

AN EXPERIMENTAL STUDY OF NEWLY DESIGNED IMPLANT WITH RBM SURFACE IN THE RABBIT TIBIA : RESONANCE FREQUENCY ANALYSIS AND REMOVAL TORQUE STUDY

Mi-Kyoung Won, D.D.S., Chan-Jin Park, D.D.S., M.S.D., Ph.D.,
Kyoung-Soo Chang, D.D.S., M.S.D., Ph.D., Chang-Whe Kim, D.D.S., M.S.D., Ph.D.,
Yung-Soo Kim, D.D.S., M.S.D., Ph.D., Zakiahbt Mohd Isa, D.D.S., M.S.D., Ph.D. *,
Yusnidar Tajul Ariffin, D.D.S., M.S.D., Ph.D.*

Department of Prosthodontics, College of Dentistry, Seoul National University, Univ. of Malaya*

Statement of problem. The importance of fixture design and surface treatment.

Purpose. The clinical success of dental implants is affected by many factors such like as degree of osseointegration, the effective load dispersion for the prostheses, and a lot of attempts have been made to overcome the difficulties. In this study, efforts were made to find the possibility of clinical acceptance of the dental implants of newly designed surface and resorbable blast media surface.

Materials and methods. In this study, two groups of custom-made, screw-shaped implants were prepared. The first with the consisting of Brånemark clone design and the other with the new design. These implants were divided into four groups according to the kinds of surface treatment. Four implants(AVANA[®], Osstem, Busan, Korea)of each group were installed in twenty rabbits. Group A was consisted of Brånemark clone implant left as machined, Group B with Brånemark clone implants with RBM(Resorbable blast media) surface, Group C with newly designed implants left as machined and Group D with newly designed implants with RBM surface. One of the twenty rabbits died from inflammation and the observation was made for six weeks. Specimens from four groups were observed using scanning electron microscopy with 40, 100, 1000 magnification power and microsurface structures were measured by white-light scanning interferometry for three dimensional surface roughness measurements(Accura 2000[®], Intek-Plus, Korea.). Removal torque was measured in 17 rabbits using digital torque gauge(MGT 12R, Mark-10 corp., NY, U.S.A.) immediately after the sacrifice and two rabbits were used for the histologic preparation(EXAKT 310[®], Heraeus Kulzer, Wehrheim, Germany) of specimens and observed under light microscope. Resonance frequency measurement(Osstell[®]) was taken with the 19 rabbits at the beginning of the implant fixation and immediately after the sacrifice.

Results. Following results were taken from the experiment.

1. The surface of the RBM implants as seen with SEM had rough and irregular pattern with reticular formation compared to that of turned specimens showing different surface topographies.
2. The newly designed implant with RBM surface had high removal torque value among four groups with no statistical significance. The average removal torque was 49.95 ± 6.70 Ncm in Group A, 51.15 ± 4.40 Ncm in Group B, 50.78 ± 9.37 Ncm in Group C, 51.09 ± 4.69 Ncm in Group D.
3. The RFA values were 70.8 ± 4.3 Hz in Group A, 71.8 ± 3.1 Hz in Group B, 70.9 ± 2.5 Hz, 72.7 ± 2.5 Hz in Group D. Higher values were noted in the groups which had surface treatment compared to the untreated groups with no statistical significance.
4. The results from the histomorphometric evaluation showed a mean percentage of bone-to-implant contact of $45 \pm 0.5\%$ in Group A, $55 \pm 3\%$ in Group B, $49.5 \pm 0.5\%$ in Group C, and $55 \pm 3\%$ in Group D. Quite amount of newly formed bone were observed at the surface RBM-treated implants in bone marrow space.

Key Words

Dental implant, Resonance frequency analysis, Removal torque, Digital torque gauge, Resorbable blast media

For the past 30 years the use of osseointegrated implants has become a scientifically accepted and well-documented treatment modality for the rehabilitation of completely and partially edentulous patients. Osseointegration is a treatment concept based on stability¹ and the rigid fixation seems to be a prerequisite for a favorable long-term clinical outcome.² The term osseointegration has been used to define a direct structural and functional connection between ordered living bone and the surface of a load carrying implant, and it has been mechanically defined as continuity between implant and adjacent hard tissue.³⁻⁵ Successful long-term stability of osseointegrated dental implants has been reported in a large number of clinical studies using commercially pure titanium implants.⁶⁻⁸

Although the implants have high survival rates,^{6,9} other clinical studies,^{9,11} however, reported increased failure rates in area with poor bone quality such as low bone density or insufficient bone volume and height, mainly in the posterior maxilla, especially for screw-type implants with a turned surface. Failures related to these situations are caused by bone loss as a result of lack of primary and secondary implant stability.

The density and quantity of the bone, the surgical technique, and the design of implant determine primary stability. Secondary stability is the stability of implant after primary healing and can be increased by bone formation and remodeling at the implant-bone interface.

Successful osseointegration of endosseous implants results from a favorable interaction between the implant geometry, surface texture and the tissues at the bone site.¹² Implant material, the macro-design and surface structure are among the crucial factors influencing the clinical outcome of an implant.¹³ Many attempts have been made over the past decade to improve bone anchorage of dental implants. Especially, surface macrostructure and microstructure have developed to establish a stable fixation be-

tween the implant and the bony tissue and to improve load transfer and to evoke favorable bone and cell response.

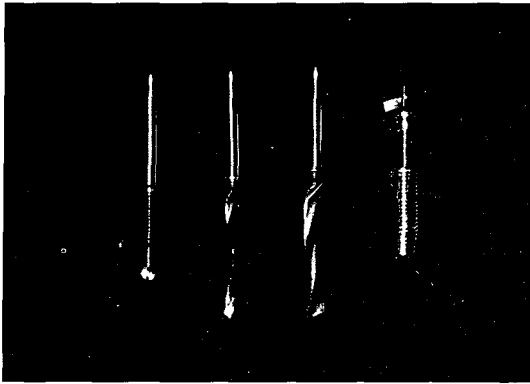
The current paper will focus on the *in vivo* investigations, especially those that were evaluated with biomechanical tests, i.e. resonance frequency analysis (RFA) and removal torque. From a mechanical standpoint, the removal torque technique measures the strength of the bone-implant interface in terms of shear, while the RFA is considered to measure the stability during bending.¹⁴

The purpose of the present study was to evaluate bone tissue reaction around a newly design threaded implants (AVANA[®], Osstem, Busan, Korea) with new surface treatment called resorbable blast media (RBM) placed in the rabbit tibia using mainly resonance frequency analysis and removal torque measurement. The resonance frequency measurement and anchorage of newly design implants with RBM surface was compared with those of conventional Brånemark clone with turned surface, Brånemark clone with RBM surface, and the new design with turned surface screws.

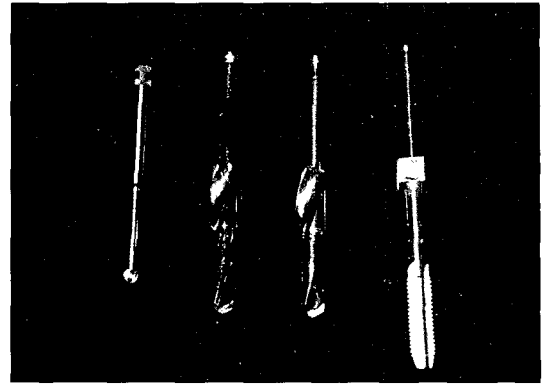
MATERIAL AND METHODS

1. Animals and Surgical technique

Twenty adult New Zealand white rabbits of both sexes weighing 3.5 to 4 kg were included in this study. Prior to surgery, animals were acclimated to the vivarium for a period of observation to insure that they were healthy and stable. The animals were anesthetized using intramuscular injections of keta (8.8mg/kg) for surgical procedure. Prior to surgery, the shaved skin was carefully washed with a mixture of iodine and 70% ethanol. 1.8mL of lidocaine 2% were injected locally into the surgical sites. The tibial metaphysis was exposed by incisions through the skin, fascia, and periosteum. By intermittent drilling using low rotary speed (not exceeding 2000r.p.m) with copious saline irrigation, 2 holes were



(A)



(B)

Fig. 1. A : drills for Brånemark clone implants. B : drills for new-design implants.

drilled 7mm apart in the central portion of each tibia and sequentially enlarged to 3mm. After tapping and slightly countersinking the sites, the implants were gently screwed into place, until the implant shoulder was leveling with the bone surface. All implants penetrated the first cortical layer only, never engaging the opposite cortical side. Each rabbit received four implants, from each of the different groups (group A, B, C, D) which were randomly assigned to their implantation sites. Then, the skin and fascia layers were closed separately using resorbable sutures. After the operation, the animals received Cefazolin IM injection per day for 7 days. The animals were kept in separate cages and immediately after surgery they were allowed to bear full weight. The follow-up time was six weeks. One week after surgery, one animal died due to an unknown inflammation. The other nineteen animals were sacrificed using an overdose of carbon dioxide.

2. Implant preparation: design and surface treatment

A total of 80 custom-made, screw-shaped implants were used. 40 implants were Brånemark clone designed, and other 40 implants had a new de-

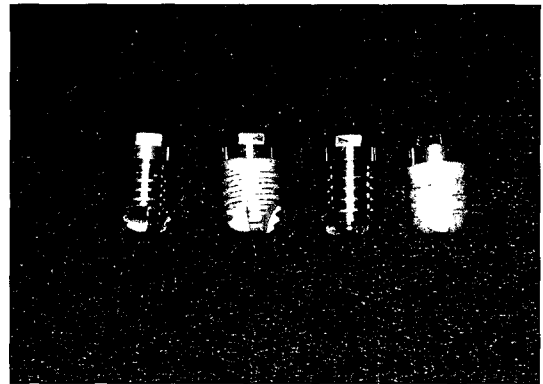


Fig. 2. Implants from each group. Group A, B, C, D

sign(AVANA®, Osstem, Busan, Korea). These implants were divided into 4 groups according to surface treatment.) Group A and B Brånemark clone implants have 0.6mm pitch, and group C, D newly design implants have 0.8mm pitch.

- Group A: 20 Brånemark clone implants left as machined.
- Group B: 20 Brånemark clone implants with RBM surface.
- Group C: 20 newly designed implants left as machined.
- Group D: 20 newly designed implants with RBM surface.

3. Scanning electron microscopy and scanning interferometry

The electron microscopy used to observe the characteristics of the microstructure of the RBM implants and to compare it with the surface of the machined implants was performed on a scanning electron microscopy. ($\times 40, 100, 1000$, JSM-840A, JEOL, Japan)

Microsurface structure (of 2 implants of each material from each group) was measured by white-light scanning interferometry for three dimensional surface roughness measurements: Acurra 2000 (INTER PLUS, Korea). In 2 samples from each of the 4 groups, 3 threads were selected randomly and scanned along their circumference in 4 different areas, yielding 12 measurements for each surface topography. Three height parameters, R_a , R_q , and R_z , were used for quantitative characterization of the surface roughness. R_a describes the arithmetic mean of departures of the roughness profile from the midline, R_q is the root mean square parameter corresponding to R_a , and R_z measures the average height difference between the 5 highest peaks and the 5 lowest valleys.

4. Resonance frequency measurement

Resonance frequency analysis (RFA), a novel technique for the clinical measurement of implant stability and osseointegration, was presented by Meredith et al.¹⁵ This method is non-invasive technique that measures the implant stability in terms of interfacial stiffness (Hz). The frequency response of the system was measured by attaching the transducer, i.e. an L-shaped cantilever beam, to a screw implant. The excitation signal was a sine wave varying in frequency from 5kHz to 15kHz with peak amplitude of 1 volt. The first flexural (bending) resonance frequency of the resulting system was measured.^{16,17} The resonance frequency was measured immediately

after implant placement (20 rabbits) and at sacrifice six weeks later (19 rabbits).

5. Removal torque

To test the implant stability, specifically the strictly interfacial shear strength, removal torque was measured at the time of animal sacrifice, i.e. six weeks after implantation. According to Roberts et al, in rabbits it takes 6 weeks for the woven bone to be replaced by lamellar bone with adequate strength for load bearing.¹⁸

The small diameter hex top was 0.7mm and too short to measure. Using resin cement (Panavia 21, Kuralay, Japan), small diameter fixture mount was attached (Fig. 3) and the force needed to unscrew the implants ($n=72$, 18 rabbits) was measured using a digital torque gauge (Model MGT12, Mark-10 Corp., 458 West John Street Hicksville, NY 11801 USA) It has the measuring torque range of 135Ncm and the accuracy of 0.5% of full scale of 1 digit, i.e. 0.3ozin (about 0.2Ncm) The implants were subjected to slowly increasing torque until loosening was detected, and the peak torque value was measured.

6. Histologic preparation of specimens

The remaining two animals were sacrificed without subjecting the implants to removal torque, and the specimens (including implants; $n=4$) and surrounding tissues were washed in saline solution and fixed in 4% paraformaldehyde and 0.1% glutaraldehyde in 0.15mol/L cacodylate buffer at 4°C and pH 7.4. The specimens were further dehydrated in ascending concentrations of alcohol rinses and infiltrated with glycolmethacrylate resin (Technovit 7200 VLC, Kulzer & Co, Wehrheim, Germany). After polymerization, the specimens were sectioned longitudinally at about 100 μ m and ground to a final thickness of about 25 μ m (EXAKT 310, GMBH & Co, Germany) as described by donath¹⁹

One section was obtained for each implant and stained with hematoxylin and eosin. The histomorphometric analysis was performed using a light microscope connected to a personal computer. The percentage of bone to implant contact around threads, which engaged only in cortical plate was observed with 40 magnification power of light microscope. No measurement was taken from the cancellous bone area in the rabbits, because not much of significance was there to measure the percentage of contact in hollow marrow space.

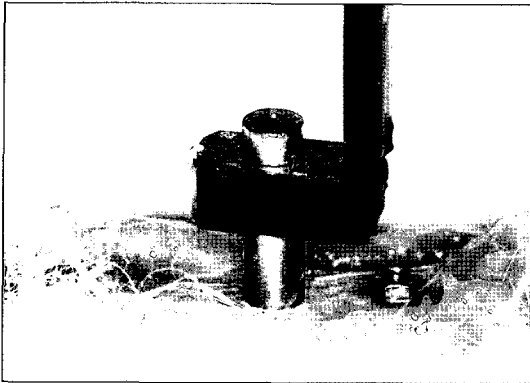


Fig. 3. Transducer attached to implant which placed into rabbit tibia.

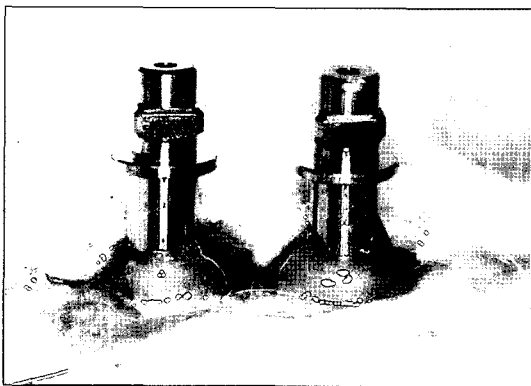


Fig. 4. Small diameter fixture mount was attached to the implant. Without this procedure, the hex top of implant would be worn-out.

7. Statistical analysis

Mean value of resonance frequency analysis were calculated and subjected to a repeated measure ANOVA to test for significant differences between 4 investigated groups. Scheffe's test between all groups was performed for removal torque evaluation. Statistical testing was carried out at the 5% significance level.

RESULTS

1. Topographic Evaluation

The scanning electron microscopy analysis and surface roughness measurements demonstrate the different surface topographies between the surface of the RBM implants and the turned implant. The surface of the RBM implants as seen with SEM had a rough and irregular pattern, while the turned specimens showed relatively smooth surface. (Fig. 6) At higher magnification, the surface appears reticulated, with undermining deformation of the metal remaining after impaction of the resorbable hydroxyapatite material blasted under pressure on the

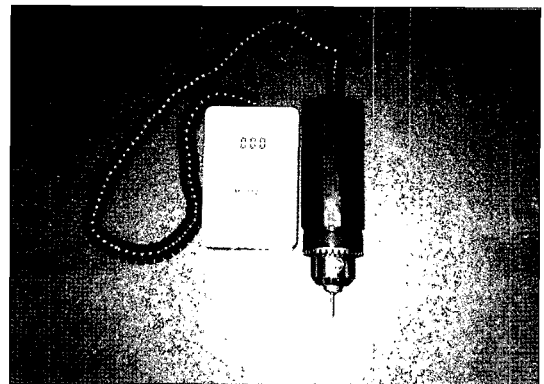
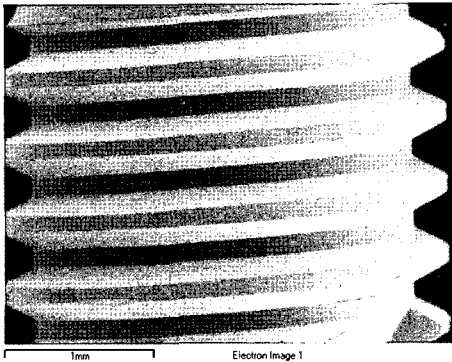


Fig. 5. Digital torque gauge for removal torque measurement.

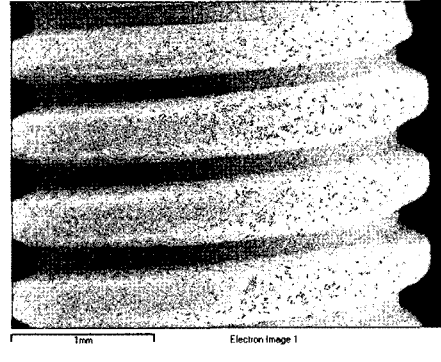
surface of the implant (Fig. 6, B3, $\times 1000$)

The Brånemark clone implant with turned surface showed an average surface roughness $0.26\mu\text{m}$; cor-

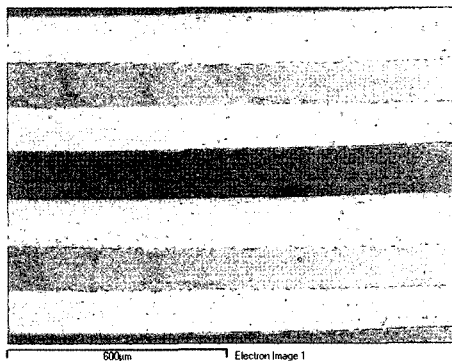
responding values for newly designed implant with RBM surface were $1.12\mu\text{m}$.



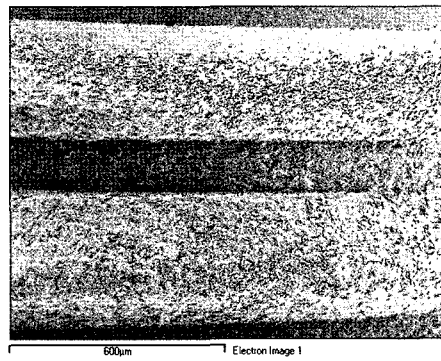
A1



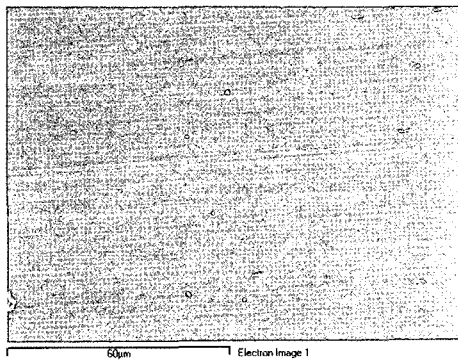
B1



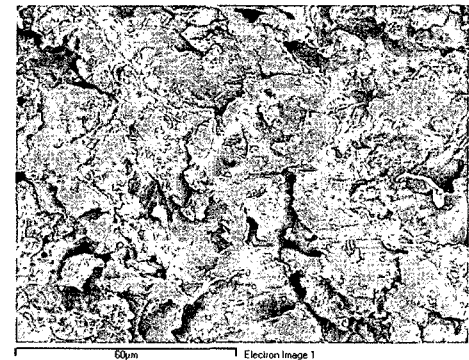
A2



B2



A3



B3

Fig. 6. Scanning electron microscopy of custom-made implants. (A1) Implant with machined surface $\times 40$ mag. (A2) Grooved pattern on machined surface $\times 100$ mag. (A3) smooth turned surface $\times 1000$ mag. (B1) Implant with RBM surface. It has coarse roughness $\times 40$ mag. (B2) $\times 100$ mag. (B3) At higher magnification, the metal presents a uniform reticulation ($\times 1000$)

2. Resonance frequency analysis (RFA)

After initial implants installation, all implants of 4 groups were measured by connecting transducer to implant and orienting it parallel to long axis of implant. The mean RFA values were 68.9 for group A implants, 66.2 for group B implants, 70.2 for group C implants, 65.9 for group D implants. The mean RFA values six weeks after implant insertion were 70.8 ± 4.3 for group A implants, 71.8 ± 3.1 for group B implants, 70.9 ± 2.5 for group C implants, and 72.7

± 2.5 for group D implants. There were no statistical significant differences between groups.

3. Removal torque measurement

Six weeks after implant placement, the average removal torque was 49.99 ± 6.70 Ncm for group A implants, 51.15 ± 4.40 Ncm for group B implants, 50.78 ± 9.37 Ncm for group C implants, and 51.09 ± 4.69 Ncm for group D implants. The removal torque results are summarized in Table II. The torque

Table I. Mean surface roughness(in μm)

Group	Ra(SD)	Rq(SD)	Rz(SD)
Group A (Brånemark clone, turned)	0.26	0.31	1.13
Group B (Brånemark clone, RBM)	1.02	1.18	4.53
Group C (New design, turned)	0.29	0.34	1.24
Group D (New design, RBM)	1.12	1.26	4.36

Table II. Mean values of RFA after six weeks of healing time.

Group	Group A	Group B	Group C	Group D
Mean RFA	70.8	71.8	70.9	72.7
SD	4.3	3.1	2.5	2.5

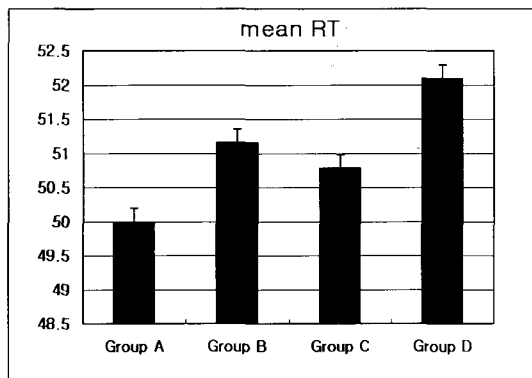


Fig. 8. Torque values for implants inserted in the tibia of rabbits 6 weeks (n=17)

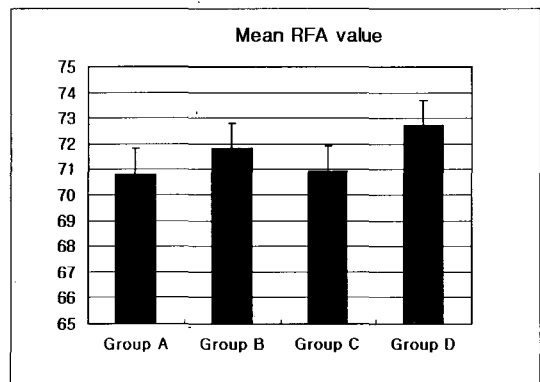


Fig. 7. Mean values of the resonance frequency measurements (Hz) after six weeks of healing time. (n=19) There were no statistically significant differences between the groups.

Table III. Torque values after 6 weeks (n= 17)

Group	Group A	Group B	Group C	Group D
Mean RT	49.99	51.15	50.78	52.09
SD	6.70	4.40	9.37	4.69

measurements yielded no statistical significance. The highest removal torque corresponded to the newly designed implant with RBM surface, while the lowest was demonstrated by the turned, Brånemark clone implants. Higher torque was needed to unscrew RBM surface implants compared to the torque needed for the turned, Brånemark clone implants.

4. Histomorphometric evaluation

A typical cross-section showed the triangular-shaped tibia and a central bone marrow cavity with the implant inserted through the superior cortex. On the microscope, all 8 implants were well inte-

grated into bone and a mature cortical bone surrounded the implants. The implants were in contact with cortical bone layer exclusively along the upper threads in the cortical region, while the threads in the bone marrow were in contact with much of mar-

Table IV. Percentage of bone-to-implant contact 6 weeks after Implant Placement

Rabbit No.	Group A	Group B	Group C	Group D
1	45	52	49	55
2	46	58	50	60
Mean	45.5	55	49.5	57.5
SD	0.5	3	0.5	2.5

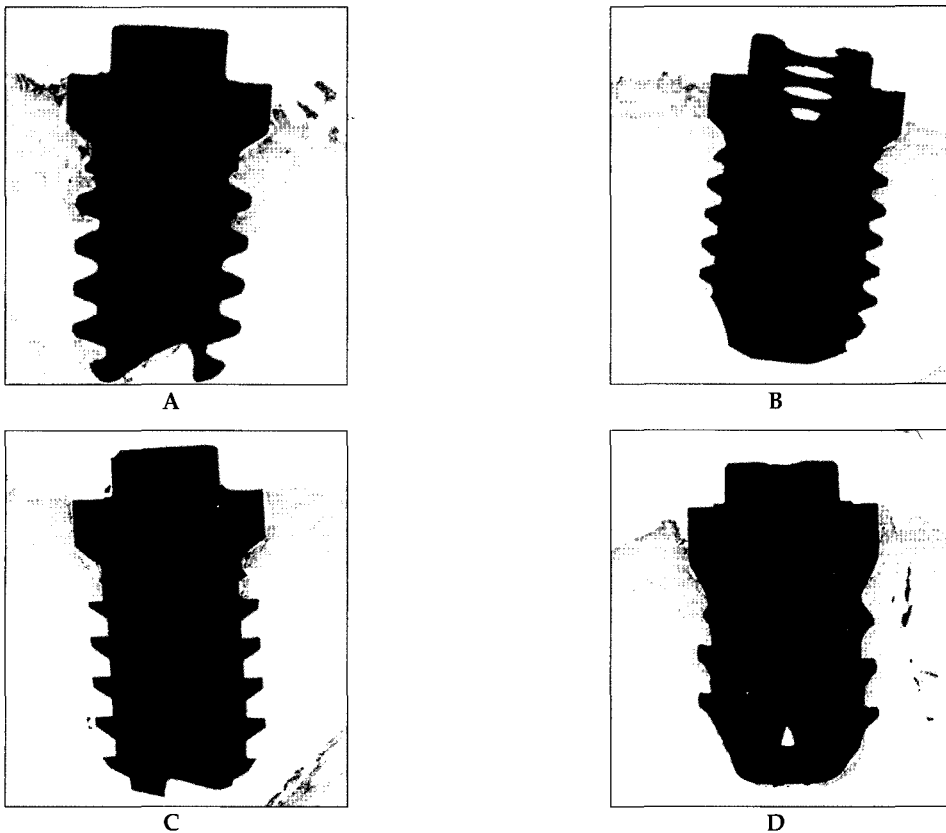


Fig. 9. 25micron ground sections of 4 implants from each group. All implants were well integrated into bone. (A) Turned Brånemark clone implant as Group A, (B) Brånemark clone implant with RBM surface as Group B, (C) Newly designed implant with turned surface as Group C, (D) Newly designed implant with RBM surface as Group D. Note newly formed bone in the marrow space. Note the bone formed at the implants body.

row tissue. Some new bone formation was observed only newly designed with RBM surface in contact with marrow space. Qualitative histologic differences among the remaining 3 groups were not seen.

Results showed a mean percentage of bone-to-implant contact was $45 \pm 0.5\%$ for the turned Brånemark clone implants, $55 \pm 3\%$ for the Brånemark clone implants with RBM surface, $49.5 \pm 0.5\%$ for the turned newly designed implants, and $57.5 \pm 2.5\%$ for the newly designed implants with RBM surface. The histomorphometric analyses were summarized in Table IV.

DISCUSSION

The quality of the implant surface is one of the six factors described by Albrektsson¹³ et al which influence the healing at the implantation site and subsequently affect the osseointegration. The roughness of the implant surfaces favors distribution of stress, retention of implants in the bone, and bone response with bone trabeculae growing in a perpendicular direction to implant surface.^{20,21} Studies have reported that adequate growth of bone in the interior of the pores or cavities left by the surface treatment requires that these must be approximately $100\mu\text{m}$ in size. The growth of bone tissue into cavities of this size allows a mechanical interlocking of the implants with bone.²² Wong et al.²⁰ have reported that bone matrix may be deposited in pore sizes of only $1\text{-}2\mu\text{m}$, which resulted in an increase in push-out force. Analysis of RBM implants using scanning electron microscopy revealed rough surface and pore size of $(2.5\text{-}4\mu\text{m})$. This allowed bone growth into these pores so that mechanical interlocking of the implant with bone was possible and resulted in an improved bone-implant interface as described by other articles.²³⁻²⁷

Implants with RBM surface had higher value than control on RFA measurement and histomorphometric evaluation. These implants also achieved higher degree of bone-to-implant contact compared

to turned implants. And newly designed implant with RBM surface had new bone formation around bone marrow space contrary to implants of other groups. This result is thought to arise from the novel design and the RBM surface. In vitro studies have also shown that the superficial roughness of the materials can influence cell function, matrix deposition, and mineralization.²⁸⁻³⁰ In a series of studies, Wennerberg et al, systematically investigated the effect of surface roughness of implants and the response in rabbit bone.³¹⁻³⁴ And the authors concluded that implants with a surface roughness of Ra 1 to $1.5\mu\text{m}$ seemed to be at an optimal roughness with regard to retention in bone as well as bone-to-implant contact as measured by histomorphometry. And this observation is also in accordance with observations by von Recum & van Kooten³⁵ that reported excellent tissue attachment without signs of inflammation when implanting filter membranes with pore size of $1\text{-}3\mu\text{m}$. Based on these results, it seems that the benefit of increasing roughness on a micrometer scale reaches a maximum level between 1.0 to $1.5\mu\text{m}$. The Ra value of RBM treated implants was within this limits seemed to have resulted in favorable bone response.

Irrespective of whether resonance frequency analysis (RFA) or removal torque (RT) was used, the biomechanical data showed the similar results. Newly designed implant with RBM surface implants had the highest value on both experiments but not much statistically significant difference. Both new design and surface characteristics of the implant enabled it. The newly designed of dental implant with half-rounded shape at the apex, flatness at the top of the threaded surface, and 45 degrees of reverse bevel at the bottom surface set the stress to the apex of the dental implant during lateral or oblique load. Forcing the stress to the top part rather than the apex part of the dental implant is not highly recommended, because there is much accumulation of stress at the top part of the dental implant. The cortical bone which integrate with the top part of the dental implant is unable to disperse the attained stress

causing the concentration of stresses at the referred area. The dental implants, which took part in the experiment, are much capable of dispersing the stress especially to the direction of apex compared with general threaded types or other models of dental implant.³⁰

RT and RFA are well-documented biomechanical techniques that evaluate the stability and stiffness of the osseointegrated implant interface. Removal torque evaluated the biomechanical bond between implant and the bone. Several studies have reported that RT techniques offer reliable test-to-control comparisons.^{12,36} RFA is a recently introduced non-invasive test method and this technique has been utilized in a number of experimental tests to record a change in implant stiffness.^{17,31,37-40} RT and RFA are quantitative method for assessment of implant stability and osseointegration and also enable us to compare different implant system, such as changes in implant geometry or surface modifications.

CONCLUSION

1. The surface of RBM implants as seen with SEM had a rough, irregular pattern than that of the turned implants that seemed to have a smoother surface. At higher magnification, the RBM surface showed reticular pattern.
2. Ra values for machined implants were $0.2-0.3\mu\text{m}$; corresponding values for newly designed implants with RBM surface were $1.0-1.2\mu\text{m}$.
3. The average removal torque was $44.99 \pm 6.70\text{Ncm}$ for group A implants, $51.15 \pm 4.69\text{Ncm}$ for group B implants, $50.78 \pm 9.37\text{Ncm}$ for group C implants, $51.15 \pm 4.40\text{Ncm}$ for group D implants. There was no statistically significant difference.
4. The mean RFA values were 70.8 for group A implants, 71.8 for group B implants, 70.9 for group C implants, 72.7 for group D implants. There was no statistically significant difference.
5. A mean percentage of bone-to-implant contact was $45 \pm 0.5\%$ for group A implants, $55 \pm 3\%$ for

group B implants, $49.5 \pm 0.5\%$ for group C implants, $57.5 \pm 2.5\%$ for group D implants. The light microscopic picture demonstrated implants with RBM surface had higher values with respect to bone-to-implant contact and group D implants showed new bone formation around implant body.

Within the limits of this 6-week experimental study, it may be stated that the overall pattern of newly designed implants with RBM surface resulted in significantly higher percentage of bone-to-implant contact and comparable removal torque and resonance frequency measurement when compared to machined, Brånemark clone implants. And the present investigation revealed the fact that RBM surface treatment has some advantage over turned surface in bone-to-implant contact, removal torque and resonance frequency measurement.

REFERENCES

1. Zarb GA, Albrektsson T. Osseointegration : a requiem for the periodontal ligament? *Int J Periodontics Restorative Dent* 1991;11:88-91.
2. Albrektsson T, Sennerby L. State of the art in oral implants. *Journal of Clinical Periodontology* 1990;18, 474-481.
3. Brånemark PI. Osseointegration and its experimental background. *J Prosthet Dent* 1983;50: 399-410.
4. Albrektsson T, Hansson HA, Ivarsson B. Interface analysis of titanium and zirconium bone implants. *Biomaterials* 1985;6:97-101.
5. Brånemark P.I, Hansson BO, Adell R . Osseointegrated implants in the treatment of the edentulous Jaw. Experience from a 10 year period. *Scand J Plast Reconstr Surg* 1997;16.
6. Adell R, Lekholm U, Rockler B, Brånemark PI. A 15-year study of osseointegrated implants in the treatment of the edentulous jaw. *J Oral Surg* 1981;10:387-416.
7. Adell R, Eriksson B, Lekholm U, Brånemark PI, Jemt T. A long-term follow-up study of osseointegrated implants in the treatment of totally edentulous jaws. *Int J Oral Maxillofac Implants* 1990;5:347-359.
8. Lekholm U, van Steenberghe D, Herrmann I, Bolender C, Folmer T, Gunne J, et al. Osseointegrated implants in the treatment of partially edentulous jaws: A prospective 5-year multicenter study. *Int J Oral Maxillofac Implants* 1994;9:627-635.

9. Adell R. Clinical results of osseointegrated implants supporting fixed prosthesis in edentulous jaw. *Jprosthet Dent* 1983; 50:251-4.
10. Adell R, Lekholm U, Rockler B, Brånemark PI, Linddhe J, Eriksson B, et al. Marginal tissue reactions at osseointegrated titanium fixtures I. A three-year longitudinal prospective study. *Int J Oral Surg* 1986;15:39-52.
11. Albrektsson B, Zarb G, Worthington P, Eriksson A. The long-term efficacy of currently used dental implants. A review and proposed criteria for success. *Int J Oral Maxillofac Implants* 1986;1:11-25.
12. Carlsson L, Rostlund T, Albrektsson B, Albrektsson T. Removal torque for polished and rough titanium implants. *Int J Oral Maxillofac Implants* 1988;3:21-24.
13. Albrektsson T, Brånemark P.I, Hansson HA, Lindstrom J. Osseointegrated titanium implants. *Acta Orthop Scand* 1981;52:155-170.
14. Rasmusson L, Meredith N, Cho I.H, Sennerby L. The influence of simultaneous versus delayed placement on the stability of titanium implants in onlay bone grafts. *Int J of Oral Maxillofac Surg* 1999;28:224-231.
15. Meredith N, Alleyne D, Cawley P. Qualitative determination of the stability of the implant-tissue interface using resonance frequency analysis. *Clin Oral Implants Res* 1996; 7:261-269.
16. Meredith N. On the clinical measurement of implant stability and osseointegration. Göteborg, Sweden: Department of Biomaterials/Handicap Research, University of Göteborg. Ph D Thesis 1997.
17. Meredith N, Shagaldi F, Alleyne D, Sennerby L, Cawley P. The application of resonance frequency measurements to study the stability of titanium implants during healing in the rabbit tibia. *Clin Oral Imp Res* 1997;8:234-243.
18. Robert RW, Smith RK, Zilberman Y, Mozsary PG, Smith R: Osseous adaptation to continuous loading of rigid endosseous implants. *Am J Orthod* 1984;86:95-111.
19. Donath K. Die Trenn-Dünnschliff-Technik zur Herstellung histologischer Präparaten von nicht schneidbaren Geweben und Materialien. *Der Praktischer* 1988;34:197-206.
20. Buser D, Schenk R, Steinemann S. Influence of surface characteristics on bone integration of titanium implants: A histomorphogenic study in miniature pigs. *J Biomed Mater Res.* 1991;25:889-902.
21. Wong M, Eulenberger J, Schenk R. Effect of surface topology on the osseointegration of implant materials in trabecular bone. *J Biomed Mater Res.* 1995;29:1567-1575
20. Johansson C.B, Albrektsson T. Integration of screw implants in the rabbit. A 1-year follow-up of removal torque of Ti implants. *Int J oral Maxillofac Implants* 1986;2:69-75.
21. Carlsson L, Röstlund T, Albrektsson B, Albrektsson T. Removal torques for polished and rough titanium implants. *Int J oral Maxillofac Implants* 1988;3:21-24.
22. Boby J, Pilliar R, Cameron H. The optimum pore size for the fixation of porous-surface metal implants by the ingrowth of bone. *Clin Orthop.* 1980;150:263-270.
23. Ericsson I, Johansson CB, Bystedt H et al. A histomorphometric evaluation of implant contact on machined-prepared and roughened titanium dental implants. *Clinical Oral Implant Res.* 1994;5:202-206.
24. Gottfredsen K, Nimb L, Hörting-Hansen E. Histomorphometric and removal torque analysis for TiO₂-blasted titanium implants. An experimental study in dog. *Clin Oral Implants Res.* 1992;3:77-84.
25. Martin JY, Schwartz, Hummer TW. Effect of titanium surface roughness on proliferation, differentiation and protein synthesis of human osteoblast-like cells(MG63). *J Biomater Res.* 1995;29:389-401.
26. Breme J, Wadewitz V, Fürbacher B. Production and mechanical properties of porous sintered specimens of the implant alloy TiAl5Fe2. *Advances in Biomaterials.* 1990;9:63-68.
27. Martin J, Schwartz Z, Hummert T. Effect of titanium surface roughness on proliferation, differentiation and protein synthesis of human osteoblast-like cells. *J Biomed Mater Res.* 1995;29:389-401.
28. Brunette D. The effects of implant surface topography on the behavior of cells. *Int J Oral Maxillofac Implants.* 1988;3:231-246.
29. Boyan BD, Hummert T, Kieswetter K. Role of material surfaces in regulating bone and cartilage cell response. *Biomaterials.* 1996;17:137-146.
30. Kim Y.S. 3-dimensional finite element study on surface characteristics and stress of implants. SNUDH Dept of Prosthodontics. Unpublished.
31. Wennerberg A, Albrektsson T, Andersson B, Krol JJ. A histomorphometric and removal torque study of screw-shaped titanium implants with three different surface topographies. *Clin Oral Implants Res* 1995;6:24-30.
32. Wennerberg A, Albrektsson T, Lausmaa J. Torque and histomorphometric evaluation of c.p.titanium screws blasted with 25- and 75-micron-sized particles of Al₂O₃. *J Biomed Mater Res* 1996;30:251-260.
33. Wennerberg A, Albrektsson T, Andersson B. Bone tissue response to commercially pure titanium implants blasted with fine and coarse particles of aluminum oxide. *Int J Oral Maxillofac Implants* 1996;11:38-45.
34. Wennerberg A, Albrektsson T, Johansson C, Andersson B. Experimental study of turned and grit-blasted screw-shaped implants with special emphasis on effects of blasting materials and surface topography. *Biomaterials* 1996;17:15-22.
35. Von Recum AF, van Kooten TG. The influence of microtopography on cellular response and the implications for silicone implants. *J Biomater Sci*

- polymer Edition 1995;7:181-198.
36. Rasmusson L, Meredith N, Sennerby L. Measurements of stability changes of titanium implants with exposed threads subjected to barrier membrane induced bone augmentation. A study in the rabbit tibia. *Clin oral Imp Res* 1997;8:103-116.
 37. Rasmusson L, Meredith N, Kahnberg K.E, Sennerby L. Stability assessments and histology of titanium implants placed simultaneously with autogenous onlay bone grafts in the rabbit tibia. *Int J Oral Maxillofac Surg* 1998;27:229-235.
 38. Park C. A study on the measurement of the implant stability using resonance frequency analysis. PhD thesis, 2000, Dankook University, Korea.
 39. Park CJ. In vitro comparative study between ISQ and Periostest® values on the implant stability measurements according to the increased effective implant length. *J Korean Academy of Prosthodontics* 2001;39:625-635.
 40. Park CJ. A study on the change of implant stability using resonance frequency analysis. Ph D thesis(Manuscript). 2000, Seoul National University, Korea.

Reprint request to:

DR. YUNG-SOO KIM, D.D.S., M.S.D., Ph.D.

DEPT. OF PROSTHODONTICS, COLLEGE OF DENTISTRY,

SEOUL NATIONAL UNIV.

28-1 YEONGJUN-DONG, CHONGNO-GU, 110-749, SEOUL KOREA

yungskim@snu.ac.kr