# Taxonomic Reappraisal of *Sphacelaria rigidula* and *S. fusca* (Sphacelariales, Phaeophyceae) Based on Morphology and Molecular Data with Special Reference to *S. didichotoma*

# Yeon-Shim Keum, Jung Hyun Oak, Stefano G.A. Draisma<sup>2</sup>, Willem F. Prud'homme van Reine<sup>2</sup> and In Kyu Lee<sup>1</sup>

Division of Life Science, Gyeongsang National University, Jinju 660-701, <sup>1</sup>School of Biological Sciences, Seoul National University, Seoul 151-742, Korea and <sup>2</sup>Universiteit Leiden Branch, Nationaal Herbarium Nederland, P.O. Box 9514, 2300 RA Leiden, The Netherlands

The taxonomic status of three closely related species of the genus *Sphacelaria, S. rigidula* Kützing, *S. fusca* (Hudson) C.F Gray and *S. didichotoma* Saunders was reassessed on the basis of morphological observations from herbarium specimens and phylogenetic analysis of RUBISCO spacer sequences. *Sphacelaria rigidula* was characterized by having only 2-armed propagules and somewhat slender filaments. Culture experiments revealed that its propagules commonly formed secondary arms, even though they were rarely produced in the field. It is concluded that *S. divaricata* Montagne characterized by dichotomously branched arms in propagules shoud be synonymized with *S. rigidula*, based on propagule morphology and measurements. *Sphacelaria fusca* clearly be separated from *S. rigidula* by having 2-4 armed propagules. These morphological differences were highly supported by the differences in the RUBISCO spacer sequences. *S. didichotoma* resembles *S. rigidula* in having propagules with two dichotomously branched arms. However, it commonly produces the secondary arms in the field. Additionally, this species has shorter primary arms than *S. rigidula*. Phylogenetic analyses supported the distinction of *S. didichotoma* from *S. rigidula*. The northwestern Pacific plants, previously recorded as *S. divaricata*, were recognized as *S. didichotoma*.

Key Words: phylogenetic analysis, propagule morphology, RUBISCO spacer sequences, *Sphacelaria didichotoma, S. fusca, S. rigidula* 

## **INTRODUCTION**

The brown algal genus *Sphacelaria* subgenus *Propagulifera* is subdivided into three sections on the basis of the shape of propagules: viz. the section *Propagulifera* with fusiform arms and apical hairs, the section *Furcigerae* with mainly cylindrical arms and no apical hairs, and the section *Tribuloides* with tribuliform propagules (Prud'homme van Reine 1982). Of these, the section *Furcigerae* is supposed to include seven to ten species (Prud'homme van Reine 1993; Draisma *et al.* 1998; Keum *et al.* 1999). Species delimitation within this section is mainly based on the distinct shape of the propagules (Kitayama 1994; Draisma *et al.* 1998). The taxonomic status of two widely recorded species, *S. rigidula* Kützing and *S. fusca* (Hudson) C.F. Gray, has

\*Corresponding author (yskeum@hotmail.com)

been discussed extensively, and these names have been considered as a being included into one single species having 2-4 armed propagules (Holmes and Batter 1891; Haas-Niekerk 1965; van den Hoek and Flinterman 1968; Russell and Fletcher 1975). However, several authors stated that *S. rigidula* only forms 2-armed propagules (as *S. furcigera*; Sauvageau 1901, 1902; Zimmerman 1925; Irvine 1956; Goodband 1971), whereas *S. fusca* produces 2-3(-4) or only 3-armed propagules (Sauvageau 1901; Setchell and Gardner 1924). Prud'homme van Reine (1982) proposed that it is necessary to separate *S. rigidula* from *S. fusca*, although the taxonomic position of *S. fusca* is still uncertain, because *S. fusca* is different from *S. rigidula* by having thicker erect filaments.

Sphacelaria divaricata Montagne was first described on the basis of repeatedly branched 2-armed propagules from Torres Strait (Montagne 1849; Sauvageau 1901) and it has been recorded from the northwestern Pacific and Australia (Lee and Kang 1963; Kitayama 1994; Phillips 1997; Draisma 1998). Prud'homme van Reine (1982) considered that this species might be a synonym of *S. rigidula*, because a similar branching of the arms frequently occurs in *S. rigidula*. Another species showing repeatedly branched 2-armed propagules, *Sphacelaria didichotoma* Saunders (1898) has been recorded a few times from the northeastern Pacific (Setchell and Gardner 1925; Abbott and Hollenberg 1976; Hansen 1997). However, Kitayama (1994) synonymized this species with *S. divaricata* having similar repeatedly branched 2-armed propagules.

The variability of propagules, including the presence or absence of branching and number of arms, has given rise to much confusion, and thus further investigations are necessary to determine the validity of the four species mentioned above. The aim of the present study was to clarify the taxonomic status of the species based on morphological observations using herbarium specimens and determination of the nucleotide sequence in the RUBISCO spacer region.

#### MATERIALS AND METHODS

The specimens examined were loaned from the herbariums coded as AD (State Herbarium of South Australia), AST (Herbarium of the Institute of Oceanology, Chinese Academy of Sciences, Qingdao, China), L (Nationaal Herbarium Nederland, Leiden University branch), MEL (National Herbarium of Victoria, Royal Botanic Gardens, Australia), PC (Muséum National d'Histoire Naturelle, France), SNU (Seoul National University, Korea), TCD (Herbarium, Trinity College, Ireland), TNS (Herbarium, Botany Department, National Science Museum, Japan), and W (Naturhistorisches Museum Wien, Austria). Morphological observations by light microscopy were made from materials preserved in 10% formalin in seawater and from resoaked (in saturated detergent solution) herbarium specimens. Materials were stained in 1% aqueous aniline-blue solution and mounted in glycerin-seawater solution (1:1) on microscope slides. The slides were photographed with an Olympus BH-2 camera system. The culture was established from apical filaments, which were cultivated in PES medium at 15°C and 12:12 h LD under 15-30  $\mu$ mol m<sup>-2</sup>s<sup>-1</sup>. The descriptive terminology used by Prud'homme van Reine (1982) and Kitayama (1994) has been followed. Morphological features of herbarium specimens of Sphacelaria are provided in Table 1.

Collection information for species used in our molecular investigations is provided in Table 2. The materials were washed with distilled water and dried on filter paper. Genomic DNA was extracted using DNeasy Plant Mini Kit (Qiagen, Hilden, Germany), according to the manufacturer's instructions. The concentration of extracted total DNA was determined by comparing the brightness of DNA band on ethidium bromide-stained 0.7% agarose gels. The RUBISCO spacer region was amplified using the primers 1 and 2, which were designed by Stache-Crain et al. (1997). PCR amplification was carried out in 50  $\mu$ L total volume composed of 0.1-20 ng template DNA, 80  $\mu$ M for each dNTP, 1 unit of Ex Taq polymerase (TaKaRa Bio, Kyoto, Japan), 1x Thermophilic buffer (TaKaRa Bio), 1.5 mM MgCl<sub>2</sub>, and 10 pmol of each primer. Amplification reactions involved 5 min at 94°C for predenaturation, 30 cycles consisting of 30s at 94°C for denaturation, 30s at 48°C for annealing, and 30s at 72°C for extension, with a final extension of 7 min at 72°C, using a GeneAmp PCR Cycler 2400 (Perkin Elmer, Foster City, CA). The reaction at 4°C after amplification. PCR products were monitored on ethidium bromidestained 1.0 % agarose gels. For automated sequencing, PCR products were directly sequenced using the BigDye Terminator Cycle Sequencing Ready Reaction Kit (Perkin Elmer Applied Biosystems). Automated sequencing was used with 377 DNA Sequencing System (Perkin Elmer Applied Biosystems).

Sequences were aligned for phylogenetic analysis using Clustal X (Thompson et al. 1999). Phylogenetic trees were inferred from the sequence data using maximum parsimony (MP), maximum likelihood (ML), and neighbor-joining (NJ) methods, as implemented in PAUP\* 4.0 (Swofford 2000). MP analysis was performed using a heuristic search with random sequence addition and Tree-Bisection-Reconnection branch swapping options. All characters were unweighted and unordered, and gaps were treated as missing data. NJ analysis (Saitou and Nei 1987) employed a Kimura (1980) twoparameter model. ML analysis was based on the HKY85 model (Hasegawa et al. 1985) with transversions weighted 2 to 1 over transitions and empirical base frequencies. Bootstrap analyses (Felsenstein 1985) were used with 1,000 replicates for all three methods to estimate the stability of tree topologies.

Sphacelaria	
ie genus	
ecies in the	
s of the sp	
al features	
orphologic	
son of me	
. Compari	
Table 1. (	

Species	Locality	Specimen	Height	Length (µm)	Width (µm) width	Ratio length / width	Length (µm)	Width of waist (µm)	Primary arm length (μm)*	Arm width (µm)
S. rigidula	Eochungdo, Korea	SNU950629	< 1 cm	52 (42-69)	31(25-41)	1.7(1.4-1.9)	326(264-391)	23(18-26)	182(172-213)	21(20-22)
	cultured plant			46(40-57)	30(24-38)	1.6(1.2-1.8)	278(220-320)	22(20-25)	386(260-460)	18(15-21)
	Kalbarri, Australia	AD051282	< 5 mm	44(32-55)	34(27-40)	1.3(1.0-1.6)	234(193-331)	24(20-29)	175(164-181)	22(18-26)
	Punarum Pass, Tahiti	L937.117-17	5  mm	45 (39-54)	39(33-44)	1.2(1.1-1.3)	265(231 - 301)	29(23-33)	261(235-286)	24(22-25)
	New Guinea	L934.263-17	5  mm	33(25-39)	38(30-44)	0.9(0.8-0.9)	255(204-293)	30(25-33)	275(248-325)	28(26-30)
	Tanzania	L200346	7 mm	28(21 - 34)	22(15-26)	1.3(1.0-1.8)	231(186-288)	21(17-24)	161(153-168)	18(16-20)
S. divaricata	Torres Strait, Australia	Type, MA9712	1-2 mm	24-28	20-24		160-200	20-30	300-400	
	Fremantle, Australia	L937.117-15	2-3 mm	46(32-63)	30(23-39)	1.5(1.1-1.8)	162(123-216)	25(22-30)	389(237-493)	24(21-26)
S. fusca	England	Neotype, TCD	$3.5~\mathrm{cm}$	65-96	44-80	1.28-1.73	356-542	36-50	320-540	30-46
	Chaesukgang, Korea	SNU930506	< 2 cm	38(28-49)	42 (34-54)	0.9(0.7 - 1.3)	290(188-290)	29(25-36)	ı	28(26-31)
	Rumoi, Hokkaido Pref., Japan Kita-61, TNS	pan Kita-61, TNS	< 1.5 cm	44(33-54)	42(35-49)	1.0(0.8-1.4)	301(236-376)	34(26-36)	ı	
. didichotom	S. didichotoma Imagoura, Hyogo Pref., Japan SNU200409	pan SNU200409	< 1cm	33(28-43)	39(33-46)	0.9(0.7-1.2)	227(146-307)	41(30-55)	168(103-269)	29(27-32)

Species	Collection sites (collectors, collection years)	Accession no
Sphacelaria didichotoma	Tsuyazaki, Fukuoka, Japan (S. Kawaguchi, 1989)	AJ287949*
S. didichotoma	Imagoura, Hyogo Pref., Japan (YS. Keum, 2000-4-9)	AY898781
S. didichotoma	Maruyama, Awajishima, Hyogo Pref., Japan (YS. Keum, 2000-4-21)	AY898782
S. fusca	Galway, Ireland (W.F. Prud'homme van Reine, 1966-25-1)	AJ287941**
S. fusca	Chaesukgang, Korea (YS. Keum, 1996-5-6	AJ287944**
S. fusca	Muroran, Hokkaido, Japan (W.F. Prud'homme van Reine, 1993-5-6)	AJ287943**
S. fusca	North America, as S. cirrosa (UTEX)	AY898783
S. rigidula	Cape Verde Islands (H. Pakker, 1991)	AJ287942
S. rigidula	O-ura, Hyogo Pref., Japan (YS. Keum, 2001-6-9)	AY898784
S. rigidula	Haengwon, Cheju Island, Korea (YS. Keum, 1995-5-7)	AY898785
S. rigidula	Eochungdo, Korea (J.H. Oak, 1997-6-12)	AY898786
S. rigidula	Beomsum, Cheju Island, Korea (J.H. Oak, 1998-2-11)	AY898787
S. yamadae	Ogi, Sado Isl., Niigata Pref., Japan (T. Kitayama, 1990)	AJ287950
S. yamadae	Imagoura, Hyogo Pref., Japan (YS. Keum, 2000-5-14)	AY898788
S. californica	Oiso, Awajishima, Hyogo Pref., Japan (W.F. Prud'homme van Reine, 1993)	AJ287951
S. californica	Dokdo Island, Korea (Y.S. Oh, 1995-8-15)	AY898789
S. tribuloides	Sesoko, Okinawa, Japan (W.F. Prud'homme van Reine, 1993)	AJ287947
S. tribuloides	Jan Thiel, Curacao (A.H. van Engelen, 1999)	AJ287948
S. cirrosa	Fife Ness, Scotland (S.G.A. Draisma, 1977)	AJ287946

Table 2. List of species, collection localities, and DNA database accession numbers (GenBank) in this study (Accession numbers for new sequences (this study) are shown in bold)

\* in Draisma 2002: as S. divaricata

\*\* in Draisma 2002: as S. rigidula

#### RESULTS

#### Morphological observations

# Sphacelaria rigidula Kützing 1843, p. 292 Figs 1-12

Holotype: Red Sea, on *Cystoseira triquetra, Schimper s.n.* (L 937.117-229; Isotype, W 19682).

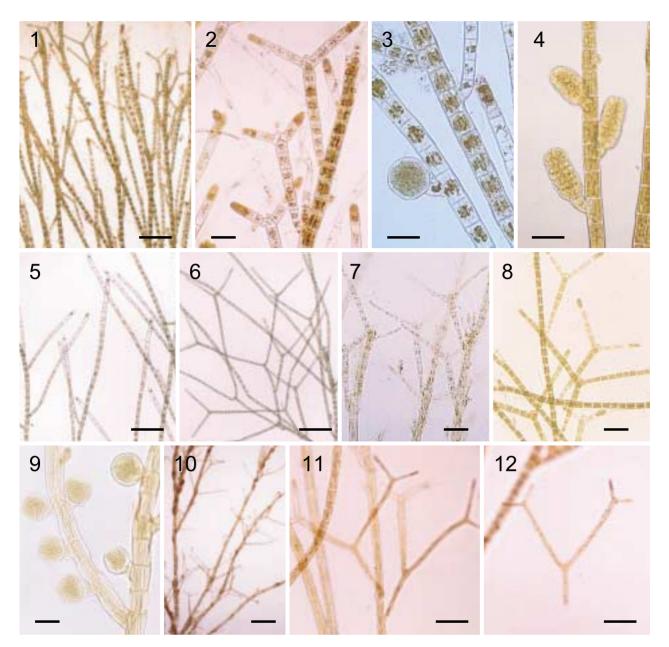
**Synonyms:** Sphacelaria furcigera Kützing 1855, p. 27, Karak Island, Persian Gulf (L 937.714-72). Sphacelaria divaricata Montagne 1849, p. 62, Torres Strait, Australia (Holotype, MA 9712, PC [Herb. Montagne], Isotype, TCD).

**Distribution:** warm temperate to tropical areas (Prud'homme van Reine 1982)

Specimens examined: KOREA. Eochungdo, 29 VI 1995, J.H. Oak (SNU950629). Haengwon, Cheju Island, 7 VI 1995, Y.-S. Keum (SNU950607). Beomsum, Cheju Island, 11 II 1998, J. H. Oak (SNU980211). JAPAN. O-ura, Hyogo Pref., 9 VI 2000, Y.-S. Keum (SNU20609). TAHITI. Punarum Pass, 2 VII 1922, Setchell and Rarks (L 937.117-17). NEW GUINEA ISLANDS. No specific locality and collector, 25 VIII 1899 (L 934.263-17). INDONESIA. No locality, date and collector (L 937.117-215). AUSTRALIA. WA. Jacques Pt, Kalbarri, 15 IX 1979, H.B.S. Womersley, (AD 51282). Point Valliant, Two People Bay, mid eulittoral pools, 30. viii. 1979, *H.B.S. Womersley* (L 471206). No specific locality and collector, 26 IV 1875 (W 19656). NEW ZEALAND. Stewart Island, 13 VI 1945, no collector (L 947.192-1). DAHOMEY. Cotonov, lower eulittoral zone, 26 III 1971, *D.M. John and G.W. Lawson* (L, no number). TANZANIA. Mafia-Wiene Island, 7 IX 1967, no collector (L 200346).

*S. divaricata.* AUSTRALIA. Torres Strait, on *Sargassum* sp., no date and collector, MA 9712 (Holotype, PC [Herb. Montagne], Isotype, TCD). Kanala, on *Turbinaria*, no collector and date (herb. Le Jolis, MEL 501112). Fremantle, W.F. Prud'homme van Reine, no date (L 937.117-15). Rockingham, no collector and date (L 937.117-18); Rockingham, no date, *Demel s.n.* (W 19651). Port Denison, no collector and date (W 19658). OCEANIA. New Caledonia, Kanala, no collector and date (L 937.117-19).

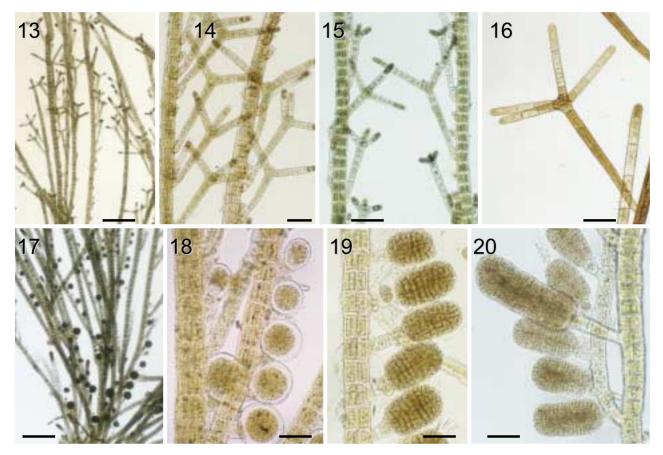
Thalli erect, 0.5-1.0 cm high, brown to olive brown, tufts to cushioned, epiphytic. Holdfasts monostromatic, discoid and stoloniferous. Erect filaments straight and cylindrical with indeterminate irregular lateral branches (Fig. 1). Segments 21-69  $\mu$ m long, 15-44  $\mu$ m broad, 0.8-1.9 in L/B ratio at main axes, divided radially by 2-3(-4) longitudinal walls. Peripheral cells rectangular, 3-10  $\mu$ m



Figs 1-9. Sphacelaria rigidula Kützing. Figs 1-6. Plant from Eochungdo, Korea. Fig. 1. Filaments bearing propagules. Fig. 2. Two-armed propagules. Fig. 3. Unilocular zoidangia. Fig. 4. Plurilocular zoidangia. Fig. 5. Filaments of cultured material. Fig. 6. Repeatedly branched two-armed propagules. Fig. 7. Two-armed propagules from Tahiti (L937. 117-17). Fig. 8. Two-armed propagules from Australian plants (AD51282). Fig. 9. Unilocular zoidangia (AD51282). Figs 10-12. Sphacelaria divaricata from Australia. Fig. 10. Filaments bearing propagules (W19651). Fig. 11. Propagules with dichotomously branched arms (L937. 117-19). Fig. 12. A propagule with dichotomously branched arms (L. 937. 117-15) (Scale bars: Figs 1, 5, 6, 10; 200 μm; Figs 2, 7, 8, 11, 12; 100 μm; Figs 3, 4, 9; 50 μm).

in diameter and scarcely divided by secondary transverse walls. Phaeophycean hairs 8-12  $\mu$ m in diameter, directly on apical cells or by oblique division of apical cells (Fig. 2). Propagules frequently in upper and middle filaments, consisting of two straight, cylindrical and symmetrical primary arms and a stalk in field-collected specimens (Fig 2). Stalks of propagules 186-391  $\mu$ m long, 17-33  $\mu$ m broad at waist. Primary arms

153-325  $\mu$ m long, 16-30  $\mu$ m broad, but these lengths varied considerably in populations (Table 1). In culture, two primary arms branched to form secondary arms, 260-460  $\mu$ m long (Fig. 6). Unilocular zoidangia produced in middle to lower parts of filaments, dark brown, spherical, 29-55  $\mu$ m in diameter, with a one-celled pedicel (Figs 3, 9). Plurilocular zoidangia occur in middle to lower parts of filaments, 45-65  $\mu$ m long, 25-40  $\mu$ m



Figs 13-20. Sphacelaria fusca from Korea. Fig. 13. Filaments bearing propagules. Figs 14, 15. 2-3 armed propagules irregularly arranged on filaments. Fig. 16. A four armed propagule. Fig. 17. Unilocular zoidangia in lower part of filaments. Fig. 18. Spherical unilocular zoidangia. Fig. 19. Macroplurilocular zoidangia. Fig. 20. Microplurilocular zoidangia (Scale bars: Figs 13, 17; 200 μm; Figs 14-16; 100 μm; Figs 18-20; 50 μm).

broad, with a one-celled stalk (Fig. 4). Loculi 3-7  $\mu$ m in diameter. No differentiation into micro- and macroplurilocular zoidangia.

The type and herbarium specimens: The type was small, about 2-4 mm high, and has a few filaments with indeterminate laterals. Only two propagules with two primary arms were found. Morphological characteristics of additional herbarium specimens of *S. rigidula* are provided in Table 1. These materials had slender filaments and propagules with always only two primary arms (Figs 7, 8).

The type and herbarium specimens of *S. divaricata*: The type specimen is up to 1-2 mm high, epiphytic on *Sargassum* sp., and has erect filaments with indeterminate, irregularly branched laterals. Segments of filaments are 24-28  $\mu$ m long, 20-24  $\mu$ m broad. Propagules are composed of two slender primary arms and a stalk with 20-30  $\mu$ m waist-width and 160-200  $\mu$ m long. The two primary arms are 2-3 times branched to form additional arms, and in those cases the primary arms grown up to 300-400  $\mu$ m in length. No data are available on unilocular and plurilocular zoidangia.

Additional herbarium specimens deposited in AD, MEL and L collected from Australia and tentatively recorded as *S. rigidula* or *S. furcigera* corresponded well to the type of *S. divaricata*. They have quite slender filaments and propagules with two arms, which are repeatedly dichotomously branched to form additional arms (Figs 10, 11, 12).

# Sphacelaria fusca (Hudson) S.F. Gray 1821, p. 333. Figs 13-22

**Type:** Sidmouth, England, June 1827, *Griffiths s.n.* (Neotype, TCD).

Basionym: Conferva fusca Hudson 1762.

**Synonyms:** *Sphacelaria subfusca* Setchell et Gardner 1924, p. 13, f. 25, Redondo, southern California, xi 1908 (Holotype specimen probably lost, is now fig. 25 of the publication).

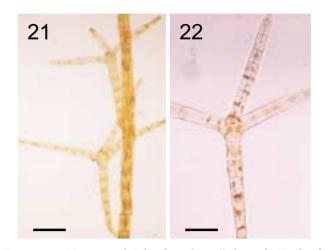
Distribution: warm temperate to tropical areas

(Prud'homme van Reine 1982)

Specimens examined: ENGLAND. Sidmouth, VI 1827, Griffiths s.n, (Neotype, TCD). Exmouth, 1851, Gulson (TCD). FRANCE. Saint Malo, Port Royal, 9 VI 1872, Sauvageau (Herb. Thuret, PC). KOREA. Chaesukgang, 6 V 1993, Y.-S. Keum (SNU930506). Jeoncheon, 6 VIII 1993, Y.-S. Keum (SNU930806). As S. rigidula. JAPAN. Rumoi, Hokkaido Pref., 2 VIII 1989, T. Kitayama, (Kita-61, TNS). Iwata, Aomori Pref., 7 IV 1991, T. Kitayama (Kita-162, TNS). Uratomi, Hyogo Pref., 7 IV 1990, T. Kitayama (Kita-104, TNS). CHINA. Qingdao, 30 VI 1994, Y.S. Oh (SNU940630). Guangdong, no date and collector (AST 55-1679). Guangdong, no date and collector (AST 55-1860).

Thalli erect, dark brown to olive brown, tufts, complanate or cushioned, epiphytic on Sargassum thunbergii, Corallina spp., Laurencia spp., and reaching up to 3 cm high. Holdfasts monostromatic to polystromatic, discoid and stoloniferous. Erect filaments cylindrical, with indeterminate irregularly branched laterals (Fig. 13). Segments 26-46  $\mu$ m long, 32-54  $\mu$ m broad, 0.65-1.10 in L/B at main axes with 2-6 longitudinal walls. Peripheral cells rectangular 5-13  $\mu$ m in diameter and secondary transverse walls often formed. Phaeophycean hairs directly on apical cells or formed by oblique division of apical cells, 8-14  $\mu$ m in diameter. Propagules irregularly but densely scattered in upper to middle filaments (Fig. 13) consisting of straight, cylindrical 2-3(-4) primary arms, a straight stalk and a lenticular apical cell (Figs 14, 15, 16). Stalks of propagules 188-330  $\mu$ m long, 23-35  $\mu$ m broad at waist. In culture, primary arms often branched to form the secondary arms. Unilocular zoidangia produced in middle to lower parts of filaments (Fig. 17), dark brown, spherical, 38-63  $\mu$ m in diameter with a one-celled pedicel (Fig. 18). Plurilocular zoidangia formed on middle to lower parts of filaments. Macroand microplurilocular zoidangia on different plants. Macroplurilocular zoidangia, with large locules (5-10  $\mu$ m in diameter), dark brown, ellipsoid, 45-102 µm long, 53-79  $\mu$ m broad, 0.85-1.48 in L/B (Fig. 19). Microplurilocular zoidangia, with small locules (2-6  $\mu$ m in diameter), yellowish brown, ellipsoid, 75-131 µm long, 41-70 µm broad, 1.39-2.57 in L/B (Fig. 20).

**The type specimen:** The plant is about 3 cm high and consists of irregularly and scarcely branched indeterminate filaments and a few 3-armed propagules. Segments of filaments are 65-96  $\mu$ m long and 44-80  $\mu$ m broad. Stalks of propagules are 356-542  $\mu$ m long, 36-50



Figs 21, 22. Neotype of *Sphacelaria fusca* (Sidmouth, England, TCD). Propagules with 3 arms (Scale bars: Fig. 21, 100  $\mu$ m; Fig. 22, 50  $\mu$ m).

 $\mu m$  broad at waist; primary arms are 320-540  $\mu m$  long (Figs 21, 22)

# Sphacelaria didichotoma Saunders 1898, p. 158 Figs 23-30

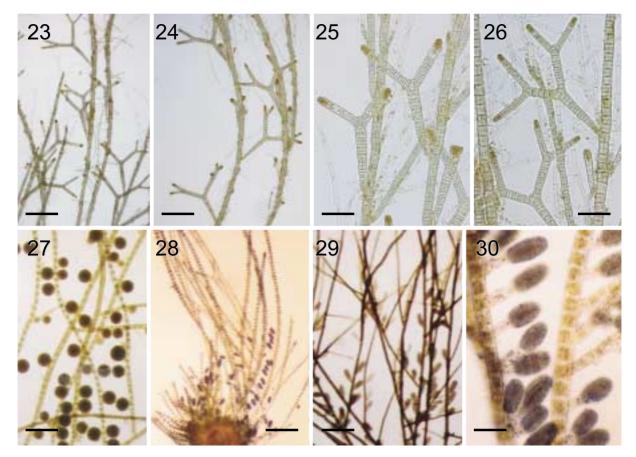
**Type:** Carmel Bay, California, Specimen probably lost, Holotype Saunders 1898, pl. XXVII, fig. 1)

**Synonyms:** Sphacelaria divaricata f. japonica Takamatsu 1943, p. 165-166, f. 6, pl. 10. Sphacelaria hizikiae Ohta et Noda 1984, p. 33-38, f. 1. Sphacelaria prostrata Takamatsu 1943, p. 160, pl. 9, f. 1, 2.

**Distribution:** Pacific Ocean; Korea (Lee and Kang 1986), Japan (Kitayama 1994) and California (Setchell and Gardner 1925, Abbott and Hollenberg 1976).

Specimens examined: KOREA. Eodal, 29 V 1993, Y.-S. Keum (SNU930529). Anin, 21 X 1993, Y.-S. Keum (SNU931021). Seongsan, Cheju Island, 21 III 1994, Y.-S. Keum (SNU940321). As S. divaricata. JAPAN. Tappi, Aomori Pref., 24 V 1989, T. Kitayama 42 (TNS). Fukaura, Aomori Pref., 25 V 1989, T. Kitayama 37 (TNS). Tsuyazaki, Fukuoka Pref., 2 IV 1989, S. Kawaguchi (TNS, Kita-37). Shimoda, Shizuoka Pref. on Hizikia fusiformis in the middle to lower intertidal zone, 20 V 1990, J. Tanaka (L 993.356-480). Imagoura, Hyogo Pref., 9 IV 2000, Y.-S. Keum (SNU200409). Maruyama, Awajishima, Hyogo Pref., 21 IV 2000, Y.-S. Keum (SNU200421). CHINA. Guangdong, no collector and date (AST 56-2970).

Thalli erect up to 1 cm high, dark brown to olive brown, tufts or cushioned, epiphytic. Holdfasts monostromatic to polystromatic, discoid and stoloniferous. Erect filaments cylindrical, with



Figs 23-30. Sphacelaria didichotoma from Japan. Figs 23, 24. Mature propagules with dichotomously branched arms (Imagoura, Hyogo Pref.). Fig. 25. Young propagules with 2 arms (Maruyama, Hyogo Pref.). Fig. 26. Propagules with dichotomously branched arms (Maruyama, Hyogo Pref.). Fig. 27. Unilocular zoidangia. Fig. 28. Plurilocular zoidangia in lower part of erect filaments. Fig. 29. Microplurilocular zoidangia. Fig. 30. Macroplurilocular zoidangia (Scale bars: Figs 23, 24, 28, 200 μm; Figs 25-27, 29, 30, 100 μm).

indeterminate irregularly branched laterals. Segments, 28-43  $\mu$ m long, 33-46  $\mu$ m broad, 0.7-1.2 in L/B at main axes, divided by 2-6 longitudinal walls. Peripheral cells rectangular 5-13  $\mu$ m in diameter, with occasion secondary transverse walls. Phaeophycean hairs 10-16  $\mu$ min diameter, directly on apical cells or formed by oblique division of apical cells. Propagules frequent in upper to middle filaments (Figs 23, 24), consisting of two dichotomously branched arms, a stalk and a lenticular apical cell (Figs 25, 26). Stalks of propagules 146-307  $\mu$ m long, 30-55  $\mu$ m broad at waist, tapering slightly and gradually to the base. Two primary arms curved slightly backward and branched to form secondary arms, 103-269  $\mu$ m long. Unilocular zoidangia produced in middle to lower parts of filaments (Fig. 27), dark brown, spherical, 30-60  $\mu$ m in diameter, with a one-celled pedicel. Plurilocular zoidangia formed on middle to lower parts of filaments (Fig. 28). Macro- and microplurilocular zoidangia on different plants. Macroplurilocular zoidangia, with large locules (4-10  $\mu$ m in diameter), dark brown, ellipsoid, 40-100  $\mu$ m long, 50-75  $\mu$ m broad, 0.7-1.3 in L/B (Fig. 30). Microplurilocular zoidangia, with small locules (2-5  $\mu$ m in diameter), yellowish brown, ellipsoid, 45-70  $\mu$ m long, 30-35  $\mu$ m broad, 1.2-2.3 in L/B (Fig. 29).

### Molecular study

The sequences of RUBISCO spacer ranged for 7 species of *Sphacelaria* from 234 to 276 base pairs. The lengths of the RUBISCO spacer were 261-264 bp in *Sphacelaria rigidula*, 252 bp in *S. fusca* and 234-238 bp in *S. didichotoma*. The sequence of RUBISCO spacer appeared to be conserved at the specific level in *S. fusca* and *S. rigidula*, except for one specimen of *S. rigidula*. The sequence of *S. rigidula* for three Korean specimens and one Japanese specimen were identical, but considerably differed from the Cape Verde Islands specimen (5.4%). The three Japanese specimens in *S. didichotoma* were showed a relatively high divergence of 0.4-2.1%. The interspecific variations of sequences among *S. rigidula*, *S.*  fusca and S. didichotoma ranged from 10% to 20.3%. Sequence divergence for seven Sphacelaria species included in this study ranged from 6.3 to 31.2%. The sequence divergences between S. cirrosa (section Propagulifera) and the other studied members in the sections Furcigerae and Tribuloides of Sphacelaria were considerably high (in the range of 21.1-31.2%). The sequence divergences between at the one side S. californica or S. tribuloides in the section Tribuloides and at the other side members of the section Furcigerae were relatively similar to those among members in the section Furcigerae.

A total of 289 nucleotide characters were in the present study available for phylogenetic analyses. Among them, 28 variable characters were phylogenetically uninformative and 86 characters were informative. Parsimony analysis resulted in a single most parsimonious tree (tree length = 176 steps, CI=0.81, RI=0.91) (Fig. 31). NJ and ML analyses generated trees of which the topology was identical with the most parsimonious tree. *S. cirrosa* was used as an outgroup based on a phylogenetic analysis of *rbcL* sequences (Draisma 2002).

Our phylogenetic tree showed that the three species, *S. rigidula, S. fusca* and *S. didichotoma* were clearly recognized. However, within *S. fusca* and the Korean and Japanese specimens of *S. rigidula*, relationships among strains could not be determined further due to identical sequences. *Sphacelaria fusca* and *S. didichotoma* formed a clade together with *S. californica*. The latter was basal within this clade, which was strongly supported by high bootstrap values (99%). *Sphacelaria rigidula* and *S. yamadae* constituted other clade (100%). *S. tribuloides* was placed as a sister group to these two clades, therefore *S. californica* and *S. tribuloides* were separated each other, implying that the sections *Furcigerae* and *Tribuloides* of the subgenus *Propagulifera* do not constitute monophyletic groups.

### DISCUSSION

Sphacelaria furcigera was first described by Kützing (1855) from Karak Island, Persian Gulf. on the basis of plant with the 2-armed propagules. Hudson (1762) described *Conferva fusca* from England, and later S.F. Gray (1821) transferred it to the genus Sphacelaria and mentioned on 3-armed propagules. However, a type specimen was not designated. Harvey (1841) recorded *S. fusca* from Sidmouth and St. Michaels Mount, England.

Prud'homme van Reine (1982) confirmed that Hudson's specimens were lost and designated Harvey's specimen collected from Sidmouth as a neotype.

Many phycologists have considered that there are no morphological differences between S. fusca and S. furcigera (Holmes and Batters 1891; De Haas-Niekerk 1965; Van den Hoek and Flinterman 1968; Russell and Fletcher 1975). De Haas-Niekerk (1965) mentioned that there is no reason to separate two species on the basis of the similarity in the production of two-armed propagules by both species and synonymized S. furcigera with S. fusca. Van den Hoek and Flinterman (1968) accepted De Haas-Niekerk's opinion, but preferred use the name S. furcigera. However, some authors noted that S. furcigera shows always only two arms in propagules, while S. fusca has two to four arms. The proportion of the number of arms in propagules in S. fusca, on the other hand, is quite variable (Sauvageau 1902; Zimmermann 1925; Goodband 1971).

Sphacelaria rigidula was established by Kützing (1843) from the Red Sea, but at that time the presence of propagules was not mentioned. Prud'homme van Reine (1982) found 2-armed propagules from the type specimen of S. rigidula and synonymized S. furcigera with S. rigidula. He regarded S. rigidula as abundant and widely dispersed plants with 2-4 arms as well as with only 2 arms in propagules. S. fusca, however, was considered by him as a rare species, consistent with the neotype showing thicker filaments and the absence of rhizoids and stolons, although it has 2-3 armed propagules. When Kitayama (1994) monographed Japanese Sphacelaria species, he recorded that the type specimen of S. rigidula has just a few 2-armed propagules (fig. 25A), but all of 53 propagules in the type of S. furcigera are only 2-armed (fig. 25B). Following Prud'homme van Reine's opinion, however, he tentatively considered the Japanese plants with 2-4 armed propagules as S. rigidula, but mentioned that the taxonomic entity of these Japanese plants needed further discussion.

In our opinion, however, *Sphacelaria rigidula* is clearly distinguished from *S. fusca* by having only 2-armed propagules. We found that the type of *S. furcigera* has 2-armed propagules as well as the type of *S. rigidula*. Thus the type materials of both taxa collected in the tropical Red Sea contained only 2-armed propagules. On the basis of the examination of herbarium specimens and our collections from Japan and Korea, we propose to use *S. rigidula* for the species occurring widely in warm

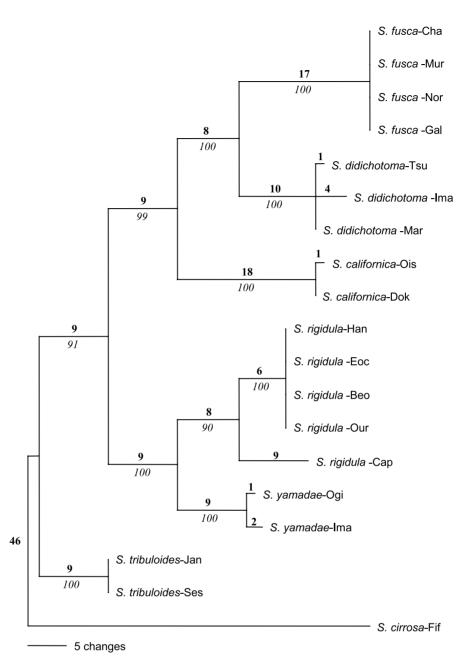


Fig. 31. The single most parsimonious tree inferred from the RUBISCO spacer sequences. For codes see table 2. The consistency index was 0.81, and the retention index was 0.91. Numbers above branches represent nucleotide substitutions. Numbers below branches indicate bootstrap values

temperate and tropical areas. *S. fusca* is characterized by 2-4 armed propagules and is also widely distributed throughout the world. In addition, *S. rigidula* has usually slightly slender erect filaments (15-44  $\mu$ m vs. 32-54  $\mu$ m), have a smaller waist width (17-33  $\mu$ m vs. 23-35  $\mu$ m), and also a smaller primary arm width (16-30  $\mu$ m vs. 26-31  $\mu$ m) than *S. fusca*. Therefore, the Japanese materials examined by Kitayama (1994) should be assigned to *S. fusca*. As well as these morphological features, *S. rigidula* was clearly separated from *S. fusca* in the sequence divergence of 17.5-18.9 %. It formed a clade withi,

showing a close genetic distance to *S. yamadae* (6.3-8.2%). These two species are commonly collected on *Sargassum* spp. from subtidal areas, whereas the other species occur both in intertidal and subtidal areas. The Korean and Japanese specimens of *S. rigidula* from various localities showed no sequence variation. However, the specimen from Cape Verde Islands showed highly different sequences (5.4%), may due to geographical distance. In *S. fusca*, the RUBISCO spacer sequence data did not resolve at population level, although various Atlantic and Pacific populations are included.

The neotype of S. fusca from Sidmouth is somewhat atypical, compared with other collections, in regards to the width of its erect filaments (almost twice as wide as those of most collections). Prud'homme van Reine (1982) also mentioned that the rare plants described as S. fusca differ from common plants described as S. rigidula by a larger diameter in erect filaments (35-80  $\mu$ m) and in the stalks of the propagules (29-47  $\mu$ m). However, he added that the differences in diameter of the erect filaments in the distal parts are inconspicuous and the absence of rhizoid is probably due to lack of sufficient materials. Additionally, we could not detect specimens in other herbarium collections that were in size agreement with the neotype. The herbarium collections of S. fusca show propagules with 2-3(4) arms and all had a similar (smaller) size in erect filaments. Thus, it is considered that the size difference of the neotype just represents a deviation from the typical form of S. fusca.

Womersley (1987) mentioned that the Australian plants of *S. fusca* show opposite and alternate branches with broad branching and prominent axes. However, these features are in accordance with the defining characters of *S. cirrosa* showing the laterals distichously and pinnately branched and the propagules with apical hairs and 3 fusiform or cylindrical arms curved backward, although occasionally propagules without apical hairs occurred. The examinations of Womersley's specimens from AD showed that some specimens have the laterals branched alternately and pinnately as well as propagules with constrictions at the base and apical hairs, thus they are considered as *S. cirrosa*.

Setchell and Gardner (1924) described S. subfusca from specimens with 2-3 armed propagules collected in California, and mentioned that it is distinguished from S. fusca in having propagules which are often bifurcate, without tapering of stalk or of arms. However, Prud'homme van Reine (1982) suggested that S. subfusca is closely related to S. fusca (as S. rigidula), and Kitayama (1994) has treated S. subfusca in the synonymy of S. rigidula. Although the type specimen of S. subfusca was probably lost, the original description and figures are consistent with S. fusca on the basis of propagule morphology. Additionally, the proportion of the number of arms in S. fusca is variable, and the stalk and arms may slightly taper or do not at all. We therefore concluded that S. subfusca should be reduced to synonymy with S. fusca.

Since the description of *S. divaricata* by Montagne (1849), that species has been recorded from the

northwestern Pacific, including Japan (Kitayama 1994), Korea (Kang 1966; Lee and Kang 1986) and China (Draisma et al. 1998). Sauvageau (1901) stated that it is characterized by repeatedly branched two-armed propagules, as exceptionally also occurring in S. rigidula (as S. furcigera). Prud'homme van Reine (1982), however, stated that the criteria to distinguish S. divaricata by Sauvageau (1901) are irrelevant, because the branching of arms of the propagules occurs frequently in S. fusca (as S. rigidula). When Kitayama (1994) recorded that Japanese Sphacelaria plants show dichotomously branched arms in the propagules, he investigated the type specimen of S. divaricata, and mentioned that the primary arms of the propagules in this type are 200 - 490  $\mu$ m in length, whereas the Japanese plants designated under that name are just up to 250  $\mu$ m. Nevertheless he identified the Japanese plants as S. divaricata and synonymized S. didichotoma with S. divaricata sharing the same arm morphology in their propagules.

Examination of the type of *S. divaricata* showed slender filaments (20-24  $\mu$ m) and long dimensions (300-400  $\mu$ m) in the primary arm length. In culture, we found that the arms of the propagules in S. rigidula commonly branched to form additional secondary arms, and that their primary arms reached up to 260-460  $\mu$ m in length. In general, in the species of the section *Furcigerae* the primary arms are able to branch again to form additional arms in propagules. In the Korean plants and other herbarium specimens of S. rigidula, however, secondary arms are rarely found in the field materials. Therefore, the type of S. divaricata is considered as an old plant producing secondary arms in its propagules before they detach. Additionally, the type and herbarium specimens of S. divaricata show slender filaments similar to those of S. rigidula. According to these characteristics and dimensions, it is reasonable that S. divaricata should be regarded as a synonym of S. rigidula.

The Japanese plants that have previously been recorded as *S. divaricata* form dichotomously branched propagules, showing shorter primary arms (103-269  $\mu$ m) than the type of *S. divaricata* and the cultured plants of *S. rigidula*. These dimensions are in accordance with the dimensions of Kitayama's plants. Abbott and Hollenberg (1976) recorded that in *S. didichotoma* from California the primary arms were 100-200  $\mu$ m in length. Moreover, *S. didichotoma* in the field usually produces secondary arms from two primary arms. Therefore, the northwestern Pacific plants are assigned to *S. didichotoma*. The type specimen of *S. didichotoma*, however, was probably lost,

but the northwestern Pacific plants are well agreement with the original features and descriptions. Thus fig.1 (pl. XXVII) of Saunders (1898) has to be considered as the holotype. Kitayama (1994) synonymized two Japanese species, S. prostrata Takamatsu (1943) and S. hizikiae Ohta and Noda (1984) with S. divaricata. Therefore, these two species should subsequently be considered as synonyms of S. didichotoma. Recently, Keum et al. (2001) proposed that S. divaricata f. japonica belongs to S. divaricata, because it has dichotomously divided two-armed and asymmetrical propagules with the secondary arms branched after the fourth secondary segment of the primary arms as seen in S. divaricata. Thus, it is also a synonym of S. didichotoma. In our phylogenetic tree, the three Japanese specimens of S. didichotoma formed a clade with S. fusca, but showed a higher sequence divergence (0.4-2.1%) than S. fusca and S. rigidula (except for the Cape Verde Islands population).

Phylogenetic relationship among the species in Sphacelaria subgenus Propagulifera based on RUBISCO spacer sequences is thus well resolved. However, S. californica and S. tribuloides do not belong to the same monophyletic group. Thus it appears that both the section Furcigerae and the section Tribuloides are not monophyletic. However, only a few strains of the two sections were investigated up to now. In previous molecular phylogenetic study, the two sections, Furcigerae and Tribuloides were also found not to be monophyletic in the tree inferred from the *rbcL* and RUBISCO spacer sequences data (Draisma 2002). Draisma (2002) mentioned that the propagule morphology, used as a traditional morphological character to separate the sections in subgenus Propagulifera, is not phylogenetically informative within the subgenus Propagulifera. Additionally, he recommended that the sections Furcigerae and Tribuloides either no longer be recognized or that they be merged into one section. Consequently, the subgenus Propagulifera still needs taxonomic revision. However, at present there are insufficient morphological and molecular data to support any useful taxonomic treatment.

### ACKNOWLEDGEMENTS

This study was supported by KOSEF grant (R03-2004-000-10022-0) to Y.-S. Keum.

## REFERENCES

- Abbott I.A. and Hollenberg G.J. 1967. *Marine algae of California*. Stanford Univ. Press. 827 pp.
- Draisma S.G.A. 2002. Calling a Class to Order. Phylogenetic Studies in the Phaeophyceae. Thesis, Universiteit Leiden, The Netherlands. 176 pp.
- Draisma S.G.A., Keum Y.-S., Prud'homme van Reine W.F. and Lokhorst G.M. 1998. The species of *Sphacelaria* (Sphacelariales, Phaeophyceae) in China with a description of a new species. *Bot. Mar.* **41**: 181-190.
- Felsenstein J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* **39**: 783-791.
- Goodband S.J. 1971. The taxonomy of *Sphacelaria cirrosa* (Roth.) Ag., *Sphacelaria fusca* (Huds.) Ag., and *Sphacelaria furcigera* (Kütz.) Sauv.: A simple statistical approach. *Ann.* Bot. **35**: 957-980.
- Gray S.F. 1821. A natural arrangement of British plants. Vol. 1. London.
- Haas-Niekerk T. de 1965. The genus *Sphacelaria* Lyngbye (Phaeophyceae) in the Netherlands. *Blumea* 13: 145-161.
- Hansen G. I. 1997. A revised checklist and preliminary assessment of the macrobenthic marine algae and seagrasses of Oregon. In: Kaye T.N., Liston A., Love R.M., Luoma D.L., Meinke R.J. & Wilson M.V. (eds), *Conservation* and Management of Native Flora and Fungi. Native Plant Society of Oregon, Corvallis, Oregon. pp. 175-200.
- Harvey W.H. 1841. A manual of the British Algae. London.
- Hasegawa M., Kishino H. and Yano T. 1985. Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. *J. Mol. Evol.* 21: 160-174.
- Hoek C. van den and Flinterman A. 1968. The life history of *Sphacelaria furcigera* Kütz.(Phaeophyceae). *Blumea* 16: 193-242.
- Holmes E.M. and Batters E.A.L. 1891. A revised list of the British marine algae. *Ann. Bot.* **5**: 63-107.
- Hudson G. 1762. Flora Anglica. London.
- Irvine D.E.G. 1956. Notes on British species of the genus Sphacelaria Lyngb. Trans. Proc. Bot. Soc. Edinb. 37: 24-45.
- Kang J.W. 1966. On the geographical distribution of marine algae in Korea. *Bull. Busan Fish. Coll.* 7: 1-125, pls. 1-12.
- Keum Y.-S., Oak J.H., Boo S.M., Prud'homme van Reine W.F. and Lee I.K. 1999. Sphacelaria californica and S. novaehollandiae, two related species of the genus Sphacelaria (Sphacelariales, Phaeophyceae). Algae 14: 23-35.
- Keum Y.-S., Oak J.H., Prud'homme van Reine W.F. and Lee I.K.
  2001. Two species of *Sphacelaria* (Sphacelariales, Phaeophyceae), *S. solitaria* (Pringsheim) Kylin and *S. recurva* sp. nov. from Korea. *Bot. Mar.* 44: 267-275.
- Kimura M. 1980. A simple method for estimating evolutionary rates of base substitution through comparative studies of nucleotide sequences. J. Mol. Evol. 16: 111-120.
- Kitayama T. 1994. A taxonomic study of the Japanese Sphacelaria (Sphacelariales, Phaeophyceae). Bull. Nat. Sc. Mus. Ser. B (Bot.) 20: 37-141.

Kützing F. T. 1843. Phycologia Generalis. Leipzig. 458 pp.

- Kützing F. T. 1855. *Tabulae Phycologicae, 5. Nordhausen* 30 pp. 100 pls.
- Lee I.K. and Kang J.W. 1986. A check list of marine algae in Korea. *Korean J. Phycol.* 1: 311-325.
- Montagne C. 1849. Sixieme centurie de plantes cellulaires nouvelles, tant indigenes qu'exofiques. *Ann. Sci. Nat. Bot. ser.* 3, **11**: 33-66.
- Ohta T. and Noda M. 1984. A new species of genus *Spacelaria* Lynbye (Phaeophyta) from the coast Tsugaru straits, northeastern Honshu, Japan. **4**: 33-38.
- Phillips J. A. 1997. Algae. In: Henderson R.J.F. (ed.), *Queensland Plants: Names and Distribution*. Queensland Herbarium, Department of Environment, Indooroopilly, Queensland. pp. 223-240.
- Prud'homme van Reine W.F. 1982. A taxonomic revision of the European Sphacelariaceae (Sphacelariales, Phaeophyceae). *Leiden Bot. Ser.* **6:** 1-193.
- Prud'homme van Reine W.F. 1993. Sphacelariales (Phaeophyceae) of the world, a new synthesis. *Korean J. Phycol.* **8**: 145-160.
- Russell G. and Fletcher R.L. 1975. A numerical taxonomic study of the British Phaeophyta. J. Mar. Biol. Ass. U.K. 55: 763-768.
- Saitou N. and Nei M. 1987. The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* **4**: 406-425.
- Saunders D.A. 1898. Phycological Memoirs. Proc. Califor. Acad. Sci. Ser. 3, Bot. 1: 147-168.
- Sauvageau C. 1900-1914. Remarques sur les Sphacelariacees.

Published 1900: J. Bot., Paris 14; 1901: *J. Bot., Paris 15*; 1902: *J. Bot, Paris 16*; 1903: *J. Bot, Paris 17*; 1904: *J. Bot, Paris 18*; 1914 and also published separately.

- Setchell W.A. and Gardner N.L. 1924. Phycological contributions, VII. Univ. Calif. Publs. Bot. 13: 1-13.
- Setchell W.A. and Gardner N.L. 1925. The marine algae of the Pacific coast of North America, part II. Melanophyceae. *Univ. Calif. Publs. Bot.* 8: 385-898.
- Stache-Crain B., Muller D.G. and Goff L.J. 1997. Molecular systematics of *Ectocarpus* and *Kuckuckia* (Ectocarpales, Phaeophyceae) inferred from phylogenetic analysis of nuclear- and plastid-encoded DNA sequences. *J. Phycol.* 33: 152-168.
- Swofford D.L. 2000. *PAUP: Phylogenetic Analysis using Parsimony*, version 4.0b3a. Illinois Natural History Survey, Champaign, Illinois.
- Takamatsu M. 1943. The species of *Sphacelaria* from Japan I. *J. Sigenkagaku Kenkyusho* 1: 153-187, pls. 8-13.
- Thompson J.D., Plewniak F. and Poch O. 1999. A comprehensive comparison of multiple sequence alignment programs. *Nucleic Acids Res.* 27: 2682-2690.
- Womersley H.B.S. 1987. The Marine Benthic Flora of Southern Australia. Part II. South Australian Government Printing Division, Adelaide, 484 pp.
- Zimmermann W. 1925. Helgolander Meeresalgen I-VI. Wiss. Meeresunters. Abt. Helgoland 16: 1-25.

Received 16 March 2005 Accepted 24 March 2005