Comparison of Analysis Methods for Ammonia from Swine Production Facilities

Ki Y, Kim*, Hong L. Choi and Chi N. Kim¹

School of Agricultural Biotechnology, Seoul National University, Seoul 151-742, Korea and ¹ Institute for Occupational Health, College of Medicine, Yonsei University, Seoul 120-752, Korea

ABSTRACT : This study was performed to evaluate the accuracy, validation and applicability of UV spectrophotometer (UV), Ion Chromatography (IC), and Detector tube (DT) methods for measuring ammonia (NH₃) concentration in a swine confinement house and swine slurry storage tank. The mean values of NH₃ emitted from the house and slurry were 5.333 ppm and 42.192 ppm for the IC method; 4.13 ppm and 36.29 ppm for the Detector tube: and 5.417 ppm and 34.193 ppm for the UV method. The accuracy and the correlation of an ammonia level analyzed by the IC method compared to the UV method were 98% and 0.998(R²) in the swine confinement house and 94% and 0.997(R²) in the swine slurry storage tank. On the other hand, those of ammonia level measured by the Detector tube compared to the UV method were 77% and 0.957(R²) in the swine confinement house and 82% and 0.941(R²) in the swine slurry storage tank. This indicated that the accuracy and the correlation of the IC method compared to the UV method were higher than those of the Detector tube method compared to the UV method. Therefore, it was concluded that the IC method was more accurate in measuring ammonia concentration in a swine house and a swine slurry storage tank. The Detector tube method should not be applied to the swine slurry storage tank in which ammonia concentration is generally higher than 30 ppm because low accuracy is caused by a gross space between scales inscribed in the Detector tube. *(Asian-Aust. J. Anim. Sci. 2004. Vol 17. No. 11 : 1608-1614)*

Key Words : UV, IC, Detector Tube, NH₃, Swine Confinement House

INTRODUCTION

Currently the intensive animal industries create large volumes of odorous and hazardous gases and wastewater and cause many environmental problems (Goopy et al., 2004). Of them, ammonia has been recognized to be an irritant to the human and animal respiratory system. Its emission from livestock production facilities causes global warming and acid rain which, in turn, increases acidity in soil. Agriculture contributes to 50% of the global NH₃ emissions (Schlesinger and Hartley, 1992). Over 70% is emitted from intensive livestock farming in Europe (Buijsman et al., 1987; Jarvis and Pain, 1990; Asman, 1992). Ammonia emission from agriculture and livestock farming has not been estimated in Korea, but could be as high as Europe. The sources of ammonia emission from livestock farming originates mainly from livestock houses, manure storages, waste management facilities, and agricultural fields sprayed with manure. High levels of ammonia result in neighbors close to the farming area complaining about the odor and often leads to lawsuits against livestock producers. Therefore, ammonia emission from livestock farming has to be abated.

Accurate and precise techniques for measurement of ammonia concentration need to be evaluated in addition to establishing a strategy for reduction of ammonia emission from livestock production sources. Table 1 shows the current methods of quantifying the concentration of ammonia emitted from livestock farming: the detector tubes, wet chemistry based on absorption in acid, fluorescence, gas chromatography, non-dispersive infrared analysers, passive diffusion device, denuder, electrochemical cell, chemiluminescence, long path optical method, and electronic nose. However, their limits of accuracy and validity for assessing emission rates of ammonia have not yet been studied.

Although the UV spectrophotometer is an official method to measure animonia concentration in aerial environments (Atmospheric Environmental Protection Law in Korea. 2000), it is known to have problems with validity and accuracy of measurement. Therefore, the purpose of this study was to compare the accuracy and validity of three analysis methods for animonia. Samples of ammonia in the swine confinement house and a swine slurry storage tank were analyzed by UV spectrophotometry, by Detector tube, and by Ion Chromatography.

MATERIALS AND METHODS

Housing and management

Experiments were performed at the swine confinement house. located at the Collegiate Livestock Experimental Station at Seoul National University. About 200 growingfattening pigs were housed in the swine confinement house $(L \times W \times H=20 \text{ m} \times 12 \text{ m} \times 3 \text{ m})$ with a concrete slatted floor. It has two rows, each of which contains ten pens on both sides. Ten crossbred (Landrace×Yorkshire×Duroc) growingfinishing pigs, which weighed about 45kg, were housed randomly in each pen. All pigs were fed a 16% crude

^{*} Corresponding Author: Ki Y. Kim. Tel: +82-2-880-4821, Fax: +82-2-781-4822, E-mail: kkysnu5@ daum.net Received March 10, 2004; Accepted July 28, 2004

| Techniques | Merit | Demerit | Reference | |
|------------------------------|---|--|--|--|
| Detector tubes | - convenient, cheap and quick | - limit of range - less precise | - Drager (1997) | |
| Wet chemistry | - simple, cheap, reliable and suitable | - high labor input | - Parkinson and Day (1979) | |
| (based onabsorption in acid) | for low concentrate of ammonia in air | - basically non-continuous | - Fehsenfeld (1995) | |
| | - very high accuracy and precision | | | |
| Fluorescence | - simple and cheap | - low accuracy | - Nakano et al. (1995) | |
| | - short sampling time | | - Vaughan et al. (1996) | |
| Gas chromatography | - very precise | - expensive | - Griffiths (1993) | |
| | - able to detect a low concentration | - requires skillful analytical technique | - Yamamoto et al. (1994) | |
| Non-dispersive | - relatively accurate | - complete selectivity for | - Janac et al. (1971) | |
| infrared analysers | | ammonia is not possible | - Hollander (1993) | |
| | | correction temperature and pressure required | | |
| Passive diffusion device | - high precision | - necessary to trap a detectable amount of gas | - Adema et al. (1991) - Blatter et al. (1992) | |
| | | - calibration for each scenario | | |
| Denuder | - very high accuracy and precision | - complex procedure | - Ferm (1979) | |
| | | | - Slanina et al. (1992) | |
| Electrochemical cell | - simple and quick | - low accuracy and precision | - Boehm (1983) | |
| | | | - Scholtens (1993) | |
| Chemiluminescence | - high accuracy | careful and frequent calibration required | - Aneja et al. (1978) - Mennen et al. (1996) | |
| Long path optical method | - advantages in averaging across a | - low accuracy and precision | - Kolb et al. (1995) | |
| | plume required in a number of flux measurement technique | | - Sommer et al. (1995) | |
| Electronic nose | - useful even in the direct | - not yet well | - Persaud and Travers (1991) | |
| | measurement of ammonia flux | characterized in accuracy and precision | | |

Table 1. The characteristics of the measurement methods of ammonia concentration



Figure 1. The plan view of the experimental confinement swine house.

protein corn-soybean meal-based diet that satisfied the NRC nutrient requirements. Pigs were fed by automatic feeder and provided with drinking water with nipples attached at the fence of the pen. Indoor air was removed by the four exhaust fans equipped at the opposite wall. Each exhaust fan has a capacity of 8.360 m³/h at maximum and was operated continuously at the minimum flow rate, recommended by MWPS (1988). to mainly control

temperature and relative humidity in the enclosed pig building.

Experimental design

Ten air samples were taken for 30 days, performed once every 3 days, in May 2003. Measurements of animonia in the house were made at the front, middle, and rear of the central alley, 0.3 m high above floor level (Figure 1). Measurements were made at the air outlet (\emptyset 2 cm) from the side wall of circular PVC column, which was filled with swine slurry at the base of the column. This was aimed to simulate the swine slurry storage, of diameter 300 mm and height 1 m. The circular column containing anaerobic swine slurry was stored in a laboratory controlled with the range of 12 to 18°C.

Air sampling method

An impinger which contained an absorption solution $(0.1 \text{ N}, \text{H}_2\text{SO}_4)$ of 10 ml was used to sample air. The impinger was connected with a moisture trap holding silica gel (200 g) to the air sampler (Gilian, No. 800519) by a polyethylene tube (Figure 2). The duration and flow rate of



Figure 2. Sampling processes of ammonia gas with air sampler.

air sampling were set at 90 min and 2 L/min in the swine confinement house and 15 min and 2 L/min in the swine shurry column. For the detector tube method, a representative value of a NH_3 concentration was taken as the average of three measurements at 30 min intervals in the swine confinement house and at 5 min intervals in the swine slurry column. This was intended to maximize the precision of values measured with the detector tube method.

Methods of NH₃ measurement

UV Spectrophotometer (UV) method : The basic principle of the UV spectrophotometry method for measuring NH3 is that the absorption solution, a strong acid solution such as sulfuric acid (H_2SO_4) or hydrochloric acid (HCl), reacts with ammonia (NH₃) in the air and NH₃ is ionized to an ammonium ion (NH_4^+) . It is adsorbed by the anion of the strong acid solution, such as sulfate (SO_4^{-2}) or chloride (CI), and then transformed into $(NH_4)_2SO_4$ or NH₄Cl. The substance deposited in the absorption solution, such as $(NH_4)_2SO_4$ or NH_4CI , is detected quantitatively by the UV spectrophotometry method. The concentration of NH₃ in air is calculated after modifying the ammonia concentration measured by the UV spectrophotometry method. A sulfuric acid (H-SO₄) solution was used as the absorption solution in this study. It reacted with NH₃ gas drawn from the swine confinement house and the swine slurry column. After sampling, the absorption solution in the impinger was carried to the laboratory and then was filtered by nuclepore filter (0.4 μ m pore size, 37 mm diameter), diluted moderately and pipetted to 3 ml. After adding Nessler's reagent (0.2ml) suggested from NIOSH (2nd Manual, 1998), it was detected by the UV spectrophotometer (UV-1601, SHIMADZU, Japan), which was set at 440 nm wavelength of UV. Before measurement, the calibration process with the six working standards (0, 2)4. 8, 12, 16 μ g/ml) was performed using (NH₄)₂SO₄ (Sigma, U.S.) as a standard solution. Application of six working standards is intended to improve an accuracy of baseline. The ammonia concentration was calculated by following Equation (1).

Table 2. Optimal conditions of the IC method for ammonia analysis

| Index | Requirement condition of the IC method | | |
|-------------------------|--|--|--|
| Column | 6.1010.000 Metrosep CATION 1-2 | | |
| Eluent | 4 mmol/l tartaric acid+l mmol/l dipicolinic acid | | |
| Flow rate | 1.0 ml/min. | | |
| Injection volume | 10 µl | | |
| Stabilization condition | Conductivity (us/cm) 650-670 Pressure (Mpa) 3.9-4.1 | | |



In order to convert the concentration of NH_3 calculated by Equation (1) into the concentration on the bases of volume ratio (V/V) unit. Equation (2) was applied.



Detector tube (DT) method : The detector tube method for analyzing NH₃ is composed of a scaled glass vial containing a chemical absorbed onto inert support granules. The basic mechanism of the method is that chemical reacts with the ammonia in the air drawn by the hand-pump, the chemical's color changes, and the degree of change in color indicates the ammonia concentration. The detector tubes applied in this study were No. 3L (1-30 ppm: GASTECH, Japan) or No. 3La (5-100 ppm: GASTECH, Japan), depending upon the strength of ammonia at the sampling sites. The air sampling with the detector tube method continued for one minute with a syringe-type vacuum pump (Pump kit No. 101, GASTECH, Japan).

Ion Chromatography (IC) method : The principle of this method is to detect and quantify ammonium ions (NH_4^+) . into which NH_3 in its gas state is converted through an absorption solution, by IC. Recently, NIOSH (2000) has suggested it as a new method for quantifying NH_3 , but until now, it has never been applied in livestock production. The IC utilized in the study was the 761 Compact IC (Metrohm, Switzerland), which was set in a laboratory. Besides no addition of Nessler's reagent. The sampling and analysis procedure for measuring ammonia by the IC method was consistent with that of the UV spectrophotometry method. Table 2 gives the required conditions of the IC for analyzing ammonia.

Statistical analyses

SAS procedure (PROC CORR) in SAS package

| Sampling site - | | IC vs. UV | | DT* vs. UV | |
|----------------------|--------|------------------|-----------------------------------|------------------|----------------------|
| | | Difference (ppm) | Rate of accuracy ¹ (%) | Difference (ppm) | Rate of accuracy (%) |
| Swine | Middle | 0.05 | 99.20 | 1.29 | 79.39 |
| confinement | | $(0.04)^2$ | (0.66) | (0.21) | (2.14) |
| house | Front | 0.14 | 97.24 | 1.17 | 76.27 |
| | | (0.06) | (1.39) | (0.11) | (1.16) |
| | Rear | 0.12 | 97.74 | 1.19 | 76.82 |
| | | (0.04) | (1.05) | (0.19) | (3.66) |
| Swine manure storage | | 2.10 | 94.22 | -6.62 | 81.76 |

(3.32)

Table 3. Evaluation of accuracy for ammonia values analyzed with the IC and the Detector tube method compared to the UV method

* Detector tube. ¹ Based on the value calculated by the UV method. ² Standard deviation.

(0.79)



Figure 3. Comparison of NH₃ concentration with the sampling sites and the analysis methods (UV, IC and Detector tube).

program (1989) was used to calculate simple correlation coefficient and the degree of correlation and to determine the significance among the values of NH_3 analyzed with the three methods.

RESULTS AND DISCUSSION

Comparison of ammonia concentrations by the sampling sites and the measurement methods

The mean ammonia concentrations in the middle of the swine confinement house were 6.252 (± 0.55) ppm with the UV method, 6.202 (± 0.56) ppm for the IC method, and 4.693 (± 0.39) ppm for the Detector tube method. At the front and rear side of the swine confinement house, the mean NH₃ concentrations were analyzed to be 5.133 (± 0.56) ppm and 4.917 (± 0.53) ppm for the UV method,

5.017 (±0.59) ppm and 4.781 (±0.57) ppm for the IC method, and 3.943 (± 0.52) ppm and 3.750 (± 0.43) ppm for the Detector tube method, respectively. The NH₃ concentration in the swine confinement house can vary with stocking density, type of shed, and ventilation rate (Groot, 1994). In addition, there are other important factors; including environmental temperature and relative humidity. wash down frequency, and feed quality, which affect NH₃ concentration in the swine house. An NH₃ concentration of 3.3 to 6.8 ppm detected in the study was very low compared with the report of Metz et al. (1998) stating that 8 to 25 ppm of ammonia was generally released in a swine house. The values are below the recommended maximum NH3 concentrations. 25 ppm, suggested by OSHA(Occupational Safety & Health Administration). This may be attributed to increased ventilation rate and superior manure management system in the swine confinement house. The mean ammonia concentrations at the front and rear of the shed were measured about 1 ppm higher than that of the middle of the shed, which probably is due to a poor air mixing. Ammonia concentration was measured 0.2 ppm lower at the front of the shed compared to the rear of the shed. This difference may be attributed to a measurement error. In swine slurry column, an initial NH₃ concentration ranged with 58 to 70 ppm was reduced at 12 to 20 ppm at the last sampling over the span of 30 days. The reduction may be caused by pH decrease in swine slurry with increasing slurry age (Rom, 1993).

(3.69)

Table 3 and Figure 4 show the accuracy rate and deviation for the values of ammonia concentration measured with the IC and the Detector tube methods compared to the UV method (the standard technique for quantifying NH₃ in the atmosphere in Korea). The difference in NH₃ concentration analyzed with the UV and the IC method was found to be 0.01-0.22 ppm in the swine confinement house and 1.32-2.93 ppm in the swine slurry storage tank. This indicated that an IC method has an accuracy of 98% in the swine confinement house and 94% in the swine slurry column, which was compared with the UV method. On the other hand, in comparison to the values of NH₃ concentrations analyzed with the UV and the

(14.98)

| Sampling site | 5 | Analysis method | correlation coefficient (R ²) | P-value |
|----------------------|--------|---------------------|---|---------|
| Swine | Center | UV*-IC ¹ | 0.998 | 0.023 |
| | | UV-DT ² | 0.957 | 0.041 |
| confinement | | IC-DT | 0.954 | 0.038 |
| house | Front | UV-IC | 0.997 | 0.018 |
| | | UV-DT | 0.973 | 0.033 |
| | | IC-DT | 0.984 | 0.048 |
| | Rear | UV-IC | 0.998 | 0.024 |
| | | UV-DT | 0.943 | 0.033 |
| | | IC-DT | 0.945 | 0.030 |
| Swine manure storage | | UV-IC | 0.997 | 0.008 |
| | | UV-DT | 0.941 | 0.036 |
| | | IC-DT | 0.966 | 0.028 |

Table 4. Statistical evaluation on degree of correlation and significance for analysis methods (UV, IC and detector tube)

* UV spectrophotometer method.

¹ Ion Chromatography method. ² Detector tube.

Detector tube method, the difference between them observed was 1.06-1.53 ppm in the swine confinement house and 2.93-10.31 ppm in the swine slurry storage tank. This shows an accuracy of 77% for the Detector tube method compared with the UV method in the swine confinement house and 82% in the swine slurry storage tank. The accuracy of the IC method compared to the Detector tube method in reference to the UV method for analyzing ammonia was about 21% higher in the swine confinement house and 12% higher in the swine shurry storage tank. It is concluded that the IC method is more accurate than the Detector tube method in both the swine confinement house and the swine shurry storage tank. Although the UV method is the standard method for measuring ammonia in Korea, it has an instrumental disadvantage to be hindered by the dust particles or alien matters suspended in the absorption solution (Skoog et al., 1999). Therefore, the IC method would probably be considered as a reliable technique for analyzing ammonia.

Correlation of the ammonia concentrations among the three analysis methods

Table 4 presents the correlation coefficients and levels of significance for the correlations between the UV. IC and Detector tube methods (also see Figures 4 and 5). The correlation coefficients for the NH₃ concentrations between UV-IC, UV-Detector tube, and IC-Detector tube were above 0.94 in the swine confinement house (middle, front, and rear) and the swine slurry storage tank and also their probability values were significant (p<0.05). In the front, middle, and rear of the swine confinement house, the correlation coefficient between the IC method and the UV method was about 0.34 higher than between the UV method and Detector tube and between the IC method and Detector tube. This could be explained by the fact that NH₃



Figure 4. Correlation relationship between the analysis methods (UV-IC and UV-Detector tube) as sampling sites in the swine confinement house.



Figure 5. Correlation relationship between the analysis methods (UV-IC & UV- Detector tube) in the swine manure storage.

concentration analyzed with the Detector tube was about 1.04-1.32 ppm lower than with the IC and UV method. This is assumed to be the influence of the dust generated in the swine confinement house or interfering substances, such as amines (R-NH₂), for detecting ammonia with the detector tube. Gas sampling by the IC and the UV method takes about an hour while the Detector tube method takes only one minute. Dust flowed into the absorption solution in impinger is expected to interfere with accuracy of measurement by the IC and UV method. It is suggested that the ammonia adsorbed on the dust surface will be desorbed and add to ammonia concentration in the adsorption water. This interpretation is supported by the statement that dust adsorbing the odorous compounds in swine houses suspends in the air or deposits on the floor (Day et al., 1965: Bart et al., 1984; Yeh et al., 2001). Also, in swine slurry, the level of correlation between the IC method and the UV method was slightly higher ($R^2=0.997$) than between the UV method and Detector tube ($R^2=0.941$) and between the IC method and Detector tube ($R^2=0.966$). This indicated that the ammonia concentration measured with the

| Analysis | Danas | Relative view | | | |
|----------------|--|--|-----------|--------|--|
| method | Kalige | Accuracy | Precision | Cost | |
| UV* | Unlimited, in principle, provided sampling period is adequate | Very high | High | Medium | |
| IC | As above | As above | Very high | High | |
| Detector tube* | Not below 1 ppm | High, provided procedure is closely followed | Low | Low | |

Table 5. Outline of the analysis methods (UV, IC and Detector tube) for NH_3 concentration

* Ref.-Phillips et al. (2001).

Detector tube method was 6.0 to 8.0 ppm higher than with the UV method and the IC method. In detecting NH₃ emitted form the swine manure stored in the cylindrical column, there was little air flow from outside of the column because the Detector tube method sucked only a small quantity of air. However, because the air sampler applied to the UV method and the IC method consistently absorbed the inner gases released from the swine slurry at the flow rate of 2 L/min for about 20 min, the comparable volume of exterior air must be supplied continuously to maintain the pressure difference between inside and outside of the column. Consequently, it is judged that the ammonia concentration quantified with the IC method and the UV method is 6.0 to 8.0 ppm lower than that of the Detector tube method due to dilution of the sample by flowing outside air into the column.

Efficiency of the analysis methods (UV, IC and Detector tube) for measuring ammonia

Table 5 gives an outline of the analysis methods (UV, IC, and Detector tube) for measuring NH3 concentration in the air. Because each method has its own instrumental limit in analyzing ammonia concentration, each method has to be applied properly according to the quantity of ammonia in livestock production facilities. The IC method has not been used to quantify NH₃ concentration in livestock systems. However, considering that the IC method has high accuracy comparable to the UV method in measuring ammonia and can detect ammonia concentration up to the 10 ppb level, the potential for the use of the IC method in the livestock production field is promising. Main disadvantages of the IC method are the high cost of the equipment and the consumption of long time acquired for obtaining stable readings. Disadvantage of the UV method is that an analyst may have difficulties in determining the titration point time to make the coupler (Nessler's reagent) and pay much attention to the analysis procedure to obtain high precision. The Detector tube method has been widely used because it is very simple, easy to treat, and cheaper than the UV and the IC. However, the analytical error by the Detector tube in measuring NH₃ concentration over 30 ppm is one reason why an analyst can not read the indicated value accurately due to wide space between each scale marked onto the

surface of Detector tube. Therefore, it is concluded that both the UV and the IC methods can be applied accurately and precisely to measure the NH_3 concentration in livestock facilities emitting above about 30 ppm of ammonia. This study was done without analysis of variance and calculation of CV (coefficient of variance) due to minimal replicates per sample (n=2) by lack of air sampler. Therefore, a supplementary study is being undertaken to statistically verify findings in this study.

REFERENCES

- Adema, E. H., V. Mejstrik and B. Binek. 1991. The determination of NH₃ concentration gradients in a spruce forest in south Bohemia, CSSR, 1998, using a passive sampling technique. Agricultural University, Wageningen. Report R-501, p. 15.
- Aneja, V. P., E. P. Stahel, H. H. Rogers, A. M. Witherspoon and W. W. Heck. 1978. Calibration and performance of a thermal converter in continuous atmospheric monitoring of ammonia. Anal. Chem. 50:1705-1707.
- Asman, W. A. H. 1992. Ammonia emission in Europe: Updated emission and emission variations. RIVM Bilthoven (The Netherlands). p. 88.
- Barth, C. L., L. F. Elliot and S. W. Melvin. 1984. Using odor control technology to support animal agriculture. Trans. Am. Soc. Agric. Eng. 27:859-864.
- Blatter, A., M. Fahrni and A. Neffel. 1992. A new generations of NH₃ passive samplers. In: Development of Analytical Techniques for Atmospheric Pollutants. Air Pollution Research Report 41, pp. 171-176.
- Buijsman, E., J. F. Mass and W. A. H. Asman. 1987. Anthropogenic NH₃ emission in Europe. Atmos. Environ. 21:1009-1022.
- Day, D. L., E. L. Hansen and S. Anderson. 1965. Gases and odors in confinement swine houses. Trans. Am. Soc. Agric. Eng. 8(1):118-121.
- Fehsenfeld, F. 1995. Measurement of chemically reactive trace gases at ambient concentrations. In: Biogenic Trace Gases: measuring Emissions from Soil and Water. Blackwell Science, Oxford, pp. 206-258.
- Ferm, M. 1986. Method for determination of ammonia. Atmos. Environ. 13:1385-1393.
- Goopy, J. P., P. J. Murray, A. T. Lisle and R. A. M. Al Jassim. 2004. Use of chemical and biological agents to improve waste of effluent discharge from Abattoirs. Asian-Aust. J. Anim. Sci. 17(1):137-145.
- Griffiths, R. F. 1993. Emissions and environmental monitoring

using energetic UV radiations: a new development in portable ambient monitoring. Proceedings of the Conference 'Monitor 93' Spring Innovations Ltd, Manchester, October. pp. 57-62.

- Groot, P. W. G. 1994. Review on emissions of ammonia from using system for laying hens in relation to sources, processes, house design and manure handling. J. Agric. Engng. Res. 59:73-87.
- Hollander, J. C. T. 1993. Non-dispersive infrared (NDIR) photometry. In: Methods for measuring ammonia emissions from animal housing. Analysis of ammonia Series. pp. 42-44.
- Janac, J., J. Catsky and P. G. Jarvis. 1971. Infrared gas analysers and other physical parameters. In: Plant Photosynthetic Production: Manual of Methods. Wjunk, The Hague (The Netherlands). pp. 111-197.
- Jarvis, S. C. and B. F. Pain. 1990. Ammonia emission from agricultural land. Proceedings Fertility Soc. No. 298. Greenhill House, Peterborough, England. p. 35.
- Kolb, C. E., J. C. Wormboudt and M. S. Zahniser. 1995. Recent advances in spectroscopic instrumentation for measuring stable gases in the natural environment. In: Biogenic Trace Gases: measuring Emissions from Soil and Water. Blackwell Science, Oxford. pp. 259-290.
- Mennen, M. G., B. G. van Elzakker, E. M. van Puten, J. W. Uiterwijik, T. A. Regts, J. van Hellemond, G. P. Wyers, R. P. Otjes, A. J. L. Verhage, L. W. Wouters, C. J. G. Heffels, F. G. Romer, L. van den Beld and J. E. H. Tetteroo. 1996. Evaluation of automatic ammonia monitors for application in an Air Quality Monitoring Network. Atmos. Environ. 30:3239-3256.
- Metz, J. H. M., G. H. Uenk and V. R. Phillips. 1998. Concentrations and emissions of ammonia in livestock houses in Norther Europe. J. Agric. Engng. Res. 70:79-95.
- MWPS. 1988. Swine housing and equipment handbook: MWPS-8. Midwest Plan Service, Iowa State University, Ames, IA 50011.
- Nakano, N., K. Sugata and K. Nagashima. 1995. Development of a monitoring tape for animonia gas in air by fluorescence detection. Anal. Chem. 302:201-205.
- NIOSH. 2000. Manual of analytical methods (4th).
- Parkinson, K. J. and W. Day. 1979. The use of orifices to control the flow rate of gases. J. Appl. Ecol. 16:623-632.

- Persaud, K. C. and P. Travers. 1991. Multi-element arrays for sensing volatile chemicals. Intelligent Instruments and Computers. July/August, pp. 147-154.
- Phillips, V. R., D. S. Lee, R. Scholtens, J. A. Garland and R. W. Sneath. 2001. A review of methods for measuring emission rates of ammonia from livestock houses and slurry or manure stores, part 2: monitoring flux rates, concentration and airflow rates. J. Agric. Engng. Res. 78(1):1-14.
- Rom, H. B. 1993. Ammonia emission from livestock buildings in Denmark. Proceedings of International Livestock Environment Symposium IV, Warwick, England, 6-9 July. ASAE 754-761.
- SAS Institute Inc. 1989. SAS/STAT User's Guide: Version 6. 4th edn. SAS Institute Inc., Carv, North Carolina.
- Schlesinger, W. H. and A. E. Hartley. 1992. A global budget for atmospheric NH₃. Biogeochem. 15:191-211.
- Skoog, D. A., D. M. West, F. J. Holler and S. R. Crounch. 1999. Analytical Chemistry. Harcourt College Publishers, pp. 575-581.
- Slanina, J., P. J. de Wild and G. P. Wyers. 1992. The applications of denuder systems to the analysis of atmospheric components. In: Gaseous Pollutants: Characterization and Cycling. John Wiley, London. pp. 129-154.
- Sommer, S. G., H. Mikkelsen and J. Mellqvist. 1995. Evaluation of meteorological techniques for measurements of ammonia loss from pig slurry. Agric. Forest Meteorol. 74:169-179.
- Vaughan, A. A., M. G. Baron and R. Naryanaswamy. 1996. Optical ammonia sensing films based on an immobilized metalloporphyrin. Analytical Communications 33:393-396.
- Yamamoto, N., H. Nishiura, T. Honjo, Y. Ishikawa and K. Suzuki. 1994. Continuous determination of atmospheric ammonia by an automated gas chromatographic system. Anal. Chem. 66:756-760.
- Yeh, Y. L., C. M. Liao, J. S. Chen and J. W. Chen. 2001. Modelling lumped-parameter sorption kinetics and diffusion dynamics of odor-causing VOCs to dust particles. Appl. Math. Modelling. 25:593-611.
- Korean Ministry of Environment. 2000. Methods of public test for air pollution. Annual Report.