

〈NOTE〉

Treatment of *Microcotyle sebastis* (Monogenea: Polyopisthocotylea) Infestation with Praziquantel under Commercial Rockfish (*Sebastes schlegeli*) Culture Conditions

Chun Soo Kim, Jae Bum Cho, Kyoung Jin Ahn, Jae Il Lee
and Ki Hong Kim*

Department of Aquatic Life Medicine, College of Fisheries Sciences,
Pukyong National University, Busan 608-737, Korea

(Received March 2002, Accepted June 2002)

The treatment efficacy of oral administration of praziquantel against the blood-sucking monogenean *Microcotyle sebastis* under the commercial rockfish-culture conditions was evaluated. The quantity of praziquantel added to the food was calculated so that the amount of praziquantel ingested daily by fish was 200 mg or 400 mg/kg body weight (bw). The fish were fed a moist-pelleted fish meal supplemented with praziquantel at an rate of 0 (control), 200 or 400 mg of praziquantel per kg bw, respectively, for 3 times at an interval of 24 h. On 1 day to 8 days after the treatment, 10 fish were taken randomly from each net-pen daily (80 fish in each group in total), and the efficacy of each treatment was confirmed. Abundance of worms were significantly reduced in groups fed praziquantel-supplemented diets. The results clearly demonstrated that feeding the praziquantel-supplemented diet was effective for controlling *M. sebastis* infestation in practical commercial rockfish-culture systems without imposing any handling stress.

Key words: Praziquantel, *Microcotyle sebastis*, Commercial rockfish culture conditions

The farming of rockfish, *Sebastes schlegeli*, has rapidly increased during the past decade, and the fish has become an important cultured marine fish species in Korea. One problem associated with the farming of rockfish in Korea is the infestation with gill monogenean parasite, *Microcotyle sebastis*. Most polyopisthocotyleans have not been reported to be overtly pathogenic (Paperna, 1987; Thoney & Hargis, 1991), but mortalities were described in various cultured fish species (Kubota & Takakuwa, 1963; Silan et al., 1985; Faisal & Imam, 1990). Polyopisthocotyleans are sanguivorous (Llewellyn, 1954; Halton & Jennings, 1965). Though a small amount of blood is ingested by each worm, an heavy infection of the parasites should cause the host fish to loose lots of blood. Thoney (1986) suggested that heavy infection of polyopisthocotyleans on individual host can be pathogenic and cause mortalities.

Praziquantel chemotherapy has been employed to control various internal helminth infections in mammals, and has recently been used to control monogenean diseases in fish by bath (Schmahl and Melhorn, 1985; Moser et al., 1986; Buchmann, 1987; Schmahl and Taraschewski, 1987; Schmahl et al., 1989; Buchmann et al., 1990; Szekely and Molnar, 1990; Thoney, 1990; Santamarina et al., 1991). Although Kim et al. (1998) and Kim & Cho (2000) recently reported the treatment efficacy of praziquantel after oral and bath treatments under the laboratory and pilot net-pen conditions, no experiment has been conducted under commercial rockfish-culture conditions. In the present study, therefore, the efficacy of praziquantel against *M. sebastis* infestation under commercial rockfish culture conditions was evaluated.

Three net-pens, each containing about 10,000 rockfish weighing about 200 g, in a local rockfish farm were used for the experiment. Before feeding prazi-

*Corresponding author: khkim@pknu.ac.kr

quantel-supplemented diets, the consumed amount of a feed given at a.m. was enumerated for 3 days, and the quantity of praziquantel added to the food was calculated so that the amount of praziquantel ingested daily by fish was 200 mg or 400 mg/kg body weight (bw). Each group of fish in the three net-pens was designated as control (C), 200 mg of praziquantel per kg bw (200P), and 400 mg of praziquantel per kg bw (400P). The fish in the group C were fed a moist-pelleted fish meal (control diet) throughout the experiment. The fish in the groups 200P and 400P were fed diets supplemented with praziquantel at an rate of 200 mg or 400 mg of praziquantel per kg bw, respectively, for 3 times at an interval of 24 h. The water temperature was 19~20°C. On 1 day to 8 days after the treatment, 10 fish were taken randomly from each net-pen daily, and the efficacy of each treatment was confirmed by the comparison of the number of parasites in each treatment group with that in the control group. All data were analyzed using Mann-Whitneys *U*-test (SPSS 7.5 for Windows, SPSS Inc.), $P < 0.05$ being taken as the minimum significance level.

Groups fed praziquantel-supplemented diets (200

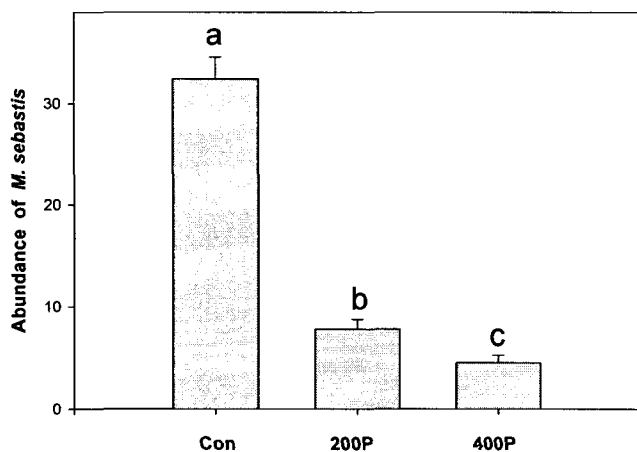


Fig. 1. Abundance (total number of parasites/number of examined fish) of *Microcotyle sebastis* in each experimental group. Values are mean \pm standard error (80 fish in each group) and different letters indicate statistical significance at $P < 0.01$ (Con, fed control diet; 200P, fed three doses of praziquantel 200 mg/kg of body weight given at 24 h intervals; 400P, fed three doses of praziquantel 400 mg/kg of body weight given at an interval of 24 h).

P and 400P) showed significantly lower ($P < 0.01$) abundances (total number of parasites/number of examined fish) of *M. sebastis* than control (Fig. 1). The abundance of group 400P was significantly lower than that of group 200P ($P < 0.01$).

The results clearly demonstrated that feeding praziquantel-supplemented diet was effective for controlling *M. sebastis* infestation in practical commercial rockfish-culture systems without imposing any handling stress. The difference in feeding amount of treatment diet among individuals in groups 200P and 400P might be a reason of the variation of treatment efficacy.

Acknowledgement

This study was supported by a grant from the Dae Sung Microbiological Labs. Co., and a grant from the Ministry of Maritime Affairs and Fisheries, Republic of Korea.

References

- Buchmann, K. 1987. The effects of praziquantel on the monogenean gill parasite *Pseudodactylogyrus bini*. Acta Vet. Scand., 28, 447~450.
- Buchmann, K., C. Szekely and J. Bjerregaard. 1990. Treatment of *Pseudodactylogyrus* infestations of *Anguilla anguilla* II. Trials with bunamidine, praziquantel and levamisole. Bull. Eur. Ass. Fish Pathol., 10, 118~120.
- Faisal, M. and E.A. Imam. 1990. *Microcotyle chrysophrii* (Monogenea: Polyopisthocotylea), a pathogen for cultured and wild gilthead seabream, *Sparus aurata*. In: F.O. Perkins and T.C. Cheng (Editors), Pathology in marine science, Academic Press, Inc., pp. 283~290.
- Halton, D.W. and J.B. Jennings. 1965. Observations on the nutrition of monogenetic trematodes. Bio. Bull., 129, 257~272.
- Kim, K.H. and J.B. Cho. 2000. Treatment of *Microcotyle sebastis* (Monogenea: Polyopisthocotylea) infestation with praziquantel in an experimental cage simulating commercial rockfish *Sebastes schlegeli* culture conditions. Dis. Aquat. Org., 40, 229~231.
- Kim, K.H., S.I. Park and B.Y. Jee. 1998. Efficacy of oral administration of praziquantel and mebendazole against *Microcotyle sebastis* (Monogenea) infestation of cultured rockfish (*Sebastes schlegeli*). Fish Pathol., 33, 467~471.
- Kubota, S.S. and M. Takakuwa. 1963. Studies on the diseases of marine cultured fishes. I. General description and preliminary discussion of fish diseases in Mie prefecture. J. Fac. Fish. Prefect. Univ. Mie-Tsu, 6, 107~124.
- Llewellyn, J. 1954. Observation on the food and gut pigment

- of the Polyopisthocotylea (Trematoda: Monogenea). Parasitology, 44, 428~437.
- Moser, M., J. Sakanari and R. Heckmann. 1986. The effects of praziquantel on various larval and adult parasites from freshwater and marine snails and fish. J. Parasitol., 72, 175~176.
- Paperna, I. 1987. Solving parasite-related problems in cultured marine fish. Int. J. Parasitol., 17, 327~336.
- Santamarira, M.T., J.L. Tojo, F.M. Ubeira, P. Quintero and M.L. Sanmartin. 1991. Anthelmintic treatment against *Gyrodactylus* sp. infecting rainbow trout *Oncorhynchus mykiss* Dis. Aquat. Org., 10, 39~43.
- Schmahl, G. and H. Mehlhorn. 1985. Treatment of fish parasites. 1. Praziquantel effective against Monogenea (*Dactylogyrus vastator*, *Dactylogyrus extensus*, *Diplozoon paradoxum*). Z. Parasitenkd., 71, 727~737.
- Schmahl, G. and H. Taraschewski. 1987. Treatment of fish parasites. 2. Effects of praziquantel, niclosamide, levamisole-HCl, and metrifonate on Monogenea (*Gyrodactylus aculea*, *Diplozoon paradoxum*). Parasitol. Res., 73, 341~351.
- Schmahl, G., H. Taraschewski and H. Mehlhorn. 1989. Chemotherapy of fish parasites. Parasitol. Res., 75, 503~511.
- Silan, P., P. Cabral and C. Maillard. 1985. Enlargement of the host range of *Polyabris tubicirrus* (Monogenea, Polyopisthocotylea) under fish farming conditions. Aquaculture, 47, 267~270.
- Szekely, C. and K. Molnar. 1990. Treatment of *Ancylodiscoides vistulensis* monogenean infestations of the European catfish (*Silurus glanis*). Bull. Eur. Assoc. Fish Pathol., 10, 74~77.
- Thoney, D.A. 1986. Post-larval growth of *Microcotyle sebastis* (Platyhelminthes: Monogenea), a gill parasite of black rockfish. Trans. Am. Microsc. Soc., 105, 170~181.
- Thoney, D.A. 1990. The effects of trichlorfon, praziquantel and copper sulphate on various stages of the monogenean *Benedeniella posterocolpa*, a skin parasite of the cownose ray, *Rhinoptera bonasus* (Mitchill). J. Fish Dis., 13, 385~389.
- Thoney, D.A. and W.J. Hargis, Jr. 1991. Monogenea (Platyhelminthes) as hazards for fish in confinement. Ann. Rev. Fish Dis., 1, 133~153.