Effects of Adenylate Cyclase, Guanylate Cyclase and K_{ATP} Channel Blockade on the Cerebral Blood Flow Response Induced by Adenosine A_{2B} Receptor Agonist in the Rats

Doo Sang Youn and In Chul SHIN*

Department of Pharmacology, College of Medicine, Hanyang University, 17 Haengdang-Dong, Sungdong-Ku, Seoul 133-791, South Korea

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Abstract – This study was performed to investigate the regulatory mechanism of cerebral blood flow of adenosine A_{2B} receptor agonist in the rats, and to define whether its mechanism is mediated by adenylate cyclase, guanylate cyclase and potassium channel. In pentobarbital-anesthetized, pancuronium-paralyzed and artificially ventilated male Sprague-Dawley rats, all drugs were applied topically to the cerebral cortex. Blood flow from cerebral cortex was measured using laser-Doppler flowmetry. Topical application of an adenosine A_{2B} receptor agonist, 5'-N-ethylcarboxamidoadenosine (NECA; 4 umol/l) increased cerebral blood flow. This effect of NECA (4 umol/l) was not blocked by pretreatment with adenylate cyclase inhibitor, MDL-12,330 (20 umol/l). But effect of NECA (4 umol/l) was blocked by pretreatment with guanylate cyclase inhibitor, LY-83,583 (10 umol/l) and pretreatment with ATP-sensitive potassium channel inhibitor, glipizide (5 umol/l). These results suggest that adenosine A_{2B} receptor increases cerebral blood flow. It seems that this action of adenosine A_{2B} receptor is mediated via the activation of guanylate cyclase and ATP-sensitive potassium channel in the cerebral cortex of the rats.

Keywords \square adenosine A_{2B} receptor, 5'-N-ethylcarboxamidoadenosine, cerebral blood flow, adenylate cyclase, guanylate cyclase, ATP-sensitive potassium channel

Adenosine is an endogenous nucleotide that modulates many physiological processes. Its actions are mediated by interaction with specific cell membrane receptors. Adenosine A2 receptors are regarded as coupling to stimulation of adenylate cyclase activity and the production of cAMP (Fredholm et al., 1994) and to mediate the dilator response of cerebral arterioles to adenosine (Edvinsson and Fredholm, 1983). Adenosine level increases in the brain with miderate hypotension, and its increment is related to the regulation of cerebral blood flow (Win et al., 1981). Adenosine is coupled to adenylate cyclase via 2 types of receptor: A1 receptor that mediates an inhibition of adenylate cyclase and A2 receptor that mediates a stimulation of the enzyme (Choca et al., 1987; Gerber and Gähwiler, 1994; Jiang et al., 1992; Van Calker et al., 1979). The A_1 and A_2 receptors have distinct distribution in the central nervous system (Bruns et al., 1987; Stone et al., 1988). The adenosine A₂ receptors are further classified as adenosine A2A receptor and

A_{2B} receptor subtypes based on the potency of agonist and the receptor affinity (Fredholm *et al.*, 1994). And adenosine may cause vasodilatation by increasing intracelluar cAMP, so adenosine A₂ receptor may produce vasodilatation in cerebral cortex. The role for adenosine in the regulation of cerebral blood flow has been proposed by a number of investigators (Dirnagl *et al.*, 1994; Wysham *et al.*, 1986). Adenosine A₂ receptor agonist produces a substantial increase in cerebral blood flow but adenosine A₁ receptor agonist has minimal effects (Coney and Marshall, 1998). Although the precise physiological functions of adenosine A_{2B} receptors remain undefined, role for the adenosine A_{2B} receptors have been suggested in the regulation of neuroglia functions (Fiebich *et al.*, 1996), myocardial contractility (Liang and Haltiwanger, 1995) and vascular smooth muscle tone (Martin, 1992).

However, little is known about the regulatory mechanism of cerebral blood flow of adenosine A_{2B} receptor agonist in the rats. This study was performed to examine the regulatory mechanism of cerebral blood flow of adenosine A_{2B} receptor agonist in the rats, and to define whether its mechanism is mediated by adenylate cyclase, guanylate cyclase and potas-

*Corresponding author

Tel: 02-2220-0651, Fax: 02-2292-6686

E-mail: icshin@hanyang.ac.kr

sium channel.

MATERIALS AND METHODS

The experimental animals, male Sprague-Dawley rats (250-300 gm), were categorized into five groups. The first group of the these groups was treated only with 5'-N-ethylcarboxamidoadenosine (NECA; 4 umol/l), an adenosine A2B receptor agonist, topically to the cerebral cortex. The second group was treated with MDL-12,330 (20 umol/l), an adenylate cyclase inhibitor, topically to the cerebral cortex 10 min before the injection of 4 umol/l of NECA. The third group was treated with LY-83,583 (10 umol/l), an guanylate cyclase inhibitor, topically to the cerebral cortex 10 min before the injection of 4 umol/l of NECA. The fourth group was treated with glipizide (5 umol/l), ATP-sensitive potassium channel inhibitor, topically to the cerebral cortex 10 min before the injection of 4 umol/l of NECA. Another group was sham-operated animal group. All drugs were purchased from RBI chemical company (USA) and SIGMA chemical company (USA). All drugs except LY-83,583 and glipizide were dissolved in artificial cerebrospinal fluid (composition: 120.00 mM NaCl, 2.8 mM KCl, 22.00 mM NaHCO₃, 1.45 mM CaCl₂, 1.00 mM Na₂HPO₄, 0.876 mM MgCl₂) prior to administration and applied topically to the cerebral cortex. LY-83,583 and glipizide were dissolved in 1 % ethanol prior to administration and applied topically to the cerebral cortex. The drug administrations were performed in pentobarbital-anesthetized (50 mg/kg, i.p.), pancuronium-paralyzed (0.1 mg/kg/min, i.v.) and artificially ventilated (Harvard, USA), male Sprague-Dawley rats (250-300 gm). Rectal temperature was maintained at 37±0.5°C with a heating pad, and the rats were placed in a stereotaxic instrument (Kopf, USA) in the prone position and the parietal bone was removed by gradually thinning the bone bilaterally between the temporal and transverse suture lines using a dental burr: a square-shape (5×5 mm) burr hole was made over the right parietal cortex and bone wax was used to achieve hemostasis. The dura mater was carefully removed from the left parietal cortex and the surface of the cortex was superfused with a artificial cerebrospinal fluid. A barrier was fixed caudal to the craniotomy so that when a drug was applied topically to the cortex. Cerebral blood flow levels were measured with a lase-Doppler flowmetry (Laserflo BPM 403A) equipped with a 1-mm-diameter needle probe through the cranial window, where prewarmed artificial CSF saturated with a gas mixture of 95% O₂-5% CO₂ was constantly suffused over the cortical surface. 'Zero' blood flows were determined in each preparation after sacrifice at the conclusion of the experiment. Blood pressure and heart rate were continuously monitored via a femoral arterial catheter (PE-50) connected to a pressure transducer (Statham P23, USA) and a polygraph (Grass, USA). PO₂, PCO₂ and pH of arterial blood were measured with blood gas analyzer before and after stereotaxic experiment. All results are expressed as mean \pm SEM with P<0.05 and P<0.01 considered as the level of significance. The statistical analysis of mean values was performed by analysis of variance (ANOVA). Student's t-test for paired data was also used for statistical evaluation of the results.

RESULTS

A representative experiment showing the effects of topical application of NECA on cerebral blood flow is illustrated in Table 1 and Fig.1. Topical application of NECA caused an increase in cerebral blood flow that reached a maximum in 30 min after application. Pretreatment with alloxazine (4 umol/l), an adenosine A2B receptor antagonist, blocked the NECAinduced cerebral blood flow responses. Topical application of NECA (4 umol/l) caused an increase in cerebral blood flow that reached a maximum in 30 min after application. The responses evoked by NECA were 136±13.7, 138±12.3, 149±15.2, 165± 18.3, 164±17.5, 165±17.6, 164±16.6 and 163±18.8% compared to the value of baseline cerebral blood flow, respectively in 5, 10, 15, 30, 45, 60, 75 and 90 min after topical application of NECA (n=10, Table 1 and Fig. 1). Baseline cerebral blood flow for these rats were 100±11.2%. Topical application of an equivalent volume of artificial cerebrospinal fluid did not affect the basal cerebral blood flow. Cerebral blood flow was not

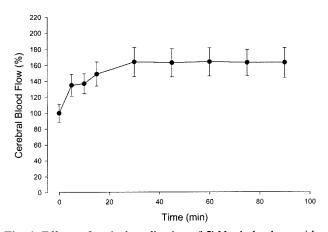


Fig. 1. Effects of topical application of 5'-N-ethylcarboxamido-adenosine (NECA; 4 umol/l) on cerebral blood flow responses in rats. Values are mean±S.E. in 10 rats

Table I. Percent changes in cerebral blood flow of 5'-N-ethylcarboxamidoadenosine (NECA; 4 umol/l) only, NECA (4 umol/l) after pretreatment with MDL-12,330 (MDL; 20 umol/l), NECA (4 umol/l) after pretreatment with LY-83,583 (LY; 10 umol/l) and NECA (4 umol/l) after pretreatment with glipizide (GL; 5 umol/l). Pretreatment with MDL-12,330, LY-83,583 or glipizide have no effects on cerebral blood flow. Cerebral blood flow is not affected in sham-operated animal group. Data are the mean±S.E. *p<0.05, compared to 0 time cerebral blood flow.

	Cerebral Blood Flow (%)								
Treatment	Minute after Treatment								
	0	5	10	15	30	45	60	75	90
NECA	100±11.3	136±13.7*	138±12.3*	149±15.2*	165±18.3*	164±17.5*	165±17.6*	164±16.6*	163±18.8*
MDL+NECA	100±13.2	129±13.6*	134±13.1*	142±16.2*	157±16.5*	159±19.5*	157±18.8*	158±17.7*	156±16.6*
LY+NECA	100±14.1	104±14.4	106±12.9	107±13.3	106±12.5	108±15.1	107±14.3	110±15.8	109±16.5
GL+NECA	100±12.8	104±14.1	106±11.8	108±13.6	108±11.7	109±14.8	109±12.1	110±14.7	109±15.8
sham-operated	100±11.6	103±12.9	106±13.6	107±16.5	108±13.2	107±12.7	109±13.6	109±14.2	110±14.9

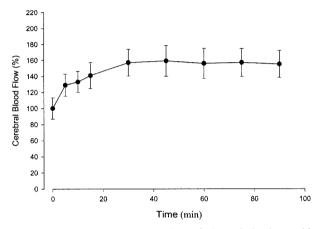


Fig. 2. Effects of topical application of 5'-N-ethylcarboxamido-adenosine (NECA; 4 umol/l) after pretreatment with MDL-12,330 (20 umol/l) on cerebral blood flow responses in rats. Values are mean±S.E. in 10 rats.

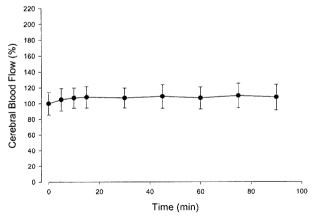


Fig. 3. Effects of topical application of 5'-*N*-ethylcarboxamido-adenosine (NECA; 4 umol/l) after pretreatment with LY-83,583 (10 umol/l) on cerebral blood flow responses in rats. Values are mean±S.E. in 10 rats.

affected in sham-operated animal group.

Pretreatment with MDL-12,330 (20 umol/l) did not attenuate the NECA-induced cerebral blood flow responses; 129±13.6,

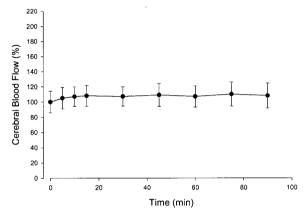


Fig. 4. Effects of topical application of 5'-N-ethylcarboxamido-adenosine (NECA; 4 umol/l) after pretreatment with glipizide (5 umol/l) on cerebral blood flow responses in rats. Values are mean±S.E. in 10 rats.

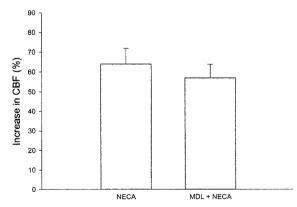


Fig. 5. Changes in cerebral blood flow (CBF) of 5'-N-ethyl-carboxamidoadenosine (NECA; 4 umol/l) only and NECA (4 umol/l) after pretreatment with MDL-12,330 (MDL; 20 umol/l) topically. Values are recorded at the end of 30 minutes after topical application of NECA. Values are the mean±S.E. in 10 rats.

134±13.1, 142±16.2, 157±16.5, 159±19.5, 157±18.8, 158±17.7 and 155±16.6% compared to the value of baseline cerebral blood flow, respectively in 5, 10, 15, 30, 45, 60, 75 and 90

min after topical application of NECA (*n*=10, Table 1, Fig. 2 and Fig. 5). Baseline cerebral blood flow for these rats were 100±13.2%. Another different groups were treated with MDL-12,330 (10 umol/l, 30 umol/l) did not affect NECA-induced cerebral blood flow responses. Therefore in this study authors chose the middle concentration.

Pretreatment with LY-3,583 (10 umol/l) significantly attenuated the NECA-induced cerebral blood flow responses; 104 ± 14.4 , 106 ± 12.9 , 107 ± 13.3 , 106 ± 12.5 , 108 ± 15.1 , 107 ± 14.3 , 110 ± 15.8 and $109\pm16.5\%$ compared to the value of baseline cerebral blood flow, respectively in 5, 10, 15, 30, 45, 60, 75 and 90 min after topical application of NECA (n=10, Table 1, Fig. 3 and Fig. 6). Baseline cerebral blood flow for these rats

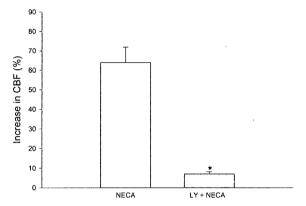


Fig. 6. Changes in cerebral blood flow (CBF) of 5'-N-ethylcarbox-amidoadenosine (NECA; 4 umol/l) only and NECA (4 umol/l) after pretreatment with LY-83,583 (LY; 10 umol/l) topically. Values are recorded at the end of 30 minutes after topical application of CPCA. Values are the mean±S.E. in 10 rats. *P<0.01, compared to NECA only group.

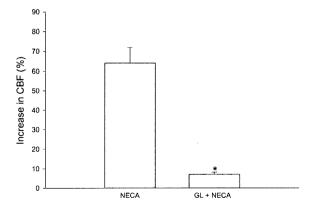


Fig. 7. Changes in cerebral blood flow (CBF) of 5'-N-ethylcarbox-amidoadenosine (NECA; 4 umol/l) only and NECA (4 umol/l) after pretreatment with glipizide (GL; 5 umol/l) topically. Values are recorded at the end of 30 minutes after topical application of NECA. Values are the mean±S.E. in 10 rats. *P<0.01, compared to NECA only group.

were 100±14.2%. Another different groups were treated with LY-83,583 (5 umol/l, 15 umol/l) affected NECA-induced cerebral blood flow responses concentration-dependently.

Pretreatment with glipizide (5 umol/l) significantly attenuated the NECA-induced cerebral blood flow responses; 104± 14.1, 106±11.8, 108±13.6, 108±11.7, 109±14.8, 109±12.1, 110 ±14.7 and 109±15.8% compared to the value of baseline cerebral blood flow, respectively in 5, 10, 15, 30, 45, 60, 75 and 90 min after topical application of NECA (*n*=10, Table 1, Fig. 4 and Fig. 7). Baseline cerebral blood flow for these rats were 100±12.8%. Another different groups were treated with glipizide (1 umol/l, 10 umol/l) affected NECA-induced cerebral blood flow responses concentration-dependently. Therefore in this study authors chose the middle concentration. Topical application of MDL-12,330 (20 umol/l), LY-83,583 (10 umol/l) and glipizide (5 umol/l) had no effects on cerebral blood flow. No significant changes in PO₂, PCO₂ and pH of arterial blood were seen following stereotaxic experiment (Table II).

DISCUSSION

Adenosine is an endogenous nucleotide that modulates many physiological processes. Its actions are mediated by interaction with specific cell membrane receptors. Four subtypes of adenosine receptors have been cloned: A₁, A_{2A}, A_{2B} and A₃. It is also recognized that adenosine A2B receptors are coupled to intracellular pathways different from those of adenosine A2A receptors, a finding that may provide the basis for their distinct physiological role. Pharmacological identification of A2B receptors, based on their affinity and characteristic order of potency for receptors. Adenosine A_{2B} receptors participate in the regulation of vascular tone (Webb et al., 1992). Therefore adenosine-induced vasodilation is mediated via adenosine A_{2R} receptors. In this present study, topical application of NECA, an adenosine A_{2B} receptor agonist, in anesthetized and artificially ventilated rats elicited an increase in cerebral blood flow. Adenosine plays an important role in many physiological processes. Its actions are mediated by specific cell surface receptors coupled to G proteins (Fredholm et al., 1994; Olah and Stiles, 1996). Adenosine vasodilates the vasculature of

Table II. Summary of blood gas analysis

Variables	Before Experiment	After Experiment
PaCO ₂ , mmHg	84.8±2.6%	88.1±4.2%
PaO ₂ , mmHg	38.9±2.3%	40.7±1.7%
pН	7.22±0.35	7.28±0.42

skeletal muscle and the brain (Mian and Marshall, 1991; Skinner and Marshall, 1996; Thomas and Marshall, 1994). Ngai (Ngai and Winn, 1993) investigated the receptors involved in adenosine-induced dilation in cerebral resistance arterioles and adenosine acts primarily via A2 receptors to elicit cerebral vasodilation. Some experimental evidences suggest that adenosine receptor plays a critical role in the mediation of cerebral blood flow responses (Van Calker et al., 1979; Hong et al., 1999), and some experimental evidences suggest that adenosine A2 receptor agonist produces a substantial increase in cerebral blood flow (Coney and Marshall, 1998; Van Wylen et al., 1989). The adenosine A2 receptor is further classified as adenosine A_{2A} receptor and A_{2B} receptor subtypes based on the potency of agonist and the receptor affinity (Fredholm et al., 1994). Cloning techniques have confirmed the existence of distinct A_{2A} receptor and A_{2B} receptor subtypes of A₂ receptor (Stehle et al., 1992). Although the precise physiological functions of the A_{2B} receptors remain undefined, roles for the A_{2B} receptor has been suggested in the regulation of intestinal chloride secretion (Strohmeier et al., 1995). Hong et al. (Hong et al., 1999) demonstrated that when the cortical surface is suffused with cAMP, the release of adenosine is increased in the artificial cerebrospinal fluid and they suggested that the cAMPadenosine pathway as a metabolic mechanism is implicated in the production of adenosine in the rat pial artery and contributes to the regulation of vasodilation in response to hypotension. But little is known about the regulatory mechanism of cerebral blood flow of adenosine A2B receptor agonist. And adenosine A2B receptor agonist produces vasodilation of rat pial artery and cerebral blood flow autoregulation (Shin et al., 2000). In the present study NECA, adenosine A_{2B} receptor agonist, produces a substantial increase in cerebral blood flow. Martin and Potts (Martin and Potts, 1994) demonstrated a similar result in that renal artery of rat contains adenosine A_{2R} receptors that are located on the endothelium.

This effect of NECA (4 umol/l) was not blocked by pretreatment with adenylate cyclase inhibitor, MDL-12,330 (20 umol/l). Hong *et al.* (Hong *et al.*, 1994; Hong *et al.*, 1996) demonstrated that the vasodilation of the pial artery is closely related with accumulation of intracellular cAMP. An involvement of the metabolic AMP-adenosine pathway was demonstrated as a likely mechanism in the production of adenosine that acted as a regulator of vasodilation in response to hypotension (Hong *et al.*, 1999). And adenosine may cause vasodilatation by increasing intracelluar cAMP, so adenosine A₂ receptor may produce vasodilatation in cerebral cortex by stimulation of adenylate

cyclase. But the role of adenosine A_{2B} receptor agonist in the cerebral blood flow response is not related to cAMP.

This effect of NECA (4 umol/l) was blocked by pretreatment with guanylate cyclase inhibitor, LY-83,583 (10 umol/l). The level of cyclic GMP in vascular smooth muscle is an important regulator of blood flow. Many vasodilators work through increases in cyclic GMP (Hyman et al., 1989; Zhou and Torphy, 1991). Generally NO production leads to increase in the level of cyclic GMP and may help couple cerebral blood flow and metabolism (Dirnagl et al., 1993). It is also believed that NO is involved in the coupling of neurotransmitter receptor stimulation with cellular cyclic GMP responses (Garthwaite, 1988). In this present study adenosine A_{2B} receptor is mediated by stimulation of guanylate cyclase. Therefore adenosine A_{2B} receptor may increase cerebral blood flow by increasing intracelluar cGMP. So adenosine A2B receptor may produce vasodilatation in cerebral cortex by stimulation of guanylate cyclase. We demonstrated that NECA-induced vasodilatation in cerebral cortex was significantly inhibited by guanylate cyclase inhibitor, LY-83,583, but not by adenylate cyclase inhibitor, MDL-12,330. Therefore the regulation of cerebral blood flow in adenosine A_{2B} receptor is mediated by guanylate cyclase. In further study we may determine the level of cGMP level to confirm this evidence.

This effect of NECA (4 umol/l) was blocked by pretreatment with ATP-sensitive potassium channel inhibitor, glipizide (5 umol/l). Some authors report that the action of adenosine is mediated via the activation of potassium channel (Trussel and Jackson, 1985; Nicoll, 1988; Gerber and Gähwiler, 1994). Adenosine receptor and GABA (Gamma-amino butyric acid) receptor act via the activation of potassium channel in substantia nigra (Watt $et\ al.$, 1995). It is known that ATP-sensitive potassium channel inhibitor is involved in the peripheral response of adenosine A_2 receptor. Therefore the regulation of cerebral blood flow in adenosine A_{2B} receptor is mediated by potassium channel.

In conclusion, our results show that adenosine A_{2B} receptor increases cerebral blood flow and this action of adenosine A_{2B} receptor is mediated by the activation of guanylate cyclase and ATP-sensitive potassium channel in the cerebral cortex of the rats.

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