

## Effect of concentrated carbon dioxide exposure on the mycelial growth and fruit body initiation of *Ganoderma lucidum*.

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### 이산화탄소 농도가 영지버섯균의 균사생장과 자실체원기 유도에 미치는 영향

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**ABSTRACT :** The effect of CO<sub>2</sub> concentration (500, 3,000, 6,000 μl/l) on the mycelial growth and fruit body primordium formation of *Ganoderma lucidum* on nutrient agar medium was examined. Optimum CO<sub>2</sub> concentration for vegetative growth was above 3,000 μl/l. Fruit body initiation was accelerated at higher than 3,000 μl/l CO<sub>2</sub> exposure but the maximum number and size of primordia, and primordium color were not influenced by CO<sub>2</sub> concentrations. Whereas each atypical fruiting structure forming stock culture showed different fruiting time under each concentration of CO<sub>2</sub> exposure.

**KEYWORDS :** *Ganoderma lucidum*, CO<sub>2</sub> concentration, mycelial growth, fruit body initiation

Carbon dioxide is one of important environmental factors that affect on growth and morphogenesis of Basidiomycetes. Excess CO<sub>2</sub> inhibits the fruiting; especially fruit body development, and sometimes generates abnormal fruit bodies in Basidiomycetes (Tschierpe, 1959; Niederpruem, 1963; Tschierpe and Sinden, 1964; Taber, 1966; Long and Jacobs, 1974; Kinugawa et al., 1986, 1994). On the contrary, primordium formation of *Sphaerobolus stellatus* is inhibited when CO<sub>2</sub> is eliminated from air (Ingold and Nawarz, 1967). Hintikka and Korhonen (1970) found that vegetative growth of lignicolous basidiomycetes was markedly stimulated under higher CO<sub>2</sub> concentration while that of litter inhabiting basidiomycetes was suppressed severely in proportion to the increment of CO<sub>2</sub> concentration.

*In vitro* culture under the ventilation and light irradiation, the colonies of *Ganoderma lucidum* form atypical fruiting structure (AFS) bearing basidiospores without the basidiocarp formation and fruit body primordium (FBP) (Shin and Seo, 1988; Seo et al., 1995). In AFS forming isolates, the colonies of *G. lucidum* formed AFS on the agar media. Basidia differentiated directly from generative hyphae on the

outside of AFS. According to light response, *G. lucidum* isolates are divided into three groups; AFS-forming isolates, FBP forming isolates and chlamyospore forming isolates without fruiting on the agar media (Seo et al., 1995). For the FBP formation, only light irradiation is essential, but for AFS formation both light irradiation and ventilation are required (Seo et al., 1995). It has been well known in commercial cultivation that the fruit body formation of *G. lucidum* is inhibited even under weak semi-anaerobic condition. A major factor that reflects the inhibitive effect by bad ventilation has not been clarified in *G. lucidum* although excess CO<sub>2</sub> is speculated to be a major inhibitive factor.

Thus the present study is conducted to examine the effect of carbon dioxide on the mycelial expansion and AFS and FBP formation of *G. lucidum*, a white rot fungus, to elucidate the major inhibitive factor under bad ventilation.

## Materials and Methods

### Isolates

The dikaryon isolates of *G. lucidum* used in this study are listed in Table 1. Two isolates, Gl-009 and Gl-012, and two isolates, Gl-010 and Gl-020 were

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used as FBP and AFS forming isolate, respectively (Seo *et al.*, 1995). Isolate GI-015 that forms AFS even under dark condition was isolated from white fruit body (Seo *et al.*, 1995).

### Culture condition

All isolates were cultured and maintained on a nutritionally complete agar medium (CM) as reported previously (Seo *et al.*, 1995). Mycelial disks (6 mm in diameter) were placed in 90 mm plastic Petri dishes containing about 30ml of CM, and were incubated at  $27 \pm 1^\circ\text{C}$  for 20 days under light and ventilated condition. As light source, white fluorescent lamp (FL 20 SD, Matsushita) was employed. The light intensity of fluorescent lamps was adjusted to about 500 to 1,000 lux by controlling the distance between lamps and the Petri dishes. The relative humidity was adjusted at  $80 \pm 5\%$  and  $\text{CO}_2$  levels in the air of growth cabinets were maintained about 550, 3,000 and 6,000  $\mu\text{l/l}$ , respectively. Oxygen concentrations in all growth cabinets were maintained at about 20% until end of experiments. To examine the effect of concentrated  $\text{CO}_2$  on mycelial growth, diameter of colony was measured by 10 days of inoculation. In each experiment, ten or fifteen replicates were applied for each  $\text{CO}_2$  exposure.

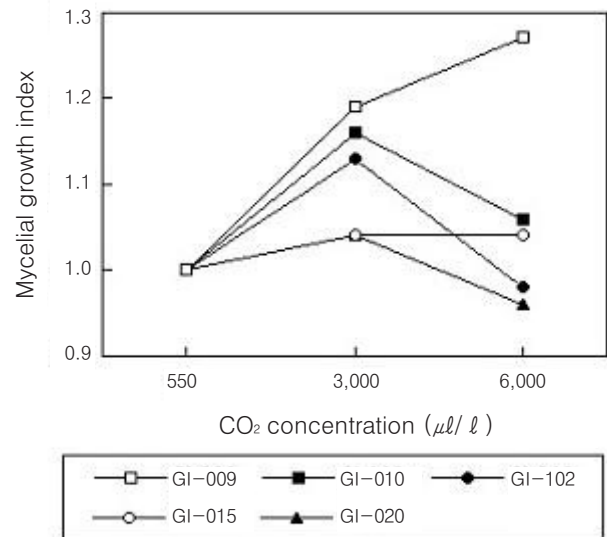
## Results and Discussion

The  $\text{CO}_2$  optimum for vegetative growth of stock cultures GI-010, GI-012 and GI-020 was 3,000  $\mu\text{l/l}$ . Whereas, that for vegetative growth of stock cultures GI-009 and GI-015 was higher than 6,000  $\mu\text{l/l}$  (Fig. 1). Hintikka and Korhonen (1970) found that vegetative growth of lignicolous basidiomycetes was markedly stimulated at around 10%  $\text{CO}_2$  but that of soil- and litter-inhabiting fungi was suppressed in proportion to the increment of  $\text{CO}_2$  concentration of atmosphere (0.03%).

**Table 1.** Isolates of *G. lucidum* used in this experiment

Isolate	Morphogenesis on agar media <sup>1)</sup>	Sources	Collected country
GI-009	FBP-forming	Wild fruit body	Japan
GI-010	AFS-forming	Cultivated fruit body	Korea
GI-012	FBP-forming	Wild fruit body	Japan
GI-015	AFS-forming	White fruit body	Korea
GI-020	AFS-forming	Wild fruit body, TMI-50087	Japan

<sup>1)</sup> Morphogenesis on agar media were confirmed by Seo *et al.*, (1995), AFS; Atypical fruiting structure bearing the basidiospores, FBP; Fruit body primordium.



**Fig. 1.** Effect of different concentration of  $\text{CO}_2$  exposure on mycelial growth of *G. lucidum*.

All cultures were incubated under light condition for 10 d on complete medium (CM; 2% glucose, 0.2% yeast extract, 0.2% peptone, 0.0046%  $\text{KH}_2\text{PO}_4$ , 0.01%  $\text{KH}_2\text{PO}_4$ , 0.005%  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ ). All cultures were replicated 10 times. The light intensity was adjusted to about 500 to 1,000 lux. Mycelial growth index = mycelial growth rate of the control/mycelial growth rate on the  $\text{CO}_2$  enriched conditions.

Mycelial expansion rates of *Flammulina velutipes*, *Pleurotus ostreatus*, *Pholiota nameko*, and *Hypsizygus marmoreus* have no significant difference between 600  $\mu\text{l/l}$  and 6,000  $\mu\text{l/l}$   $\text{CO}_2$  exposure (Kinugawa *et al.*, 1986). This suggests that the  $\text{CO}_2$  optimum for vegetative growth *G. lucidum* is very low comparing with those of other wood rotting fungi. There may be an inverse relationship between vegetative growth and fruiting in fungi. The obtained data, however, indicate that this relationship cannot be always observed in *G. lucidum* (Fig. 1; Table 2).

**Table 2.** Effect of different concentration of CO<sub>2</sub> ( $\mu\text{l}/\ell$ ) exposure on the formation of AFS and FBP of *G. lucidum*<sup>1)</sup>

Isolate	AFS formation at different CO <sub>2</sub> concentration ( $\mu\text{l}/\ell$ )			FBP formation at different CO <sub>2</sub> concentration ( $\mu\text{l}/\ell$ )		
	550	3,000	6,000	550	3,000	6,000
GI-009	-	-	-	-	++	++
GI-012	-	-	-	+	++	+++
GI-010	-	+	+	-	-	-
GI-020	+++	++	+	-	-	-
GI-015	+++	+++	+++	-	-	-

<sup>1)</sup> All cultures were incubated under light condition for 20 d. -, only vegetative growth, +; 10~39% forming (AFS- or FBP-forming Petri-dish / total replicate X 100), ++; 40~69% forming, +++; 70~100% forming.

When *G. lucidum* cultured under semi anaerobic condition, i. e., cultured in the Petri dish double sealed with Para film(American National Can), the mycelial growth was accelerated. This stimulation may base on the gaseous environments such as decrease of O<sub>2</sub> concentration, and increase of air humidity and various volatile substances, and not be caused merely by the increment of CO<sub>2</sub> concentration. FBP forming stock cultures formed FBP earlier under the higher CO<sub>2</sub> concentration than atmospheric condition(Table 2). No significant difference was observed on the number, size and color of FBP among different CO<sub>2</sub> concentrations(Table 2). AFS formation of stock culture G1-015 was not influenced under among different CO<sub>2</sub> concentrations (Table 2). AFS formation of stock culture G1-020 decreased with the increment of CO<sub>2</sub> concentration (Table 2). Stock culture G1-010 formed primordia at 3,000 and 6,000  $\mu\text{l}/\ell$  CO<sub>2</sub> exposure, not at 550  $\mu\text{l}/\ell$  CO<sub>2</sub> exposure(Table 2). In previous study(Seo *et al.*, 1995), the AFS formation of *G. lucidum* was inhibited but the mycelial growth was rather accelerated by preventing aeration, i. e., cultivation in the Petri dish double-sealed with Para film. The difference in the results between present research and previous one suggests that this may be derived from excess CO<sub>2</sub> accompanying other gaseous environments, and this can not be caused merely by the increment of CO<sub>2</sub> concentration.

Fruiting of *G. lucidum* also greatly affected by light and CO<sub>2</sub> level in cultivation house(Hemmi and Tanaka, 1936; Stamet, 1993). It has been observed that a long stipe and a small pileus, so called antler type fruit body were formed in aeration prevented artificial cultivation of *G. lucidum*, but has not been confirmed that whether those abnormal fruit bodies derived from higher CO<sub>2</sub> level or genetic backgrounds. In many basidiomycetes,

stipe elongation and inhibition of pileus development by higher level of CO<sub>2</sub> exposure have been also reported(Taber, 1966; Kinugawa *et al.*, 1994). FBP forming isolates formed FBP earlier under the higher CO<sub>2</sub> levels than atmospheric condition (Table 2). However, no significant differences were observed on the number, size and color of FBP among different CO<sub>2</sub> levels. In previous study (Seo *et al.*, 1995), the AFS formation of *G. lucidum* was inhibited by preventing aeration, while the mycelial growth was rather accelerated. Whereas, the FBPs were formed regardless of aeration. These results might that higher concentration of CO<sub>2</sub> does not affect fruit body primordium initiation and stipe elongation. AFS-forming isolates, GI-010 and GI-015 did not show significant differences on the AFS formation among different CO<sub>2</sub> levels. However, the exposure of higher concentration of CO<sub>2</sub> inhibited AFS formation in isolate GI-020. Although we have disregard for effect of oxygen concentration(in this experiment maintained about 20%) in the growth cabinet, high concentrated CO<sub>2</sub> is effective for induction of fruit body primodium, but not AFS formation including sporulation of *G. lucidum*.

In conclusion, *G. lucidum* is very sensitive to excess CO<sub>2</sub> than other wood rotting fungi that have been examined and it shows somewhat similar tendency in the reaction to CO<sub>2</sub> concentration with litter and soil inhabiting fungi.

## References

- Hemmi, T. and Tanaka, I. 1936. Experiments for developing sporophores of *Ganoderma japonicum*. Bot. and Zoo. 4: 13-23. (in Japanese.)  
 Hintikka, V. 1982. The colonisation of litter and wood by basidiomycetes in Finnish forests. In: "Decomposer

- basidiomycetes," (ed. by Frankland, J. C. *et al.*), pp. 227-239. Cambridge Univ. Press, London.
- Hintikka, V. and Korhonen, K. (1970). Effect of carbon dioxide on the growth of lignicolous and soil-inhabiting Hymenomycetes. *Communicationes Instituti Forestalis Fenniae*, 62:5, 1-29.
- Ingold, C. T. and Nawaz, M. 1967. Carbon dioxide and fruiting in *Sphaerobolus*. *Ann. Bot. (N. S.)* 31: 351-357.
- Kinugawa, K. Takamatsu, Y. Suzuki, A. Tanaka, K. and Kondo, N. 1986. Effect of concentrated carbon dioxide on the fruiting of several cultivated basidiomycetes. *Trans. mycol. Soc. Japan*, 27: 327-340. (In Japanese.)
- Kinugawa, K. Suzuki, A. Takamatsu, Y. Kato, M. and Tanaka, K. 1994. Effect of concentrated carbon dioxide on the fruiting of several cultivated basidiomycetes (II). *Mycoscience*, 35: 345-352.
- Long, P. E. and Jacobs, L. 1974. Aseptic fruiting of the cultivated mushroom *Agaricus bisporus*. *Trans. Br. Mycol. Soc.* 63: 99-107.
- Niederpruem, D. J. 1963. Role of carbon dioxide in the control of fruiting of *Schizophyllum commune*. *J. of Bacteriol.* 85: 1300-1308.
- Seo, G. S., Shin, G. C., Otani, H., Kodama, M. and Kohmoto, K. 1995. Formation of atypical fruiting structures in *Ganoderma lucidum* isolates on a nutrition agar media. *Mycoscience*, 36: 1-7.
- Shin, G. C. and Seo, G. S. 1988. Formation of the nonbasidiocarpous basidiospore of *Ganoderma lucidum*. *Kor. J. Mycol.* 16: 230-234. (In Korean.)
- Stamets, P. 1993. The polypore mushrooms of the genera *Ganoderma*, *Grifola* and *Polyporus*. In: "Growing gourmet and medical mushrooms". pp. 351-369. Ten Speed Press, Berkely, California.
- Taber, W. A. 1966. Morphogenesis in basidiomycetes. In: "The Fungi," (ed. by Ainsworth, G. C. and Sussman, A. S.) Vol. 2, pp. 387-412. Academic Press, New York.
- Tschierpe, H. J. 1959. Der Einfluss von Kohlendioxyd auf die Fruchtkorperbildung und die Fruchtkorperform des Kulturchampignons. *Mush. Sci.* 4: 235-250.
- Tschierpe, H. J. and Sinden. 1964. Weitere Untersuchungen uer die Bedeutung von Kohlendioxyd fur die Fructifikation des Kulturchampignons, *Agaricus campestris* var. *bisporus* (L) Lge. *Arch. Mikrobiol.* 49: 405-425.