

# Effects of Hydrolysis Time on the Molecular Weight Distribution of Alginates Prepared from Sea Tangle Laminaria japonicus

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To prepare oligouronic acids from high-molecular-weight alginates, sea tangle *Laminaria japonicus* alginates were hydrolyzed at 80°C for various hydrolysis times (HT). The effects of hydrolysis time on the distribution ratios (DR) of the molecular weights (MW) of the hydrolyzed alginates were in-vestigated. As HT increased, the DR of the alginates with MW>500 kDa decreased ex-ponentially, while those with MW=50-100 kDa or MW<50 kDa increased exponentially. For MW=300-500 kDa, DR increased exponentially until HT reached 60 min, and then decreased exponentially. Similarly, for MW=100-300 kDa, DR increased exponentially until HT reached 90 min, and then decreased.

Key words: Alginates, Molecular weight, Distribution ratio, Laminaria japonicus, Hydrolysis time

### Introduction

Alginates are binary copolymers of D-mannuronic acid and L-guluronic acid residues arranged in a block-wise pattern (Haug et al., 1967). This material is a linear polymer composed of  $(1\rightarrow 4)$ -linked  $\beta$ -D-mannuronopyranosyl and α-L-guluronopyranosyl units arranged in both homo- and heteropolymeric sequences (Haug et al., 1966, 1967; Penman and Sanderson, 1972; Sime, 1990). Recently, many studies of alginates have examined their physiological functions (Hidaka et al., 1986; Fujihara and Nagumo, 1993; Hideki et al., 1993; Suzuki et al., 1993; Fujiki et al., 1994; Hajime et al., 1994; Kobayashi et al., 1997), which are affected by their molecular weights and uronic acid composition (Kobayashi et al., 1997; You et al., 1997). Numerous studies have been devoted to the analysis of their uronic acid composition. In contrast, there are few studies of the effects of the distribution ratio (DR) of alginate molecular weight (MW) on the physicochemical properties of alginates. Both the molecular weight and the uronic acid composition vary with algal species, and the type and age of the tissues used to prepare the alginates (You and Lim, 2003; You et al., 2004). In addition, the molecular weights depend on the extraction conditions used for their preparation. Therefore, control of the molecular weight of alginates is important in the alginates industry.

There are many methods of measuring the molecular weights of alginates, including light scattering (Turquois and Gloria, 2000), high-performance sizeexclusion chromatography (Nagasawa et al., 2000), gel permeation chromatography (Hein et al., 2000), and calculating molecular weight by measuring the intrinsic viscosity of the alginate solution (Mancini et al., 1996; Chen et al., 1997; You and Lim, 2003; You et al., 2004). Frequently, these methods require expensive equipment beyond the means of small-tomedium-sized enterprises. Moreover, there are many difficulties in deducing the molecular weight of alginates from the intrinsic viscosity, because other components (e.g., ash, metals) contained in alginates can affect the viscosity of an alginate solution (You et al., 2004). Therefore, determining the degree of polymerization (DP) of alginates by measuring the reducing power of uronate residues is one way to evaluate the molecular weights. Very recently, we reported that determining the DP is very useful for evaluating the average molecular weights of alginates (You et al., 2004).

In this study, to prepare oligouronic acids from high-molecular-weight alginates, sea tangle alginates were hydrolyzed for various times and the distributions of the molecular weights of the resulting alginates were measured.

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## Materials and Methods Extraction and purification of alginates

The procedure used to extract alginates from sea tangle was based on the method described by You and Lim (2003). Dried powdered (60 mesh) sea tangle (*Laminaria japonica*) was suspended in 50x 0.5% Na<sub>2</sub>CO<sub>3</sub> solution (w/v). The suspension was shaken in a water bath at 60°C for 6 hours, after which the mixture was filtered through a cloth. To precipitate alginates, the same volume of methanol was added to the filtrate. The precipitated alginates were washed three times with methanol and lyophilized. The dried alginates were ground and stored at 5°C.

#### Hydrolysis of alginates

The 1% alginates solution made from the prepared alginates was adjusted to pH 5.0 with citric acid. Then, it was hydrolyzed in a shaking water bath at 80°C for hydrolyzing times (HT) of 10, 20, 30, 40, 60, 90, or 120 minutes.

### Fractionating the alginates by molecular weight (MW)

The hydrolyzed alginates were fractionated using ultrafiltration membranes (Millipore Corp., Billerica, MA, USA) that established five MW cutoffs: below 50, 50-100, 100-300, 300-500, and over 500 kilo-Daltons.

### Determining the average molecular weight (AMW) of the hydrolyzed alginates

The AMWs of the hydrolyzed alginates were determined using the method described by You and Lim (2003).

### Measuring the ash, uronic acid, and water contents

The ash and water contents of the hydrolyzed alginates were measured using AOAC (1995) methods. The uronic acid content was evaluated using a previously reported method (You et al., 2004).

#### **Results and Discussion**

The physical properties of the alginates prepared from sea tangle are shown in Table 1. The ash content was 21.5%, the uronic acid content was 77.3%, and the AMW was 1,013 kDa. Table 2 shows the distribution ratios (DR) of the MW classes of the alginates before hydrolysis. Of the alginates, 67.4% had MW>500 kDa, with an AMW of 1,307 kDa, followed by 20.2% with MW=300-500 kDa and an AMW of 499 kDa.

The DRs of the MW classes of the alginates hy-

Table 1. The physical properties of alginates used in this study 1,2)

| Ash (g/100 g) | Uronic acid (g/100 g) | AMW (kDa) |
|---------------|-----------------------|-----------|
| 21.5±0.3      | 77.3±0.7              | 1,013±12  |

<sup>1</sup>The alginates were extracted from sea tangle with 0.5% sodium carbonate solution for 6 hr at 60%.

Table 2. The distribution ratio of MW cut-off sizes in alginates used in this study <sup>1,2)</sup>

| _ '              |                               |                       |
|------------------|-------------------------------|-----------------------|
| MW cut-off (kDa) | Distribution (%) <sup>3</sup> | AMW (kDa)             |
| >500             | 67.4±3.7ª                     | 1,307±13 <sup>a</sup> |
| 300-500          | 20.0±2.2 <sup>b</sup>         | 499± 8 <sup>b</sup>   |
| 100-300          | 10.2±1.5°                     | 299± 5°               |
| 50-100           | 1.8±0.5 <sup>d</sup>          | 99± 7 <sup>₫</sup>    |
| <50              | 0.6±0.2 <sup>e</sup>          | 49± 6 <sup>e</sup>    |
|                  |                               |                       |

The alginates were extracted from sea tangle with 0.5% sodium carbonate solution for 6 hr at  $60^{\circ}$ C.

<sup>2</sup>All data (mean±SE with 5 replications) within a column with difference superscripts were signify-cantly difference at p<0.05.

<sup>3</sup>The distribution ratios were expressed as the percentage of content (mg/100 g) of alginates having a certain MW cut-off size to total alginates content (mg/100 g).

drolyzed at 80°C are listed in Table 3. After the alginates were hydrolyzed for 10, 20, 30, or 40 min, the alginates with MW>500 kDa constituted 58.4, 49.4, 40.3, and 31.4%, respectively, with AMWs of 1,204, 1,101, 997, and 894 kDa. For HT of 10 to 40 min, as HT increased, the DRs of alginates with MW >500 kDa and their AMWs decreased gradually. For HT of 60 min, the alginates with MW=300-500 kDa had the highest DR (32.5%) and an AMW of 387 kDa. For HT of 90 min, the highest DR (30.8%) was for MW=100-300 kDa, with an AMW of 190 kDa. After hydrolysis for 120 min, the alginates with MW=50-100 kDa had the highest DR (34.8%), with an AMW of 80 kDa. For HT of over 60 min, as HT increased, the MW cutoff size of the alginates with the highest DR decreased.

The changes in the AMW of alginates hydrolyzed at 80°C for various times before or after fractionation by MW are shown in Fig. 1. Before fractionation, as HT increased, the AMW of the hydrolyzed alginates decreased exponentially. The change in the AMW of the alginates fit equation (1):

AMW=1,016.6 exp(-0.019 HT), 
$$r^2$$
=0.996, (1)

where  $r^2$  is the correlation coefficient.

<sup>&</sup>lt;sup>2</sup>All data were expressed as mean±SE of dry basis with 5 replications.

Table 3. The changes of distribution ratios of MW cut-off sizes in alginates hydrolyzed at  $80\,^{\circ}\mathrm{C}$  and pH 5.0 for various HTs  $^{1,2)}$ 

| J.0 101 variou | 19 1119    |                        |                       |
|----------------|------------|------------------------|-----------------------|
| Hydrolysis     | MW cut-off | Distribution           | AMW                   |
| time (min)     | (kDa)      | ratio¹ (%)             | (kDa)                 |
| 10             | Before     | 100.0                  | 851±17                |
|                | >500       | 58.4±7.1 <sup>a</sup>  | 1,204±11 <sup>a</sup> |
|                | 300-500    | 22.1±4.0 <sup>b</sup>  | 480±15 <sup>b</sup>   |
|                | 100-300    | 12.5±2.3 <sup>c</sup>  | 287± 9°               |
|                | 50-100     | 4.5±0.8 <sup>d</sup>   | 98± 5 <sup>d</sup>    |
|                | <50        | 2.5±0.5 <sup>d</sup>   | 47± 6 <sup>e</sup>    |
|                | Before     | 100.0                  | 705±10                |
|                | >500       | 49.4±8.3ª              | 1,101±15 <sup>a</sup> |
|                | 300-500    | 24.2±3.0 <sup>b</sup>  | 462±12 <sup>b</sup>   |
| 20             | 100-300    | 14.8±4.7°              | 275± 8°               |
|                | 50-100     | 7.2±0.4 <sup>cd</sup>  | 97±19 <sup>d</sup>    |
|                | <50        | 4.4±0.8 <sup>d</sup>   | 45±11 <sup>e</sup>    |
|                |            |                        |                       |
|                | Before     | 100.0                  | 576± 4                |
|                | >500       | 40.3±5.5 <sup>a</sup>  | 997±17 <sup>a</sup>   |
| . 30           | 300-500    | 26.3±4.0 <sup>b</sup>  | 443±10 <sup>b</sup>   |
|                | 100-300    | 17.1±2.2°              | 263± 9°               |
|                | 50-100     | 10.0±2.7 <sup>d</sup>  | 96±13 <sup>d</sup>    |
|                | <50        | 6.3±1.0 <sup>d</sup>   | 43± 9 <sup>e</sup>    |
|                | Before     | 100.0                  | 466± 9                |
|                | >500       | 31.4±3.6°              | 894±15°               |
| 40             | 300-500    | 28.4±4.7 <sup>a</sup>  | 424±17 <sup>b</sup>   |
| 40             | 100-300    | 19.4±2.6 <sup>b</sup>  | 252±10°               |
|                | 50-100     | 12.7±3.1°              | 95± 4 <sup>d</sup>    |
|                | <50        | 8.1±0.6°               | 41± 6 <sup>e</sup>    |
|                | Before     | 100.0                  | 294± 6                |
|                | >500       | 13.3±1.3 <sup>d</sup>  | 688±16 <sup>a</sup>   |
|                | 300-500    | 32.6±0.7 <sup>a</sup>  | 387±19 <sup>b</sup>   |
| 60             | 100-300    | 24.1±1.1 <sup>b</sup>  | 228±10°               |
|                | 50-100     | 18.2±1.0°              | 93±13 <sup>d</sup>    |
|                | <50        | 11.8±1.2 <sup>d</sup>  | 37± 5 <sup>e</sup>    |
|                | Before     | 100.0                  | 201±12                |
|                | >500       | 8.9±0.8 <sup>d</sup>   | 529± 8 <sup>a</sup>   |
|                | 300-500    | 21.8±3.1 <sup>b</sup>  | 323±14 <sup>b</sup>   |
| 90             | 100-300    | 30.8±4.4 <sup>a</sup>  | 190±12°               |
|                | 50-100     | 23.1±2.0 <sup>b</sup>  | 84±10 <sup>d</sup>    |
|                | <50        | 15.4±3.8°              | 34± 8 <sup>e</sup>    |
| 120            | Before     | 100.0                  | 138±12                |
|                | >500       | 3.9±0.5 <sup>d</sup>   | 504±17 <sup>a</sup>   |
|                | 300-500    | 14.5±3.8°              | 307±11 <sup>b</sup>   |
|                | 100-300    | 20.5±2.2 <sup>bc</sup> | 181±16°               |
|                | 50-100     | 34.8±2.9 <sup>a</sup>  | 80±10 <sup>d</sup>    |
|                | <50        | 26.3±5.5 <sup>b</sup>  | 32± 6 <sup>e</sup>    |
|                |            | 20.010.0               |                       |

<sup>&</sup>lt;sup>1,2</sup>Refer to Table 2.

The changes in the AMWs of the alginates after fractionation by MW are shown in Fig. 1. As HT increased, the AMW of the fractionated alginates decreased exponentially. The changes were fitted to equations (a)-(e), respectively.

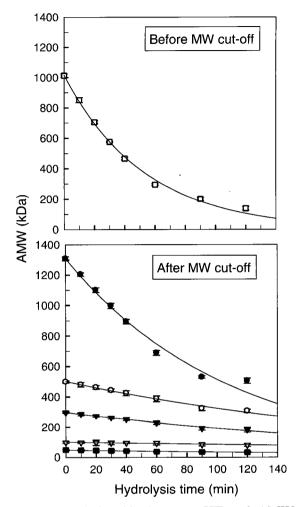


Fig. 1. The relationship between HT and AMW in alginates hydrolyzed at  $80\,^{\circ}\text{C}$  and pH 5.0

- $\square$  Before AMW=1,016.6 exp(-0.019 HT),  $r^2$ =0.996
- MW 500 kDa over AMW=1,308.6 exp(-0.009 HT), r<sup>2</sup>=0.983
- O MW 300-500 kDa AMW=501.7 exp(-0.005 HT), r<sup>2</sup>=0.989
- ▼ MW 100-300 kDa AMW=300.0 exp(-0.004 HT), r<sup>2</sup>=0.989
- $\triangledown$  MW 50-100 kDa AMW=100.6 exp(-0.002 HT),  $r^2$ =0.952
- MW 50 kDa below AMW=48.4 exp(-0.001 HT), r²=0.981

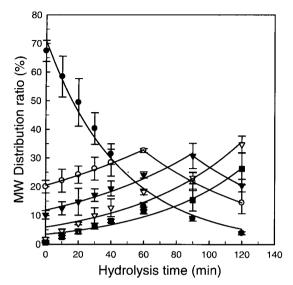


Fig. 2. The relationship between HT and MW distribution ration in alginates hydrolyzed at  $80\,^{\circ}$ C and pH 5.0.

MW 500 kDa over

10-120 min: DR=70.9 exp(-0.022 HT),  $r^2$ =0.980

O MW 300-500 kDa

10-60 min: DR=20.5 exp(-0.008 HT),  $r^2$ =0.994 60-120 min: DR=73.2 exp(-0.014 HT),  $r^2$ =0.999

▼ MW 100-300 kDa

10-90 min: DR=11.8 exp(-0.011 HT),  $r^2$ =0.976 90-120 min: DR=104.5 exp(-0.014 HT),  $r^2$ =0.999

∇ MW 50-100 kDa

10-120 min: DR=6.0 exp(-0.015 HT),  $r^2$ =0.950

■ MW 50 kDa below

10-120 min: DR=3.4 exp(-0.017 HT),  $r^2$ =0.962

The exponent for the decrease in the AMW of the hydrolyzed alginates before fractionation by MW was -0.0019. After fractionation, the rates were -0.009, -0.005, -0.004, -0.002, and -0.001 for decreasing MW, respectively. These data indicate that the smaller the MW of the alginates, the less the decrease in their AMW and the more difficult it was to hydrolyze them.

Figure 2 shows the variation in the DR of the fractionated alginates hydrolyzed at 80°C for various times. As HT increased, the DR of the alginates with MW>500 kDa decreased exponentially, accordion to equation (2).

DR=70.9 exp(-0.222 HT), 
$$r^2$$
=0.980, (2)

where HT is the hydrolysis time (min) at 80°C. The DR for the MW fractions 50-100 kDa and below 50 kDa increased exponentially, following equations (3) and (4), respectively.

DR=6.0 exp(0.015 HT), 
$$r^2$$
=0.950 (3)

DR=3.4 exp(0.017 HT), 
$$r^2$$
=0.962 (4)

The decrease in the DRs of the high-MW alginates with increasing HT agrees with the results of Haug and Larsen (1962), who also reported a loss of uronic acids during hydrolysis due to the breakdown of free uronic acids, which led to the formation of products with reducing power that formed colors with phenolsulfuric acid. Therefore, a considerable portion of the material in alginates with MW<50 kDa was products formed by the breakdown of free uronic acids.

The DR for MW 300-500 kDa increased exponentially until HT reached 60 min, and then decreased exponentially for longer HTs. The increase and decrease fit equations (5) and (6), respectively:

DR=20.5 exp(0.008 HT), 
$$r^2$$
=0.994 (5)

DR=73.2 exp(-0.014 HT), 
$$r^2$$
=0.999. (6)

The DR for MW=100-300 kDa followed a trend similar to the case MW=300-500 kDa: the DR increased exponentially until HT reached 90 min, and then decreased for longer HTs. The increase and decrease fit equations (7) and (8), respectively:

DR=11.8 exp(0.011 HT), 
$$r^2$$
=0.976 (7)

DR=104.5 exp(-0.014 HT), 
$$r^2$$
=0.999. (8)

Overall, the lower the MW of the alginates was, the greater the increase in the DR of the MW.

For all equations, the correlation coefficient (r<sup>2</sup>) exceeded 0.95. The equations derived from the experimental data are very important for the production of alginates of a certain MW, because the manufacturer can use them to determine the appropriate hydrolysis conditions and the HT.

The MW cutoffs of alginates with the maximum DR for hydrolysis at 80°C are listed in Table 4. For HTs from 0 to 40 min, the MW cutoff of alginates with the greatest DR was over 500 kDa. For HT=60, 90, and 120 min it was 300-500, 100-300, and 50-100 kDa, respectively.

Table 4. The changes of maximum MW distribution in alginates hydrolyzed at  $80\,^{\circ}\mathrm{C}$  and pH 5.0

| Hydrolysis time (min) | MW cut-off<br>(kDa) | Distribution ratio <sup>1</sup> (%) | AMW<br>(kDa) |
|-----------------------|---------------------|-------------------------------------|--------------|
| before                | >500                | 67.4±3.7                            | 1,307±13     |
| 10                    | >500                | 58.4±7.1                            | 1,204±11     |
| 20                    | >500                | 49.4±8.3                            | 1,101±15     |
| 30                    | >500                | 40.3±5.5                            | 997±17       |
| 40                    | >500                | 31.4±3.6                            | 894±15       |
| 60                    | 300-500             | 32.6±0.7                            | 387±19       |
| 90                    | 100-300             | 30.8±4.4                            | 190±12       |
| 120                   | 50-100              | 34.8±2.9                            | 80±10        |

<sup>&</sup>lt;sup>1</sup>Refer to Table 2.

### Acknowledgments

This research was supported by a grant from East Coastral Marine Bioresources Research Center in 2005.

#### References

- AOAC. 1995. Official Methods of Analysis, 16<sup>th</sup> ed., Chapter 44, p. 3. Association of Official Analytical Chemists. Arlington, VA, USA.
- Chen, S.L., A.I. Yeh and S.B. Wu. 1997. Effects of particle radius, fluid viscosity and relative velocity on the surface heat transfer coefficient of spherical particles at low Reynolds numbers. J. Food Eng., 31, 473-484.
- Fujihara, M. and T. Nagumo. 1993. An influence of the structure of alginate on the chemotactic activity of macrophages and the antitumor activity. Carbohydr. Res., 243, 211-215.
- Fujiki, K., H. Matsuyama and T. Yano. 1994. Protective effect of sodium alginates against bacterial infection in common carp, *Cyprinus carpio*. L. J. Fish Dis., 17, 349-354.
- Hajime, O., S. Yasushi, Y. Kanto, U. Isamu and K. Koichi. 1994. Possible antitumor promoting properties of marine algae and *in vitro* activity of Wakame seaweed extract. Biosci. Biotech. Biochem., 56, 994-999.
- Haug, A. and B. Larsen. 1962. Quantitative determination of uronic acid composition of alginates. Acta Chem. Scand., 16, 1908-1918.
- Haug, A., B. Larsen and O. Smidsrød. 1966. A study of the constitution of alginic acid by partial acid hydrolysis. Acta Chem. Scand., 20, 183-190.
- Haug, A., B. Larsen and O. Smidsrød. 1967. Studies on the sequence of the uronic acid residues in alginic acid. Acta Chem. Scand., 21, 691-704.
- Hidaka, H., T. Eida, T. Takizawa, T. Tokuzawa and Y. Tashiro. 1986. Effect of fructooligosaccharide on intestinal flora and human health. Bifido. Micro., 5, 37-50.
- Hideki, O., S. Jitsuo and K. Yoshinari. 1993. Direct control of the constituents ratio in a wide range in alginate produced by *Azobacter vinelandii*. Biosci. Biotech. Biochem., 57, 332-336.
- Hein, N.Q., N. Nagasawa, L.X. Tham, F. Yoshii, V.H.

- Dang, H. Mitomo, K. Makuuchi and T. Kume. 2000. Growth-promotion of plants with depolymerized alginates by irradiation. Rad. Phys. Chem., 59, 97-101.
- Kobayashi, N., Y. Kanazawa, S. Yamabe, K. Iwata, M. Nishizawa, T. Yamagishi, O. Nishikaze and K. Tsuji. 1997. Effects of depolymerized sodium alginate on serum total cholesterol in healthy women with a high cholesterol intake. J. Home Econ. Japan, 48, 255-230.
- Mancini, M., M. Moresi and F. Sappino. 1996. Rheological behaviour of aqueous dispersions of algal sodium alginates. J. Food Engr., 28, 283-295.
- Nagasawa, N., H. Mitomo, F. Yoshii and T. Kume. 2000. Radiation-induced degradation of sodium alginate. Polymer Degrad. Stabil., 69, 279-285.
- Penman, A. and G.R. Sanderson. 1972. A method for the determination of uronic acid sequence in alginate. Carbohydr. Res., 25, 273-282.
- Sime, W.J. 1990. Alginates. In: Food Gels, Harris, P., ed. Elsevier, London, UK, pp. 53-78.
- Suzuki, T., K. Nakai, Y. Yoshie, T. Shirai and T. Hirano. 1993. Digestibility of dietary fiber in brown alga, kombu, by rats. Nippon Suisan Gakkaishi, 59, 879-
- Turquois, T. and H. Gloria. 2000. Determination of the absolute molecular weight average and molecular weight distributions of alginates used as ice cream stabilizers by using multiangle laser light scattering measurements. J. Agric. Food Chem., 48, 5455-5458.
- You, B.J., Y.S. Im, I.H. Jeong and K.H. Lee. 1997. Effect of extraction conditions on bile acids binding capacity in vitro of alginate extracted from sea tangle (*Laminaria* spp.). J. Kor. Fish. Soc., 30, 31-38.
- You, B.J. and Y.S. Lim. 2003. Effects of extracting and drying method on physical properties of alginates from sea tangle, *Laminaria japonica*. J. Kor. Fish. Soc., 36, 340-345.
- You, B.J., Y.S. Lim and H.S. Ryu. 2004. Effects of hot water treatment and dialysis on measuring the average molecular weight of alginates. J. Kor. Fish. Soc., 37, 1-6.

(Received June 2005, Accepted September 2005)