The structure of the plankton community and the cyanobacterial bloom during the rainy season in mesoeutrophic lake (Lake Juam), Korea

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Abstract

강우가 집중하고 남조류가 대발생하던 시기의 중영양호소인 주암호에서 박테리아, 식물플랑크톤, ANF, HNF, 섬모충플랑크톤, 동물플랑크톤과 환경요인들의 변화에 대해 조사하였다. 조사기간동안 우점종은 Cyclotella meneghinana, Aulacoseira granulata, Cryptomonas tetrapyrenoidsa로서 총현존량의 67% 이상을 차지하였다. 조사기간동안 남조류 대발생의 구성원은 현존량 및 종조성에서 크게 변화 하였다. 여름철 강우에 의한 엽록소 a, 수질 (COD, TP)의 변동이 심하였으며, 특히 C. tetrapyrenoidsa는 HNF, nauplii, rotifer, 섬모충의 변동에 매우 밀접한 관계를 보였다. 포식자인 rotifer는 박테리아, 피코플랑크톤이나 ANF같은 소형플랑크톤의 변동과 높은 관계를 보였다.

Key words: cyanobacterial bloom, rainfall, phytoplankton, physicochemical variables

I INTRODUCTION

The cyanobacterial bloom, triggered by stagnation of water body and high and/or continuous organic loadings from watersheds and tributaries, is one of the most obvious indicators of eutrophication in freshwater lakes. ^{1,2)} The nuisance causing cyanobacterium *Microcystis* spp. decreases dissolved oxygen and light penetration, releases microcystin that threatens herbivores, planktivores and other biota, and ultimately deteriorates an aquatic ecosystem. ^{3,4,5,6)}

Lake Juam was constructed in 1992, and since then, a bloom of *M. aeruginosa* has been occurring every summer. A dense cyanobacterial bloom occurred at the inflow site

of the lake and gradually extended towards the lake center and dam.⁹⁾ In 1993, M. aeruginosa, species genera. of the Coelasphaerium, Ceratium, Coelastrum, Eudorina, and Aulacoseira dominated the phytoplankton community of the lake.71 In 1994 and 1995, new including Chroococcus. Peridinium, Dictyosphaerium, Scenedesmus, Staurastrum, Fragilaria and Melosira, appeared in the inflow region of the lake along with Microcystis aeruginosa.8 Finally, abundance and species composition of phytoplankton community shifted from Microcystis and Anabaena to Cryptomonas, Coelastrum and Peridinium. 9,10) Aulacoseira. There are no published works that adequately discuss the mechanism of bloom formation or other aspects related to algal ecology of Lake

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Juam.

The present study was carried out in Lake Juam to (i) learn the relationships between abundance and distribution of plankton and physicochemical variables, and (ii) to understand the seasonal succession patterns and predator prey relationships.

II. METHODS AND MATERIALS

Lake Juam (lat. 3500N, long. 12715 E) impounds two upper streams of the Seomjin River, Korea. The lake has the surface area of 1,010km² and is surrounded by mountainous valleys 400 m at sea level). The lake is connected to another small lake, Lake Sangsa, through an underground tunnel. In Lake Juam, thermal stratification has been occurring near the dam every year, and irregularly in the central and inflow regions.

Sampling was carried out at three stations. Sta J1 was the upstream of the dam; Sta J2 was in the middle of the lake; Sta J3 was inflow site near the confluence of the two tributaries, Posung and Dongbok streams.

Water temperature, dissolved oxygen (DO), conductivity, and turbidity were directly with a YSI meter (Model 610) equipped with a pH meter. A transparency was measured by Secchi disc. Water samples were collected with a 5 L Van Dorn sampler. Samples were GF/F Whatman filtered with a filter precombusted at 450°C for 3 h. Dissolved reactive phosphate phophorus was measured by the ascorbic acid method, and nitrite and nitrate by the cadmium reduction method. 11) Ammonium was determined using the salicylate hexacyanoferrate dichloroisocyanurate system. (2) Total phosphorus (TP) and total nitrogen (TN) were measured by the methods as above following persulphate digestion at 120°C for 45 min. For chlorophyll *a* (Chl *a*) measurement, 100 mL of water sample were filtered with a Nucleopore filter under gentle vacuum(<40 kPa). The amount of chl *a* was measured by high performance liquid chromatography(LC Module 1, Waters, Inc.). Other limnological variables, such as biochemical oxygen demand(BOD), chemical oxygen demand(COD), suspended solids(SS) and SiO₂, were analyzed using standard methods.¹¹⁾

For the enumeration of a pico and nanoplankton, the surface water samples were preserved with 1% glutaraldehyde solution and kept refrigerated at 4°C in the dark. Bacteria, picocyanobacteria (PC), eukaryotic picoplankton (EPP), autotrophic nanoflagellates (ANF) and heterotrophic nanoflagellates (HNF) were counted with a fluorescence microscope after being stained with DAPI and FITC. 13,14,15)

Other phytoplankton ciliate and were enumerated from samples preserved with 1% and examined with an microscope after sedimentation for 12 hrs.16) Zooplankters were counted with a dissecting microscope, after a 5 10 L surface water sample filtered through a 40µm net and preserved with 5% sugar formalin. Carbon content of phytoplankton was estimated from the volume of live algae (1.33x fixed cells) with the following equations:

Carbon (pg cell^{-1})=0.12×Vol^{1.051} (Montagenes *et al.* 1994). 17)

Plankton volume was determined from the geometric shape or gross volume of sample.

A sequential multiple regression analysis was carried out to statistically explain the various plankton component abundances (changes) with changes in all the measured physicochemical variables. The statistical significance of the series of correlation coefficients was determined

Table 1. Correlation among the planktonic components during rainy season in Lake Juam

Kinds	HNF	Copepoda	Nauplius	Cladocera	Rotifera	Ciliates
Bacteria	0.38*	0.41*	0.65***	0.41*	0.70***	0.36*
Picoplankton	0.29	0.14	0.58**	0.04	0.64***	0.54**
ANF	0.59**	0.18	-0.02	-0.10	0.71***	0.82***
Cryptomonas	0.44*	0.10	0.48*	0.03	0.74***	0.70***
Pediastrum	0.60***	0.18	0.21	-0.09	0.86***	0.86***
Anabaena	-0.12	0.10	-0.12	-0.04	-0.14	-0.01
Aphanizomenon	0.10	-0.15	0.38*	0.04	0.11	-0.10
Microcystis	-0.04	-0.01	-0.12	-0.16	0.15	0.21
Oscillatoria	-0.20	-0.22	0.74***	0.01	0.14	-0.1
Synedra	-0.10	0.09	0.10	-0.13	0.10	0.25
Aulacoseira	0.30	-0.07	-0.32	-0.13	0.09	0.30
Cyclotella	-0.31	-0.24	0.53**	0.23	-0.06	-0.2
Melosira	-0.03	0.75***	0.08	0.83***	0.17	-0.1
Nitzschia	0.68***	0.25	-0.02	-0.07	0.82***	0.92***

ANF; autotrophic nanoflagellates, HNF; heterotrophic nanoflagellates

by the sequential Bonferroni's procedure with the significance level at $\alpha = 0.05^{18)}$. Differences in each plankton density between bloom and non bloom periods were tested by t-test.

III RESULTS

The lake was stratified at 4~27 m depth, Sta J1 and Sta J2 from April to November. Water quality was strongly altered by rain when a typhoon occurred in August; turbidity and total phosphate(TP) suddenly increased, but gradually returned to normal levels.

During the study period, TP influenced COD (r=0.42, p<0.001) and the concentration of Chl α (r=0.40, p<0.001), and COD was correlated with Chl a (r=0.75, p<0.0001). There were no other significant correlations among the physicochemical variables.

Bacteria and picocyanobacteria averaged 10^{5~6} and 10⁴ cells/mL (Fig. 1). Ciliates were relatively low (< 10² cells/mL) and showed one peak in July. Rotifers gradually decreased from July, while one peak of naupli showed in September at Sta J2. During the summer, most zooplankton showed high abundance until September but thereafter showed a sudden decrease. In this lake, high abundance of planktivore predators dropped after October, however, HNF were sustained in the colder

^{*} Phytoplankton: Microcystis (M. aeruginosa), Aulacoseira (A. granulata), Anabaena (A. flos aquae), Peridinium (P. bipes), Cryptomonas (C. tetrapyrenoidsa), Cyclotella (C. meneghiniana), Melosira (M. varians), Staurastrum (S. floriferum), Oscillatoria (O. agardhii), Gleocystis (G. gigas), Synedra (S. acus), Nitzschia (N. pleae), Pediastrum (P. duplex), Aphnizomenon (A. sp) n=24, * p<0.1, ** p<0.01, *** p<0.001

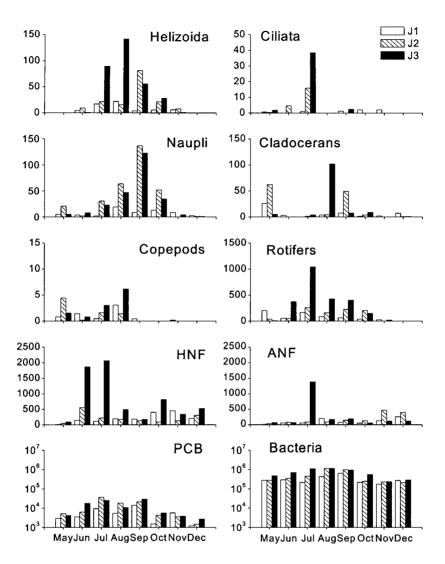


Fig. 1. Abundances of total bacteria, picocyanobacteria, ANF, and HNF in units of cells, and abundances of rotifers, copepods, cladocerans, naupli, and ciliates in individuals at three study sites of Lake Juam from May to December, 2000.

During the season. study period, seven phytoplankton species including diatoms Aulacoseira granulata and Cyclotella meneghniana, cryptomonads Cryptomonas tetrapyrenoidsa contributed significantly (67.3%) to the total phytoplankton biomass (Table 1, Fig. 2), and M. aeruginosa density was below about 7% of the total. A distinct shift in the cyanobacterial community was observed with the passage of time: Anabaena flos aquae in June August; M. aeruginosa in June October, Oscillatoria agardhii in August November; and Aphanizomenon sp. in August November. In spite of small quantity, Microcystis, Oscillatoria and Aphanizomenon were high in abundance until late summer, and Staurastrum, Aphanizomenon and Synedra still persisted even in December. Phytoplankton biomass was usually the highest at Sta J3

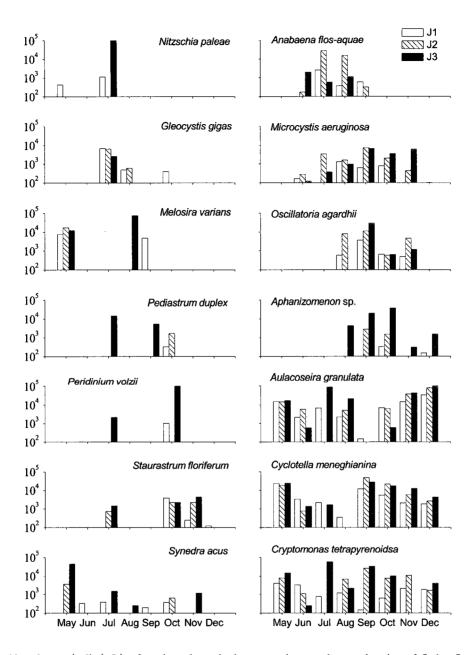


Fig. 2. Abundance (cells/mL) of major phytoplankton species at the study site of Lake Juam from May to December, 2000.

regardless of species composition, while Anabaena flos aquae was most abundant at Sta J2. All predators were closely related to bacteria (p<0.1). Of these, ciliates, rotifers and nauplius showed positive relation with highly picoplankton (p<0.01). For the cyanobacterial

members, such as A. flos aquae, Aphnizomenon sp., O. agardhi and Synedra acus, most of the predators did not show any relationship; only naupli showed a relationship with Aphanizomenon (p<0.1) and Oscillatoria (p<0.001) (Table 1). For all the phytoplankton, no predator showed a specific relation with the abundance of its prey.

IV DISCUSSION

Although the high abundance of the epilithic diatom C. meneghiniana at the dam station was not expected, cyanobacterial components of the bloom remarkably changed in biomass and species composition. The filamentous cyanobacterium Aphanizomenon sp. was newly introduced with Cryptomonas to Lake Juam. In particular, the former was mainly concentrated in the inflowing region of the lake during the study period, and occurred in small number in the center of the lake. It has been commonly known that the seasonal changes of species composition and/or succession in the phytoplankton community are triggered by shift in nutrient ratios such as C:P19) N:P20,21,22) and Si:P22,23) Also, low N:P ratio (~5) and/or falling nitrogen in phosphorus rich lakes usually induces the proliferation of nitrogen fixing cyanobacteria such as Anabaena spp. and Aphanizomenon spp. 24,25) while high nitrate and ammonia promote green algae belonging to Chloroococcales.²⁶⁾

In general, silicate limitation is followed by a decline in diatom biomass.271 Although the high abundance of Cyclotella at the dam site cannot be explained with the available data, changes in cyanobacterial population observed in the present study are contrary to the previous studies. The N:P ratio of Lake Juam has been increasing year after year, while the relative abundance of cvanobacteria was decreased in phytoplankton community. The silicate level, increased by the heavy rains during the study period, remained relatively high throughout the lake. However, lack of data for the past does not permit us to relate increase in abundance of six diatoms, regardless of sampling sites, to rise in silicate concentration. Therefore, the species succession

and/or changes of phytoplankton community in Lake Juam may be attributed to the nutrient gradient influenced by summer heavy rain, and there is the change of relative abundance of each species, rather than species composition in the phytoplankton community.

It is not surprising that most plankton communities flourished in the inflowing region of the lake, due to the high and/or continuous input of organic loadings from its watershed and tributaries. In the last two decades, the changes of dominant phytoplankton species are not rare in a majority of Korean lakes. 28,29) abundance of predators such as rotifers. cladocera, ciliates, copepods and HNF was particularly evident in the lake. However, due to lack of adequate data for the present as well as the past it cannot be fully understood whether it was induced by the flourishing summer as, Agranulata, phytoplankton, such meneghiniana and C. tetrapyrenoidsa. However, according to several indirect evidence, for instance, positive and negative correlations among the prey and predators, it may be the dominant pico and suggested that nanoplankton are mainly fed on by copepod naupli, while rotifer zooplankton and copepod naupli fed on phytoplankton.

V ABSTRACT

The rainy season and a cyanobacterial bloom, abundance and distribution of bacteria, picocyanobacteria(PC), autotrophic nanoflagellates (ANF), heterotrophic nanoflagellates (HNF), ciliates, and zooplankton were examined physicochemical variables relation in to mesotrophic Lake Juam, Korea. Three species dominated the phytoplankton community even in the cold season; Cyclotella meneghinana and Aulacoseira granulata (Bacillariophyceae), and

tetrapyrenoidsa (Cryptophyceae) Cryptomonas contributed over 67% to the total phytoplankton biomass. The cyanobacterial component of the algal bloom changed remarkably in biomass and species composition. The phytoplankton abundance (Chl a) and water quality (COD and TP), were markedly influenced by heavy rain abundance summer. The of the during Cryptomonas was closely related to that of HNF. nauplii. rotifers and ciliates. The abundance of rotifers was highly correlated to that of bacteria, PC and ANF.

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VII. LITERATURE CITED

- Wetzel RG (1963) A comparative study of the primary productivity of higher aquatic plants, periphyton and phytoplankton in a saline lake.
 Ph. D. Diss. Univ. California. Davis
- Harper D (1992) Eutrophication of freshwaters principles, problems and restoration. Chapman and Hall. 327 p.
- Carmichael WW (1981) Freshwater blue green algae (Cyanobacteria) toxins a review.
 In Carmichael WW (ed.) The Water Environment. Algal toxins and Health. Plenum Press, New York, pp. 1~13
- Carmichael WW (1988) Toxins of freshwater algae. In Tu AT (Ed.) Handbook of Natural Toxins, Vol. 3, Marine Toxins and Venoms. Marcel Dekker, New York, pp. 121 47
- Watanabe MM, Kaya K, Takamura N (1992)
 Fate of the toxic cyclic heptapeptides, the microcystins, from blooms of *Microcystis* (Cyanobacteria) in a hypertrophic lake. J

- Phycol 28:761 767
- Falconer IR, Beresford A, Runnegar MTC (1983) Evidence of liver damage by toxin from a bloom of the blue green alga, Microcystis aeruginosa. Med J Aust. 1:511~5116
- Choi MK, Kim BH, Chung YT, Wui IS (1994)
 Occurrence and dynamics of phytoplankton in Lake Chuam. Kor J Limnol 27:79~91
- Kim BH (1996) An Ecological Study on the Phytoplankton in Lake Chuam. Ph. D. Thesis, Chonnam Univ. 255 p.
- 9. KOWACO (1988~1999) Annual Report on Water Quality Monitoring in Lake Juam.
- KOWACO (1999~2000) Annual Report on Water Quality Monitoring in Lake Juam.
- APHA (1995) Standard methods for the examination of water and wastewater. 19th Ed. APHA, AWWA, WPCF, Washington, 1134 p.
- Otsuki A, Sekiguchi K (1983) Automated determination of ammonia in natural freshwaters using salicylate hexacyanoferrate dichloroisocyanurate system. *Analytical Letters*, 16:979~985
- 13. Hoff KA (1993) Total and specific bacterial counts by simultaneous staining with DAPI and fluorochrome labeld antibodies. Handbook of methods in aquatic microbial ecology (eds. Kemp PF, Sherr BF, Sherr EB, Cole JJ), pp. 149 154. Lewis Publishers.
- Porter KG, Feig YS (1980) The use of DAPI for identifying and counting aquatic microflora. Limnol Oceanogr 25:943~948.
- 15. Sherr EB, Caron DA, Sherr BF (1993) Staining of heterotrophic protists for visualization via epifluorescence microscopy. *In*: Handbook of methods in aquatic microbial ecology (eds. Kemp PF, Sherr BF, Sherr EB, Cole JJ), Lewis Publishers, pp. 21 3~228.

- Utermöhl H (1958) Zur Vervollkommnung der quantitativen Phytoplankton Methodik.
 Int Verein Theor Ang Limnol Mitt 9: 1 38
- Montagnes DJS, Berges JA, Harrison PJ, Taylor FJR (1994) Estimating carbon, nitrogen, protein, and chlorophyll a from volume in marine phytoplankton. *Limnol Oceanogr* 39:1044~1060
- 18. Holm S (1979) A simple sequentially rejective multiple test procedure. Scan J Stat 6:65~70
- Harris GP (1986) Phytoplankton ecology, structure, function and fluctuation. Chapman and Hall. 384 p.
- 20. Takamura N, Ishikawa Y, Mikami H, Mikami H, Fujita Y, Higuchi S, Murase H, Yamanaka S, Nanjyo Y, Igari T, Fukushima T(1996) Abundance of bacteria, picophytoplankton, nanoflagellates and ciliates in relation to chlorophyll a and nutrient concentrations in 34 Japanese waters. *Jpn J Limnol* 57:245~259.
- 21. Takamura N, Mikami H, Mizutani H, Nagasaki K (1999) Did a drastic changes in fish species from kokanee to pond smelt decrease the Secchi disc transparency in the oligotrophic Lake Towada, Japan? Arch Hydrobiol 144:283 304.
- 22. Han MS, Auh YY, Bang SW, Ki SS (2000) Ecological studies on Paltang River reservoir systems in Korea. 4. Dynamics on inorganic nutrients. POM and phytoplankton succession in the lower stream Kyungan. Int Sym. Ecotechnology in environmental protection and freshwater lake management. Paechai, Korea. pp. 74~88.
- 23. Tilman D, Kilham SS, Kilham (1982) Phytoplankton community ecology: the role of limiting nutrients. Annu Rev Ecol Syst 13:340 372
- 24. Schindler JE (1971) Carbon, nitrogen and

- phosphorus and the eutrophication of freshwater lakes. *J Phycol* 7:321~329.
- 25. Reynolds CS (1980) Phytoplankton assemblages and their periodicity in stratifying lake systems. *Holarctic Ecol* 3:141~159.
- 26. Reynolds CS (1978) The plankton of the northen west Midland meres. Occasional Papers of the Caradoc and Severn Valley Field Club. No. 236+xxiii pp.
- Reynolds CD (1985) The ecology of freshwater phytoplankton. Cambridge Univ Press. 384 p.
- 28. Cho KS, Kim BC, Heo WM, Cho SJ (1989)
 The succession of phytoplankton in Lake
 Soyang. Kor J Limnol 22:179~189.
- Kim BC, Cho KS, Shim JH (1985) Temporal and spatial variation of chlorophyll a concentration in Lake Soyang. Kor Wat Poll Res Contr 1:18~23
- 30. Choi CK, Hwang YJ (1991) On the fish communities of the Posong river. *Kor J Limnol* 24:199~206
- Hutchinson GE (1957) A treatise on Limnology. Geography, physics and chemistry, Vol I. John Wiley & Sons, Inc. New York. 1015 p.
- KOWACO (1990) Eutrophication protection research using an ecosystem model in Daechung Reservoir. KOWACO 90 54, 122p.
- KOWACO (1994) Report of the Pollutant Loading of Water in the Multi purpose Dam Reservoirs. 100p.
- 34. KOWACO (1996) Studies on the distribution and effect of exotic fishes in Dam reservoir. 264p.
- 35. Lee OM (1994) The annual dynamics of standing crops and distribution of phytoplankton in Juam lake in 1992. Kor J Limnol 27:327~ 337
- 36. Lee OM, Song HY (1995) The annual

- dynamics of standing crops and distribution of phytoplankton in Juam lake in 1993. Kor I Limnol $28:427 \sim 436$
- McLeod JA, Bondar GF (1952) A case of suspected algal poisoning in Manitoba. Can J Pub Health 43:347~350
- 38. MOC, Ministry of Construction, Korea (1985)
 Environmental Impact Assessment on the
 Construction of Juam Multipurpose Dam.
 414 p.
- 39. Nah CS, Shin SS (1992) A study on the fish fauna after the construction of Chuam Dam. *Korean J Ichthyol* 4:55~62
- OECD (1982) Organization for Economic Cooperation and Development. Eutrophication of Waters. OECD, Paris.
- 41. Rocha O, Duncan A (1985) The relationship between cell carbon and cell volume in freshwater algal species used in zooplankton

- studies. J Plankton Res 7:279~294
- Sandgren CD, Smol JP, Kristiansen J (1995)
 Chrysophyte algae. Ecology, phylogeny and development. Cambridge University Press. 399 p.
- 43. Takamura N, Hanazato T, Iwakuma T, Nojiri Y, Otsuki A, Aizaki M (1998) Long term Monitoring of Nutrients, Plankton and Benthos in Lake Kasumigaura. *In*: Iwakuma T (Ed.) Long term ecological research in the east Asia pacific region: biodiversity and conservation of terrestrial and freshwater ecosystems. Center for Global Environmental Research, Tsukuba, Japan, pp. 155~165
- 44. Wetzel RG, Likens GE (1991) Limnological Analyses. 2nd ed. Springer Verlag, New York. NY