

## Rare 2-Phenoxychromones from *Ononis serrata* Growing in Egypt

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**Abstract** – Two 2-phenoxychromones; a rare group of natural products, were isolated from the combined ethyl acetate and chloroform fractions of the ethanolic extract of *Ononis serrata* frossk (Leguminosae) growing in Egypt. Structures were elucidated by chemical, spectroscopic methods as well as single X-ray diffraction analyses. The cytotoxic and antimicrobial effects of the two compounds were examined. It is worth to mention that this is the first report on the isolation of 2-phenoxychromones from the family Leguminosae.

**Key words** – *Ononis serrata*, Leguminosae, 2-phenoxychromones, X-ray, cytotoxicity, antimicrobial.

### Introduction

Chemical investigation of *Ononis* species growing in Egypt is one of the points of our interest. Previous study conducted in our laboratory resulted in the isolation of several plant phenolics belonging to different classes of secondary metabolites. (Amer *et al.*, 1989; 2001; Abdel-Kader, 1997; 2001; Amer, 2001).

We previously reported on the isolation of seven flavonoids from the combined ethyl acetate and chloroform fractions of the ethanol extract of *O. serrata* (Amer *et al.*, 2001). In this paper we report on the identification of two phenoxychromones from the same fraction. Antimicrobial and cytotoxic activities of the two isolates were evaluated.

### Experimental

**Plant materials** – The whole plants of *Ononis serrata* Forssk collected near Rosetta, Egypt were described earlier (Amer *et al.*, 2001).

**Extraction and Isolation** – The detailed fractionation procedures were described earlier (Amer *et al.*, 2001). The ethanol extract of the whole dried powdered plants (1.5 kg) was fractionated with C<sub>6</sub>H<sub>14</sub>, CHCl<sub>3</sub> and EtOAc. A part (20 g) of the combined CHCl<sub>3</sub> and EtOAc extracts (32 g)

was chromatographed over silica gel column (400 g). The early fraction (7-12) of the column eluted with 2% MeOH in CH<sub>2</sub>Cl<sub>2</sub> (0.7 g) were rechromatographed over silica gel column (50 gm) eluted with 1% MeOH in CH<sub>2</sub>Cl<sub>2</sub>. 25 Fractions, 75 ml each were collected, screened by TLC and similar fractions were combined. Crystallization of fractions 5-9 from MeOH afforded 45 mg of 1 (R<sub>f</sub> value = 0.62, CHCl<sub>3</sub>/MeOH 9:1), while crystallization of fractions 12-18 from acetone-benzene afforded 400 mg of 2 (R<sub>f</sub> value = 0.43, CHCl<sub>3</sub>/MeOH 9:1).

**Capillarisin [2-(*p*-hydroxyphenoxy)-6-methoxy-5,7-dihydroxy-chromone] 1** – C<sub>16</sub>H<sub>12</sub>O<sub>7</sub>, creamy crystals, mp 225-226°C (MeOH). UV λ<sub>max</sub><sup>MeOH</sup> nm: 232, 289, IR ν<sub>KBr</sub><sup>cm<sup>-1</sup></sup> : 3260, 1654, 1616, 1564, 1234, 1170. <sup>1</sup>H- and <sup>13</sup>C-NMR data (Table 1). EI-MS (rel. int., %) : 317 (M<sup>+</sup>+1, 50), 316 (M<sup>+</sup>, 85), 301 ((M<sup>+</sup> - CH<sub>3</sub>), 25), 298 (27), 273 (35), 110 (22), 93 (46), 69 (97), 65 (100), 63 (24), 53 (25).

**6-Demethoxycapillarisin [2-(*p*-hydroxyphenoxy)-5,7-dihydroxy-chromone] 2** – C<sub>15</sub>H<sub>10</sub>O<sub>6</sub>, colourless crystals, mp >280°C. UV. λ<sub>max</sub><sup>MeOH</sup> nm: 230, 287, 310 (sh), IR ν<sub>KBr</sub><sup>cm<sup>-1</sup></sup> : 3500, 3150, 1649, 1618, 1569, 1500, 1431, 1369. <sup>1</sup>H- and <sup>13</sup>C-NMR data (Table 1). EIMS (rel. int., %): 287 (M<sup>+</sup>+1, 40), 286 (M<sup>+</sup>, 100), 153 (82), 134 (59), 121 (30), 110 (56), 106 (43), 93 (22), 81 (34), 69 (94), 65 (75), 63 (27), 53 (35).

**Antimicrobial and Cytotoxicity testing** – The two compounds were subjected to antimicrobial testing against *Bacillus subtilis* (ATCC), *Micrococcus luteus* (ACTT10240), *Staphylococcus aureus* (ATCC6538), *Bordetella bronchiseptica*

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(ATCC4617), *Escherichia coli* (ATCC8739), *Salmonella typhi* (ATCC3112), *Pseudomonas aeruginosa* (ATCC9027), and *Candida albicans* (ATCC8731). Cytotoxicity were tested utilizing Human Ovarian Cancer Cell (A 2780).

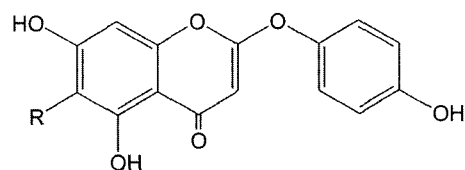
#### Single-Crystal X-ray Analysis of 6-demethoxycapillarisin

**2**—Diffraction data of  $C_{15}H_{10}O_6$ , were collected from colourless crystals of dimensions  $0.2 \times 0.25 \times 0.3$  mm, using the automatic diffractometer Enraf Nonius FR 590. The crystal system is monoclinic with  $a = 5.2464$  (2) Å,  $b = 29.2043$  (11) Å,  $c = 10.7650$  (5) Å,  $v = 1632.01$  (12) Å<sup>3</sup>,  $P_{calc} = 1.259$  gm/cm<sup>3</sup> for  $z = 4$ . The space group is  $P_{2,2,2}$ . Diffraction data : 3234 reflections were collected using MO radiation  $\lambda$  (K $\alpha$ ) = 0.71073 Å. Computing data collection: Kappa CCD.

### Discussion

Compounds **1** and **2** were isolated from the non-polar fractions of the combined  $CHCl_3/EtOAc$  extracts along with seven other flavones (Amer *et al.*, 2001). These two compounds were different from other flavonoids in both TLC behaviours and spectral data. They develop an orange red colour with anisaldehyde/ $H_2SO_4$  spray reagent. Both **1** and **2** give very faint yellow colour with ammonia and Marquis reagent, violet colour with Mandalin's reagent. Erdman's reagent develops orange red colour with **1** and yellow colour with **2**.

<sup>1</sup>H- and <sup>13</sup>C-NMR data of **1** (Table 1) looks like a simple flavone with 5,6,7,4'-tetraoxygenation including one methoxyl group. The singlet at  $\delta$  6.40 assigned for H-8 in **1** was replaced by two meta-coupled doublets at  $\delta$  6.04, 6.13 ( $J$



1: R = OCH<sub>3</sub>  
2: R = H

= 2.1 Hz) in **2** assigned for H-8 and H-6 respectively. The disappearance of the methoxyl singlet from the <sup>1</sup>H-NMR indicated that **2** is the demethoxy derivative of **1**. However, UV, <sup>1</sup>H-, <sup>13</sup>C-NMR (Table 1) and MS data (experimental) did not match a regular flavone skeleton. The odd values were the <sup>1</sup>H-NMR singlets at  $\delta$  4.98 and 4.97 in **1** and **2** respectively. The chemical shifts of some characteristic carbons in flavones such as C-2, C-3, C-1', C-2' and C-6' could be changed in **1** and **2**. Comparing the MS of **1** ( $M^+$  at 316  $m/z$ ) and **2** ( $M^+$  at 286  $m/z$ ) with sorbifolin ( $M^+$  at 300  $m/z$ ) and apigenin ( $M^+$  at 270  $m/z$ ), the two simple flavones have same substitution pattern, revealed that **1** and **2** have an additional oxygen atom.

Compound **2** was crystallized from acetone/benzene and the crystals were subjected to single X-ray diffraction analysis (Fig. 1) which proved that the extra oxygen atom is located between C-2 and C-1'. Consequently **2** were identified as 6-demethoxy capillarisin previously isolated from three Compositae plants (Kijjoa *et al.*, 1999; Tanko and Hashimoto, 1981; Komiy *et al.*, 1976) and *Rosa rugosa* (Rosaceae) (Hashidoko, 1991). Compound **1** was identified as the more rare capillarisin only isolated from *Artemisia capillaris* (Komiy, 1975).

**Table 1.** <sup>1</sup>H- and <sup>13</sup>C-NMR data of compounds **1** and **2**.

Position	<b>1</b> <sup>a</sup>		<b>2</b> <sup>a</sup>	
	<sup>13</sup> C	<sup>1</sup> H	<sup>13</sup> C	<sup>1</sup> H
<b>2</b>	168.3		168.3	
<b>3</b>	87.0	4.98 (1H, s)	87.0	4.97 (1H, s)
<b>4</b>	183.8		184.0	
<b>5</b>	150.2		161.3*	
<b>6</b>	132.1		99.4	6.04 (1H, d, $J = 2.1$ )
<b>7</b>	153.4		163.7*	
<b>8</b>	94.5	6.40 (1H, s)	94.1	6.13 (1H, d, $J = 2.1$ )
<b>9</b>	156.5*		155.3**	
<b>10</b>	102.7		102.5	
<b>1'</b>	143.5		143.6	
<b>2'</b>	122.2	7.13 (2H, d, $J = 9$ )	121.4	6.82 (2H, d, $J = 8$ )
<b>3'</b>	116.9	6.84 (2H, d, $J = 9$ )	116.4	6.67 (2H, d, $J = 8$ )
<b>4'</b>	157.2*		155.6**	
<b>5'</b>	116.9	6.84 (2H, d, $J = 9$ )	116.4	6.67 (2H, d, $J = 8$ )
<b>6'</b>	122.2	7.13 (2H, d, $J = 9$ )	121.4	6.82 (2H, d, $J = 8$ )
<b>OCH<sub>3</sub></b>	60.4	3.70 (3H, s)		

<sup>a</sup>Spectra were measured in  $CDCl_3$  in 400 MHz NMR Spectrometer.

\*, \*\* Signals can be interchanged.

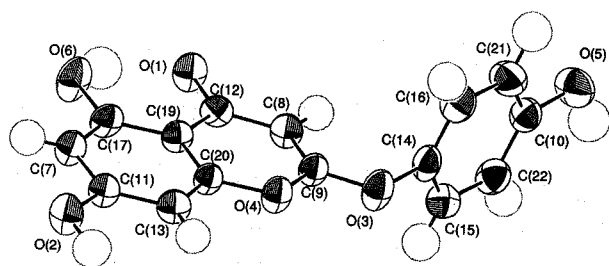


Fig. 1. X-ray structure of compound 2.

$^{13}\text{C}$ -NMR data of **1** was not previously reported. However, the results obtained from an HMQC experiments indicated that the  $^1\text{H}$ -NMR singlets at  $\delta$  4.98 and 4.97 are correlated to carbons at 86.96 and 86.97 ppm in **1** and **2** respectively. These findings clearly prove that the assignments of C-3 and C-8 (Hashidoko, 1991; Kijjoa, 1999) must be revised. These new assignments of C-3 and C-8 fit with the calculated values by means of ACD Lab simulation program.

In the antimicrobial assay the two compounds were active against *Bacillus subtilis* with an MIC = 32 and 48  $\mu\text{g}/\text{ml}$  respectively. Compound **1** was also active against *Bordetella bronchiseptica* with an MIC = 64  $\mu\text{g}/\text{ml}$ . Compounds **1** and **2** showed weak cytotoxic effect against A 2780 (Human Ovarian Cancer Cell) with an  $\text{IC}_{50}$  = 23 and 21  $\mu\text{g}/\text{ml}$  respectively.

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### References

- Abdel-Kader, M. S., Two new norphenylpropanoid glucosides and hemipholin from the flowers of *Ononis vaginalis*. *J. Braz. Chem. Soc.*, **8**, 637 (1997).
- Abdel-Kader, M. S., Phenolic constituents of *Ononis vaginalis* roots" *Planta Medica*, **67** (4), 388 (2001).
- Amer, M. E., Abdel-Kader, M. S., Mahmoud, Z. F., Abdel-Salam, N. A., Yang, S. S. and Mabry, T. J., Flavonoids of *Ononis vaginalis* Vahl. *Symb. Rev. Latinoamer. Quim.*, **20**, 152- 3 (1989).
- Amer, M. E., Kassem, F. F. and Abdel-Kader, M. S., Flavonoids from *Ononis serrata* growing in Egypt. *Alex. J. Pharm. Sci.*, **15** (2), 99 (2001).
- Hashidoko, Y., Tahara, S. and Mizutani J., 2-Phenoxychromones and a structurally related flavone from leaves of *Rosa rugosa*. *Phytochemistry*, **30**(11), 3837-8 (1991).
- Kijjoa, A., Vieira, L. M., Pereira, J. A., Silva, A. M. S. and Herz, W., further constituents of *Achillea ageratum*. *Phytochemistry*, **51**, 555- 8 (1999).
- Komiya, T., Tsukui, M. and Oshio, H., Capillarisin, a constituent from *Artemisia capillaries* Herba. *Chem. Pharm. Bull.*, **23**(6), 1387- 8 (1975).
- Lorian, V., "Antibiotics in laboratory medicine" Williams and Wilkins, London, 1980.
- Takeo, H. and Hashimoto, M., Synthesis of demethoxycapillarisin, a naturally occurring 2-phenoxychromone, and related compounds. *J. C. S. Chem. Comm.*, 474- 5 (1981).

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