

Indolyl Alkaloid Derivatives, N_b -Acetyltryptamine and Oxaline from a Marine-Derived Fungus

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Indolyl alkaloids, N_b -acetyltryptamine (1) and the known oxaline (2) have been isolated from the organic extract of the broth of an unidentified fungus collected from the surface of the marine red alga *Gracilaria verrucosa*. The structure of N_b-acetyltryptamine (1) was assigned on the basis of comprehensive spectroscopic analyses.

Key words: Marine fungus, Indolyl alkaloid, N_b-Acetyltryptamine, Oxaline, Gracilaria verrucosa

INT RODUCTION

Marir e microorganisms are receiving increasing attention as sources of bioactive compounds, and expanded research can be expected in this area. Marine natural product research is now focusing more on marine microorganisms, mairly pacteria and fungi that can be cultured (Faulkner, 2002; Fietra, 1997). The marine fungi, particularly those associated with marine animals and plants, appear to be an unusually rich resource for secondary metabolites. As part of a program to explore the bioactive metabolites produced by fungi isolated from marine habitats (Son et al., 2002), we investigated the chemical constituents of an unidentified marine algicolous fungus, which was separated from the red alga Gracilaria verrucosa, and isolated indol/l alkaloids, N_b -acetyltryptamine (1) and the known oxaline (2) (Nagel et al., 1976). This paper deals with the structure elucidation of these alkaloids having an indolyl moie:y.

MATERIALS AND METHODS

General experimental

IR spectrum was recorded on a Bruker FT-IR model IFS-88 spectrometer. ¹H (400 MHz) and ¹³C NMR (100 MHz) specira were obtained on a JEOL JNM-ECP 400 NMR spectrometer, using TMS or solvent peaks as reference standard. MS spectra were obtained on a JEOL JMS-700 spectrometer. UV/visible spectra were measured on a Hitachi U-2001 UV/Vis spectrometer.

Fungal isolation and culture

An unidentified fungal strain (culture # MFA 936) was isolated from the surface of the marine red alga Gracilaria verrucosa collected in Hamdeok Beach, Cheju Island in 2001. The fungus was cultured (20 L) for 30 days (static) at 29°C in SWS medium : soytone (0.1%), soluble starch (1.0%), and seawater (100%).

Isolation of N_b -acetyltryptamine (1) and oxaline (2)

The mycelium and broth were separated by filtration. The broth was extracted twice with EtOAc. The combined extract (1.9 g) was subjected to silica gel flash column chromatography, eluting with n-hexane/EtOAc (from 100% to 0%), to obtain 8 fractions. Further purification of fraction 7 (400 mg) by silica gel column chromatography using CH₂Cl₂ / MeOH (20:1), followed by HPLC (YMC ODS-A, $10\times250 \text{ mm}$) (MeOH) yielded a N_b -acetyltryptamine (1, 13 mg) and oxaline (2, 6.0 mg).

1 : Yellowish oil; IR (neat): 3400 (NH), 1638 (amide), 1384, 744 cm⁻¹; UV (MeOH): 222 (log ε 4.0), 283 (2.3) nm; HREIMS m/z 202.1060 (calcd for $C_{12}H_{14}N_2O$, 202.1106); LREIMS m/z 202[M]⁺ (rel. int., 7), 154 (47), 143 (62), 130 (58), 86 (18), 70 (100).

See Table I for NMR spectral data.

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Table I. 1 H (δ , mult, J) and	13 C (δ , mult) NMR data of N_b -
acetyltryptamine (1) ^a	

Carbon No.	δ_{H}	$\delta_{ extsf{C}}$	HMBC (H to C)
1	8.14 (br, s)		
2	7.04 (d, 2.2)	122.0 (d)	8, 9
3		113.1 (s)	
4	7.60 (d, 8.0)	118.7 (d)	9 ,11
5	7.13 (dd, 8.0 8.0)	119.5 (d)	7, 9
6	7.21 (dd, 8.0 8.0)	122.2 (d)	4, 8
7	7.38 (d, 8.0)	111.2 (d)	5, 9
8		136.4 (s)	
9		127.3 (s)	
10	2.98 (t, 6.5)	25.3 (t)	2, 3, 9, 11
11	3.60 (dt, 6.5, 6.0)	39.8 (t)	3, 10, 13
12	5.53 (br, s)		
13		170.0 (s)	
14	1.92 (s)	23.4 (q)	11, 13

^aRecorded in CDCl₃ at 400 MHz (¹H) and 100 MHz (¹³C).

2: Yellowish oil; IR (neat): 3186 (NH), 3089, 3013, 1702 (δ-lactam), 1632 (amide) cm⁻¹; UV (MeOH): 208 (log ε 4.4), 228 (4.0), 284 (3.6), 344 (4.0), 360 (4.0) nm; HRFABMS m/z 448.1985 (calcd for C₂₄H₂₆N₅O₄, 448.5015); LRFABMS m/z 448[M+H]⁺ (rel. int., 100), 379 (34), 154 (12), 136 (23); ¹H NMR (400 MHz, CDCl₃) δ_{H} 7.58 (1H, d, J = 7.6 Hz, H-4), 7.09 (1H, dd, J = 7.6, 7.5 Hz, H-5), 7.29 (1H, dd, J =7.6, 7.5 Hz, H-6), 6.98 (1H, d, J = 7.6 Hz, H-7), 5.13 (1H, s, H-8), 12.88 (1H, br. s, 14-NH), 8.37 (1H, s, H-15), 7.58 (1H, br. s, H-18), 7.23 (1H, br. s, H-20), 6.11 (1H, br. s, H-22), 5.10 (1H, br. d, J = 18.3 Hz, H-23a), 5.06 (1H, br. d, J= 14.0 Hz, H-23b), 1.25 (3H, s, H_3 -24), 1.32 (3H, s, H_3 -25), 3.64 (3H, s, H₃-26), 3.73 (3H, s, H₃-27); ¹³C NMR (100 MHz, CDCl₃) δ_C 101.3 (C-2), 52.4 (C-3), 146.4 (C-3a), 124.7 (C-4), 123.4 (C-5), 128.5 (C-6), 112.1 (C-7), 146.5 (C-7a), 106.8 (C-8), 125.9 (C-9), 157.4 (C-10), 122.8 (C-12), 165.7 (C-13), 110.0 (C-15), 126.2 (C-16), 136.8 (C-18), 134.6 (C-20), 42.4 (C-21), 142.6 (C-22), 114.1 (C-23), 24.1 (C-24), 23.7 (C-25), 55.7 (C-26), 65.2 (C-27).

RESULTS AND DISCUSSION

 N_b -Acetyltryptamine (1) was isolated as a yellowish oil and was found to have an elemental composition $C_{12}H_{14}N_2O$ on the basis of HREIMS and ^{13}C NMR methods. The IR spectrum of 1 revealed absorption bands for amine (3400 cm⁻¹) and amide (1638 cm⁻¹). The ^{1}H and ^{13}C NMR data for 1, including results from DEPT, TOCSY, HMQC, and HMBC experiments, showed indol and ethylamine acetate moieties (Table I). These moieties were further supported

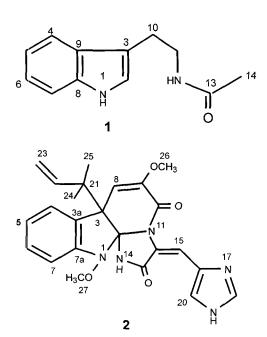


Fig. 1. Structures of N_b -acetyltryptamine (1) and oxaline (2)

by UV spectral data [222 (log ε 4.0), 283 (2.3) nm], and by MS fragment m/z 143 [M-CH₃CONH₂]⁺. The position of the ethylamino acetate group was deduced by HMBC correlations from H-4 to C-3, and H₂-10 to C-2, C-3 and C-9, and by TOCSY correlations between H-2 and H-1 and H₂-10, and between H₂-10 and H-2, H₂-11 and H-12.

Based on all of the foregoing evidence, the structure of N_b -acetyltryptamine was determined to be 2-(3-indolyl) ethylamine acetate (1) (Fig. 1). N_b -Acetyltryptamine (1) has not previously been found to occur in nature but had been obtained as a biotransformed-metabolite derived from tryptamine by *Streptomyces staurosporeus* (Yang and Cordell, 1997).

Tryptamine and its derivatives are widely distributed in animals, plants and fungi (Buckingham *et al.*, 1994; Morales-Rios *et al.*, 1987; Salmoun *et al.*, 2002). Tryptamine has been discovered to be present in several edible fruits, namely, tomato, plum, and eggplant, and also in traces in oranges (Saxton *et al.*, 1965). *N,N*-Dimethyltryptamine was first identified as a constituent of the seeds and pods of *Piptadenia peregrina* and *P. macrocarpa* (Leguminosae) during an attempt to isolate the hallucinogenic principles present in the narcotic snuff prepared from these plants by certain American Indian tribes.

In later investigations, *N,N*-dimethyltryptamine occurs more widely in nature, and is the simplest of several naturally occurring tryptamine derivatives which exhibit psychotomimetic activity (Fish *et al.*, 1955)

On account of the activity of N,N-dimethyltryptamine, the physiological activity of N_b -acetyltryptamine in humans is of interest.

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