

The Effect of Boron Supplementation on Bone Strength in Ovariectomized Rats Fed with Diets Containing Different Calcium Levels

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Abstract The effect of calcium and boron supplementation on bone strength was determined in growing and ovariectomized (OVX) Sparague-Dawley rats. Rats were divided into 9 groups and fed diet with different intake levels of calcium and boron for 4 weeks. About fifty percentages of rats in each group were OVX and the others were sham-operated. The rats were fed same diets after operation for 8 weeks. The feed intake, body weight gain, and FER were significantly higher in OVX rats than those in sham-operated ones. Serum osteocalcin, bone formation biomarker, was significantly increased with increment in calcium and boron intakes. Serum estradiol was lower in OVX rats than in sham-operated ones. Bone mineral density of femur was significantly lower in OVX rats than in other group. The breaking forces of bones were not significantly different among the groups. The urinary excretion of deoxypyridinoline, osteolytic marker was significantly increased with increment in calcium intake and ovariectomy. The urinary calcium excretion was significantly increased with increment in calcium intake, but decreased with increment in boron intake. According to theses results, the boron supplementation resulted in higher serum osteocalcin and lower urinary calcium excretion. Therefore, it could be suggested that the boron supplementation may be complementary and useful to calcium nutrition for bone health.

Keywords: boron, calcium, bone strength, ovariectomized rat

Introduction

Incidence of osteoporosis is increasing with increment in elderly population in Korea. And thus, developments of strategies to prevent this disease and maintain bone health are on the rising trend. For instance, the research for isoflavone as phytoestrogen was raised as a goal for preventing osteoporosis (1). Boron is an essential ultratrace element in plants (2). There are circumstantial evidences to prove that boron may also affect bone mineral homeostasis in various animal models, including humans. Hunt and Nielsen (3) reported that boron deprivation depressed growth and elevated plasma alkaline phosphatase activity in chicks fed with inadequate levels of cholecalciferol. Working with boron as a micronutrient, Nielsen et al. (4) found that adding boron to boron-deficient diets had salutary effects on indices of bone metabolism and mineral homeostasis in humans. These and other data have contributed to the preliminary conclusion that boron is 'probably essential' for human nutrition (5).

Some of the reported observation could be due to changes in macromineral metabolism. Hegsted *et al.* (6) found that apparent absorption and balance of calcium, magnesium, and phosphorus were higher in boron-supplemented than in boron-deprived rats fed with cholecalciferol-deficient diet. Brown *et al.* (7) observed that supplemental boron increased the apparent absorption and retention of calcium in sheep. Nielsen *et al.* (8) found

that boron repletion in 12 postmenopausal women, who were previously on a low-boron diet for 119 days, had markedly reduced levels of urinary excretion of calcium. Although the preceding studies indicate that boron affects calcium metabolism, only limited reports are available on the interaction between boron and calcium.

Past studies in postmenopausal women, who were previously on a low-boron diet, showed that dietary boron repletion increased serum 17β -estradiol (E2) and testosterone levels (8). A similar increase in serum E2 levels was seen in healthy males after 4 weeks of dietary boron supplementation (9). Because boron increases the serum E2 level, it has been suggested that boron may reduce some of the adverse effects of E2 deficiency associated with menopause and/or cessation of ovarian function (4, 10).

The purpose of the present work was to confirm whether boron supplementation affects the bone status of the rats fed on diets with different calcium levels. Therefore, we designed a study to determine the effects of boron supplementation on bone strength and bone metabolism indicators in OVX rats and sham-operated control rats consuming diets with different calcium and boron intakes. The experiments were part of an overall plan to obtain further evidence that boron might be involved in bone metabolism, thus indicating that boron is an essential nutrient for bone health.

Materials and Methods

Animals and diets One hundred-and eighty 6-wk-old Sprague-Dawley female rats were purchased from Chungang Experimental Animal Inc. (Seoul, Korea). After

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7 days of acclimatization, the animals were systematically assigned to 9 weight-matched treatment groups (20 rats per group). The animals were housed in a room maintained at 24°C on 12 hr light/12 hr dark cycles and were fed diets. ad libitum containing different content of calcium (0.1, 0.5, 1.5%) and boron (0.5, 50, 100 ppm) for 4 weeks. The chow diet fed to the animals in this experiment was the AIN-93 diet (11) and its ingredients are as shown in Table 1. At 12 weeks of age, the half of rats in each group were bilateral OVX and the others were sham operated under anesthesia with 15 mg of ketamine per kg of body weight (Ketalar, Yuhan Co., Korea). Deionized water was supplemented as drinking water. Feed cups and drinking bottles were soaked overnight in conc. HNO₃ prior to each use to remove trace metal contamination.

Experimental protocol The rats were divided into 18 groups (10 rats per group) as: (1) low calcium (0.1%) + adequate boron (0.5 ppm); (2) low calcium + high boron (50 ppm); (3) low calcium + very high boron (100 ppm); (4) adequate calcium (0.5%) + adequate boron; (5) adequate calcium + high boron; (6) adequate calcium + very high boron; (7) high calcium (1.5%) + adequate boron; (8) high calcium + high boron; (9) high calcium + very high boron. 9 groups were OVX rats and 9 groups were sham-operated animals. Treatment diet with adequate nutrition as recommended by the American Institute of Nutrition (11) except calcium and boron was available ad libitum to the rats. The daily food intake in each group was measured and the rats were weighed weekly. The experiment was continued for 8 weeks. For 3 days before the termination of the experiment, the rats were placed in metabolic cages, and 24 hr urine was collected. At the termination of the experiment, animals were anesthetized

Table 1. Composition of diets

Ingredients ¹⁾	Low Ca	Adequate Ca	High Ca
		g/kg	
Corn starch	517.1504	507.2597	475.3273
Casein	200.0000	200.0000	200.0000
Sucrose	100.0000	100.0000	100.0000
Soybean oil	70.0000	70.0000	70.0000
Cellulose	50.0000	50.0000	50.0000
Mineral mixture (Ca/P/B free) ¹⁾	35.0000	35.0000	35.0000
Vitamin mixture ²⁾	10.0000	10.0000	10.0000
L-Cystine	3.0000	3.0000	3.0000
Choline bitartrate	2.5000	2.5000	2.5000
t-butylhydroquinone	0.0140	0.0140	0.0140
Calcium phosphate	3.4000	6.8770	33.3040
Calcium carbonate	0.0000	7.4200	12.9254
Potassium phosphate	2.7156	0.0000	0.0000
Potassium citrate	6.2200	7.9293	7.9293
Boron, Adequate	0.0005	0.0005	0.0005
High	0.05	0.05	0.05
Very high	0.1	0.1	0.1

¹⁾Low Ca: 0.1% Ca, 0.3% P; Adequate Ca: 0.5% Ca, 0.3% P; High Ca: 1.5% Ca, 0.9% P.
²⁾AIN-93 mineral mixture (Ca/P/B free).

³⁾AIN-93 vitamin mixture.

with ether and blood was collected. The scapulae, tibiae, and femurs of each animal were dissected and defleshed by sterile saline. The right bones were immediately soaked in 70% ethanol in a glass vial, kept in the dark at room temperature, and was used for histomorphometric measurements. The left bones were stored in a freezer at -70°C until examination. The animals were treated in accordance with the guidelines of the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

Analytical procedures Serum concentrations of total protein and albumin were measured spectrophotometrically by using commercial kits in a dry-chemistry analyzer (Fuji Dri-Chem 3000, Fujifilm, Japan). Serum alkanine phosphatase (ALP), osteocalcin, estradiol, parathyroid hormoneintact (PTH-intact), and sex hormone binding globulin (SHBG) were measured with commercial radioimmunoassay at gamma counter (Cobra II, Packard, USA). For histological observation of femur, the specimens were unfrozen before biomechanical tests and kept moist during all handling and test procedures. Microphotographs of a cross-section of the femoral neck were determined by scanning electron microscope (PSEM-75 MICGOPSM, RJ Lee Instruments Ltd., USA). Bone mineral density (mg/cm³) was calculated by dividing the bone wet weight from bone volume. Bone volume was measured by the principle of Archimedes (12). The 'three-point bending test' at fracture was used to determine the breaking force of bone. The force applied to the midpoint of the bone until breaking, was determined with a universal testing machine (CR-100D, Sun Scientific Co., Japan). Urinary deoxypyridinoline (DPD) excretions were measured with commercial radioimmunoassay in an immulite analyzer (DPC, USA). Food and urine were hydrolyzed with nitric acid by microwave digestion system (Ethos touch control, Milestone Inc., Italy). The calcium and boron concentrations of food and urine were measured by inductively coupled plasma spectrometer (Atomscan advantage axial sequential plasma spectrometer, Thermo Jarrell Ash Co., USA).

Statistical analysis Results are shown as means ± standard deviation. Statistical significance was determined with three-way analysis of variance (ANOVA) using the SAS software. When the ANOVA indicated significant difference among the means, the differences were evaluated using Duncan's multiple range test. The difference was significant when p<0.05.

Results and Discussion

Feed consumption, weight gain, and feed efficiency The initial body weight at baseline among the groups was not significantly different. However, OVX rats fed with high-calcium diet consumed significantly more feed and had greater weight gain when compared to sham-operated ones fed with low-calcium diet (Table 2). Feed efficiency of OVX rats was significantly higher than that of shamoperated animals. Dietary boron supplementation had no effects on body weight gain and feed efficiency. This finding is consistent with previous reports (13, 14), that OVX or calcium supplemented animals frequently gained more body weight as a result of overeating.

Table 2. Feed intake, body weight gain, and feed efficiency of the experimental rats

	Groups		Feed intake	Body weight gain	Feed efficiency
	Calcium	Boron	g/day	g/wk	g/g
Sham operation	0.1%	0.5 ppm	12.19 ± 0.18^{j1}	3.80 ± 1.46^{d}	0.04 ± 0.02^{b}
	0.1%	50 ppm	12.34 ± 0.17^{j}	5.06±2.41 ^d	0.06 ± 0.02^{b}
	0.1%	100 ppm	11.22 ± 1.05^{k}	4.09 ± 2.00^{d}	0.05 ± 0.02^{b}
	0.5%	0.5 ppm	12.66 ± 0.08^{ij}	4.01 ± 1.35^{d}	0.05 ± 0.02^{b}
	0.5%	50 ppm	12.91±0.28 ⁱ	4.73±0.85d	0.05 ± 0.01^{b}
	0.5%	100 ppm	13.17±0.63 ^{hi}	5.19 ± 1.45^{d}	0.06 ± 0.02^{b}
	1.5%	0.5 ppm	13.64±0.23gh	4.40 ± 1.01^{d}	0.05 ± 0.01^{b}
	1.5%	50 ppm	13.93±0.49 ^{fg}	4.34±1.11 ^d	0.04 ± 0.01^{b}
	1.5%	100 ppm	14.11 ± 0.15^{fg}	4.75±1.44 ^d	0.05 ± 0.01^{b}
Ovariectomy	0.1%	0.5 ppm	14.35 ± 0.65^{ef}	10.38±4.51°	0.10 ± 0.05^{a}
	0.1%	50 ppm	14.75 ± 0.79^{de}	12.28±3.63abc	0.12 ± 0.04^{a}
	0.1%	100 ppm	14.67 ± 0.43^{de}	11.45±4.46 ^{bc}	0.11±0.04 ^a
	0.5%	0.5 ppm	15.07 ± 0.36^{cd}	12.95 ± 2.56 abc	0.12 ± 0.03^{a}
	0.5%	50 ppm	14.73±0.63 ^{de}	12.69 ± 4.20^{abc}	0.12 ± 0.04^{a}
	0.5%	100 ppm	15.35±0.86°	12.39±2.44 ^{abc}	0.12 ± 0.02^{a}
	1.5%	0.5 ppm	15.94 ± 0.32^{b}	13.34±2.69ab	0.12 ± 0.02^{a}
	1.5%	50 ppm	16.51±0.50 ^a	14.61±2.37a	0.13±0.01 ^a
	1.5%	100 ppm	16.47±0.71 ^a	13.97±2.17 ^{ab}	0.12 ± 0.02^{a}
	Significance		p<0.001	p<0.001	p<0.001
Calcium			p<0.001	p<0.05	N.S.
Boron			p<0.01	N.S.	N.S.
Operation			p<0.001	p<0.001	p<0.001
Calcium×Boron			p<0.001	N.S.	N.S.
Calcium×Operation	a		p<0.001	p<0.05	N.S.
Boron×Operation			N.S. ²⁾	N.S.	N.S.
Calcium×Boron×C	peration		N.S.	N.S.	N.S.

¹⁾Values with different superscripts within a column are significantly different.

²⁾Not significant.

Serum bone metabolism indicators There were no significant difference in serum ALP and PTH levels among any of animals in the test groups (Table 3). However, serum osteocalcin, osteogenic marker, was significantly higher in high-calcium, boron supplementation, and OVX groups than those in other groups. Serum E_2 level of OVX rats was significantly lower than that of sham-operated ones, confirming the success of the surgery. One hundred ppm of boron supplementation significantly increased serum SHBG in low-calcium fed rats.

The mechanism whereby boron influences bone status has not been determined. However, some studies (14, 15) have shown that dietary boron supplementation increases serum E₂ and testosterone levels. Accordingly, it is possible that boron may modify bone status by increasing the synthesis mechanism of sex hormones. Unlike past studies, we observed that dietary boron supplementation does not affects serum level of E2, but increases osteocalcin in low-calcium/sham-operated group and SHBG in low-calcium/OVX group to improve bone quality in OVX rats. Osteocalcin is the most abundant noncollagenous protein in bone, and serum concentrations of osteocalcin can provide a measure of osteoblast activity or bone remodeling (16). Therefore, the reported data suggests that boron supplementation may prevent bone loss through osteogenic effect in OVX rats.

Bone measures Figure 1 shows histological evidence on the difference in femur loss in OVX rats and sham-

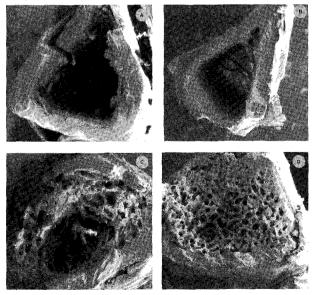


Fig. 1. Histological observation of the tibia and femur in ovariectomized rat by scanning electron microscope. A: Tibia ovariectomized. B: Tibia sham-operated. C: Femur ovariectomized. D: Femur sham-operated.

Table 3. Biochemical values of serum indicators of the bone metabolism in the experimental rats

	roups		Total protein	Albumin	ALP	Osteocalcin	Estradiol	PTH	SHBG
	Calcium	Boron	g/dL	g/dL	U/L	ng/mL	pg/mL	pg/mL	nmol/dL
Sham operation	0.1%	0.5 ppm	5.66 ± 0.15^{ab1}	3.98±0.04 ^{ab}	33.86±12.92	0.12±0.03 ^{bcd}	35.70±11.46 ^a	8.73 ± 2.85	0.06 ± 0.01^{d}
•	0.1%	50 ppm	6.04 ± 0.48^{a}	4.20±0.33a	57.18±32.58	0.12 ± 0.02^{abcd}	33.00±5.99 ^a	6.06±1.31	0.06 ± 0.03^{cd}
	0.1%	100 ppm	4.38±0.39 ^{cde}	2.94±0.30°	29.76±7.68	0.20 ± 0.05^{a}	31.24±5.00 ^{ab}	8.91±1.92	0.11 ± 0.03^{ab}
	0.5%	0.5 ppm	5.52±0.97 ^{abc}	3.78 ± 0.73^{abc}	26.08±10.53	0.13±0.08 ^{abcd}	32.42±6.93 ^a	7.99±1.56	0.11 ± 0.05^{ab}
	0.5%	50 ppm	5.00±0.38 ^{abcde}	3.56±0.30 ^{abc}	34.16±8.03	0.13±0.04 ^{abcd}	27.08±4.52 ^{abc}	8.01±2.15	0.09 ± 0.01^{abcd}
	0.5%	100 ppm	$4.94{\pm}0.84^{abcde}$	3.68±0.50 ^{abc}	31.32±21.02	0.13 ± 0.04^{abcd}	26.92±3.29 ^{abc}	5.55±1.27	0.09 ± 0.02^{abcd}
	1.5%	0.5 ppm	4.26 ± 0.96^{de}	3.20 ± 0.81^{bc}	40.22±17.98	0.16 ± 0.04^{abc}	33.40±8.29 ^a	8.42±3.15	0.07 ± 0.01^{cd}
	1.5%	50 ppm	4.20±0.73 ^e	3.08 ± 0.48^{c}	29.94±21.12	0.16 ± 0.11^{abc}	33.48±13.22 ^a	8.23±2.54	0.07 ± 0.01^{cd}
	1.5%	100 ppm	4.50±0.72 ^{bcde}	3.42±0.53abc	41.22±23.37	0.16 ± 0.03^{abc}	36.82±10.94 ^a	5.64±1.75	0.06 ± 0.01^{cd}
Ovariectomy	0.1%	0.5 ppm	5.28±0.78 ^{abcde}	3.55±0.54abc	37.85±12.32	0.14 ± 0.05^{abcd}	22.52±3.93 ^{bcd}	9.20±3.53	0.07 ± 0.00^{cd}
·	0.1%	50 ppm	5.44 ± 0.70^{abcd}	3.50±0.51abc	34.18±19.01	0.10 ± 0.04^{cd}	18.18±2.45 ^{cd}	8.91±1.76	0.06 ± 0.01^{cd}
	0.1%	100 ppm	4.65±1.02 ^{bcde}	2.95±0.85°	23.62±9.35	0.09 ± 0.02^{cd}	21.64±10.13 ^{cd}	8.61±2.57	0.12 ± 0.02^{a}
	0.5%	0.5 ppm	5.32±0.91 abcde	3.54±0.67 ^{abc}	35.08±30.79	0.08 ± 0.06^{d}	21.36±4.23 ^{cd}	8.57±1.38	0.07 ± 0.03^{cd}
	0.5%	50 ppm	4.55 ± 0.60^{bcde}	2.92±0.40°	30.92±14.04	0.10 ± 0.04^{cd}	18.86±2.38 ^{cd}	7.40 ± 2.24	0.10 ± 0.03^{abc}
	0.5%	100 ppm	5.50±1.22abc	3.73±0.78abc	36.32±25.32	$0.15 {\pm} 0.05^{abcd}$	18.72±2.58 ^{cd}	8.89±3.35	0.09±0.03 ^{abcd}
	1.5%	0.5 ppm	4.98±0.86 ^{abcde}	3.52±0.59abc	41.82±20.60	$0.15{\pm}0.06^{abcd}$	18.82±5.10 ^{cd}	5.64±1.39	0.08 ± 0.03^{bcd}
	1.5%	50 ppm	5.12±1.13 ^{abcde}		40.10±19.52	0.11 ± 0.06^{cd}	19.70±4.36 ^{cd}	6.07±3.94	0.07 ± 0.03^{cd}
	1.5%	100 ppm	4.92±1.04 ^{abcde}	3.47±0.71 ^{abc}	31.67±16.58	0.20 ± 0.05^{ab}	15.70 ± 2.17^{d}	8.25±2.59	0.07 ± 0.01^{cd}
Sign	nificance		p<0.05	p<0.05	N.S. ²⁾	p<0.05	p<0.001	N.S.	p<0.001
Calcium			p<0.05	N.S.	N.S.	p<0.05	N.S.	N.S.	p<0.01
Boron			N.S.	N.S.	N.S.	p<0.05	N.S.	N.S.	N.S.
Operation			N.S.	N.S.	N.S.	N.S.	p<0.001	N.S.	N.S.
Calcium×Bo	ron		p<0.05	p<0.01	N.S.	N.S.	N.S.	N.S.	p<0.001
Calcium×Op	eration		N.S.	N.S.	N.S.	N.S.	N.S.	N.S.	N.S.
Boron×Opera	ation		N.S.	N.S.	N.S.	N.S.	N.S.	N.S.	N.S.
Calcium×Bo	ron×Oper	ation	N.S.	N.S.	N.S.	p<0.05	N.S.	N.S.	N.S.

¹⁾ Values with different superscripts within a column are significantly different.

2) Not significant.

operated animals. It can be seen that femur loss in OVX rats was greater than sham-operated animals. These findings confirmed the success of the OVX. The OVX rat has been considered as a useful animal model for studying the effects of different osteoporosis treatment diets on the skeleton (17, 18). The data on bone status with boron supplementation are provided in Table 4 and 5. There was no significant difference in bone mineral density of scapular, tibia, and femur among boron treatment groups. Breaking forces of these bones also were not significantly different with boron supplementation in OVX and shamoperated rats fed diets with various calcium levels. However, breaking force of scapular increased with increment in calcium intake and OVX. This result may be explained by the possibility that OVX or calcium supplemented animals gained more body weight and scapular weight as a result of overeating.

Past studies in postmenopausal women (8, 10) and animals (19, 20) showed that dietary boron supplementation independently could promote positive bone mineral balance. These findings led to the speculation that dietary boron might also have a beneficial effect on bone quality and strength in E_2 -deficient subjects. However, the present study showed that 100 ppm of dietary boron supplementation did not improve bone mineral density and bone strength in OVX rats. Sheng *et al.* (15) emphasized that a positive response of boron on bone status was seen only in

those postmenopausal women who were deficient in boron. Chapin *et al.* (21) reported that supplementation of higher levels of boron will not bring about any further changes in bone strength because of saturation of a relatively small pool of boron binding sites. Therefore, to address these findings, additional studies are needed to investigate the effects of higher boron intake levels.

Urinary DPD and calcium excretion Table 6 shows that urinary DPD excretion, osteolytic marker was significantly diminished with increasing levels of dietary calcium and was higher in OVX rats when compared to shamoperated control rats. However, boron supplementation had no effect on urinary DPD excretion. Urinary calcium excretion was significantly increased with increasing levels of dietary calcium, but diminished with boron supplementation, particularly in high-calcium/OVX group.

The effect of boron supplementation on calcium metabolism most likely involved some biochemical or endocrine mechanism. It seems unlikely that urinary calcium excretion was reduced through chemical or complexing actions of boron. That is, it suggests that boron supplementation prevents urinary calcium loss and bone demineralization through endocrine mechanism of osteocalcin. However, this hypothesis must be confirmed by further experiments.

In conclusion, the boron supplementation resulted in higher serum osteocalcin in low-calcium group and lower

Table 4. Bone mineral density of the experimental rats

	Groups		Scapula	Tibia	Femur
	Calcium	Boron	mg/cm ³	mg/cm ³	mg/cm ³
Sham operation	0.1%	0.5 ppm	2.45±0.26	2.62±0.39	2.94±0.38 ^{a1)}
•	0.1%	50 ppm	2.74±0.59	2.72±0.26	2.94 ± 0.11^{a}
	0.1%	100 ppm	2.54±0.36	2.78±0.11	2.92 ± 0.16^{a}
	0.5%	0.5 ppm	2.65±0.28	2.68±0.11	2.95±0.21°
	0.5%	50 ppm	2.63±0.51	2.14±1.91	2.80 ± 0.20^{a}
	0.5%	100 ppm	2.51±0.44	2.45±0.19	2.90 ± 0.16^{a}
	1.5%	0.5 ppm	2.42 ± 0.20	2.62 ± 0.17	2.78 ± 0.22^{a}
	1.5%	50 ppm	2.40 ± 0.24	2.65±0.17	2.84 ± 0.15^{a}
	1.5%	100 ppm	2.24 ± 0.17	2.67±0.10	2.83 ± 0.15^{a}
Ovariectomy	0.1%	0.5 ppm	2.67±0.41	2.67±0.25	2.69 ± 0.26^{ab}
·	0.1%	50 ppm	2.55±0.34	2.80 ± 0.21	2.73 ± 0.26^{a}
	0.1%	100 ppm	2.40±0.31	2.74 ± 0.37	2.84 ± 0.11^{a}
	0.5%	0.5 ppm	2.58±0.13	2.89 ± 0.23	2.77±0.14 ^a
	0.5%	50 ppm	2.90 ± 1.02	2.63±0.18	2.36 ± 1.22^{b}
	0.5%	100 ppm	2.61±0.25	2.79±0.17	2.71 ± 0.18^{ab}
	1.5%	0.5 ppm	2.48±0.13	1.63±3.09	2.69 ± 0.09^{ab}
	1.5%	50 ppm	2.41±0.19	2.69±0.15	2.64 ± 0.17^{a}
	1.5%	100 ppm	2.48 ± 0.14	2.54±0.38	2.71 ± 0.27^{ab}
	Significance		N.S. ²⁾	N.S.	N.S.
Calcium			N.S.	N.S.	N.S.
Boron			N.S.	N.S.	N.S.
Operation			N.S.	N.S.	p<0.001
Calcium×Boron			N.S.	N.S.	N.S.
Calcium×Operation			N.S.	N.S.	N.S.
Boron×Operation			N.S.	N.S.	N.S.
Calcium×Boron×Op	eration		N.S.	N.S.	N.S.

¹⁾Values with different superscripts within a column are significantly different. ²⁾Not significant.

Table 5. Breaking force of bone in the experimental rats

<u> </u>	Groups		Scapula	Tibia	Femur
	Calcium	Boron	kg	kg	kg
Sham operation	0.1%	0.5 ppm	2.94 ± 0.39^{c1}	10.90±2.84	14.64±3.04
	0.1%	50 ppm	3.49 ± 0.42^{abc}	10.19±3.61	12.71±2.13
	0.1%	100 ppm	3.03 ± 0.50^{bc}	12.47±1.93	13.07±1.56
	0.5%	0.5 ppm	3.62 ± 0.92^{abc}	12.62 ± 2.13	14.85±1.74
	0.5%	50 ppm	3.60 ± 0.54^{abc}	12.83±2.36	13.97±1.61
	0.5%	100 ppm	3.51 ± 0.50^{abc}	11.08±2.74	12.63±1.44
	1.5%	0.5 ppm	3.60 ± 0.68^{abc}	12.44±3.17	14.20±2.86
	1.5%	50 ppm	4.00 ± 0.66^{a}	11.11±3.67	14.64±2.00
	1.5%	100 ppm	3.44 ± 0.73^{abc}	10.62±3.28	13.43±1.01
Ovariectomy	0.1%	0.5 ppm	3.60 ± 0.33^{abc}	13.64±3.35	14.57±1.80
•	0.1%	50 ppm	3.48 ± 0.63^{abc}	13.43±2.64	13.76±4.38
	0.1%	100 ppm	3.36 ± 0.64^{abc}	12.09±2.44	14.20±2.33
	0.5%	0.5 ppm	3.95 ± 1.24^{a}	13.21±2.43	15.67±1.46
	0.5%	50 ppm	3.80 ± 0.63^{abc}	13.76±2.66	15.67±2.47
	0.5%	100 ppm	4.08 ± 1.06^{a}	14.21±1.56	14.40±2.81
	1.5%	0.5 ppm	3.87 ± 0.61^{ab}	12.51±2.66	13.74±1.93
	1.5%	50 ppm	3.99 ± 0.48^{a}	13.42 ± 2.70	14.76±2.29
	1.5%	100 ppm	3.92±0.96 ^a	12.68±2.67	13.19±3.96
S	Significance		N.S. ²⁾	N.S.	N.S.
Calcium			p<0.01	N.S.	N.S.
Boron			N.S.	N.S.	N.S.
Operation			p<0.05	N.S.	N.S.
Calcium×Boron			N.S.	N.S.	N.S.
Calcium×Operation			N.S.	N.S.	N.S.
Boron×Operation			N.S.	N.S.	N.S.
Calcium×Boron×Op	peration		N.S.	N.S.	N.S.

¹⁾Values with different superscripts within a column are significantly different. ²⁾Not significant.

Table 6. Urinary deoxypyridinoline and mineral excretions in the experimental rats

	Groups		Deoxypyridinoline	Ca	В
	Calcium	Boron	nmol/mmol Cr	mg/day	μg/day
Sham operation	0.1%	0.5 ppm	114.05±21.81 ^{ef1)}	0.60 ± 0.34^{ab}	0.13 ± 0.14^{b}
	0.1%	50 ppm	104.43±28.61 ^f	0.33 ± 0.18^{b}	0.34 ± 0.39^{ab}
	0.1%	100 ppm	136.73±50.57 ^{def}	0.42 ± 0.52^{b}	0.65±0.69a
	0.5%	0.5 ppm	84.53±15.33 ^f	0.60 ± 0.46^{ab}	0.39 ± 0.44^{ab}
	0.5%	50 ppm	86.93±11.33 ^f	0.44 ± 0.36^{b}	0.24 ± 0.17^{ab}
	0.5%	100 ppm	99.88 ± 6.32^{f}	0.49 ± 0.47^{b}	0.35 ± 0.36^{ab}
	1.5%	0.5 ppm	92.08±21.58 ^f	1.05 ± 0.86^{a}	0.20 ± 0.13^{ab}
	1.5%	50 ppm	93.18 ± 20.49^{f}	0.40 ± 0.17^{b}	0.41 ± 0.52^{ab}
	1.5%	100 ppm	87.05±13.03 ^f	0.69 ± 0.39^{ab}	0.35 ± 0.33^{ab}
Ovariectomy	0.1%	0.5 ppm	198.13±33.74 ^{bcd}	0.34 ± 0.09^{b}	0.11±0.12 ^b
	0.1%	50 ppm	261.23±86.71 ^a	0.40 ± 0.20^{b}	0.23 ± 0.17^{ab}
	0.1%	100 ppm	252.48±37.93 ^{ab}	0.35 ± 0.08^{b}	0.48 ± 0.41^{ab}
	0.5%	0.5 ppm	177.53±17.28 ^{cd}	0.52 ± 0.23^{b}	0.48 ± 0.79^{ab}
	0.5%	50 ppm	227.03±91.74 ^{abc}	0.60 ± 0.48^{ab}	0.36 ± 0.36^{at}
	0.5%	100 ppm	220.63 ± 31.12^{abc}	0.46 ± 0.41^{b}	0.53 ± 0.25^{ab}
	1.5%	0.5 ppm	167.15±22.74 ^{cde}	1.03 ± 0.62^{a}	0.10 ± 0.07^{b}
	1.5%	50 ppm	172.95±14.93 ^{cde}	0.78 ± 0.51^{ab}	0.41 ± 0.32^{ab}
	1.5%	100 ppm	187.80 ± 53.73^{cd}	0.34 ± 0.26^{b}	0.27 ± 0.29^{at}
Si	gnificance		p<0.001	p<0.01	N.S. ²⁾
Calcium			p<0.01	p<0.01	N.S.
Boron			N.S.	p<0.05	N.S.
Operation			p<0.001	N.S.	N.S.
Calcium×Boron			N.S.	N.S.	p<0.05
Calcium×Operation	1		N.S.	N.S.	N.S.
Boron×Operation			N.S.	p<0.05	N.S.
Calcium×Boron×O	peration		N.S.	N.S.	N.S.

¹⁾Values with different superscripts within a column are significantly different.

2)Not significant.

urinary calcium excretion in high-calcium group. Therefore, it could be suggested that the boron supplementation may be complementary and also useful to calcium nutrition for bone health.

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