); ¹³C NMR (100 MHz, CDCl₃) δ 212.56, 186.88, 186.45, 173.87, 171.59, 161.38, 156.14, 155.59, 135.49, 134.78, 133.86, 133.57, 120.70, 119.68, 119.23, 118.32, 111.25, 111.01, 100.56, 76.23, 69.45, 68.21, 68.08, 66.05, 56.50, 44.82, 35.58, 33.59, 33.11, 29.42, 29.25, 20.42, 17.29; UV (CH₂Cl₂): λ_{max} (log ε) = 252 (0.24), 265 (0.22), 485 (0.11); Mass (FAB⁺, Na) *m/z* 711 (M + Na)⁺.

Daunomycin-14-stearate hydrochloride (6). 14-Bromo-daunomycin hydrochloride (**1c**, 0.40 g, 0.62 mmol) and stearic acid (0.35 g, 1.24 mmol) were dissolved in acetone (300 mL). To the mixture was added triethyl amine (0.17 mL, 1.24 mmol), and the mixture was then stirred at room temperature for 5 hr. After removing the solvent by a rotary evaporator, an ethereal HCl in dry THF (200 mL) was added to the reaction mixture. The resulting mixture was stirred at -20 °C for 2 hr and further stirred at room temperature for 3 hr, and then the solvent was removed under reduced pressure. Purification of the residue by column chromatography (CH₂Cl₂/CH₃OH/HCO₂H/H₂O = 100 : 15 : 2 : 1) gave daunomycin-14-stearate hydrochloride (**6**, 0.51 g, 97%) as a red powder: mp 100-102 °C; [α]_D²⁰ +112.04° (c 0.1, CH₃OH); IR (KBr) 3442, 2927, 2853, 1743, 1621, 1582, 1414, 1286, 1205, 1118, 984, 815 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.86 (s, 1H, PhOH), 13.10 (s, 1H, PhOH), 8.33 (s, 2H, NH₂), 7.87 (m, 2H, ArH), 7.67 (m, 1H, ArH), 5.63 (d, 1H, *J* = 6.3 Hz, C₄H), 5.54 (s, 1H, C₉OH), 5.32 (d, 1H, *J* = 18.5 Hz, C₁₄H), 5.16 (d, 1H, *J* = 3.9 Hz, C_{7eq}H), 5.09 (d, 1H, *J* = 18.5 Hz, C₁₄H), 4.91 (m, 1H, C₁H), 4.20 (q, 1H, *J* = 6.3 Hz, C₅H), 3.97 (s, 3H, C₄OCH₃), 3.58 (m, 1H, C₄OH), 3.34 (m, 1H, C₃H), 3.01 (d, 1H, *J* = 18.0 Hz, C_{10eq}H), 2.86 (d, 1H, *J* = 18.0 Hz, C_{10ax}H), 2.36 (m, 2H, C₁₆H), 2.26 (m, 2H, C₈H), 1.52-1.92 (m, 4H, C₂H, C₁₇H), 1.19 (m, 31H, C₁₈₋₃₁CH₂, C₅CH₃), 0.82 (m, 3H, C₃₂CH₃); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 208.01, 186.30, 185.48, 173.42, 174.41, 165.34, 160.79, 135.67, 135.09, 134.20, 133.84, 121.34, 119.94, 118.99, 114.10, 110.60, 99.42, 75.13, 69.56, 66.24, 65.45, 56.59, 46.50, 36.11, 35.24, 34.43, 33.43, 31.38, 31.30, 30.99, 29.69, 29.67, 29.63, 29.51, 29.47, 29.37, 29.14, 29.05, 28.72, 28.32, 24.44, 22.55, 22.10, 16.68, 13.94; UV (CH₃OH): λ_{max} (log ε) = 233 (0.81), 250 (0.51), 482 (0.19); Mass (FAB⁺, Na) *m/z* 833 (M-HCl + Na)⁺.

Daunomycin-3'-*N*-stearicarboamide (7). The mixture

of stearic acid (0.36 g, 1.27 mmol) and EDCI (0.24 g, 1.27 mmol) in dry DMF (100 mL) was stirred on an ice bath for 30 min and allowed to reach to room temperature. To the stirred solution was added daunomycin hydrochloride (**1a**, 0.60 g, 1.06 mmol) and catalytic amounts of 4-pyrrolidino-pyridine, and the mixture was then stirred for 3 hr. The resulting mixture was diluted with CH₂Cl₂ (200 mL), washed with water (2 × 200 mL) and brine (2 × 200 mL), dried over MgSO₄, and the solvent was removed under reduced pressure. The residue was purified by column chromatography (CH₂Cl₂/Hexane/CH₃OH = 12 : 4 : 1) to give daunomycin-3'-*N*-stearicarboamide (**7**, 0.88 g, 98%) as a red powder: mp 89-90 °C; [α]_D²⁰ +142.02° (c 0.1, CH₂Cl₂); IR (KBr) 3469, 2939, 2853, 1738, 1658, 1621, 1584, 1547, 1443, 1412, 1289, 1209, 1129, 1129, 1030, 990, 815 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 13.91 (s, 1H, PhOH), 13.61 (s, 1H, PhOH), 7.95 (d, 1H, *J* = 7.8 Hz, ArH), 7.72 (dd, 1H, *J* = 7.8, 8.3 Hz, ArH), 7.33 (d, 1H, *J* = 8.3 Hz, ArH), 5.51 (d, 1H, *J* = 6.8 Hz, C₄H), 5.29 (s, 1H, C₉OH), 5.18 (d, 1H, *J* = 4.4 Hz, C_{7eq}H), 5.14 (m, 1H, C₁H), 4.26 (m, 1H, C₃H), 4.09 (q, 1H, *J* = 6.8 Hz, C₅H), 4.02 (s, 3H, C₄OCH₃), 3.61 (m, 1H, C₄OH), 3.33 (m, 1H, C₃H), 3.13 (d, 1H, *J* = 18.5 Hz, C_{10eq}H), 2.80 (d, 1H, *J* = 18.5 Hz, C_{10ax}H), 2.38 (s, 3H, C₁₄CH₃), 2.24 (d, 1H, *J* = 14.6 Hz, C_{8eq}H), 2.08 (dd, 1H, *J* = 4.4, 14.6 Hz, C_{8ax}H), 2.01 (m, 2H, NstC₂CH₂), 1.80 (m, 2H, NstC₃CH₂), 1.39-1.64 (m, 2H, C₂H), 1.20 (m, 28H, NstC₄₋₁₇CH₂), 1.12 (d, 3H, *J* = 6.3 Hz, C₅CH₃), 0.84 (m, 3H, NstC₁₈CH₃); ¹³C NMR (100 MHz, CDCl₃) δ 212.27, 186.92, 186.46, 173.39, 172.29, 160.89, 156.38, 155.77, 135.63, 135.42, 134.50, 133.94, 120.74, 119.75, 118.28, 111.30, 111.12, 100.67, 76.53, 70.49, 69.99, 66.29, 56.58, 43.67, 36.62, 35.13, 34.22, 33.32, 31.88, 31.54, 30.46, 29.66, 29.62, 29.50, 29.44, 29.33, 29.27, 29.18, 25.53, 25.20, 24.90, 22.65, 22.61, 16.94, 14.08; UV (CH₂Cl₂): λ_{max} (log ε) = 235 (0.33), 252 (0.21), 485 (0.10); Mass (FAB⁺, Na) *m/z* 863 (M + Na)⁺.

Doxorubicin-3'-*N*-stearicarboamide (8). The mixture of stearic acid (0.41 g, 1.45 mmol) and EDCI (0.28 g, 1.45 mmol) in dry DMF (100 mL) was stirred on an ice bath for 30 min and allowed to reach to room temperature. To the stirred solution was added doxorubicin hydrochloride (**1b**, 0.70 g, 1.21 mmol) and catalytic amounts of 4-pyrrolidino-pyridine, and the mixture was then stirred for 3 hr. The resulting mixture was diluted with CH₂Cl₂ (300 mL), washed with water (2 × 300 mL) and brine (2 × 300 mL), dried over MgSO₄, and the solvent was removed under reduced pressure. The residue was purified by column chromatography (CH₂Cl₂/Hexane/CH₃OH = 12 : 4 : 1) to give doxorubicin-3'-*N*-stearicarboamide (**8**, 0.96 g, 98%) as a pale red powder: mp 97-99 °C; [α]_D²⁰ +97.01° (c 0.1, CH₂Cl₂); IR (KBr) 3482, 2927, 2853, 2738, 1627, 1584, 1449, 1412, 1289, 1209, 1123, 1018, 987, 815, 790 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 13.80 (s, 1H, PhOH), 13.00 (s, 1H, PhOH), 7.87 (d, 1H, *J* = 7.8 Hz, ArH), 7.65 (dd, 1H, *J* = 7.8, 8.3 Hz, ArH), 7.25 (d, 1H, *J* = 8.3 Hz, ArH), 5.86 (d, 1H, *J* = 6.8 Hz, C₄H), 5.37 (s, 1H, C₉OH), 5.07 (d, 1H, *J* = 4.4 Hz, C_{7eq}H), 4.98 (m, 1H, C₁H), 4.58 (s, 2H, C₁₄H), 4.01-

4.12 (q, 1H, $J = 6.8$ Hz, C₅H), 3.92 (s, 3H, C₄OCH₃), 3.53 (m, 1H, C₄OH), 3.23 (m, 1H, C₃H), 3.09 (d, 1H, $J = 18.5$ Hz, C_{10eq}H), 2.75 (d, 1H, $J = 18.5$ Hz, C_{10ax}H), 2.36 (d, 1H, $J = 14.6$ Hz, C_{8eq}H), 2.00 (dd, 1H, $J = 4.4, 14.6$ Hz, C_{8ax}H), 1.95 (m, 2H, NstC₂CH₂), 1.72 (m, 2H, C₂H), 1.60 (m, 2H, NstC₃CH₂), 1.11-1.23 (m, 31H, C₅CH₃, NstC₄₋₁₇CH₂), 0.78 (m, 3H, NstC₁₈CH₃); ¹³C NMR (100 MHz, CDCl₃) δ 206.74, 186.74, 186.27, 173.18, 172.54, 160.83, 156.18, 155.56, 135.63, 135.28, 133.88, 133.64, 120.57, 119.71, 118.32, 111.29, 111.11, 100.63, 77.06, 69.73, 69.61, 67.25, 65.84, 56.48, 45.07, 36.71, 33.87, 33.43, 31.87, 31.53, 29.86, 29.65, 29.64, 29.60, 29.45, 29.31, 29.26, 29.22, 29.08, 25.66, 24.90, 22.63, 22.60, 16.66, 14.07; UV (CH₂Cl₂): λ_{\max} (log ϵ) = 235 (0.37), 252 (0.22), 482 (0.10); Mass (FAB⁺, Na) m/z 833 (M + Na)⁺.

Doxorubicin-14, 3'-disteariccarboamide (9). The mixture of stearic acid (0.75 g, 2.65 mmol) and EDCI (0.58 g, 3.02 mmol) in dry DMF (100 mL) was stirred in an ice bath for 30 min and allowed to reach to room temperature. To the stirred solution was added doxorubicin hydrochloride (**1b**, 0.70 g, 1.21 mmol) and catalytic amounts of 4-pyrrolidino-pyridine, and the mixture was stirred for 8 hr. The resulting mixture was extracted with CH₂Cl₂ (300 mL), washed with water (2 × 300 mL) and brine (2 × 300 mL), dried over MgSO₄, and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel (CH₂Cl₂/Hexane/CH₃OH = 12 : 6 : 1) to give compound **9** (1.12 g, 86%) as a pale red solid; mp 82-84 °C; [α]_D²⁰ +187.05° (c 0.1, CH₂Cl₂); IR (KBr) 3494, 2927, 2853, 1738, 1640, 1584, 1541, 1467, 1418, 1289, 1215, 1123, 1018, 987 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 13.93 (s, 1H, PhOH), 13.16 (s, 1H, PhOH), 7.99 (d, 1H, $J = 7.8$ Hz, ArH), 7.74 (dd, 1H, $J = 7.8, 8.3$ Hz, ArH), 7.35 (d, 1H, $J = 8.3$ Hz, ArH), 5.53 (d, 1H, $J = 6.8$ Hz, C₄H), 5.29 (s, 1H, C₉OH), 5.24 (d, 1H, $J = 18.0$ Hz, C₁₄H), 5.18 (d, 1H, $J = 18.0$ Hz, C₁₄H), 5.07 (d, 1H, $J = 4.4$ Hz, C_{7eq}H), 4.51 (m, 1H, C₁H), 4.12 (q, 1H, $J = 6.8$ Hz, C₅H), 4.09 (s, 3H, C₄OCH₃), 3.62 (m, 1H, C₄OH), 3.24 (m, 1H, C₃H), 3.23 (d, 1H, $J = 19.0$ Hz, C_{10eq}H), 2.94 (d, 1H, $J = 19.0$ Hz, C_{10ax}H), 2.43 (d, 1H, $J = 14.6$ Hz, C_{8eq}H), 2.14 (dd, 1H, $J = 4.4, 14.6$ Hz, C_{8ax}H), 2.02 (m, 4H, C₁₆CH₂, NstC₃CH₂), 1.82 (m, 2H, C₂H), 1.66 (m, 4H, C₁₇CH₂, NstC₃CH₂), 1.05-1.40 (m, 59H, C₅CH₃, C₁₈₋₃₁CH₂, NstC₄₋₁₇CH₂), 0.84 (m, 6H, C₃₂CH₃, NstC₁₈CH₃); ¹³C NMR (100 MHz, CDCl₃) δ 206.79, 18.05, 186.52, 182.99, 174.02, 173.44, 173.20, 172.34, 160.98, 156.24, 155.78, 149.64, 135.71, 135.47, 135.07, 134.07, 133.62, 132.56, 120.79, 119.83, 118.32, 118.24, 113.37, 111.42, 111.25, 100.61, 95.42, 77.11, 75.14, 75.02, 70.56, 69.77, 66.52, 65.90, 60.37, 56.62, 43.69, 36.61, 35.52, 34.24, 33.88, 33.58, 31.90, 31.56, 30.46, 29.68, 29.64, 29.52, 29.45, 29.34, 29.27, 29.18, 29.09, 25.50, 25.23, 24.90, 22.67, 22.63, 21.64, 21.02, 16.93, 14.17, 14.09; UV

(CH₂Cl₂): λ_{\max} (log ϵ) = 235 (0.36), 252 (0.22), 482 (0.11); Mass (FAB⁺, Na) m/z 1099 (M + Na)⁺.

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