

Tyrosinase Inhibitory Activity of 80 Plant Extracts (II)

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(Received Feb. 18, 2003 ; accepted Mar. 11, 2003)

Abstract – The purpose of this study was to evaluate tyrosinase inhibitory activity of plant extracts for cosmetic use. When 80 plant extracts were tested, the methanol extracts of *Allium thunbergi*, *Asparagus oligoclonos*, *Ixeris dentate*, *Salvia plebeia*, *Sophora flavescens* and *Sophora japonica* showed more than 30% inhibition of mushroom tyrosinase activity at 100 µg/mL. Although less active than the reference compound, kojic acid (IC₅₀=7.0–16.3 µg/mL), these plant extracts may be used as tyrosinase inhibitors in cosmetics.

Keywords – tyrosinase, plant, skin whitening, *Salvia plebeia*, *Sophora flavescens*.

INTRODUCTION

The color of human skin is determined mainly by melanin content. Melanin is synthesized from tyrosine by enzymatic oxidative as well as non-enzymatic autooxidation processes. Among these sequential reactions, the most important and rate-limiting step is oxidation of tyrosine to 3,4-dihydroxyphenylalanine (DOPA) by a mixed function oxidase, tyrosinase (EC 1.14.18.1) (Sanchez-Ferrer *et al.*, 1995). Therefore, the inhibition of tyrosinase activity may lead to reduction of skin darkness. Especially, in East Asia, skin whitening is an important issue for cosmetic industry. Several tyrosinase inhibitors including arbutin, kojic acid and Glycyrrhizae radix extract have been widely used. But, there is always a need for new skin whitening agents. Thus, as our continual efforts to search useful skin whitening agents (Lee K.T. *et al.*, 1997), the methanol extracts from 80 plants were prepared and evaluated for their tyrosinase inhibitory activity in this investigation. It is found that the several plant materials such as *Salvia plebeia* and *Sophora flavescens* possess considerable inhibitory activity and may be used as skin whitening ingredients in cosmetics.

MATERIAL AND METHODS

Preparation of Plant Extracts

The plant materials used in this study were obtained from the Eui-Sung botanical garden of herbal drug, Kyungbuk Province,

Korea and identified by Dr. Kyu Young Chung (Dept. Resources and Environment, Andong National University). They were dried and finely chopped. Each material (100 g) was soaked in 300 mL methanol at room temperature for 3 days. After filtration, the filtrates were evaporated to dryness under vacuum and used throughout this study.

Tyrosinase Assay

Tyrosinase activity was determined essentially based on the previously described procedure (Vanni *et al.*, 1990) with slight modification. In brief, the test reaction mixture comprised of each plant extract, mushroom tyrosinase (105 unit, Sigma-Aldrich) and 0.15 mg L-tyrosine in 0.05 mM sodium phosphate buffer (pH 6.8). The reaction mixture (1.5 mL) was incubated at 37°C for 10 min, and the absorption at 475 nm was measured. And the absorbance of the same mixture without tyrosinase was used as the control.

RESULTS AND DISCUSSION

Eighty plant extracts were tested for their inhibitory activity against tyrosinase. Table 1 summarized the experimental results. Majorities of the extracts did not possess tyrosinase inhibitory activity, or possessed weak activity if any at the concentrations tested. Among the extracts tested, some plant extracts such as *Actinidia arguta*, *Allium thunbergi*, *Angelica gigas*, *Asparagus oligoclonos*, *Chrysanthemum zawadskii*, *Dianthus chinensis*, *Ixeris dentate*, *Salvia plebeia*, *Sophora flavescens* and *Sophora japonica* showed considerable inhibition at 10 and 100 µg/mL.

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Table 1. Tyrosinase inhibitory activity of the plant extracts

Plant Name	Part Used	% Inhibition of Tyrosinase ^{a)}	
		10 µg/mL	100 µg/mL
<i>Achillea sibirica</i>	whole	–	11.1
<i>Aconitum carmichaeli</i>	whole	3.0	–
<i>Acorus calamus</i>	leaf	8.5	18.6
<i>Actinidia arguta</i>	leaf	9.0	25.0
<i>Ademophora triphylla</i>	radix	–	–
<i>Allium thunbergii</i>	whole	8.7	31.3
<i>Allium tuberosum</i>	whole	–	1.8
<i>Althaea rosea</i>	whole	16.6	17.9
<i>Aneilema keisak</i>	whole	7.7	7.2
<i>Angelica gigas</i>	whole	16.5	22.3
<i>Arctium lappa</i>	fruit	–	–
<i>Artemisia capillaris</i>	whole	–	13.4
<i>Asparagus cochinchinensis</i>	whole	–	–
<i>Asparagus oligoclonus</i>	whole	21.6	16.8
	stem	32.3	32.8
<i>Aster koraiensis</i>	whole	10.6	20.8
<i>Aster tataricus</i>	whole	–	7.8
<i>Campsis grandiflora</i>	leaf	–	5.6
<i>Cassia tora</i>	whole	15.3	17.8
<i>Cassia occidentalis</i>	aerial	–	20.0
<i>Cedreal sinensis</i>	leaf	11.9	0.6
<i>Chrysanthemum zawadskii</i>	whole	–	25.7
<i>Clamatis manshurica</i>	radix	–	–
<i>Crataegus pinnatifida</i>	fruit	–	–
<i>Dianthus chinensis</i>	whole	–	29.2
<i>Dictamnus dasycarpus</i>	whole	16.9	18.6
<i>Disporum sessile</i>	whole	–	14.0
<i>Ecinopsis setifer</i>	whole	–	12.5
<i>Ficus carica</i>	aerial	–	–
	leaf	–	5.6
<i>Forsythia viridissima</i>	whole	–	–
	fruit	–	–
<i>Galium verum</i>	whole	–	–
	seed	–	–
<i>Glycyrrhizae uralensis</i>	seed	–	7.1
<i>Gossypium indicum</i>	leaf	18.4	9.2
<i>Hemerocallis aurantiaca</i>	whole	–	8.4
<i>Hemerocallis fulva</i>	whole	20.7	21.2
<i>Hemerocallis lilioasphodelus</i>	whole	–	10.8
<i>Hibiscus manihot</i>	whole	4.2	9.8
<i>Houttuynia cordata</i>	aerial	9.8	–
<i>Hosta plantaginea</i>	whole	–	–
<i>Inula helenium</i>	aerial	–	–
<i>Iris koreana</i>	whole	–	–
<i>Iris pallasi</i>	leaf	9.3	9.8
<i>Iris sanguinea</i>	whole	–	–

Table 1. Continued

Plant Name	Part Used	% Inhibition of Tyrosinase ^{a)}	
		10 µg/mL	100 µg/mL
<i>Ixeris dentate</i>	whole	14.0	35.1
<i>Leonurus sibiricus</i>	whole	28.9	13.8
<i>Lotus corniculatus</i>	whole	18.1	16.8
<i>Lysimachia davurica</i>	whole	17.6	12.4
<i>Mentha arvensis</i>	whole	–	–
<i>Ostericum koreanum</i>	whole	–	7.7
<i>Patarinia scabiosaefolia</i>	whole	1.4	25.6
<i>Patrinia villosa</i>	whole	–	–
<i>Peucedanum japonicum</i>	whole	7.5	19.7
<i>Physalis alkekengi</i>	leaf	6.1	22.3
<i>Pleuroptems multiflorus</i>	whole	–	–
<i>Polygonatum stenophyllum</i>	whole	7.5	21.2
<i>Potentilla discolor</i>	flower	4.7	–
<i>Prunus persica</i>	flower	–	9.2
<i>Pteridium aquilinum</i>	whole	–	9.3
<i>Pulsatilla koreana</i>	whole	–	12.1
<i>Reynoutria elliptica</i>	leaf	–	5.9
<i>Rhodea japonica</i>	leaf	–	–
<i>Rhododendron mucronulatum</i>	whole	–	12.5
<i>Rumex acetocella</i>	whole	1.7	13.3
<i>Rumex crispus</i>	whole	2.6	0.6
<i>Ruta graveolens</i>	aerial	2.0	1.2
<i>Salvia miltiorrhiza</i>	radix	–	–
<i>Salvia plebeia</i>	whole	8.1	36.1
<i>Saururus chinensis</i>	whole	5.6	11.1
<i>Scilla chinensis</i>	aerial	–	–
<i>Sedum kamtschaticum</i>	whole	–	7.9
<i>Selaginella tamariscina</i>	whole	13.3	18.2
<i>Sophora flavescens</i>	root	24.1	56.3
<i>Sophora japonica</i>	flower	–	38.7
<i>Sophora subprostrata</i>	radix	–	–
<i>Symphytum officinale</i>	whole	13.1	20.6
<i>Trachelospermum asiaticum</i>	whole	–	10.0
<i>Veronica linariaefolia</i>	whole	8.5	16.7
<i>Viola mandshurica</i>	whole	4.8	1.7
<i>Zanthoxylum schinifolium</i>	whole	–	11.0
	Leaf	2.6	14.0
<i>Zingiber mioga</i>	whole	–	–
<i>Zizyphus jujuba</i>	seed	–	–
<i>Kojic acid</i> ^{b)}		63.5	98.4

^{a)}Tyrosinase (105 U) showed 0.38-0.53 abs. at 475 nm by 10 min incubation in our experiments. ^{b)}IC₅₀ values of kojic acid ranged at 7.0-16.3 µg/mL from five separate experiments. % Inhibition shown here represented one typical result of five experiments.

Among the extracts tested in this study, 11 plant extracts from *Aconitum carmichaehi*, *Actinidia arguta*, *Angelica gigas*, *Artemisia capillaries*, *Asparagus cochinchinesis*, *Crataegus pinnatifida*, *Forsythia viridissima*, *Leonurus sibiricus*, *Ostericum koreanum*, *Sophora flavescens* and *Zizyphus jujuba* were previously evaluated for their tyrosinase inhibitory activity and showed similar results as ours (Lee S.-H. *et al.*, 1997; Choi *et al.*, 1998; Choi *et al.*, 2001). Especially, the plant materials including *Allium thunbergi*, *Asparagus oligoclonos*, *Ixeris dentate*, *Salvia plebeia*, *Sophora flavescens* and *Sophora japonica* showed more than 30% inhibition at 100 µg/mL. These plant materials merit for further investigation, although no one showed the comparable activity with kojic acid ($IC_{50} = 7.0 - 16.3$ µg/mL). In particular, our results of *Sophora flavescens* were well correlated with the previous investigation describing that the methanol extract from *Sophora flavescens* possessed potent tyrosinase inhibitory activity (Lee S.-H. *et al.*, 1997). And some parts of active principles from the same plant material have been recently identified as kurarinone and kushnol F (Ha *et al.*, 2001). For further isolating active principles, *Salvia plebeia* and *Sophora flavescens* are now being under investigation.

From the results, it is clear that some plant extracts were able to inhibit melanin formation, at least *in vitro*. This study sug-

gests that several plant extracts have a potential as skin whitening agents in cosmetics.

REFERENCES

- Chi, B.W., Lee, B.H., Kang, K.J., Lee, E.S. and Lee, N.H. (1998). Screening of the tyrosinase inhibitors from marine algae and medicinal plants. *Kor. J. Pharmacogn.* **29**, 237-242.
- Choi, S.-S., Noh, H.-S., Cho, S.-H. and Kong, K.H. (2001). Screening of inhibitors against tyrosinase activity from natural products. *Yakhak Hoeji* **45**, 522-528.
- Ha, T.J., Yang, M.S., Jang, D.S., Choi, S.U. and Park, K.H. (2001). Inhibitory activities of flavanone derivatives isolated from *Sophora flavescens* for melanogenesis. *Bull. Korean Chem. Soc.* **22**, 97-99.
- Lee, K.T., Kim, B.J., Kim, J.H., Heo, M.Y. and Kim, H.P. (1997). Biological screening of 100 plant extracts for cosmetic use (I): inhibitory activities of tyrosinase and DOPA auto-oxidation. *Int. J. Cosmetic Sci.* **19**, 291-298.
- Lee, S.-H., Park, J.S., Kim, S.Y., Kim, J.J. and Chung, S.R. (1997). The screening of the inhibitory compounds on tyrosinase activity from the natural product. *Yakhak Hoeji* **41**, 456-461.
- Sanchez-Ferrer, A., Rodriguez-Lopez, J.N., Garcia-Canova, F. and Garcia-Carmona, F. (1995). Tyrosinase: a comprehensive review of its mechanism. *Biochim. Biophys. Acta* **1247**, 1-11.
- Vanni, A., Gastaldi, D. and Giunatu, G. (1990). Kinetic investigation on the double enzymatic activity of the mushroom tyrosinase. *Annali di Chimica* **80**, 35-60.