

## Genomic Structure of the Luciferase Gene and Phylogenetic Analysis of the Firefly, *Pyrocoelia rufa*

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(Received 16 October 2003; Accepted 27 October 2003)

We describe here the complete nucleotide sequence and the exon-intron structure of the luciferase gene of the firefly, *Pyrocoelia rufa*. The luciferase gene of the *P. rufa* firefly consisted of six introns and seven exons coding for 548 amino acid residues. From the translational start site to the end of last exon, however, the genomic DNA length of the *P. rufa* luciferase gene from the Korean and Chinese samples spans 1,968 bp and 1983 bp, respectively, and 3 amino acid residues were different to each other. Additionally, we also analyzed mitochondrial cytochrome oxidase I (COI) gene of the Chinese *P. rufa* fireflies. Analysis of DNA sequences from the mitochondrial COI protein-coding gene revealed 4 mitochondrial DNA sequence-based haplotypes with a maximum divergence of 0.7%. With the 20 *P. rufa* haplotypes found in Korea, phylogenetic analyses using PAUP and PHYLIP subdivided the *P. rufa* into three clades, termed clades A and B for the Korean sample, and clade C for the Chinese sample.

**Key words:** Firefly, *Pyrocoelia rufa*, Luciferase gene, Mitochondrial DNA, Cytochrome oxidase I gene, Phylogenetic analysis

### Introduction

The Korean firefly, *Pyrocoelia rufa*, is one of the abundant

firefly species in Korea and is also found in China and only at Tsushima in case of Japan (Kim and Nam, 1981; Suzuki, 1997, 2001). The *Pyrocoelia*-group in the firefly is divided into two lineages. The first one consists of *P. rufa*, *P. miyako* and *P. atripennis*, and the second of *P. fumosa*, *P. oshimana*, *P. matsumurai matsumurai*, *P. m. kumejimensis*, *P. discicollis* and *P. abdominalis* (Suzuki, 1997, 2001). The body sizes of the former group are larger than those of the other Lampyrine species, and members of the group are characterized by the continuous broadcast of strong light. *P. rufa* has the largest luminescent organs among the former group (Suzuki, 1997, 2001). In fall (August – September), *P. rufa* females oviposit about 40 ~ 120 eggs under the rocks and roots of glasses where enough moisture is available, and the eggs are hatched on May ~ June (Kim *et al.*, 2003). Larval fireflies are subjected to ecdysis four times, and thereafter are subjected to metamorphosis approximately on August. Ten- to twelve-days after metamorphosis, pupal fireflies become adult fireflies, and their lifespan continues for 15 – 20 days until they finish their lives (Kim *et al.*, 2003).

The firefly luciferase gene is widely used as a genetic marker or as a reporter gene in a variety of organisms including bacteria, plants and animals (De Wet *et al.*, 1987; DiLella *et al.*, 1988; Howard *et al.*, 1988; Kondo *et al.*, 1992; Miller *et al.*, 1992; Jacobs *et al.*, 1993; Bailey *et al.*, 1994; Vikas *et al.*, 1995). Today, more than 10 luciferase genes have been isolated from various firefly species (Tatsumi *et al.*, 1992; Cho *et al.*, 1999; Masuda *et al.*, 1989; Devine *et al.*, 1993; Ohmiya *et al.*, 1995; Choi *et al.*, 2002). The nucleotide sequences of a cDNA encoding the luciferase of *P. rufa* have been reported (Lee *et al.*, 2001).

Choi *et al.* (2003) cloned and sequenced the genomic

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structure of the luciferase from the *Hotaria*-group fireflies and elucidated their phylogenetic relationships using luciferase and mitochondrial cytochrome oxidase I (COI) genes. Kim *et al.* (2004) also elucidated the genomic structure of the luciferase from *Luciola lateralis* and their phylogenetic relationships using luciferase and mitochondrial COI genes. The genetic divergence, population genetic structure, and possible speciation of *P. rufa* were investigated on the midsouthern Korean mainland, coastal islets, a remote offshore, Jeju - do, and Tsushima Island in Japan (Lee *et al.*, 2003).

In this study, we analyzed the genomic structure of the luciferase from *P. rufa* collected in Korea and China, and investigated the phylogenetic analysis from the mitochondrial COI gene of *P. rufa* between the Korean and Chinese samples.

## Materials and Methods

### Insects

The firefly, *Pyrocoelia rufa* used in this study was collected in Korea and China. The firefly samples in Korea were previously described (Lee *et al.*, 2003). The firefly samples in China were caught at Wuhan, Hubei, China from June to July 2003.

### PCR of *P. rufa* luciferase genomic DNA

Genomic DNA was extracted from the larvae of *P. rufa* by Wizard™ Genomic DNA Purification Kit, according to the manufacturers instructions (Promega). The primers used for amplification of the genomic DNA of the luciferase from the *P. rufa* were 5'-ATGGAAGATGATAGTAAACATATTATGCAT-3' for the translational start sequence region and 5'-TTACAATTTGGATTTTTGTC-CCATTGTAAGG-3' for the 3' coding region, based on the luciferase cDNA of *P. rufa* (Lee *et al.*, 2001). After a 35-cycles amplification (94°C for 1 min; 55°C for 1 min; 72°C for 1 min), PCR products were ethanol precipitated, centrifuged at 10,000 × g for 15 min, and rinsed with 70% ethanol. These DNAs were analyzed 1.0% agarose gel electrophoresis. The PCR products for sequencing were cloned into pGem-T vector (Promega, Madison, WI).

### Amplification of *P. rufa* mitochondrial COI gene

Total DNA was extracted following the standard Proteinase K method (Kocher *et al.*, 1989). A part of the COI gene was amplified by PCR using primers CI-J-1751 (5'-GGAGCTCCTGACATAGCATTCCC-3') and CI-N-2191 (5'-CCCGGTAAAATTTAAATATAAACTTC-3') (Simon *et al.*, 1994). PCR conditions were as follows: after an initial denaturation step at 94°C for 5 min, 40 cycles of 94°C

for 30 s, 50 for 40 s, and 72 for 45 s, and a final extension step at 72°C for 7 min. To confirm the successful DNA amplification, electrophoresis was carried out using 0.5 × TAE buffer in 1% agarose gel. The PCR product was then purified using a PCR purification Kit (QIAGEN, Germany) following manufacturers instruction.

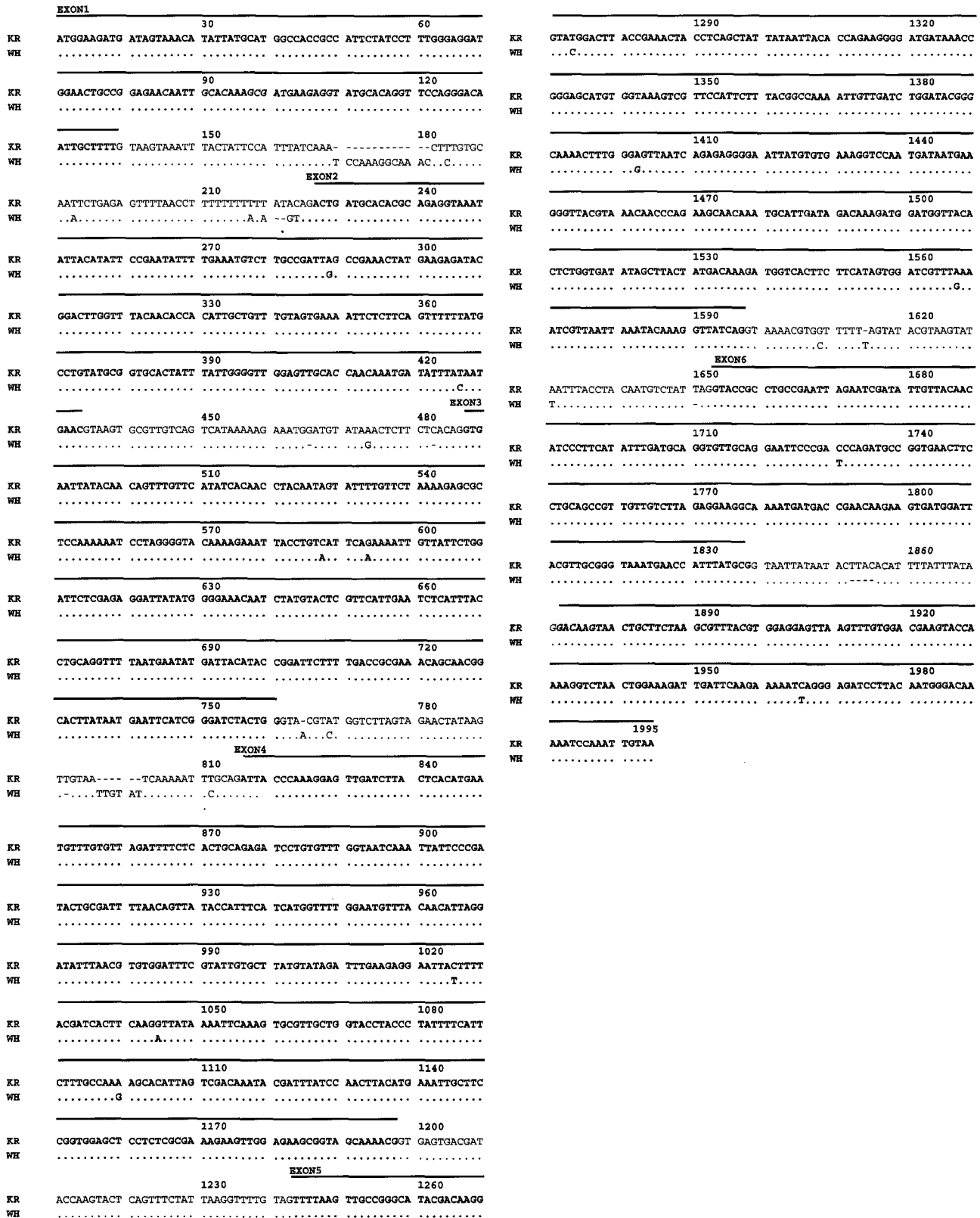
### DNA sequencing and data analysis

DNA sequencing was performed using an automatic sequencer (model 310 Genetic Analyzer; Perkin-Elmer Applied Biosystems, Foster City, CA). Sequence alignment was performed using IBI MacVector (ver. 6.5). PAUP (Phylogenetic Analysis using Parsimony) ver. 4.0b8 (Swofford, 2000) was used to infer possible phylogenetic relationships among the matrilineal of *P. rufa*. The homologous mtDNA sequences from one individual of *Hotaria unmunisana*, collected in Busan was used as an outgroup (Choi *et al.*, 2003). The analysis was performed using an equal weighting of transitions and transversions by heuristic search as well as several ratios up to and including 1:20. The reliability of the trees was tested by 1,000 iterations of bootstrapping (Felsenstein, 1985). As an alternative to the parsimony analysis, we used Neighbor-Joining (NJ) method and maximum likelihood (ML) method incorporated in PHYLIP (Phylogeny Inference Package) ver. 3.5c (Felsenstein, 1993). To obtain a phylogenetic tree, the data set was first iterated 1,000 times using the subprogram SEQBOOT. Next, the iterated data set was run using the subprogram DNADIST to obtain a distance matrix between pairs of haplotypes with the option of Kimura's 2-parameter method (Kimura, 1980), which attempted to correct observed dissimilarities for multiple substitution in sequences evolving with a transition bias. Individual trees from each distance matrix were obtained using the subprogram NEIGHBOR. The *H. unmunisana* sequence was included in the analysis to root the tree. Finally, a consensus tree representing reliability at each branch in the trees was obtained using the subprogram CONSENSE.

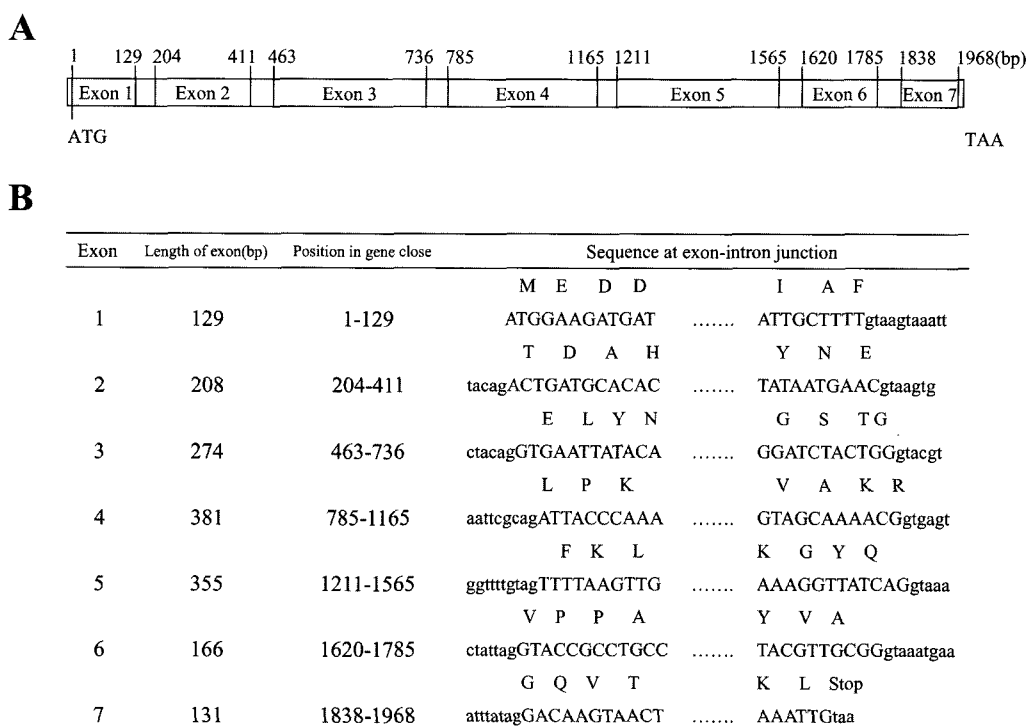
## Results and Discussion

### Genomic structure of the luciferase gene of *P. rufa* firefly

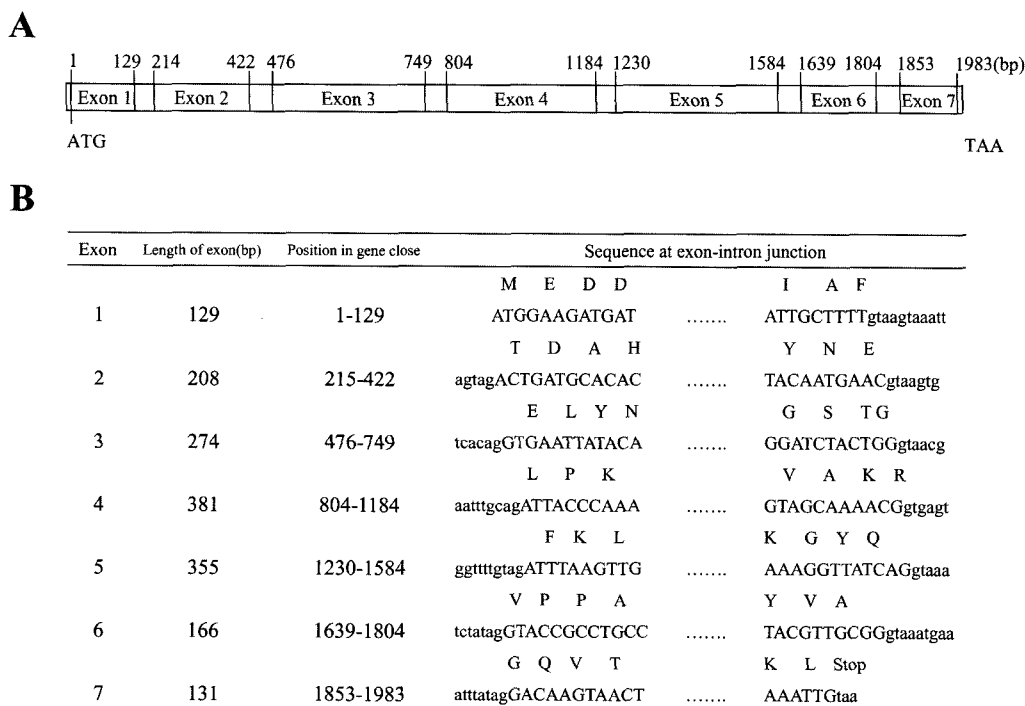
To identify the genomic DNA of the luciferase gene of the firefly, *P. rufa*, we have designed the PCR primer set based on the sequences of the luciferase cDNA of *P. rufa* already known in our laboratory (Lee *et al.*, 2001). The genomic DNA of the luciferase gene from the Korean and Chinese samples of the *P. rufa* firefly was cloned and sequenced (Fig. 1).



**Fig. 1.** The nucleotide sequence and genomic organization of *P. rufa* luciferase gene. The firefly used in this study was collected in the Korea (KR) and China (WH). Nucleotide numbers are presented on the upper, and the first base of initiation codon of the ORF is defined as +1. Exons are labeled with bold-lines.



**Fig. 2.** Genomic organization of the luciferase gene of the Korean *P. rufa* firefly. (A) Exon/intron structures. Numbers indicate the length (bp) of exons and introns. (B) Lengths of exons and exon/intron boundaries.



**Fig. 3.** Genomic organization of the luciferase gene of the Chinese *P. rufa* firefly. (A) Exon/intron structures. Numbers indicate the length (bp) of exons and introns. (B) Lengths of exons and exon/intron boundaries.

Comparison of the genomic sequence with the sequence of cloned luciferase cDNA in the *P. rufa* revealed the pres-

ence of seven exons (Fig. 1 and 2). From the known cDNA 5' site to the end of exon 7, the gene is 1,968 bp

			30		60	
KR	MEDDSKIMH	MEDDSKIMH	GTAGEQLHKA	MKRYAQVPGT	IAFTDAHAEV	NITYSEYFEM
WH	.....	.....	.....	.....	.....	.....
			90		120	
KR	SCRLAETMKR	YGLGLQHHLA	VCSSENSLQFF	MPVCGALFIG	VGVAPTNDIY	NRELYNSLF
WH	.....	.....	.....	.....	.....	.....
			150		180	
KR	ISQPTIVFCS	KRALQKILGV	HKKLPVIQXI	VILDSREDYM	GKQSMYSFIE	SHPAGPNEY
WH	.....	.....	Q.....	.....	.....	.....
			210		240	
KR	DYIPDSFDRE	TATALIMNS	GSTGLPKGVD	LTHMNVCFRF	SHCRDPVFGN	QIIPDTAILT
WH	.....	.....	.....	.....	.....	.....
			270		300	
KR	VIFPHGFGM	FTTLGYLTCG	FRIVLMYRFE	EELFLRSLQD	YKIQSALLVP	TLFSPFAKST
WH	.....	.....	.....	.....	.....	.....
			330		360	
KR	LVDKYDLSNL	HEIASGGAPL	AKEVGEAVAK	RFKLPQIRQG	DGLTETTSAL	IITPBGDDKP
WH	.....	.....	.....	.....	Y.....	.....
			390		420	
KR	GACGKVVPPF	AAKIVDLDTG	KTGLVNRQGE	LCVKGPMIMK	GYVMNPEATN	ALIDKDWLH
WH	.....	T.....	.....	.....	.....	.....
			450		480	
KR	SGDIAYYDEK	GHPFLVDRLK	SLIKYKGYQV	PPAELESILL	QHPFIDFAGV	AGIPDPDAQE
WH	.....	.....	.....	.....	.....	.....
			510		540	
KR	LPAAVVVLEE	GKMTQEQVM	DYVAGQVTAS	KRLRGVKFV	DEVPKLGTGK	IDSRKIRELL
WH	.....	.....	.....	.....	.....	.....
			548			
KR	TMGQSKSL					
WH	.....					

**Fig. 4.** The deduced amino acid sequence of the luciferase gene of *P. rufa* firefly. The firefly used in this study was collected in the Korea (KR) and China (WH). Residues are numbered according to the *P. rufa* luciferase cDNA sequences previously known (Lee *et al.*, 2001), and identical residues are dotted.

long and consisted of 548 amino acid residues. On the other hand, genomic structure of luciferase gene from the Chinese *P. rufa* sample was identical in their exon number and coding sequence size (548 amino acid residues), but the genomic DNA length differed from the Korean *P. rufa* sample (Fig. 1 and 3). Furthermore, the amino acid sequence of the luciferase gene of the Chinese *P. rufa* sample differed from the Korean sample by three amino acid residues (Fig. 4). The intron boundaries are listed in Fig. 2B and 3B. The consensus sequences, including an invariant GT at the intron 5 boundary and an invariant AG at its 3 boundary were very well conserved in two samples.

When compared with the Korean *P. rufa* firefly sample, the Chinese *P. rufa* luciferase gene differed by three amino acid residues in the coding region. Also, the length of introns differed between two samples, which resulted in genomic DNA length difference. However, the genomic structure of the luciferase gene in both samples consisted of six introns and seven exons coding for 548 amino acid residues. This is the same as that of luciferases from other fireflies, such as *L. lateralis* (Kim *et al.*, 2004) and *Hotaria*-group fireflies (Choi *et al.*, 2003).

### Phylogenetic analysis using the mitochondrial COI gene

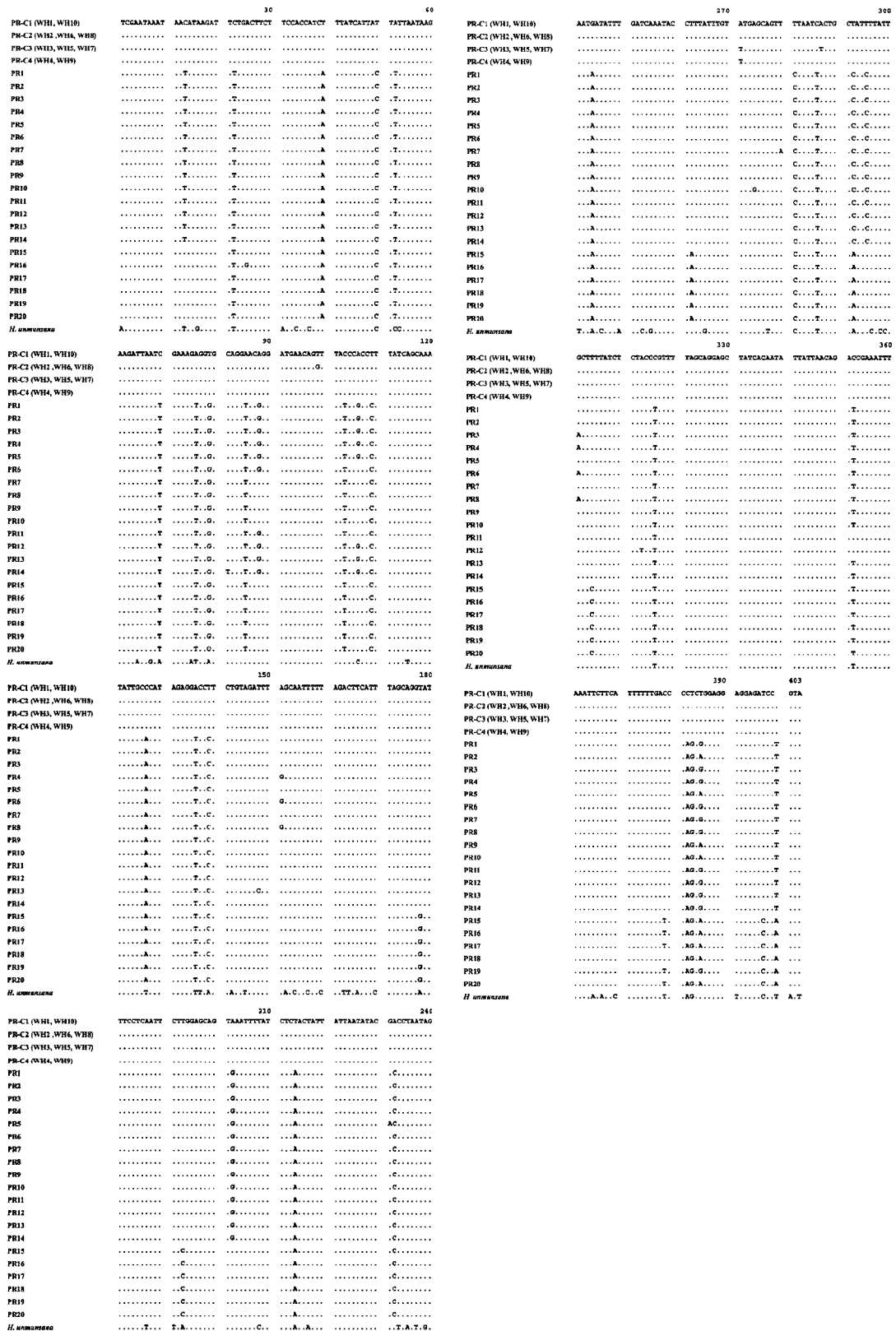
Fig. 5 shows the nucleotide sequences of the 403 bp region of the COI gene from the Chinese samples of the *P. rufa* firefly. The nucleotide sequences of the COI gene from the Chinese *P. rufa* samples were compared with those of the Korean *P. rufa* firefly already known in our laboratory (Lee *et al.*, 2003). A total of 4 haplotypes (PR-C1 – PR-C4) was obtained from the 403 bp of the COI gene from 10 individuals of the *P. rufa* firefly collected from Wuhan in China.

Table 1 shows the nucleotide sequence divergence among the *P. rufa* haplotypes. Sequence divergence among 4 haplotypes from the Chinese samples by pairwise comparisons ranged from 0.2 to 0.7% (one bp three bp). Sequence divergence among 24 haplotypes, including 20 haplotypes found in Korean *P. rufa* firefly, by pairwise comparisons ranged from 0.2 to 8.4% (one bp 34 bp), and the largest sequence divergence was observed when PR4 found in Korean sample was compared with PR-C4 found in three individuals from the Chinese sample.

Phylogenetic relationships among haplotypes are depicted in Figure 6. Because analyses run with transition: transversion weightings of 1:0, 1:1, 1:5, 1:10, and 1:20 did not affect the topology of the tree, only the result obtained by unordered analysis is presented (Fig. 6). Twenty-four haplotypes used in this study were subdivided into three groups (termed clades A and B for Korean sample, and clade C for Chinese sample), although haplotype relationships within each clade were mostly not resolved, possibly by small nucleotide difference (*e.g.*, one or two bp). Each clade was somewhat different in haplotype composition. For example, clade A consists of a large number of haplotypes (14 haplotypes among 24) and is larger in maximum sequence divergence (1.5%) than clade B (Lee *et al.*, 2003). Clade B contained six haplotypes with a maximum sequence divergence of 0.7% (Lee *et al.*, 2003). Also, clade C for Chinese sample contained four haplotypes with a maximum sequence divergence of 0.7% (Fig. 6 and Table 1).

Fig. 7 represents the result of probability-based phylogenetic analyses using the neighbor-joining (NJ) method. Each monophyletic group designated in the PAUP analysis included identical haplotypes, suggesting that *P. rufa* can be subdivided into three distinct genetic groups.

Phylogenetic analysis of *P. rufa* showed that the fireflies are subdivided largely into three clades and distribution of each clade is concordant with geographic separations: clade B exclusively from Jeju-do Island, clade A from all localities but Jeju-do Island (Lee *et al.*, 2003), and clade C from Wuhan, China.



**Fig. 5.** Sequence alignment of 4 mitochondrial haplotypes (designated as PR-C1 PR-C4) obtained from 403 bp COI sequences of *P. rufa* fireflies collected in Wuhan, China. Sequences of 20 mitochondrial haplotypes (PR1 PR20) are taken from the Korean *P. rufa* firefly (Lee *et al.*, 2003).

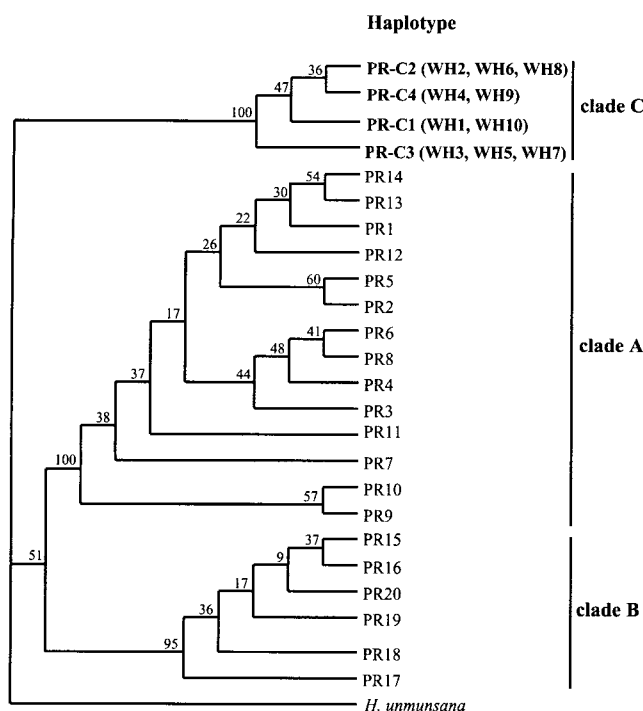
**Table 1.** Pairwise comparisons among nucleotide sequences of COI genes obtained from this study and known *P. rufa* COI genes obtained through GenBank search

	1	2	3	4	5	6	7	8	9	10	11	12	13
1. PR-C1 (WH1,WH10)	–	0.002	0.002	0.005	0.069	0.072	0.072	0.072	0.072	0.074	0.074	0.074	0.074
2. PR-C2 (WH2, WH6, WH8)	1	–	0.005	0.007	0.072	0.074	0.074	0.074	0.074	0.077	0.077	0.077	0.077
3. PR-C3 (WH4, WH9)	1	2	–	0.002	0.072	0.074	0.074	0.074	0.074	0.077	0.077	0.077	0.077
4. PR-C4 (WH3, WH5, WH7)	2	3	1	–	0.074	0.077	0.077	0.077	0.077	0.079	0.079	0.079	0.079
5. PR9	28	29	29	30	–	0.005	0.002	0.005	0.037	0.007	0.005	0.007	0.037
6. PR7	29	30	30	31	2	–	0.007	0.005	0.042	0.007	0.010	0.007	0.042
7. PR10	29	30	30	31	1	3	–	0.007	0.040	0.010	0.007	0.010	0.040
8. PR11	29	30	30	31	2	2	3	–	0.042	0.002	0.005	0.007	0.042
9. PR18	29	30	30	31	15	17	16	17	–	0.045	0.042	0.045	0.005
10. PR1	30	31	31	32	3	3	4	1	18	–	0.002	0.010	0.045
11. PR2	30	31	31	32	2	4	3	2	17	1	–	0.012	0.042
12. PR8	30	31	31	32	3	3	4	3	18	4	5	–	0.045
13. PR15	30	31	31	32	15	17	16	17	2	18	17	18	–
14. PR17	30	31	31	32	15	17	16	17	2	18	17	18	2
15. PR19	30	31	31	32	16	16	17	16	3	17	18	17	1
16. PR20	30	31	31	32	16	18	17	18	1	19	18	19	1
17. PR3	31	32	32	33	4	4	5	2	19	1	2	3	19
18. PR5	31	32	32	33	3	5	4	3	18	2	1	6	18
19. PR6	31	32	32	33	4	4	5	2	19	3	4	1	19
20. PR12	31	32	32	33	5	5	6	3	18	2	3	6	18
21. PR13	31	32	32	33	4	4	5	2	19	1	2	5	19
22. PR14	31	32	32	33	4	4	5	2	19	1	2	5	19
23. PR16	31	32	32	33	17	19	18	19	2	20	19	20	2
24. PR4	32	33	33	34	5	5	6	3	20	2	3	2	20
25. <i>Hotaria unmunšana</i>	85	86	86	87	80	81	81	81	76	81	81	80	76

**Table 1.** continud

	14	15	16	17	18	19	20	21	22	23	24	25
1. PR-C1 (WH1,WH10)	0.074	0.074	0.074	0.077	0.077	0.077	0.077	0.077	0.077	0.077	0.079	0.211
2. PR-C2 (WH2, WH6, WH8)	0.077	0.077	0.077	0.079	0.079	0.079	0.079	0.079	0.079	0.079	0.082	0.213
3. PR-C3 (WH4, WH9)	0.077	0.077	0.077	0.079	0.079	0.079	0.079	0.079	0.079	0.079	0.082	0.213
4. PR-C4 (WH3, WH5, WH7)	0.079	0.079	0.079	0.082	0.082	0.082	0.082	0.082	0.082	0.082	0.084	0.216
5. PR9	0.037	0.040	0.040	0.010	0.007	0.010	0.012	0.010	0.010	0.042	0.012	0.199
6. PR7	0.042	0.040	0.045	0.010	0.012	0.010	0.012	0.010	0.010	0.047	0.012	0.201
7. PR10	0.040	0.042	0.042	0.012	0.010	0.012	0.015	0.012	0.012	0.045	0.015	0.201
8. PR11	0.042	0.040	0.045	0.005	0.007	0.005	0.007	0.005	0.005	0.047	0.007	0.201
9. PR18	0.005	0.007	0.002	0.047	0.045	0.047	0.045	0.047	0.047	0.005	0.050	0.189
10. PR1	0.045	0.042	0.047	0.002	0.005	0.007	0.005	0.002	0.002	0.050	0.005	0.201
11. PR2	0.042	0.045	0.045	0.005	0.002	0.010	0.007	0.005	0.005	0.047	0.007	0.201
12. PR8	0.045	0.042	0.047	0.007	0.015	0.002	0.015	0.012	0.012	0.050	0.005	0.199
13. PR15	0.005	0.002	0.002	0.047	0.045	0.047	0.045	0.047	0.047	0.005	0.050	0.189
14. PR17	–	0.007	0.002	0.047	0.045	0.047	0.045	0.047	0.047	0.005	0.050	0.186
15. PR19		–	0.005	0.045	0.047	0.045	0.042	0.045	0.045	0.007	0.047	0.189
16. PR20	1		–	0.05	0.047	0.050	0.047	0.05	0.050	0.002	0.052	0.186
17. PR3	19	18		–	0.007	0.005	0.007	0.005	0.005	0.052	0.002	0.199
18. PR5	18	19	19		–	0.012	0.010	0.007	0.007	0.050	0.010	0.203
19. PR6	19	18	20	2		–	0.012	0.010	0.010	0.052	0.002	0.201
20. PR12	18	17	19	3	4		–	0.007	0.007	0.050	0.010	0.196
21. PR13	19	18	20	2	3	4		–	0.005	0.052	0.007	0.203
22. PR14	19	18	20	2	3	4	3		–	0.052	0.007	0.203
23. PR16	2	3	1	21	20	21	20	21		–	0.055	0.189
24. PR4	20	19	21	1	4	1	4	3	3	22	–	0.201
25. <i>Hotaria unmunšana</i>	75	76	75	80	82	81	79	82	82	76	81	–

Numbers above the diagonal are mean distance values; numbers below the diagonal are absolute distance values.



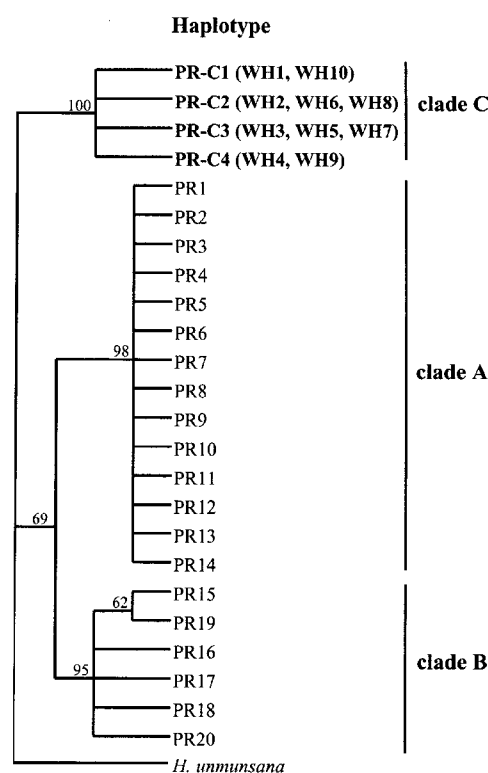
**Fig. 6.** A PAUP analysis of mitochondrial COI gene sequences of *P. rufa* using homologous sequence of another firefly, *Hotaria unmunšana*, as an outgroup. The tree shown is an unordered tree obtained with the option of “retain groups with frequency >50%” by majority-rule consensus of three equally parsimonious trees from the heuristic search. The numbers shown on the branches represent bootstrap values for 1,000 replicates. Tree length is 120 steps long, Consistency Index is 0.800, and retention index is 0.975.

## Acknowledgments

This paper was supported by the Dong-A University Research Fund.

## References

- Bailey, J., M. Benard and T. G. Burland (1994) A luciferase expression system for *Physarum* that facilitates analysis of regulatory elements. *Curr. Genet.* **26**, 126-131.
- Cho, K. H., J. S. Lee, Y. D. Choi and K. S. Boo (1999) Structural polymorphism of the luciferase gene in the firefly, *Luciola lateralis*. *Insect Mol. Biol.* **8**, 193-200.
- Choi, Y. S., J. S. Bae, K. S. Lee, S. R. Kim, I. Kim, J. G. Kim, K. Y. Kim, S. E. Kim, H. Suzuki, S. M. Lee, H. D. Sohn and B. R. Jin (2002) Genomic structure of the luciferase gene and phylogenetic analysis in the *Hotaria*-group fireflies. *Comp. Biochem. Physiol.* **134B**, 199-214.
- Choi, Y. S., K. S. Lee, J. S. Bae, K. M. Lee, S. R. Kim, I. Kim, S. M. Lee, H. D. Sohn and B. R. Jin (2002) Molecular cloning and expression of a cDNA encoding the luciferase from



**Fig. 7.** PHYLIP analysis of mitochondrial COI gene sequences of *P. rufa*. The tree was obtained using the subprogram NEIGHBOR incorporated in PHYLIP with the option of Kimura's 2-parameter method (1980). The tree was rooted using *H. unmunšana*. The numbers shown on the branches, which represent bootstrap values for 1,000 replications, were obtained using the subprogram CONSENSUS.

the firefly, *Hotaria unmunšana*. *Comp. Biochem. Physiol.* **132B**, 661-670.

- Devine, J. H., G. D. Kutuzova, V. A. Green, N. N. Ugarova and T. O. Baldwin (1993) Luciferase from the east European firefly *Luciola mingrellica*: cloning and nucleotide sequence of the cDNA, overexpression in *Escherichia coli* and purification of the enzyme. *Biochim. Biophys. Acta* **1173**, 121-132.
- DiLella, A. G., D. A. Hope, H. Chen, M. Trumbauer, R. J. Schwartz and R. G. Smith (1988) Utility of firefly luciferase as a reporter gene for promoter activity in transgenic mice. *Nucleic Acids Res.* **16**, 4159.
- Felsenstein, J. (1985) Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* **29**, 783-791.
- Felsenstein, J. (1993) PHYLIP (Phylogenetic Inference Package). Department of Genetics, Univ. of Washington, Seattle.
- Howard, P. K., K. G. Ahern and R. A. Firtel (1988) Establishment of a transient expression system for *Dictyostelium discoideum*. *Nucleic Acids Res.* **16**, 2613-2623.
- Jacobs, W. R., R. G. Barletta, R. Udami, J. Chan, G. Kalkut, G. Sosne, T. Kieser, G. J. Sarkis, G. F. Hatfull and B. R. Bloom (1993) Rapid assessment of drug susceptibilities of *Mycobacterium tuberculosis* by means of luciferase reporter



- phages. *Science* **260**, 819-822.
- Kim, C. W. and S. H. Nam (1981) Present status of the Korean fire-flies and their conservation. *Bull. Korean Asso. Conser. Nature Ser.* **3**, 311-324.
- Kim, J. G., Y. S. Choi, K. Y. Kim, J. S. Bae, I. Kim, H. D. Sohn and B. R. Jin (2004) Genomic structure and phylogenetic analysis of luciferase gene of the firefly, *Luciola lateralis* (Coleoptera: Lampyridae). *Eur. J. Entomol.* In press.
- Kim, J. G., K. Y. Kim, Y. C. Choi, J. Y. Choi, S. E. Kim, B. R. Jin, S. M. Lee, J. E. Lee, K. Y. Lee and S. H. Lee (2003) Developmental characteristics and life history of the Korean native firefly, *Pyrocoelia rufa*. *Int. J. Indust. Entomol.* **6**, 69-73.
- Kimura, M. (1980) A simple method for estimating evolutionary rate of base substitution through comparative studies of nucleotide sequences. *J. Mol. Evol.* **16**, 111-120.
- Kocher, T. D., W. K. Thomas, A. Meyer, S. V. Edwards, S. Pääbo, F. X. Villablanca and A. C. Wilson (1989) Dynamics of mitochondrial DNA evolution in animals: amplification and sequencing with conserved primers. *Proc. Natl. Acad. Sci. USA* **86**, 6196-6200.
- Kondo, T., N. Takahashi and M. Muramatsu (1992) The regulation of the murine Hox-2.5 gene expression during cell differentiation. *Nucleic Acids Res.* **20**, 5729-5735.
- Lee, S. C., J. S. Bae, I. Kim, H. Suzuki, S. R. Kim, J. G. Kim, K. Y. Kim, W. J. Yang, S. M. Lee, H. D. Sohn and B. R. Jin (2003) Mitochondrial DNA sequence-based population genetic structure of the firefly, *Pyrocoelia rufa* (Coleoptera: lampyridae). *Biochem. Genet.* **41**, 427-452.
- Lee, K. S., H. J. Park, J. S. Bae, T. W. Goo, I. Kim, H. D. Sohn and B. R. Jin (2001) Molecular cloning and expression of a cDNA encoding the luciferase from the firefly, *Pyrocoelia rufa*. *J. Biotechnol.* **92**, 9-19.
- Masuda, T., H. Tatsumi and E. Nakano (1989) Cloning and sequence analysis of cDNA for luciferase of a Japanese firefly, *Luciola cruciata*. *Gene* **77**, 265-270.
- Millar, A. J., S. R. Short, N. H. Chua and S. A. Say (1992) A novel circadian phenotype based on firefly luciferase expression in transgenic plants. *Plant Cell* **4**, 1075-1087.
- Ohiyama, Y., N. Ohba, H. Toh and F. I. Tsuji (1995) Cloning, expression and sequence analysis of cDNA for the luciferases from the Japanese fireflies, *Pyrocoelia miyako* and *Hotaria parvula*. *Photochem. Photobiol.* **62**, 309-313.
- Simon, C., F. Frati, A. Beckenbach, B. Crespi, H. Liu and P. Flook (1994) Evolution, weighting, and phylogenetic utility of mitochondrial gene sequences and a composition of conserved polymerase chain reaction primers. *Ann. Entomol. Soc. Am.* **87**, 651-701.
- Suzuki, H. (1997) Molecular phylogenetic studies of Japanese fireflies and their mating systems (Coleoptera: Cantharodea). *TMU Bull. Natl. Hist.* **3**, 1-53.
- Suzuki, H. (2001) Studies on biological diversity of firefly in Japan. *Int. J. Indust. Entomol.* **2**, 91-105.
- Swofford, D. L. (2000) PAUP\*. Phylogenetic Analysis Using Parsimony (\*and Other Methods), version 4, Sinauer Sunderland, MA.
- Tatsumi, H., N. Kajiyama and E. Nakano (1992) Molecular cloning and expression in *Escherichia coli* of a cDNA clone encoding luciferase of a firefly, *Luciola lateralis*. *Biochim. Biophys. Acta* **1131**, 161-165.
- Vikas, B. P., S. Sumathy and P. G. Karumathil (1995) Baculovirus mediated high-level expression of luciferase in silkworm cells and larvae. *BioTechniques* **19**, 97-104.
- de Wet, J. R., K. W. Wood, M. DeLuca, D. R. Helinski and S. Subramani (1987) Firefly luciferase gene: structure and expression in mammalian cells. *Mol. Cell Biol.* **7**, 725-737.