Expression of Id-1 Gene in Mouse Uterus

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생쥐 자궁에서의 Id-1 유전자의 발현

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연구 목적: Microarray data Id hormonal effect implantation process .

연구재료 및 방법: estrogen 6 , 12 sample . RNA laser capture microdissection (LCM) , , RNA semi-quantative RT-PCR Id . 4.5 Chicago blue dye RNA Id semi-quantitative RT-PCR .

결 과: Estrogen cDNA microarray Id-1 mRNA Id-2 mNRA . Microarray semi-quantitative RT-PCR , Id-1 estrogen 6 12 4 , Id-2 mRNA estrogen 6 12 4 . LCM , , estrogen Id-1 , estrogen 6 12 . , Id-2 mRNA estrogen 6, 12 , estrogen 6 12 . , estrogen 6, 12 Id-2 mRNA . Id-1 mNRA .

결 론: Id-1, -3 estrogen , Id-2 . LCM Id-2 Id-2

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.   Id   Id-1

                      Id   estrogen   ,

                                  .

Steroid hormone actions in target tissues are normally mediated via binding to nuclear receptors that are ligand-inducible transcription factors.[1,8,15] The uterus is composed of heterogenous cell types that undergo dynamic changes to support embryo development and implantation by harmony of ovarian steroid hormones. Implantation is a complex process involving various molecular "cross-talk" between the blastocyst and the uterus. The sites of blastocyst attachment in uterus is visualized by increased stromal vascular permeability and undergo morphological and biochemical changes to receive the blastocyst under the control of ovarian steroid hormone.[7,19,20] These changes suggest that cell proliferation and differentiation at this sites is associated with up- and down-regulation of a many of genes, which are poorly understood. We have recently performed cDNA microarray to screen estrogen-responsive genes in the mouse uterus and so shown that Id genes were differentially expressed in the estrogen-treated ovariectomized mouse uterus. Id, inhibitors of basic helix-loop-helix (bHLH), are positive and negative regulators of proliferation, differentiation, cell cycle, apoptosis and angiogenensis.[3,14] Four members, Id1 through Id4, have been identified in mammals.[2,4,17,18]

In the present study, we examined the expression patterns of Id genes according to uterine specific cell type and the difference between implantation sites and interimplantation sites using laser capture microdissection (LCM) and RT-PCR.

MATERIALS AND METHODS

1. Animals and Uterus preparation

ICR mice were housed within temperature- and light-controlled conditions under the supervision of a licenced veterinarian. Mice were maintained on a 12L:12D photoperiod and provided with food and water ad libitum. Female mice (6~ 7 weeks of age) were ovariectomized (OVX) and rested for 14 days before receiving estrogen treatment. They were injected with oil (0.1 ml) and 17β-estradiol (Sigma, St. Louis, MO; 300 ng/mouse) which was dissolved in sesame oil and injected (0.1 ml/mouse) subcutaneously. Mice (n=9) were killed and uterine horns were collected at 6 h or 12 h after injection.

To obtain pregnant mice, adult female mice (6~ 8 weeks old) were placed with fertile males of the same strain, and the day that a vaginal plug was found was considered as day 1 of pregnancy. On the evening of day 4 (22:00~ 23:00) at the time of blastocyst attachment, implantation sites were visualized by intravenous injection (tail vein) of Chicago Blue B solution (1% in saline) 10 min before killing the mice. Implantation segments containing implanting embryo were finely separated from non-implantation segments, and both segments were then snap-frozen in liquid nitrogen until utilized for total RNA extraction. All animal experiments were performed in accordance with the Guide of Ulsan University for Care and Use of Laboratory Animals.

2. cDNA microarray and Data analysis

Uterine tissues from nine female mice were pooled, snap frozen and homogenized by mortar in liquid nitrogen. For cDNA microarray analysis, total RNA was extracted using TRIZOL reagent (InVitrogen, Carlsbad, CA) and purified using RNeasy total RNA isolation kit (Qiagen, Valencia,

CA) according to the manufacturers instructions. Total RNA was quantified by spectrophotometer and its integrity was assessed by running on a denaturing 0.8% agarose gel. Prior to use in cDNA microarray analysis, each RNA sample via OVX/ estrogen treatment/6 h/12 h protocol was validated by assaying for up-regulation (asparagine synthetase and lactoferrin) or down-regulation genes (glutathione S-transferase and secreted frizzled-related sequence protein 2) as markers of estrogen efficacy. Profiling of estrogen-regulated gene expression was analyzed with a TwinChip Mouse-7.4 K (Digital Genomics, Seoul) consisting of 7616 mouse cDNA clones. Twenty μg RNA was reverse-transcribed with Cy3- or Cy5-conjugated dUTP (Amersham Pharmacia Biotech, Piscataway, NJ) respectively, using SuperScript II (Gibco BRL, Rockville, MD) and oligo(dT)$_{18}$ primer (Ambion, Austin, TX) in a reaction volume of 20 μl according to the method suggested by the manufacturer. After the labeling reaction for 1 h at 42 , unincorporated fluorescent nucleotide was cleaned up using Microcon YM-30 column (Millipore, Bedford, MA). The Cy3- and Cy5-labeled cDNA probes were mixed together and hybridized to a microarray slide. After overnight at 65 , the slide was washed twice with 2× SSC containing 0.1% SDS for 5 min at 42 , once with 0.1× SSC containing 0.1% SDS for 10 min at room temperature, and finally with 0.1× SSC for 1 min at room temperature. Slide was dried by centrifugation at 650 rpm for 5 min. Hybridization images on the slide were scanned by Scanarray lite (Packard Bioscience, Boston, MA) and analyzed by GenePix Pro3.0 software (Axon Instrument, Union City, CA). Three independent experiments were performed, and the ratio of Cy3 and Cy5 signal intensity was calculated for each spot. These ratio was log$_2$-transformed and normalized by subtracting the average of log$_2$(Cy3/Cy5) values for internal control genes using Excel (Office 2000, Microsoft Corp.). For each gene, the mean values

were then calculated and a difference of two-fold was applied to select up- or down-regulated genes by estrogen.

3. Laser capture microdissection

Uterine horns were embedded in Tissue-Tek (Sakura Finetek, Torrance, CA) and frozen in liquid nitrogen. Cryosections (thickness, 6 μm) were cut and mounted onto clean glass slides. After the sections were conterstained with Mayers hematoxylin, each population of uterine cells (luminal epithelial, muscle and stromal cells) was isolated from these sections using P.A.L.M. Robot-Microbeam version 4.0 (P.A.L.M. Microlaser Technologies AG, Bernried, Germany). For each cell, an average of 150 laser shots were transferred onto 0.5 ml tube cap and stored at -70 until utilized for total RNA extraction.

4. Confirmation of microarray data with semiquantitative RT-PCR analysis

Total RNA was extracted from whole uterine tissues or each population of uterine cells using TRIZOL reagent and purified using RNeasy total RNA isolation kit following the manufacturers instructions. One μg of RNA was reverse-transcribed at 42 for 60 min in 20 μl reaction mixture consisting of oligo(dT)-adapter primer (Takara, Shiga, Japan) and AMV reverse transcriptase XL (Takara). The following PCR was performed in a total volume of 40 μl with 2 μl of the RT reaction mixture, 2 μl 25 mM MgCl$_2$, 4 μl 10× PCR buffer (100 mM Tris-HCl, 500 mM KCl, pH 8.3), 4 μl 2.5 mM dNTPs, 10 pmole forward and reverse primer and 1.25 U Taq polymerase (Takara). The sequences of the primers used were described in Table 1. An increasing number of cycles was tested to assess the best conditions of achieve linear amplification. The thermal cycling parameters were consisted of 22~26 cycles of denaturing (94 , 30 sec); annealing (60 , 30 sec); and extension

Table 1. Primer sequences for RT-PCR

Gene		Forward (F) and Reverse (R) primer sequences	Product size (bp)
Id-1	F	GATCATGAAGGTCGCCAGTG	703
	R	TCCATCTGGTCCTCAGTGC	
Id-2	F	GTGACCAAGATGGAAATCCT	523
	R	TTTATTTAGCCACAGAGTAC	
Id-3	F	GAAGGCGCTGAGCCCGGTGC	383
	R	GTTCGGGAGGTGCCAGGACG	
Id-4	F	GCGATATGAACGACTGCTAC	616
	R	TCACCCTGCTTGTTCACGGC	
HB-EGF	F	CTCCTCCAAGCCACAAGGCC	389
	R	CTAGCCACGCCCAACTTCAC	
rpL7	F	TCAATGGAGTAAGCCCAAAG	246
	R	CAAGAGACCGAGCAATCAAG	

Table 2. Change of Id genes by estrogen in ovariectomized mouse uterus

| Gene name	Fold change		Genbank No.
	6 H	12 H	
Inhibitor of DNA binding 1 (Id-1)	1.23	2.01	U43884
Inhibitor of DNA binding 2 (Id-2)	-1.26	-2.26	AF077860
Inhibitor of DNA binding 3 (Id-3)	-1.48	-1.69	NM008321
Inhibitor of DNA binding 4 (Id-4)	1.17	1.07	AF077859

(72 , 30 sec). The PCR products were separated by electrophoresis on a 1.2% TBE agarose-ethidium bromide gels and visualized under UV light. The images were quantified by densitometric scanning followed by BioID image analysis software (Vilber-Lourmat, Mama La Vallee, Cedex, France) and gene expression was normalized against the density of the corresponding ribosomal protein L-7 (rpL7) PCR product as internal control.

RESULTS

1. Changes of Id genes by estrogen in ovariectomized mouse uterus

We have performed cDNA microarray to screen estrogen-regulated genes in the mouse uterus (data not shown) and then unexpectedly, Id genes were showed differential expression pattern (Table 2). In particular, the expression level of Id-1 mRNA was increased in proportion as estrogen-treated time in the estrogen-induced uterus and Id-2 decreased. Although there is no significance, the level of Id-3 mRNA was decreased in the estrogen-induced uterus. However, the expression of Id-4 gene made a no difference between the groups. To confirm microarray data, the expression patterns of Id-1 and -2 using semi-quantitative RT-PCR were shown in Figure 1. All groups were mixed with five mice to each other. The RT-PCR data was displayed just Id-1 and -2 which were detected signi-
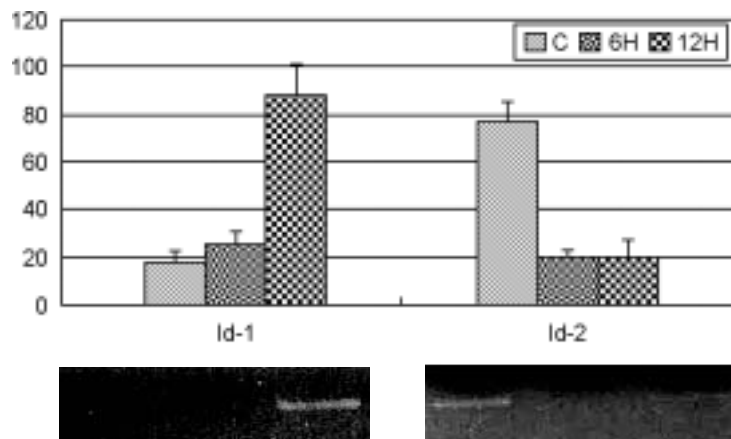
Figure 1. Semiquantitative RT-PCR analysis confirming Id-1 and -2 expression level in cDNA microarray result. Values of each band were normalized to rpL7 for the same sample. Data were mean± SEM from three replicate experiments.
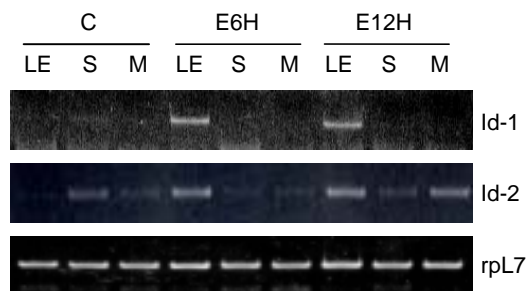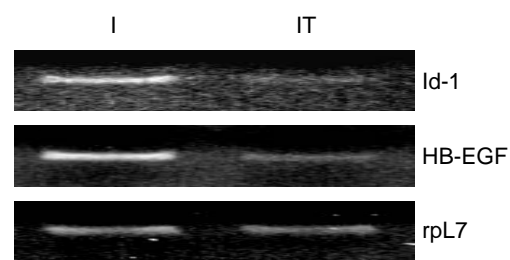

Figure 2. Expression patterns of Id-1 and -2 mRNA in uterine specific cell types obtained with LCM. C, oil-treated group; E6H, post-estrogen 6 h; E12H, post-estrogen 12 h; LE, luminal epithelial cells; S, stromal cells; M, muscle cells.

Figure 3. RT-PCR analysis of Id-1 mRNA obtained from implantation (I) and interimplantation (IT) sites of pregnancy day 4 night. The expression level of HB-EFG mRNA was examined as marker gene.

ficantly difference. After injection of estrogen, the level of Id-1 mRNA were few difference at 6 h, increased more than four-fold at 12 h. In contrast, the expression of Id-2 mRNA was remarkably decreased than control both 6 h and 12 h after injection of estrogen. In comparison with Id-1 and Id-2, Id-2 mRNA was rapidly responded to estrogen, but Id-1 was relatively showed slow response to estrogen.

2. Differential Expression of Id genes in the uterine specific cell types

Id genes were displayed differential expression according to cell type in the estrogen-treated OVX mouse uterus (Figure 2). Surprisingly, Id-1 gene was showed the specific expression in luminal epithelial cell. However, the expression of Id-2 mRNA in the uterine specific cell types was showed different pattern in contrast with the whole uterus data.

Although the level of Id-2 mRNA in the stromal cells was similar to the whole uterus data, expression of Id-2 in the luminal epithelial cells and muscle cells were the very reverse to the data. In the mouse uterus, stromal cells exist broadly compared with epithelial cells. Therefore, these results suggest that low expression of Id-2 mRNA

in the broad stromal cell may be compensates for highly expression in the epithelial cell.

3. Expression patterns of Id-1 in the Implantation and the Interimplantation Sites

We compared the mRNA expression pattern of implantation and interimplantation sites on day 4.5 pregnant (vaginal plug = day 1) in mouse using RT-PCR. Total RNA was isolated in individual uterine tissues obtained from sixteen female mice on day 4.5 pregnant and submitted to RT-PCR amplification (Figure 3). The expression level of Id-2 mRNA (0.89± 1.79) was shown to make a slightly difference between the two. Id-4 mRNA was almost never detected. However, the expression levels of Id-1 and -3 mRNA (1.96± 0.75 and 1.83± 0.77) were shown to upregulate at the sites of implantation in all mice and similar to that of heparin-binding EGF-like growth factor (HB-EGF, 2.05± 0.62). The amplification of the rpL7 mRNA was served as an internal control in the RT-PCR.

DISCUSSION

In the mouse, the main distinguished signs of blastocyst attachment sites are an angiogenesis and apoptosis. With regard to the angiogenesis, recent work has shown that the expression of Id-1 and 3 is required for the proper angiogenesis in the neuroectoderm during embryo development and particularly the partial reduction of Id-1 dosage also results in an angiogenic defect in adult mice which blocks the vascularization of tumour xenografts.[13] More interestingly, the thickening of the extracellular matrix surrounding endothelial cells in Id knockout mice was observed and thus Id expression may regulate the expression of integrin and metalloproteinase MMP2 which are required for tumour angiogenesis. VEGF system is important for uterine vascular permeability and angiogenesis during implantation. Others have also shown the expression of VEGF and its receptors in the uterus as a whole during pregnancy and in response to steroid hormones.[10] Estrogen rapidly stimulates uterine vascular permeability and VEGF expression, and because vascular permeability is considered a prerequisite for angiogenesis. It is widely believed that estrogen is potent stimulator of uterine angiogenesis during normal reproductive process in vivo.[10,11] However, The role for estrogen in uterus angiogenesis are still unclear.

Id-1 mediates the effects of sex steroid hormones on T47D cells, particularly on hormonal control of cell proliferation. Estrogen stimulates Id-1 expression and activates cell proliferation. Progesterone represses Id-1 expression and inhibits cell growth.[5] Loss of Id function leads to a decrease in VEGF expression.[13] Mobilization of CFPs is regulated by VEGF/VEGFR2 signaling pathway and also proteins of Id-1 and -3 is controlled the regulation of mobilization of CEPs. Perhaps the most unsuspected activity of Id proteins has been their essential role in angiogenesis, both in the forebrain during development and during the growth and metastasis of tumors in adults.[2] In the mammary glands, Id-1 mRNA is highly expressed in the developing gland and also during involution, a phase of extensive remodeling characterized by degradation of ECM and epithelial cell death by apoptosis.[12,16] Some of the activities of Id-2, such as induction of apoptosis, have been shown to reside in the N-terminal domain and are independent of HLH-mediated dimerization.[9] Implantation process is similar to tumor that (which) angiogenesis and apoptosis act in the capacity of key feature.

In this present study, we hypothesized that Id genes under the estrogenic control might be important in early implantation process, in specially related angiogenesis. The expression patterns of Id family were analyzed in uterine tissues obtained from sixteen pregnant mice considering the variation in each mouse by semiquantitative RT-PCR.

These previous data provide the possibility that the steroid-regulated and site-restricted expression of Id mRNAs is contributed to the differential expression between the implantation and the interimplantation sites in each mouse. Id-1 and -3 was showed the differential expression up-regulated in the implantation sites compare to the interimplantaiton sites and the expression levels of them was similar to that of HB-EGF.

The expression level of HB-EGF mRNA was examined as control because HB-EGF gene was demonstrated to induced in the mouse uterine luminal epithelium temporally by the blastocyst soley at the site of its attachment on day 4 of pregnancy.[6]

In addition, estrogenic effect of Id genes was revealed it using estrogen-treated OVX mouse model. After estrogen injection, Id-1 was shown to gradually up-regulation and Id-2 presented opposite results.

These results strongly suggest that endometrial angiogenesis during implantation period may be directly or indirectly regulated by Id expression under the control of ovarian steroid hormone.

The observations presented in the current study implicate that Id genes may have a distinct function in uterine physiological events, such as implantation process and estrous cycle, contrary to the hypothesis that Id genes have redundant functions and may play a critical role for blastocyst attachment or angiogenesis at implantation process. However, we do not know the molecular mechanisms which regulate the expression of Id genes between the uterus and the blastocyst and how steroid hormone might regulate the expression of Id genes is not clear.

Therefore, further work is needed to localize Id mRNA and proteins in uterus, to define factors which control expression of Id proteins and to identify target genes whose expression is regulated by Id at the site of blastocyst attachment in implantation process.

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