

Genetic Study of the Class Dinophyceae Including Red Tide Microalgae Based on a Partial Sequence of SSU Region: Molecular Position of Korean Isolates of *Cochlodinium polykrikoides* Margalef and *Gyrodinium aureolum* Hulburt

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The nucleotide sequence for a nuclear-encoded small subunit rDNA (SSU rDNA) was determined for 43 species of the class Dinophyceae, including harmful algae *Cochlodinium polykrikoides* and *Gyrodinium aureolum*. These sequences and data analyses were performed by parsimony, distances and maximum likelihood methods in PHYLIP (Phylogenetic Inference Package) version 3.573c. The species *Noctiluca scintillans*, *Gonyaulax spinifera* and *Cryptocodinium cohnii* occupied a basal position within the Dinophyceae in our analyses. The genera *Alexandrium* and *Symbiodinium* were monophyletic (supported by a bootstrap value of >70%), whereas the genera *Gymnodinium* and *Gyrodinium* formed polyphyletic nodes, for which bootstrap support was strong (>70%) in the neighbor-joining and maximum likelihood methods except for the PHYLIP parsimony analysis (=59%). The sequence divergence between *G. aureolum* and *G. dorsum*/*G. galathenum* was the largest at 7.4% (45 bp), whereas *G. aureolum* and *G. mikimotoi* showed an extremely low value of genetic divergence of 0.9% (5 bp). The genetic divergence between *C. polykrikoides* and *G. aureolum* was a low value of 5.2% (31 bp). In the phylogenetic analysis, the placement of *G. aureolum* and *C. polykrikoides* was closer to the genus *Gymnodinium* than to the genus *Gyrodinium*, which was supported by a moderate bootstrap value.

Key words – *Cochlodinium polykrikoides*, Dinophyceae, Genetic study, *Gyrodinium aureolum*, morphology, phylogeny, red tide, SSU

Harmful algal blooms (HABs) are increasing in frequency or severity in many coastal environments and worldwide. The blooms of unicellular marine algae can cause mass mortality in a variety of marine organisms and can cause illness and even death in humans who consume contaminated seafood. At present, about 57 HAB species are capable of producing potent toxins and causing serious problems globally [7]. Most HAB species are of the class Dinophyceae, which can be divided into five types (Gymnodinioids, Gonyaulacoids, Dinophysoids, Procentroids, and Peridinoids) according to the morphological features described by Taylor [29]. Furthermore, the identification of dinoflagellate species is sometimes painstaking and the discrimination of each is sometimes confusing because of closely related morphology in a few species and of possible variability in morphological features depending on environmental conditions [1,22]. For example, the genera *Alexandrium*, *Gymnodinium* and *Prorocentrum* have ongoing considerable studies and discussions on the morphological characteristics, which are subjects of interest to taxonomists.

Recent advances in DNA amplification and sequencing provide powerful tool to examine phylogenetic relationships of both closely and distantly related organisms. Nuclear ribosomal RNA genes are attractive candidates for molecular studies at taxonomic levels. The sequence analysis of the small subunit (SSU) is one method that can be used to classify populations and to identify specific genetic markers of *Alexandrium* [9,24-26]. In addition, the sequence data from SSU region provide information on heterogeneity within the genus *Prorocentrum* [12]. The dinoflagellate *Cochlodinium polykrikoides* occurred for the first time in 1982 in Korean coastal waters, and was associated with massive fish kills [18]. Impacts of *C. polykrikoides* appear to have increased in frequency, intensity and geographic distribution. Previously, we have studied the molecular structures and systematics of *C. polykrikoides* using only internal transcribed spacers (ITS) regions [3-6]. However, little has been investigated of the sequence analysis of *C. polykrikoides* targeted to the SSU region.

Recently, Dr. K. J. Cho, Inje University, Korea, isolated *Gyrodinium aureolum* from Chinhae Bay, Korea, which is a species closely related to *Gymnodinium mikimotoi* [2]. Despite the fact that the morphological characteristics of *G. aureolum*

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have been much studied and debated, some genetic characteristics remain to be determined and understood. As mentioned above, the sequence analysis of SSU has been proved to be useful for the study of intra and interspecific genetic variation in Dinophyceae. This study sequenced the SSU to investigate the genetic divergence of *C. polykrikoides* and *G. aureolum* from related species, and also obtained other sequences through a GenBank search to form a combined data set for the analysis of genetic divergence and phylogenetic relationships.

Materials and Methods

Cultures

Cochlodinium polykrikoides was isolated from the coast of Tongyong, Korea, in 1997 and has been grown in f/2-Si medium [13] containing an antibiotic mixture [15]. *Gyrodinium aureolum* was provided by Dr. K. J. Cho, Inje University, Kimhae, Korea. These species have maintained at 20°C under a photon flux from cool white fluorescent tubes of 50 $\mu\text{mol m}^{-2} \text{s}^{-1}$ in a 14: 10 h L:D cycle.

Isolation of genomic DNA

Cultures in exponential phase were harvested by centrifugation. Pelleted cells were frozen immediately and stored at -20°C until required. Cells were thawed in 500 μL of extraction buffer (100 mM Tris-HCl, pH 8.0; 40 mM EDTA) and 150 μL of 10% sodium dodecyl sulphate (SDS), and incubated at 55°C for 30 min. The supernatant was extracted twice with phenol : chloroform : isoamylalcohol (25 : 24 : 1, v/v/v) and RNase (1 mg mL^{-1}) was added. The algal cells were pelleted and the supernatant mixed with 40 μL of 3 M sodium acetate (pH 4.0) and incubated on ice for 1 h. The DNA was precipitated from the tube at room temperature for 10 min by adding 2.5 volumes of 100% ice-cold ethanol. The DNA pellet was washed with 2 volumes of 70% ethanol and was then dried by desiccator. The DNA was dissolved in 20 μL TE buffer (10 mM Tris, 0.1 mM EDTA, pH 7.5) and was kept at -20°C. Gel electrophoresis was performed at voltage of 100 V for 20 min at room temperature. After electrophoresis, the 1.5% agarose gel was visualized by embedding in ethidium bromide (2 $\mu\text{L mL}^{-1}$) and irradiating by ultraviolet light (312 nm).

DNA amplification and sequencing

The partial region of nuclear SSU rDNA was amplified from genomic DNA using the polymerase chain reaction

(PCR) with the two primers [17], NS1 (5'-GTAGTCATA-TGCTTGCTC-3') and NS2 (5'-GGCTGCTGGCACCAGAC-TTGC-3'). PCR was performed in a 20 μL reaction (Korea Biotech. Inc., Ltd., Taejon, Korea) mixture using Perkin-Elmer 2400 Thermocycler under the following cycle conditions; pre-denaturing at 95°C for 30 sec, followed by 35 cycles of denaturing at 95°C for 1 min, primer annealing at 50°C for 1 min, and 5 min chain extension at 72°C for 5 min. The PCR product after the amplification was subjected to preparative electrophoresis in a 1.5% agarose gel in TBE buffer. All PCR products yielded only a single visible band. Target fragments were cut off from the gel and withdrawn using a QIAGEN gel elution kit (Qiagen, Wartworth CA) according to the instructions by the manufacture. Purified rDNA region was suspended in distilled water and stored at -20°C until use. Amplified PCR products were sequenced directly on the Perkin-Elmer Applied Biosystems (ABI) 377A DNA sequencer using a ABI PRISM Big Dye™ Terminator Ready Reaction Kit (Perkin Elmer) following the manufacture's protocol.

Sequence analysis

Sequence data were aligned using the multiple alignment program Clustal W [30] and were determined by parsimony, distances and maximum likelihood (ML) methods (<http://bioweb.pasteur.fr/intro-uk.html#phylo>). To understand the possible genetic relationships, PHYLIP (Phylogenetic Inference Package) version 3.573c [10] was used in this study. PHYLIP was used dinoflagellates such as *Symbiodinium corcolorum* L13717, *S. pilosum* X62650, *S. meandrinae* L13718, *S. microadriatic* M88521, *Gymnodinium beii* U41087, *G. simplex* U41086, *Symbiodinium* sp. M88509, *Gyrodinium impudicum* AF022197, *Lepidodinium viride* AF022199, *G. catenatum* AF022193, *Gymnodinium* sp. AF022196, *G. fuscum* AF022194, *Heterocapsa triquetra* AF022198, *G. sanguineum* U41085, *Scrippsiella nutricula* U52357, *Pentapharsodinium tyrrhenicum* AF022201, *Prorocentrum micans* M14649, *G. breve* AF172714, *G. mikimotoi* AF022195, *Gloeodinium viscum* L13716, *Peridinium* sp. AF022202, *Gyrodinium dorsum* AF274261, *G. uncatenum* AF274263, *Ceratococys horrida* AF022154, *Gonyaulax cochlea* AF274258, *Pyrodinium bahamense* AF274275, *Gyrodinium galatheanum* AF274262, *Scrippsiella trochoidea* AF274277, *Prorocentrum minimum* Y16238, *P. lima* Y16235, *Noctiluca scintillans* AF022200, *Alexandrium fundyense* U09048, *A. tamarense* X54946, *A. margaelefii* U27498, *A. minutum* U27499, *A. ostenfeldii* U27500, *Ceratium tenue* AF022192, *Pyrocystis*

noctiluca AF022156, *Amphidinium belauense* L13719, *Gonyaulax spinifera* AF022155 and *Cryptothecodinium cohnii* M64245. This search for parsimony analysis was repeated several times from different random starting points using the stepwise addition option to make certain the most parsimonious tree was found. For distance analysis, subprogram DNADIST in PHYLIP was used to obtain a matrix of Kimura's two-parameter distance [19]. Distance matrix was analyzed by subprogram NEIGHBOR in PHYLIP with algorithms based on Saitou and Nei's neighbor-joining (NJ) method [21]. All nucleotide substitutions were equally weighted and unordered alignment gaps were treated as missing information. As an indication of confidence in the branching order, a bootstrap analysis (100 replications) was completed for both distance and parsimony methods. Reliability of the tree was constructed using subprogram CONSENSE in PHYLIP after pairwise sequence distances were estimated by Kimura's two-parameter method, which attempts to correct observed dissimilarities for multiple substitutions in sequences evolving with a transition bias.

Results

The SSU rRNA gene sequence was determined for 41 species except for Korean *C. polykrikoides* and *G. aureolum*, which were retrieved from the GenBank database (Fig. 1). A Kimura two-parameter NJ analysis and a substitution rate calibration analysis each produced phylogenetic trees, and the substitution rate calibration tree was presented in Fig. 2. Three groups were inferred. Group I contained three different taxa (*Noctiluca*, *Gymnodinium*, and *Symbiodinium*). *Noctiluca scintillans* played the role of a sister to Group I. Several species of *Symbiodinium* had a monophyletic relationship. Group II consisted of various genera and was trisected with Group II-1 (*Prorocentrum*, *Gyrodinium*, *Scripsiella*, *Pyrodinium*, *Gonyaulax*, and *Ceratocorys*), Group II-2 (*Peridinium*, *Gloeodinium*, and *Gymnodinium*), and Group II-3 (*Heterocapsa*, *Cochlodinium*, *Lepidodinium*, and *Pentaparsodinium*). Group II was fragmented with *P. minimum*/*P. lima* as a sister to Group II-1 (bootstrap 74%), *P. micans* as a sister to Group II-2 (bootstrap 92%) and *P. tyrrhemicum* as a sister to Group II-3 (bootstrap 58%). In particular, *S. nutricula* and *G. sanguineum* had an independent subgroup, which was supported by a weak bootstrap value of 51%. The phylogenetic placement of Korean *G. aureolum* was closer to *G. mikimotoi* and *G. breve* than that of *C. polykrikoides*, which joined *Gymnodinium* sp. The position of *C. polykrikoides* located the branch

with *G. impudicum*, and *G. catenatum*. Group III contained six taxa (*Ceratim*, *Pyrocystis*, *Alexandrium*, *Amphidinium*, *Gonyaulax*, and *Cryptothecodinium*). *Alexandrium* formed the most basal clade, a sister to the remaining taxa and had a monophyletic relationship. However, Group I and II were supported by a moderate bootstrap value of 72%, while Group III was supported by a very weak bootstrap value of 51%. The PHYLIP NJ and ML analyses produced a tree that was similar to the substitution rate calibration tree, and all groups were recovered (Figs. 3, 4). The essential aspects of topology were retained. For example, the tree members of each group according to parsimony and NJ analyses formed their own groups, exactly distinctive monophyletic groups. The unrooted phylogenetic tree obtained from ML PHYLIP was slightly changed which tree members were inserted or not. The essential aspects of topology were that *G. aureolum* formed an extremely strong monophyletic group with *G. mikimotoi* and *G. breve* (bootstrap >90%) and *C. polykrikoides* appeared to be affected according to the topology of the trees. The DNA similarity between taxa was shown in Table 1. Among the species used in this study, the DNA similarity was high, over 70%. *Cochlodinium polykrikoides* had a high similarity (88-90%) to the genus *Gymnodinium* including *G. aureolum* and *G. impudicum*. However, *G. aureolum* was higher value of similarity between taxa than did *C. polykrikoides*. *Gonyaulax cochllea* had the lowest value of similarity among all tested species. To further illustrate the genetic relationships between the genera *Gymnodinium* and *Gyrodinium* including *C. polykrikoides*, we constructed a matrix of Kimura genetic distances between pairs of taxa, as shown in Table 2. The sequence divergence in pairwise comparisons ranged from 0.0% to 9.5% (0 to 58 bp) and the largest sequence divergence occurred in a comparison of *G. fuscum* and *C. polykrikoides*. The sequence divergence between *G. aureolum* and *G. dorsum*/*G. galathenum* was the largest at 7.4% (45 bp), while *G. aureolum* and *G. mikimotoi* showed an extremely low value of genetic divergence of 0.9% (5 bp). The genetic divergence between *C. polykrikoides* and *G. aureolum* was a low value of 5.2% (31 bp), compared with other taxa. The genetic relationships between the genera *Gymnodinium* and *Gyrodinium* including *C. polykrikoides* sequences were shown in PHYLIP and were in Fig. 5. In the pairwise analysis, *G. aureolum* and *C. polykrikoides* consisted of a member of Group I and of Group II, respectively; the two groups were supported by a moderate bootstrap value of 84% (Fig. 5). Group III was supported by a weak bootstrap value of 62%.

1 -AACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
21 -----GGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
7 -AACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
9 CAACCTGGT-GATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
10 -AACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
12 -AACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
13 -AACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
11 -AACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
19 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
33 -----GGTTGATCCTGCCAGTAGTCATATGCTTGTCTCAAAGATTAAGCCATGCATGT
14 --ACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
18 --ACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
32 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
22 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
29 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
27 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
26 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
24 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
28 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
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34 -AACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
4 -AACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
6 -AACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
8 --ACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
5 -AACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
3 -----GGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
17 --ACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
16 --ACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
2 -----GGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
30 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
20 -----AAGATTAAGCC-TGCATGT
43 -----AGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
39 -----AGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
40 -----AGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
42 -----AGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
38 -----AGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
41 -----AGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
36 -----TCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
37 -----TCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
31 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT
15 -CACCTGGTTGATCCTGCCAGTAGTCATAT-GCTTGTCTCAAAGATTAAGCCATGCATGT

1 CTGAGTCTATGGCT-TTACACGGTTAAGCTGCGAATGGCTCATT-ATAACAGTAAATGATC
21 CTCAGAATAAGCTT-TT-CATGGCAAGCTGCGAATGGCTCATT-AAACAGTTATAGTT
7 CTCAGCATAAGCGT-TT-CACAGCAAGCTGCGAATGGCTCATT-AAACAGTTATAGTT
9 CTCAGTATAAGCTT-TCATTTTGTGAAACTGCGAATGGCTCATT-AAACAGTTATAGTT
10 CTTAGTATAAGCTT-CAATTTTGTGAAACTGCGAATGGCTCATT-AAACAGTTACAAAG
12 CTCAGTATAAGCAT-TCATTTGGCGAAACTGCGAATGGCTCATT-AAACAGTTATAATG
13 CTCAGTATAAGCAT-TCATTTGGCGAAACTGCGAATGGCTCATT-AAACAGTTATAATG
11 CTCAGTATAAGCAT-TTATTTGGCGAAACTGCGAATGGCTCATT-AAACAGTTATAATG
19 CTGAGTTAAGCTA-ATTTTGGTGAAGCTGCGAATGGCTCATT-ATATCAGTTATAATA
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18 CTCAGTATAAGCTT-CTACACGGCAAACTGCGAATGGCTCATT-AAACAGTTATAGTT
32 CTCAGTATAAGCTT-CTATACGGCAAACTGCGAATGGCTCATT-AAACAGTTATAGTT
22 CTCAGTATAAGCTT-CTACACGGCGAAACTGCGAATGGCTCATT-AAACAGTTATCGTT
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27 CTCAGTATAAGCTT-TCACACGGCGAAACTGCGAATGGCTCATT-AAACAGTTATCGTT
26 CTCAGTATAAGCTT-CTACACGGCGAAACTGCGAATGGCTCATT-AAACAGTTATCGTT
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35 CTCAGTATAAGCTTCTATACGGCGAAACTGCGAATGGCTCATT-AAACAGTTATAGTT
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6 CTCAGCATAAGCTT-CTACACGGCGAAACTGCGAATGGCTCATT-AAACAGTTATAAAT
8 CTCAGTATAAGCTT-CTACACGGCGAAACTGCGAATGGCTCATT-AAACAGTTATAAAT

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31 CTCAGTACAAACTT-GTATATGGTGAAGCTGCGAATGGCTCATT-AAATCAGTTACAGCT
15 CTCAGTATAAGCTT-CTACACGGCGAAACTGCGAATGGCTCATT-AAAAAGTTATCGTT
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9 TATTTGACAATCAACT--CTATATGG--ATATCTGTGGAAATTTCTATAGTTAATACATGCA
10 CATTTGGCGATCAAIT--CTAAATGG--ATATCTGTGGAAATTTCTATAGTTAATACATGCA
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18 CATTTGATGGT-CTCTCACTACATGG-ATAACTGTGGTAATTTCTAGAGCTAATACATGCG
14 TATTTGATGGT-CAATCCTTACATGG-ATAACTGTGGTAATTTCTAGAGCTAATACATGCG
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43 GATCCTTGATGGTTGTTTTCTGAAATGGATATCTTGGTAATTTCTAGAGCTAATACATGCT
39 TATTTGGTGGT-CATTCTTACATGG-ATAACTGTGGTAATTTCTATAGCTAATACATGCT
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37 TATTTGATGGTCAITCTTT-ACATGG-ATAACTGTGGTAATTTCTAGAGCTAATACATGCG
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15 TATTTGGTGGT-CATTCACTACATGG-ATAACCGTGGTAATTTCTAGAGCTAATACATGCG
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1 TCAACACCAACTTATTGGA---G-GGTGGTCTTACCAA-ATTCAGAACCAAT-CCAAG
21 TGCATACCCGACTTTCCGGAA---G-GGTGGTCTTATCAG-CTCCAGAACCATT-CCTGG
7 GACATACCTGACTTTTGGGGA---ATGGTGTCTATCATGAG-CCACAGAATCGGC-GCAGG
9 TTGAAACCGGATCTTTGGGGA---AAGGTTGTGGTCTGTTAG-TTACAGAACCAAT-TCAGG
10 CTAAACCTTATCTTTGGGGA---AAGGTTGTGGTCTGTTAG-TTACAGAACCAAT-TCAGG
12 TCTAACCTGACTTTTGG-A---AGGTTGTGGTCTTATAG-TTACAGAACCAAC-CCAAG
13 TCCAACCTGACTATTGGGA---AGGTTGTGGTCTTATAG-TTACAGAACCAAC-CCAAG

11 ACCAAGCCKGACTTGTGGG---AGGGCTGAAGTCATTAG-TTACAGAACCAC-CCAAG
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33 ACCAAACCTGACTTGTGGGAAGATAAAGTTGTGGTCTTAGCTTTTGAATCAAC-CCAAG
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* * *

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** *

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* * * * *

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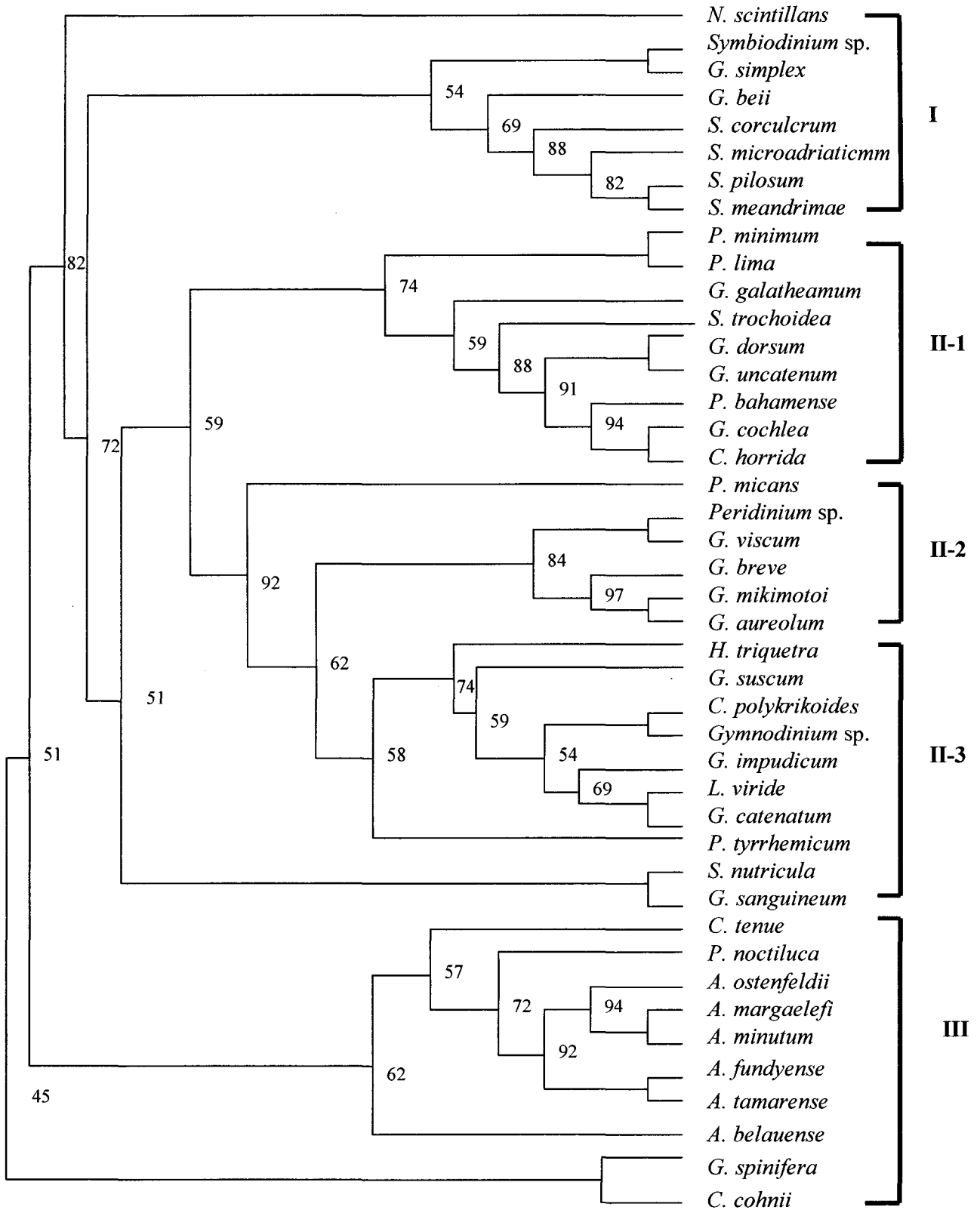


Fig. 2. PHYLIP analysis of 41 species obtained from GenBank database including *C. polykrikoides* and *G. aureolum*. The tree was obtained using subprogram DNAPARS in PHYLIP. The topology represents the consensus tree from an heuristic search yielding two equally most parsimonious tree. Bootstrap values (100 replications) are given above the internal nodes. This is an unrooted tree.

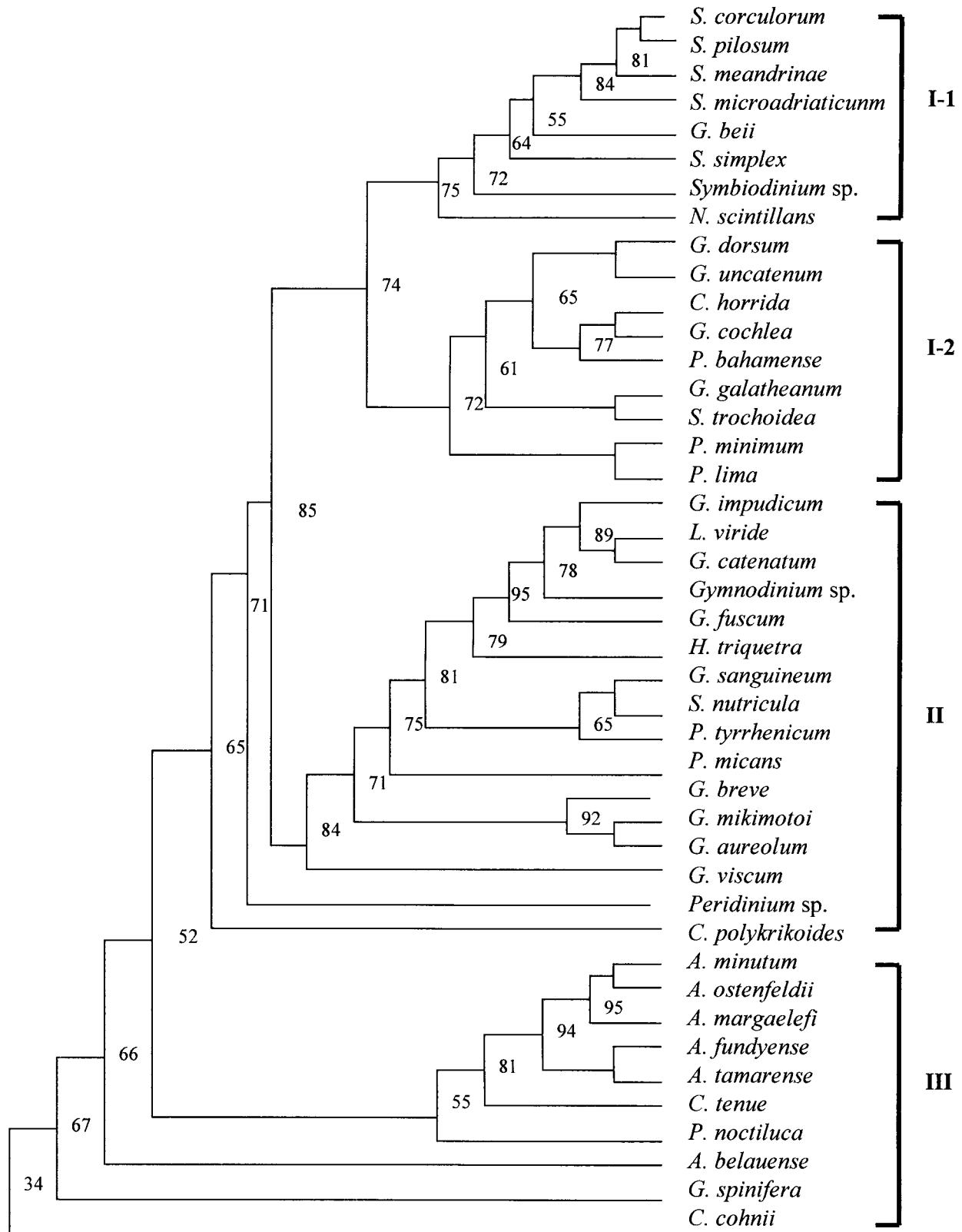


Fig. 3. PHYLIP analysis of 41 species obtained from GenBank database including *C. polykrikoides* and *G. aureolum*. The tree was obtained using subprogram NEIGHBOR in PHYLIP with the option of Kimura's two-parameter method. Bootstrap values (100 replications) are given above the internal nodes. This is an unrooted tree.

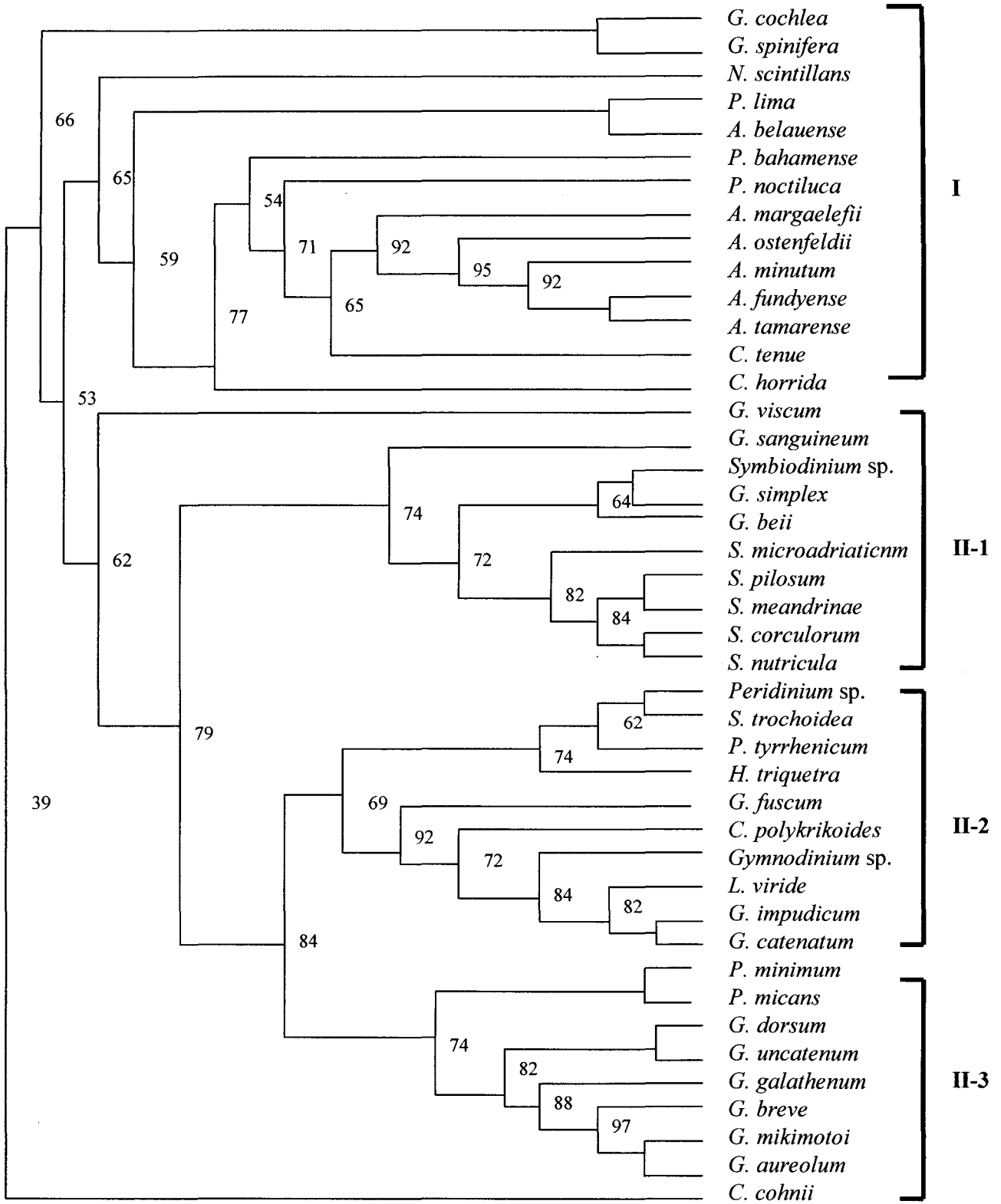


Fig. 4. PHYLIP analysis of 41 species obtained from GenBank database including *C. polykrikoides* and *G. aureolum*. The tree was obtained using subprogram DNAMLK in PHYLIP with the option of Kimura's two-parameter method. Bootstrap values (100 replications) are given above the internal nodes. This is an unrooted tree.

Table 1. Percentage of similarity of nucleotide sequences of Dinophyceae

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43											
1	-																																																					
2	84	-																																																				
3	85	94	-																																																			
4	84	88	90	-																																																		
5	86	93	98	91	-																																																	
6	85	93	98	91	81	-																																																
7	79	81	81	81	82	81	-																																															
8	85	93	98	91	99	99	81	-																																														
9	81	83	84	84	84	84	80	84	-																																													
10	81	83	84	83	85	84	80	85	84	-																																												
11	82	86	86	85	87	86	80	86	88	82	-																																											
12	88	87	86	88	88	87	82	87	82	87	92	-																																										
13	82	88	87	86	88	87	82	87	82	87	92	83	93	-																																								
14	85	92	92	92	93	92	83	93	84	85	85	87	88	89	-																																							
15	81	84	85	85	86	86	80	86	80	81	82	83	84	86	86	-																																						
16	86	94	95	91	95	85	85	87	89	88	93	88	93	94	85	96	-																																					
17	86	94	96	92	96	96	80	96	85	85	86	88	88	94	86	93	86	-																																				
18	86	92	93	93	93	93	83	93	83	83	86	88	89	90	96	86	93	86	84	-																																		
19	82	84	84	83	85	85	79	85	84	83	85	84	83	85	84	83	86	85	86	85	84	-																																
20	81	81	79	81	80	72	80	77	76	78	80	79	82	76	81	81	81	81	81	81	75	-																																
21	84	86	85	85	85	80	85	81	81	83	85	84	85	81	81	86	85	86	86	82	81	81	-																															
22	86	91	92	93	93	92	83	93	85	86	87	89	94	88	92	93	96	84	80	84	80	84	85	-																														
23	85	90	92	92	92	91	81	92	85	85	87	89	88	95	90	92	93	95	84	80	84	80	84	85	95	-																												
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25	86	91	91	92	92	92	82	92	86	86	88	89	89	95	88	93	95	84	80	84	80	84	80	85	96	99	95	95	-																									
26	86	90	91	91	91	91	82	91	84	84	86	90	87	93	88	91	92	94	84	81	84	81	84	84	97	94	94	95	94	-																								
27	85	90	91	91	91	91	82	91	84	84	86	90	87	93	88	91	92	94	84	81	84	81	84	84	97	94	94	96	96	96	-																							
28	86	91	92	93	93	93	82	92	85	85	88	89	95	85	88	94	94	94	84	77	85	99	95	97	95	97	97	97	97	96	-																							
29	86	91	92	93	93	93	83	93	85	86	87	83	88	94	88	92	93	96	84	77	85	99	95	97	95	97	97	97	96	88	-																							
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34	85	90	91	92	92	92	81	82	91	85	85	87	89	88	95	87	92	93	95	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80			
35	84	91	92	92	92	92	81	82	91	85	85	87	89	88	94	89	94	94	94	85	79	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80	84	80					
36	81	88	89	87	88	88	81	88	83	83	83	83	87	84																																								

Table 2. Pairwise distances between fourteen species including *C. polykrikoides* of the genus *Gymnodinium* and *Gyrodinium* from the aligned SSU rDNA gene sequences on the basis of Kimura two-parameter estimates

	1	2	3	4	5	6	7	8	9	10	11	12	13	14
1	-	19	50	50	35	42	42	49	46	45	41	41	43	49
2	0.0319	-	42	42	31	34	36	45	38	39	39	39	38	52
3	0.0828	0.0695	-	0	43	40	42	53	48	42	46	42	45	50
4	0.0826	0.0694	0.0000	-	43	40	42	53	48	42	46	42	45	50
5	0.0583	0.0510	0.0711	0.0710	-	24	26	36	29	32	20	19	20	53
6	0.0693	0.0563	0.0659	0.0658	0.0403	-	30	40	34	31	25	24	26	54
7	0.0695	0.0601	0.0698	0.0697	0.0440	0.0494	-	17	12	16	27	24	26	43
8	0.0805	0.0745	0.0870	0.0869	0.0604	0.0656	0.0285	-	20	26	36	34	36	42
9	0.0770	0.0638	0.0793	0.0792	0.0477	0.0566	0.0200	0.0337	-	18	31	29	12	42
10	0.0751	0.0655	0.0698	0.0697	0.0532	0.0514	0.0268	0.0441	0.0302	-	32	29	30	58
11	0.0681	0.0640	0.0759	0.0757	0.0334	0.0426	0.0443	0.0603	0.0514	0.0532	-	6	6	39
12	0.0681	0.0640	0.0702	0.0701	0.0316	0.0409	0.0407	0.0566	0.0477	0.0476	0.0100	-	5	36
13	0.0721	0.0639	0.0745	0.0743	0.0335	0.0432	0.0430	0.0598	0.0504	0.0504	0.0105	0.0094	-	31
14	0.0812	0.0864	0.0825	0.0823	0.0882	0.0897	0.0721	0.0703	0.0695	0.0956	0.0654	0.0598	0.0521	-

Note: 1. *G. simplex* U41086; 2. *G. beii* U41087; 3. *G. uncatenum* AF274263; 4. *G. dorsum* AF274261; 5. *G. galathenum* AF274262; 6. *G. sanguineum* U41085; 7. *G. catenatum* AF022193; 8. *G. impudicum* AF022197; 9. *Gymnodinium* sp. AF022196; 10. *G. fuscum* AF022194; 11. *G. breve* AF172714; 12. *G. mikimotoi* AF022195; 13. *G. aureolum*; 14. *C. polykrikoides* Numbers above the diagonal are absolute distance values; numbers below the diagonal are mean distance values.

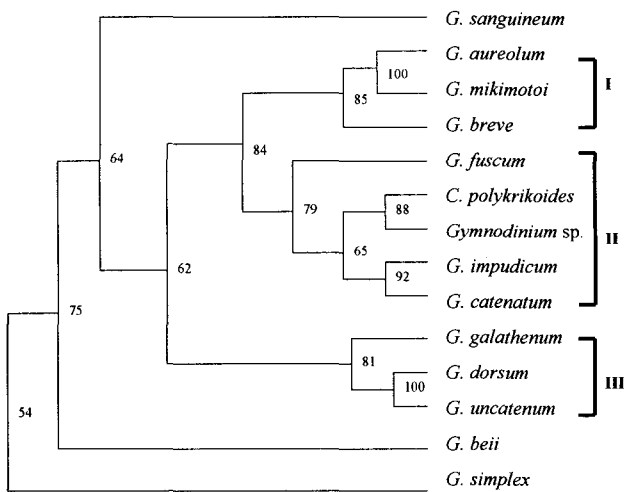


Fig. 5. PHYLIP analysis 12 species within the genus *Gymnodinium* and *Gyrodinium* obtained from GenBank database including *C. polykrikoides* and *G. aureolum*. The tree was obtained using subprogram DNAPARS in PHYLIP. The topology represents the consensus tree from an heuristic search yielding two equally most parsimonous tree. Bootstrap values (1000 replications) are given above the internal nodes. This is an unrooted tree.

Gymnodinium beii and *G. simplex* were sister groups to Group I and to Group II, respectively. The unrooted phylogenetic trees obtained from the NJ and ML methods were similar to the result obtained from the parsimony analysis (Figs. 6, 7). Likewise, *C. polykrikoides* and *G. aureolum* were closer to the genus *Gymnodinium* than to the genus *Gyrodinium*, and

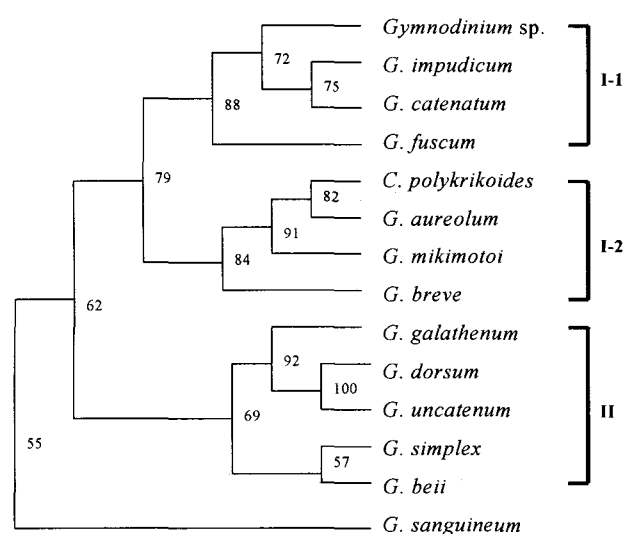


Fig. 6. PHYLIP analysis 12 species within the genus *Gymnodinium* and *Gyrodinium* obtained from GenBank database including *C. polykrikoides* and *G. aureolum*. The tree was obtained using subprogram NEIGHBOR in PHYLIP with the option of Kimura's two-parameter method. Bootstrap values (1000 replications) are given above the internal nodes. This is an unrooted tree.

were supported by a moderate bootstrap value of 79%.

Discussion

Dinoflagellates

Fensome *et al.* [11] separated *N. scintillans* from the class

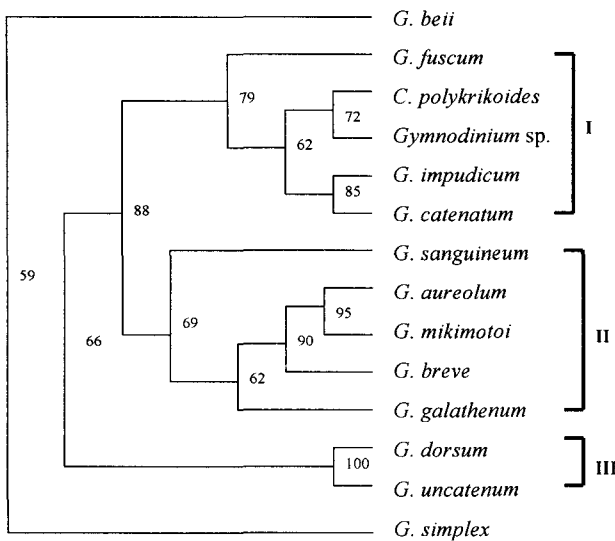


Fig. 7. PHYLIP analysis 12 species within the genus *Gymnodinium* and *Gyrodinium* obtained from GenBank database including *C. polykrikoides* and *G. aureolum*.

The tree was obtained using subprogram DNAMLK in PHYLIP with the option of Kimura's two-parameter method. Bootstrap values (1000 replications) are given above the internal nodes. This is an unrooted tree.

Dinophyceae and placed it in the class Noctiluciphyceae. In the present study, *N. scintillans* played a role as a primitive dinoflagellate, but also *G. spinifera* and *C. cohnii*, which were a member of the gonyaulacoid dinoflagellates reflected many genetic divergences of other taxa. Since *C. cohnii* appears intermediately between Noctiluciphycidae and Balstodiniophycidae, some studies have found that this species diverged early in dinoflagellate evolution [12,23]. This study showed that *N. scintillans*, *C. cohnii* and *G. spinifera* did not enough to support the dinoflagellates. Fensome *et al.* [11] suggested *C. cohnii* was able to contribute to its own family within the Gonyaulacales. The phylogenetic position of *C. cohnii* in our study was possibly responsible for its joining to the order Gonyaulacales. Fensome *et al.* [11] reported that *G. viscum* was not assigned to a subclass, which has been done uncertainly, but that gave strong bootstrap support to *Gymnodinium* (Figs. 2-4). Thus, the three species of *N. scintillans*, *G. spinifera* and *C. cohnii* represent a separate lineage outside the Dinophyceae according to this SSU phylogeny. According to the taxonomic scheme presented in Fensome *et al.* [11], the genus *Prorocentrum* was considered to belong to the independent subclass Prorocentrophycidae, but was closely related and diverged relatively to the subclass Gymnodiniophycidae in our analysis, with strong bootstrap support (Figs. 2-4).

Gymnodinium/Gyrodinium

The classification of naked microalgae bearing a resemblance to the *Gymnodinium* or *Gyrodinium* genera is difficult, as the unique distinction between these two genera is the degree of displacement of the cingulum separating the epicone and hypocone. Cingular displacement must be less than one fifth of the cell's body length for the genus *Gymnodinium* and greater than one fifth of the body length for the genus *Gyrodinium* [16]. Such an arbitrary criterion for determining genera does not account for the variations between individual cells of a unialgal isolate. Important attention has been paid to *G. mikimotoi* and *G. aureolum*, whose circumscriptions has been confused for a long time. Since two very similar dinoflagellates have been cultured, both of these species were called *G. nagasakiense* [28], prior to the name change to *G. mikimotoi* [27]. Some researchers have considered these to synonymous species, while others have proposed to remain uncertain [20]. However, the name *G. aureolum* is still widely used in Europe [14].

This study indicated that Korean *G. aureolum* and *G. mikimotoi* were genetically different targeted to the SSU, although the genetic divergence was extremely low (Table 2). The result obtained from the large subunit (LSU) gene sequence was that the genetic divergence between them was a higher value of approximately 18.7% compared with that of 0.9% when targeted to the SSU. This indicates that it is likely that the LSU is expected to be desirable as a genetic marker for discriminating *G. aureolum* from *G. mikimotoi* and for determining the clear genetic positions of the two species. Thus, study based on only molecular data has supported the distinction of *G. mikimotoi* from Korean *G. aureolum*. Although *G. aureolum* and *G. impudicum* belong to the genus *Gyrodinium*, the two species are closer to the genus *Gymnodinium* than to the genus *Gyrodinium* based on the analysis of morphological characters and gene sequences in the present work. Thus, they did not move to the taxa of *G. galathenum*, *G. dorsum* and *G. uncatenatum* with the trees produced by the parsimony, distance, and ML methods (Figs. 5-7). We suggest that *G. aureolum* and *G. impudicum* are limited to the genus *Gyrodinium*, but transferred to the genus *Gymnodinium*. More recent study has defined the need for improving the classification of gymnodinioid dinoflagellates by the analysis of morphological and molecular data [8,31]. Although Korean *G. aureolum* probably belongs to the genus *Karenia*, it did not play a role as a member of the genus *Gyrodinium*.

Cochlodinium polykrikoides

One example of the typically different character between *C. polykrikoides* and the genus *Gymnodinium* of morphological features is that a girdle in *C. polykrikoides* makes 1.8-1.9 turns around the cell [4]. Because of this problem, three species, *C. polykrikoides*, *G. impudicum*, and *G. catenatum*, are difficult to discriminate and often are misidentified as each other under a light microscope [3]. Previously, we analyzed gene sequences targeted to internal transcribed spacer (ITS) regions and the phylogenetic placement of them indicated that *C. polykrikoides* was distant from *G. impudicum* and *G. catenatum* [3,4]. At present, the genetic distance between *C. polykrikoides* and *G. impudicum*/*G. catenatum* based on the SSU is also similar to the result obtained from the ITS region, that *G. impudicum* was close to *G. catenatum* than to *C. polykrikoides* (Table 2). However, as can be seen in Figs. 5-7, *C. polykrikoides* did not transfer more to the genus *Gymnodinium* than to the genus *Gyrodinium*, under any type of topology, and also was supported by a moderate bootstrap. Although cigulum plays an important role in the potential taxonomic identification of *C. polykrikoides* and the genus *Gymnodinium*, the genetic studies on sequence divergence and phylogenetic analyses give us assurance that *C. polykrikoides* may be closer to the genus *Gymnodinium* than to the genus *Gyrodinium*.

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초록 : SSU 부위의 유전자 염기서열 분석에 의한 한국연안에서 분리한 *Cochlodinium polykrikoides* Margalef와 *Gyrodinium aureolum* Hulburt 적조생물의 분자생물학적 연구

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유해성 적조생물 *Cochlodinium polykrikoides*/*Gyrodinium aureolum*을 포함한 43 종류의 외편조류를 대상으로 SSU 부위 유전자를 분석했다. 유전자 염기서열에 의거한 상호 계통수는 parisomny, distance, maxium 방법으로 실행했다. *Noctiluca scintillans*, *Gonyaulax spinifera*와 *Crythecodinium cohnii* 종 들은 외편모조류 중 가장 유전적으로 먼 것으로 보였다. *Alexandrium*과 *Symbiodinium* 종 간의 bootstrap는 70% 이상의 상호 단일 계통도를 보인 반면에, *Gymnodinium*과 *Gyrodinium*은 근립절약계수와 최대 유사도 방법에서 다형 계통도를 나타내었다. *Gyrodinium aureolum*과 *G. dorsum*/*G. galathenum*의 유전적 분화율은 7.4% (45 bp) 였고, *G. aureolum*과 *G. mikimotoi* 상호간에는 0.9% (5bp) 밖에 나타나지 않았다. 또한 *C. polykrikoides*와 *G. aureolum*도 5.2% (31bp)로 낮은 유전적 분화율을 보였다. 계통도를 분석한 결과 *G. aureolum*과 *C. polykrikoides*는 *Gyrodinium* 보다 *Gymnodinium* 속에 훨씬 더 근접하게 나타났다.