

## Change in Photosynthesis, Proline Content, and Osmotic Potential of Corn Seedling under High-Saline Condition

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**ABSTRACT :** To identify salt-tolerance characteristics of corn seedling was treated in solution of 0, 50 and 100 mM NaCl of hydroponic cultivation. In photosynthesis of corn seedling, there was no large difference between 50mM and 0 mM NaCl solution, however, in 100 mM NaCl solution, the tolerance gradually decreased to 76%, 49%, and 31% after one day, four days, and seven days, respectively, in comparison to 0 mM NaCl solution. Osmotic potential of corn in seedling period was significantly decreased with increasing saline level, however, free proline content in the plant on the ground was significantly increased with increasing saline level and with the lapse of time. In terms of correlation among major characteristics, there was a highly significant positive difference between osmotic pressure potential and photosynthesis, However, highly negative correlation was found between osmotic pressure potential and free proline content. In addition, it was expected that young seedling of corn with saline tolerance may be utilized in the transplantation in salt-accumulated land. Based on above-shown result, in terms of saline tolerance of Chalok-2 variety, growth suppression was serious with 100 mM NaCl solution. However, growth was expected that seedling growth would be favorable under 50 mM NaCl solution.

**Keywords:** seedling corn, saline, NaCl concentration, photosynthetic rate, osmotic potential, free proline content

In general, synthesis of chlorophyll, leaf area, affected photosynthesis and respiration, and enzyme activation was affected under saline condition (Beatriz *et al.*, 2001; Cho & Kim, 1998; Lee *et al.*, 1996; Lee & Kim, 1995; Volkmar *et al.*, 1998). This saline stress decreased productivity and quality of farm products. However, it was known that an appropriate increase in saline level results in improvement in quality such as increased sugar contents in tomato. Saline damage accelerate the aging of farm products and, especially, photosynthetic capacity was significantly influenced in intracellular ion balance and zoning; Volkmar *et al.* (1998) and Cho *et al.* (2002) reported that optimum levels of

Na<sup>+</sup> and K<sup>+</sup> are unbalanced under saline stress; Mohammad (1994) reported that stomatal conductance is decreased due to suppressed water absorption; and Abou-Hadid (1992) reported photosynthesis of plants is affected due to decreased stomatal transpiration. While, there was also a report that affection of water absorption was resulted due to decreased water potential and excessive soil caused by toxic activity of specific ions existing as high levels or salts existing in plant medium (Megel & Kirkby, 1978). Salt gives a large influence on metabolism process of plant. In case of farm products with high salt-tolerant property, accumulation of free amino acid, especially proline, it was apparent under the condition with insufficient water. According to Xiaomu *et al.* (1995), although proline was the amino acid that was shown in small quantity under the favorable plant cultivation condition, it was accumulated in large quantities under the unfavorable physiological condition such as saline damage or drought damage. James *et al.* (1996) reported that proline acted on membrane osmotic pressure between cytoplasm and intercellular space, enzyme defense, and pH of cytoplasm and accumulation of proline is related to tolerance capacity. Corn is C<sub>4</sub> plant with large sizes and excellent carbon dioxide assimilation capacity with light and grows fast with absorption of large quantity of fertilizers with strong fertilizer absorption capacity (Marschner, 1986); Choi *et al.* (1997) reported that the larger photosynthetic capacity and rooting power in species, the higher salt-tolerance. Frensch & Hsiao (1994) reported that corn roots exposed to mannitol or KCl show immediate decrease in water potential and growth rate within 30 minutes; Neumann *et al.* (1994) reported that low growth of root under salts in seedling period of corn wasn't affected influence in the change of osmotic pressure but makes cell walls hard; and Evlagon *et al.* (1992) reported that Ca<sup>2+</sup> accumulation in the root causes an important role to NaCl reaction. Zidan *et al.* (1990) reported that initial growth of roots in corn seedling period was significantly suppressed by treatment with high-level NaCl. The purpose of these studies were to get the data of salt-tolerance capacity and physiological characteristics of saline reaction for transplantation during seedling period in salt-accumulated cultivation land and reclaimed

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land through determination of photosynthesis, osmotic pressure potential, free proline in plant under different concentrations of NaCl in water culture of corn seedling.

### Materials and Methods

Chalok-2 corn (*Zea mays* L.) was cultivated in a growing seedling box for 25 days and, after transplantation in a water culture box with salt concentration of 0 mM, 50 mM and 100 mM NaCl. this process was repeated three times after one, four, and seven days. Leaf photosynthetic rates (LPS) were measured with a photosynthesis measuring apparatus LAC-4 (Analytical Development Company, Ltd. UK). Stomatal conductance (Gs) and intercellular CO<sub>2</sub> concentrations (Ci) were also concurrently measured. Those leaves were ground; 0.8 µl was taken into a chamber and stabilized for 25 minutes; and then, the chamber was connected with water potential measuring apparatus (HR-33T Dew point Microvoltmeter, INC. USA) to measure osmotic potential of leaves. Photometric method was used in measurement of free proline according to the method of Troll & Lindsley (1955). 1.0 g of living body of above ground plant was ground in centrifuge tube with liquid nitrogen. 10 ml of MCW solution (methanol : chloroform : water = 12 : 5 : 1) was added to the ground solution and centrifuged at 5,000 rpm with 4 °C for 10 minutes. 2 ml of upper layer in centrifuged tube was taken after centrifuge and, 3 ml of glacial acid and 3 ml of ninhydrine reagent (ninhydrine : glacial acid : 6M-phosphoric acid = 6.25 g : 150 ml : 100 ml) was added and boiled in water bath for 1 hour; after cooling of the solution at normal temperature, 5 ml of toluene was added and shaken vertically for overnight; absorption of spectrophotometer was measured at 520 nm. L-proline was used to get standard curve of proline.

### RESULTS AND DISCUSSION

Table 1 showed the differences of photosynthetic rate, stomatal conductance, and intercellular CO<sub>2</sub> concentration by saline level in water culture of corn seedling. Photosynthetic rate and stomatal conductance decreased significantly as the level of saline increased and, although no apparent difference was shown between 0 mM and 50 mM NaCl in which photosynthetic rate in 50 mM treatment was 86 - 90% in comparison to 0 mM with the lapse of time. But, in case of 100 mM NaCl solution, photosynthetic rate was 76% of 0 mM at one day after saline treatment and decreased significantly to 31% in comparison to 0 mM at seven days after saline treatment. In case of 100 mM, stomatal conductance was 75% of 0 mM at one day after treatment and decreased significantly with 33% in photosynthetic rate in comparison to 0 mM at seven days after saline treatment. Photosynthetic rate and stomatal conductance were closely related and showed positive correlation. Stomatal conductance is used in diagnosis of opening of stoma on the leaves. Stomata were not only the pathway of CO<sub>2</sub> dispersion but also the pathway of stoma transpiration. Decrease in stomatal conductance with increase of saline was a protective mechanism to suppress water loss caused by stomatal transpiration, so stomatal conductance is related to photosynthesis. In this study, in case of 50 mM which was low-saline, plant did not receive large saline damage. However, in case of 100 mM which was high-saline treatment, photosynthetic rate is significantly decreased due to closing of stoma. Xu *et al.* (1994) reported that decreased photosynthesis under high saline level is the cause of the decrease of stomatal conductance. The results of this study were consistent with result of Lee *et al.* (1998) in which photosynthesis of tobacco decreased significantly with elevated saline level under

**Table 1.** Changes in leaf photosynthetic rate of corn seedling grown under different NaCl treatment

Days after treatment	NaCl concentration (mM)	A <sup>1</sup>	Gs	Ci
		(µmol m <sup>-2</sup> s <sup>-1</sup> )	(µmol m <sup>-2</sup> s <sup>-1</sup> )	(ppm)
1	0	21.55 ± 0.55(100)*	0.16 ± 0.01	148.2 ± 14.3
	50	19.35 ± 0.70(90)	0.12 ± 0.01	110.7 ± 17.4
	100	16.45 ± 0.66(76)	0.12 ± 0.01	140.6 ± 17.6
4	0	19.00 ± 1.08(100)	0.16 ± 0.02	114.2 ± 9.4
	50	16.29 ± 0.14(86)	0.12 ± 0.00	99.6 ± 6.6
	100	9.38 ± 0.78(49)	0.07 ± 0.00	90.2 ± 1.7
7	0	15.81 ± 0.66(100)	0.12 ± 0.01	105.7 ± 10.7
	50	14.16 ± 0.24(90)	0.10 ± 0.01	109.3 ± 13.7
	100	4.95 ± 0.44(31)	0.04 ± 0.00	120.1 ± 12.7

A<sup>1</sup>, Leaf photosynthetic rate, Gs, Stomatal conductance

Ci, Intercellular CO<sub>2</sub> concentration

\*, Figures in the parentheses are percentage ratio to the control

saline stress condition. Fig. 2 showed the changes of osmotic pressure potential of leaves juice by saline level in water culture of corn seedling. Osmotic pressure potential of leaves juice of 50 mM decreased to 20% of 0 mM at one day after treatment and it decreased to 58% in case of 100 mM NaCl solution. Luttus *et al.* (1995) reported that significant decrease in water potential of leaves with saline stress was caused by intracellular accumulation of organic or inorganic solute. Katsuhiko *et al.* (1996) reported that when water potential of plant decreased to -2.0 or lower MPa, there were suppression of cell division, suppression of cell wall and

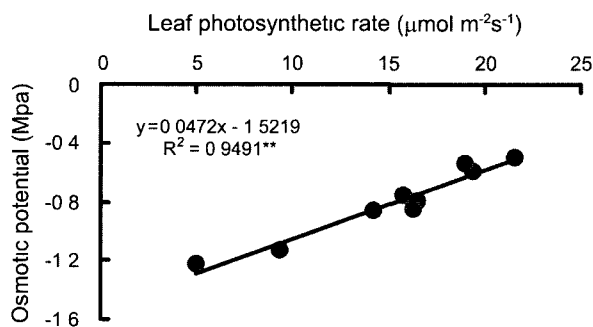


Fig. 1. Relationship between leaf osmotic potential and leaf photosynthetic rate of corn seedling under different NaCl treatment.

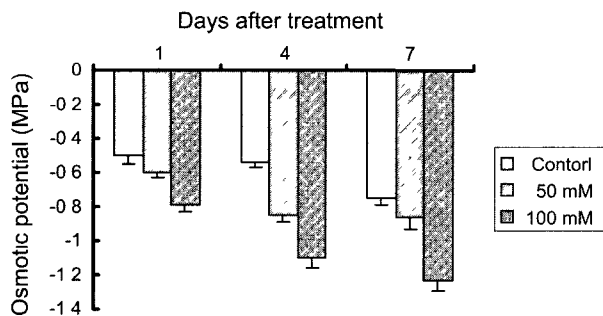


Fig. 2. Changes of osmotic potential of leaf of corn seedling under different NaCl treatment.

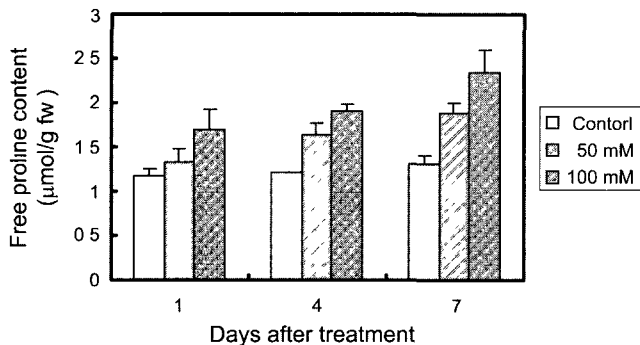


Fig. 3. Changes of free proline content of corn seedling under different NaCl treatment

protein synthesis, accumulation of solute, closing of stoma, and suppression of photosynthesis were resulted with withering to die finally. In this study, photosynthesis in low-saline as much as 50 mM showed slight decrease comparison to 0 mM due to salt tolerance of corn but high-saline as much as 100 mM large decrease which exceeded the limit of salt tolerance of corn. Fig. 3 showed changes of the content of free proline of above ground corn seedling by saline level in water culture. The content of free proline was increased 1.14 time of 0 mM in case of 50 mM at one day after saline treatment and to 1.45 time in case of 100 mM. The content of free proline increased to 1.45 time of 0 mM in case of 50 mM at seven days after saline treatment add to 1.80 time in case of 100 mM. The content of free proline increased with higher level of saline and increased by the lapse of time also. This result was consistent with the report of Lee *et al.* (1992) in which proline content increased significantly with elevated NaCl level in rice plant, and consistent with the report of Heuer & Nadler (1998) in which proline was accumulated and adjusted osmotic pressure in the plant under saline damage in potatoes. Hanson *et al.* (1979) reported that carboxy group of glutamic acid which the precursor of proline, was reoxidated to aldehyde group to synthesize proline under water insufficient situation, and the synthesized proline was translocation to other site. Huber *et al.* (1974) reported that NaCl promoted the activation of  $\Delta$ -pyrroline-5-carboxylase that induce proline accumulation, and Choi *et al.* (1997) reported that increase of free amino acid under drought damage or saline damage was caused by complicated factors of metabolism process with resolution of protein, suppression of protein synthesis in amino acid, and suppression of photosynthesis. He also reported that in terms of metabolism of free amino acid, it might be considered as adaption to inappropriate environment. In this study also, it was found that content of proline increased with the elevated saline level of corn by the lapse of time. Osmotic pressure potential and photosynthetic rate had highly positive correlation (Fig. 1). That means that the higher osmotic pressure potential was, the more photosynthetic rate increased; the lower the osmotic pressure potential was, the less photosynthetic rate decreased. This phenomenon may be considered as the result of suppressed photosynthetic rate due to insufficient water condition (Katsuhiko *et al.*, 1996). Stomatal conductance has highly positive correlation with osmotic pressure potential. Bethke *et al.* (1992) reported that low water potential closes stoma. Stomatal conductance is presumed to be changed by cellular turgor pressure caused by osmotic pressure potential. Osmotic pressure potential and free proline content shows highly positive correlation; this was consistent with the report by James *et al.* (1996) in which osmotic pressure adjust is made in the plant body

through accumulation of chloride and proline under saline damage on potatoes, and with consistent the report by McNulty (1985) that proline was synthesized by supplement cellular water stress caused by salt and it took an important role in osmotic pressure adjustment.

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### REFERENCES

- Abou-Hadid, A. F. 1992 Protected cultivation for tomato under salt affected soils. *Acta Hort.* 323 : 443-449.
- Balibrea, M. E., E. Cayuela, F. Artes, and F. Perz-Alfocea 1997. Salinity effects on some postharvest quality factors in a commercial tomato hybrid. *J. Hort. Sci.* 72 : 885-892.
- Beatriz, G., Neves-Piestun, and Nirit Bernsten. 2001. Salinity-induced inhibition of leaf elongation in maize is not mediated by changes in cell wall acidification capacity. *Plant Physiol.* 125 : 1419-1428.
- Bethke, P. C. and M. C. Drwe. 1992 Stomatal and nonstomatal components to inhibition of photosynthesis in leaves of *Capsicum annuum* during progressive exposure to NaCl salinity. *Plant Physiol.* 99 : 219-226.
- Cho, J. W. and C. S. Kim. 1998 Effect of NaCl concentration on photosynthesis and mineral content of barley seedlings under solution culture. *Korean J. Crop Sci.* 43(3) : 152-156.
- Cho, J. W., C. S. Kim, and J. D. So. 2002. Effect of NaCl stress on inorganic ion, L-proline, sugar and starch content of soybean seedlings. *Korean J. Crop Sci.* 47(2) : 75-79.
- Choi, W. Y., J. H. Park, and Y. W. Kwon 1997. Physiological responses of barley to salt stress at reproductive stage. *Korean J. Crop Sci.* 42(6) : 687-692.
- Evlagon, D., I. Ravina, and P. M. Neumann 1992 Effects of salinity stress and calcium on hydraulic conductivity and growth in maize seedling roots. *J. Plant Nutr.* 15 : 795-803.
- Frensch, J. and T. C. Hsiao 1994. Transient responses of cell turgor and growth of maize roots as affected by changes in water potential. *Plant Physiol.* 104 : 247-254.
- Hanson, A. D., C. E. M. Nansen, A. R. Pedersen, and E. H. Everson. 1979. Capacity for proline accumulation during water stress in barley and its implication for drought resistance. *Crop Sci.* 19 : 489-493.
- Heuer, B. and A. Nadler. 1998 Physiological response of potato plants to soil salinity and water deficit. *Plant Sci.* 137 : 43-51.
- Huber, W., P. N. Rustagi, and N. Sankhria. 1974 Effect of sodium chloride and gibberellic acid on the activity of the enzyme of carbohydrate metabolism in leaves of *pennisetum typhoides*. *Ecophysiological studies on indian arid zone plants.* Bot. 77-84.
- James, R. D. and M. L. Binzel 1996. NaCl reduces indol-3-acetic acid levels in the roots of tomato plants independent of stress-induced abscisic acid. *Plant Physiol.* 112 : 379-384.
- Katsuhiko, W., H. Tadashi, and I. Kuni. 1996. Analysis of photosynthesis depression under low leaf water potential by comparison of CO<sub>2</sub> exchange and O<sub>2</sub> evolution rates. *J. J. Crop Sci.* 65(4) : 590-598.
- Lee, K. S., J. S. Lee, and S. Y. Choi. 1992. Changes in contents of chlorophyll and free proline as affected by NaCl in rice seedling. *Korean J. Crop Sci.* 37(2) : 178-184.
- Lee, S. G., J. S. Shin, Y. S. Seo, and G. K. Bae. 1998 Effects of salt stress on photosynthesis free proline content in tobacco. *Korean J. Environmental Agriculture.* 17(3) : 215-219.
- Lee, S. Y. and C. S. Kim. 1995. Cellular structural change of barley seedling on different salt concentration under hydroponic culture. *Korean J. Crop Sci.* 40 : 481-486.
- Lee, S. Y., C. S. Kim, J. W. Cho, and Y. G. Kang 1996. Physiological response of barley seedlings to salt stress. *Korean J. Crop Sci.* 41 : 665-671.
- Lutts, S., J. M. Kinet, and J. Bouharmont 1995 Changes in plant response to NaCl during development of rice (*Oryza sativa* L.) varieties differing in salinity resistance. *J. Exper. Bot.* 46(293) : 1843-1852.
- Marschner. H. 1986. Mineral nutrition in higher plants. Academic Press Inc Ltd. London PP. 674.
- McNulty, I. B. 1985. Rapid osmotic adjustment by a succulent halophyte to saline shock. *Plant Physiol.* 78 : 100-103.
- Megel, K. and E. A. Kirkby. 1978 'In principles of Plant Nutrition' International Potash Institute Switzerland. 193-210.
- Mohammad, P. 1994. Handbook of plant and crop stress Marcel Dekker press. New York p. 235-250.
- Neumann, P. M., H. Azaizeh, and D. Leon. 1994. Hardening of roots cell walls; a growth inhibitory response to salinity stress. *Plant Cell Environ.* 16 : 15-24.
- Shalhevet, J., M. G. Huck, and B. P. Schroeder 1995. Root and shoot growth responses to salinity in maize and soybean. *Agron. J.* 87 : 512-516.
- Troll, W. and J. Linsley. 1955 A photometric method for the determination of proline. *J. Biol. Chem.* 215 : 655-660.
- Volkmar, K. M., Y. Hu, and H. Steppuhn. 1998 Physiological response of plant to salinity: A review. *Can. J. Plant Sci.* 78 : 19-27.
- Xiaomu, Niu, A. B. Ray, M. H. Paul, and M. P. Jose. 1995 Ion homeostasis in NaCl stress environments. *Plant Physiol.* 109 : 735-742.
- Xu, H. L., L. Gauthier, and A. Gosselin. 1994 Photosynthetic responses of greenhouse tomato plants to high solution electrical conductivity and low soil water content. *J. Hort. Sci.* 69 : 821-832.
- Zidan, I., H. Azaizeh, and P. M. Neumanu. 1990. Does salinity reduce growth in maize root epidermal cells by inhibiting their capacity for cell wall acidification? *Plant Physiol.* 93 : 7-11.