

Studies on Fungi Isolated from Dermatomycoses Patients in Egypt

A. H. M. El-Said*

Botany Department, Faculty of Science, South Valley University, Qena, Egypt

(Received December 6, 2001)

Fifty cases of dermatomycoses were recorded from adult male and female at Qena Governorates. These included tinea capitis (62% of total cases), tinea corporis (20%), tinea versicolor (12%) and tinea unguium (6%). Males are more susceptible to all cases of tinea than females. Thirty-one species and 2 varieties belonging to 16 genera were recovered from several infection sites. These were identified as *Aphanoascus fulvescens*, *A. terreus*, *Arthroderma fulva*, *A. obtusa*, *Trichophyton rubrum* and *T. soudanense*. Several saprophytes were also found. These were: *Aspergillus flavus*, *A. fumigatus*, *A. niger*, *A. terreus*, *Cochliobolus lunatus*, *Mycosphaerella tassiana*, *Penicillium chrysogenum* and *P. citrinum*. Twenty-one isolates were able to hydrolyze gelatin with variable capabilities. *T. rubrum* was the most active protease producer. The maximum production of protease was obtained at 8 days of incubation at 30°C in Sabouraud's basal medium with maltose as a carbon source and pepton as a nitrogen source. The optimal pH for the maximum production of protease was pH 6.

KEYWORDS: Dermatomycoses, Protease

Dermatophytes and other pathogenic fungi have been isolated from keratinized part of body in animals, birds and human (Gugnani *et al.*, 1975; Hubalek and Hornick, 1977; Abdel-Hafez, 1987; Ali-Shtayeh *et al.*, 1988a, b; Nicholls and Midgley, 1989; Lee *et al.*, 1990; Abdel-Hafez *et al.*, 1995; and others).

It was reported that protease had an important role in the pathogenicity of a number of microorganisms (Meevootison and Niederpruem, 1979). The ability of dermatophytes and other moulds to produce this enzyme was investigated by many researchers (Drucker, 1972; Cohen, 1973; Takiuchi *et al.*, 1982; Sanyal *et al.*, 1985; Abdel-Hafez *et al.*, 1995 and other). The optimum conditions for protease activity were studied by others (Mohamed and Turner, 1983; Afzal and Chadhary, 1991; Abdel-Hafez *et al.*, 1995).

The present investigation is aimed to study the prevalence of dermatophytic diseases and associated fungi in adult male and female in Qena Governorates (Egypt) and their capability of protease production.

Materials and Methods

Collection of specimens. Fifty specimens were collected from various parts of patient bodies (adult male and female) from march to july in 2000 at dermatology hospital in Qena Governorates. The patients were clinically diagnosed as tinea capitis, tinea corporis (tinea circinata), tinea versicolor and tinea unguium (onychomycosis). Skin scraping was collected using sterile scalpel or glass slides and was placed in sterile labelled petridishes.

Examination and culturing of dermatophytic specimens.

1-Microscopic examination: Specimens were mounted in 20% potassium hydroxide on a glass slide and covered with a slip. The preparation was warmed gently and left for 20 min before examination (Rhode and Hartmann, 1980). One drop of lactophenol cotton blue was then added and the specimens were examined using a light microscope (in positive cases the fungal hyphae, arthrospores or budding yeast cells were observed).

2-Isolation and identification of fungi: When microscopic examination was positive, the dermatophytic specimen was deposited on the surface of Sabourauds dextrose agar medium. Plates were incubated at 25°C for 2-3 weeks. The growing fungi were isolated by baiting technique. The identification was carried out on the basis of macro- and microscopic characteristics (Carmichael, 1962; Raper and Fennell, 1965; Frey *et al.*, 1979; Domsch *et al.*, 1980; Pitt, 1985).

Proteolytic activity.

1-Screening of fungal isolates for protease production: Twenty-one isolates recovered from tinea capitis (14 isolates), tinea corporis (5) and tinea unguium (onychomycosis) (2) were screened for their abilities to produce protease. Using a sterile cork borer (9 mm diameter), the agar block inoculum was prepared. Also, holes were punched into the gelatin peptone agar. Three plates were used for each organism. The inoculum of each culture was put in the hole of gelatin peptone agar medium. Plates were incubated for 8 days at 25°C.

Plates were flooded with mercuric chloride solution (HgCl₂, 15 g; conc. HCl, 20 ml; dist. water, 100 ml). Plates were examined for the appearance of uncolored zone, indicating protease activity. The average diameter of clear

*Corresponding author <E-mail: husseinsaid@yahoo.com>

Table 1. Distribution of dermatophytic diseases according to sex, number and percentage of positive cases shown by direct microscopic examination and protease production

Clinical diagnosis	Number of cases	%	Sex				**	
			Male		Female		+ ve cases	% of + ve cases
			No.	%	No.	%		
Tinea capitis	31	62	20	64.5	11	35.5	28	90.3
Tinea corporis	10	20	7	70	3	30	7	70
Tinea versicolor	6	12	5	83.3	1	16.7	4	66.7
Tinea unguium	3	6	2	66.7	1	33.3	3	100
Total number of cases	50	100	34	68	16	32	42	84

**Direct microscopic examination.

zones (in mm) for each isolate was measured.

2-Factors affecting protease production: The effect of physiological and nutritional factors on protease production by *Trichophyton rubrum* were studied.

a-Effect of temperature and time course: Three flasks containing 30 ml of Sabouraud liquid medium (pH 6.8), were incubated at 20, 25, 30 and 40°C. Flasks were removed at 2, 4, 6, 8, 10, 12 and 14 days and the mycelia were harvested by filtration using Whatmann filter paper. Filtrates from triplicate samples were combined and the combined filtrate was assayed for protease activity.

b-Effect of pH values: The test organism was cultured in the same medium which was initially adjusted to different pH values ranging from 2 to 12. Adjustment of pH was made by 0.1 N HCl or NaOH. After incubation at 30°C for 8 days, cultures were filtered and tested for enzyme activity.

c-Effect of different carbon sources: The culture medium was supplemented with 1% of each of the following carbon sources: cellulose, glucose, maltose, sucrose, wheat bran and wheat straw. Cultures were incubated at 30°C for 8 days. Filtrate was assayed for protease activity.

d-Effect of various nitrogen sources: NaNO₃, KNO₃, NH₄NO₃, casein, gelatin, peptone, urea and yeast extract were used as a sole nitrogen source in the culture medium. Cultures containing peptone were used as a control. After incubation at 30°C for 8 days, cultures were filtered and assayed for enzyme activity.

Assay for protease activity. Protease activity was determined according to the method described by Rick (1963). This method is based on hydrolytic action. The release of soluble tyrosine was estimated by Folin reagent. The activity was calculated from the difference between control and experimental titration value. Enzyme activity was expressed by units of L-tyrosine liberated during the reaction. The units were determined by a standard curve of L-tyrosine.

Results and Discussion

Tinea capitis. In this study, tinea capitis was the most

common dermatophytes and was emerged in 62% of total cases (Table 1). This is in a good agreement with the previous results (Chadegani *et al.*, 1987; Ekanem and Gugnani, 1987; El-Gendy, 1988; Zohdi *et al.*, 1988; Mahmoud, 1991; El-Shanawany, 1993; Abdel-Hafez *et al.*, 1995). Male was more affected by tinea capitis (64.5% of total cases) than female (35.5%) (Table 1). This result is in harmony with the finding of Zaini and Chagari (1989). *Arthroderma* (anamorph: *Nannizzia*) and *Trichophyton* were two main dermatophytes found in tinea capitis, comprising 29.03% and 22.58% of cases, respectively. Of the above two genera, two most prevalent species were *Arthroderma obtusa* (anamorph: *Nannizzia obtusa*, 22.58% of cases) and *T. soudanense* (16.13%). *Arthroderma fulva* (anamorph: *Nannizzia fulva*) and *Trichophyton rubrum* were less common (6.45% and 6.45%, respectively) (Table 2). These dermatophytes were isolated previously from cases of tinea capitis in Egypt (El-Gendy, 1988; Mahmoud, 1991; El-Shanawany, 1993; Abdel-Hafez *et al.*, 1995), as well as in other parts of the world (Al-Sogair *et al.*, 1989; El-Benhawi *et al.*, 1991). Several saprophytes were associated with cases of tinea capitis. These were *Aphanoascus fulvescens*, *A. terreus*, *Aspergillus flavus*, *A. fumigatus*, *A. niger*, *A. terreus*, *Cochliobolus luntus*, *Mycosphaerella tassiana*, *Penicillium chrysogenum* and *P. funiculosum* (Table 2).

Tinea corporis. Tinea corporis (tinea circinata) ranked second in incidence and distribution among patient of dermatophytes and was recovered from 20% of total cases (Table 1). This is almost in harmony with the results of Chadegani *et al.* (1987), Ekanem and Gugnani (1987), El-Gendy (1988) and Abdel-Hafez *et al.* (1995). Results revealed that male (70% of total cases) is more susceptible to this disease than female (30%). This is almost in accordance with the previous results from Egypt (El-Shanawany, 1993; Abdel-Hafez *et al.*, 1995) as well as from Libya (Elghoul *et al.*, 1989). *Arthroderma obtusa* and *T. rubrum* were two main causative species of tinea corporis and were recovered from 50% and 20% of cases, respectively (Table 2). The above dermatophytes were encoun-

tered previously in several cases of tinea corporis from patients in some Egyptian Governorates (Zohdi *et al.*, 1988; Mahmoud, 1991; El-Shanawany, 1993 and Abdel-Hafez *et al.*, 1995) as well as from many cases in other

parts of the world (Shekelakov *et al.*, 1984; El Ghoul *et al.*, 1989; Katoh *et al.*, 1991 and others). *A. flavus*, *A. fumigatus* and *M. tassiana* were rarely recovered in cases of tinea corporis. Abdel-Hafez *et al.* (1995) found that

Table 2. Incidence (I) and percentage incidence (I) of dermatophytes, related fungi and other fungi associated with the different mycotic diseases

Clinical diagnosis	Tinea capitis		Tinea corporis		Tinea unguium		Tinea versicolor		Total cases	
	I	I %	I	I %	I	I %	I	I %	I	I %
I-Dermatophytes and related fungi										
<i>Aphanoascus</i>	3	10							3	6
<i>A. fulvescens</i>	1	3							1	2
<i>A. terreus</i>	2	7							2	4
<i>Arthroderma</i>	9	29	5	50			1	17	15	30
<i>A. fulva</i>	2	6							2	4
<i>A. obtusa</i>	7	23	5	50			1	17	13	26
<i>Trichophyton</i>	7	23	2	20	1	33			10	20
<i>T. rubrum</i>	2	7	2	20	—				4	8
<i>T. soudanense</i>	5	16			1	33			6	12
II-Other fungi										
<i>Aspergillus</i>	8	26	2	20					10	20
<i>A. flavus</i>	4	13	1	10					5	10
<i>A. fumigatus</i>	2	7	1	10					3	6
<i>A. niger</i>	1	3							1	2
<i>A. terreus</i>	1	3							1	2
<i>Cochliobolus lunatus</i>	1	3			2	67	2	33	5	10
<i>Mycosphaerella tassiana</i>	1	3	1	10			3	50	5	10
<i>Penicillium</i>	2	6							2	4
<i>P. chrysogenum</i>	1	3							1	2
<i>P. funiculosum</i>	1	3							1	2
Total isolates	31	100	10	100	3	100	6	100	50	100
Number of genera	7	—	4	—	2	—	3	—	7	—
Number of species	14	—	5	—	2	—	3	—	14	—

Table 3. Proteolytic activities (diameter in mm. of clear zones hydrolyzed) and degree of proteolysis of fungal isolates recovered from ringworm

Organisms	Tinea capitis		Tinea corporis		Tinea unguium		Tinea versicolor	
	M + SD	PA	M + SD	PA	M + SD	PA	M + SD	PA
I-Dermatophytes and related fungi								
<i>Aphanoascus</i>								
<i>A. fulvescens</i>	19 + 0.82	W	—	—	—	—	—	—
<i>A. terreus</i>	24 + 0.82	W	—	—	—	—	—	—
<i>Arthroderma</i>								
<i>A. fulva</i>	35 + 0.82	M	—	—	—	—	—	—
<i>A. obtusa</i>	32 + 0.82	M	37 + 0.82	M	—	—	27 + 0.82	W
<i>Trichophyton</i>								
<i>T. rubrum</i>	-ve	—	54 + 0.82	H	—	—	—	—
<i>T. soudanense</i>	47 + 0.82	M	—	—	19 + 0.82	W	—	—
II-Other fungi								
<i>Aspergillus</i>								
<i>A. flavus</i>	58 + 0.82	H	27 + 0.82	W	—	—	—	—
<i>A. fumigatus</i>	22 + 0.82	W	18 + 0.82	W	—	—	—	—
<i>A. niger</i>	33 + 0.82	M	—	—	—	—	—	—
<i>A. terreus</i>	28 + 0.82	W	—	—	—	—	—	—
<i>Cochliobolus lunatus</i>	-ve	-ve	—	—	-ve	-ve	-ve	-ve
<i>Mycosphaerella tassiana</i>	-ve	-ve	-ve	-ve	—	—	-ve	-ve
<i>Penicillium</i>								
<i>P. chrysogenum</i>	44 + 0.82	M	—	—	—	—	—	—
<i>P. funiculosum</i>	20 + 0.82	W	—	—	—	—	—	—

M = mean; SD = standard deviation; PA = degree of proteolysis activity : H = high proteolysis, 50–70; M = moderate proteolysis, 30–49; W = weak proteolysis, less than 30 mm; -ve = nonproteolytic isolates; — = isolates not appear.

A. flavus and *A. fumigatus* were recovered with low incidences in cases of tinea corporis.

Tinea versicolor. Tinea versicolor occurred in 6 cases out of 50 (12%) (Table 1). This is almost in agreement with the previous results (El-Gendy, 1988; Inwidthaya *et al.*, 1989; Mahmoud, 1991; El-Shanawany, 1993; Abdel-Hafez *et al.*, 1995). Male was more susceptible than female (83.3% and 16.7% of cases, respectively). Similar findings were recorded in patients (El-Shanawany, 1993; Abdel-Hafez *et al.*, 1995). *Malassezia furfur*, the causative fungus of tinea versicolor, was detected by direct microscopic examination and was recovered in 66.7% of cases of tinea versicolor (Table 1).

Tinea unguium (onychomycosis). Three cases of tinea unguium were found (6% of total cases) (Table 1). Abdel-Hafez *et al.* (1995) found that this type of infection of the skin was least common (4.3% of total cases). In Egypt, onychomycosis was infrequent among patients of der-

matophytic disease (El-Gendy, 1988; Mahmoud, 1991; El-Shanawany, 1993; Abdel-Hafez *et al.*, 1995). In present study, one dermatophytic fungus, *T. soudanense* was recovered from 33.3% of total cases of tinea unguium. *C. lunatus* emerged in 66.7% of cases of tinea unguium. It was reported that some onychomycosis were caused by saprophytes such as members of *Alternaria*, *Aspergillus*, *Fusarium*, *Penicillium*, *Scopulariopsis* and some unidentified dermatiaceous species (Velez and Diaz, 1985; Wadhawani and Srivastava, 1985). On the other hand, several of the above fungi were frequently recovered from skin (Abdel-Hafez *et al.*, 1995).

Proteolytic activity of dermatophytes and some other moulds. Isolates were screened for their ability to produce protease. Most of isolates had the ability to produce protease with variable degree of activity (Table 3). High proteolytic activity was exhibited by *A. flavus* and *T. rubrum*. Isolates of *A. niger*, *A. fulva*, *A. obtusa*, *T. soudanense* and *P. chrysogenum* showed moderate pro-

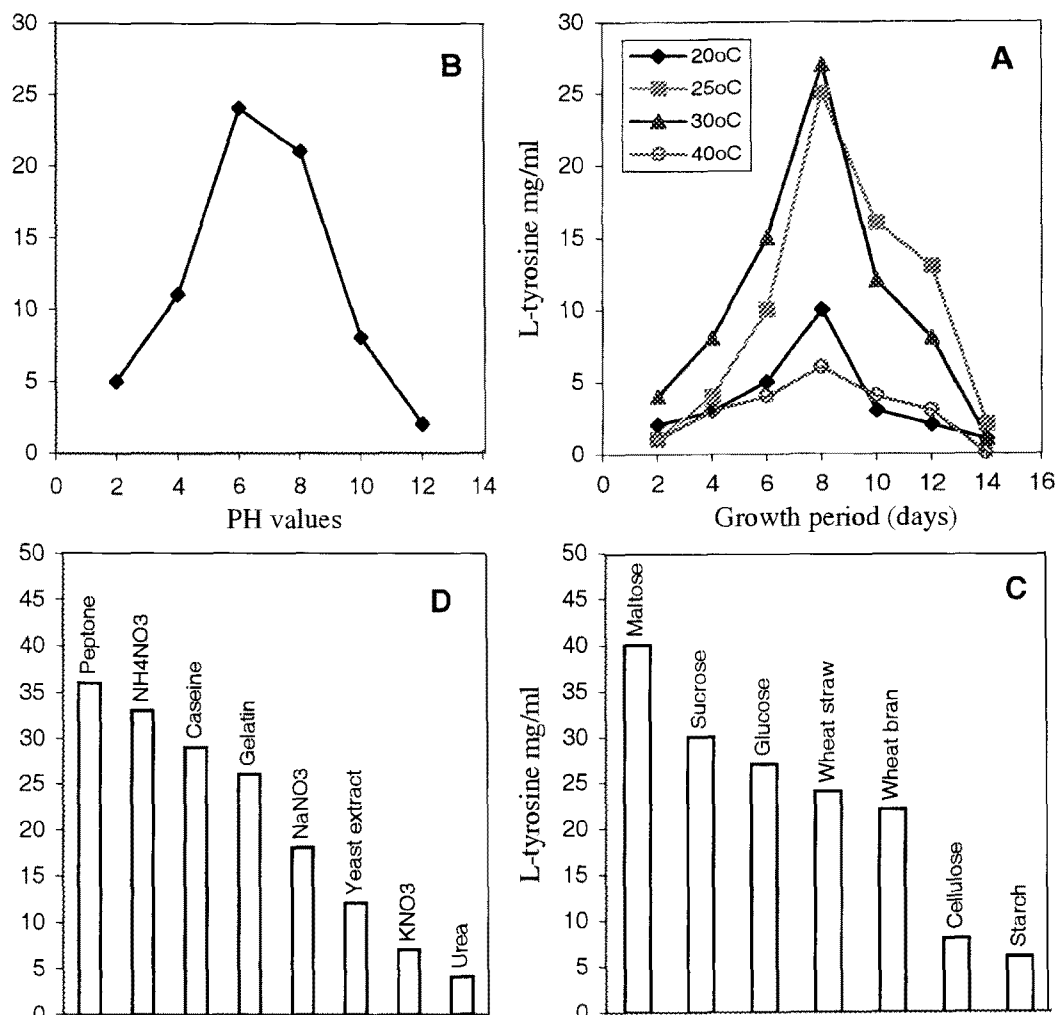


Fig. 1. Effect of time course and temperature (A), pH values, (B), different carbon (C) and nitrogen sources (D) on production of protease by *Trichophyton rubrum*.

teolytic activity. The remaining isolates were of weak or non-proteolytic activity. Abdel-Hafez *et al.* (1995) found that some isolates of *Aspergillus*, *Nannizzia*, *Penicillium*, *Scopulariopsis* and *Trichophyton* showed good protease activity.

T. rubrum was chosen to study the effects of different physiological and nutritional factors on protease production, since it was the most active dermatophytic producer (Table 3). The maximum protease production by *T. rubrum* was observed after 8 days of incubation at 30°C (Fig. 1). Abdel-Hafez *et al.* (1995) found that the protease production by *T. soudanense* was maximum after 8 days of incubation at 30°C.

The optimal pH for protease production by *T. rubrum* was recorded within a pH 6 (Fig. 1). Groninger and Eklund (1966), Lenney and Dalbec (1967) and Olutiola and Nwaogwugwu (1982) found that proteases from *Trichosporon* sp., *Saccharomyces cerevisia* and *Aspergillus aculeatus* were highly active at pH 7. In addition, Abdel-Hafez *et al.* (1995) found that the maximum protease production by *T. soudanense* was within the range of pH 6-8.

Maltose was the best carbon source for proteolytic activity of *T. rubrum* (Fig. 1). Maximum protease production was obtained by the incorporation of sucrose as carbon source in the growth medium of *A. aculeatus* (Olutiola and Nwaogwugwu, 1982), *A. niger* (Chopra and Mehta, 1985), *P. chrysogenum* (Mahmoud, 1988). Abdel-Hafez *et al.* (1995) found that maltose followed by sucrose were the the best carbon sources for proteolytic activity by *T. soudanense*.

Peptone followed by ammonium nitrate and casein were the best nitrogen sources for protease production by *T. rubrum* (Fig. 1). Abdel-Hafez *et al.* (1995) also found that casein followed by gelatin and ammonium nitrate were the best nitrogen sources for protease production by *T. soudanense*.

In conclusion, dermatomycoses recorded in this investigation were tinea capitis, tinea corporis, tinea versicolor and tinea unguium. Males are more susceptible to tinea than females. Fungal isolates of dermatophytes and other moulds were able to produce protease enzyme with variable capabilities. *T. rubrum* was the most active protease producer and the maximum production were obtained 8 days after incubation at 30°C with the incorporation of maltose as a carbon source and pepton as a nitrogen source in Sabourauds basal medium initially adjusted to pH 6.

References

- Abdel-Hafez, A. I. I. 1987. Survey on the mycoflora of goat and sheep hairs from Gaza Strip. *Bull. Fac. Sci. Assiut Univ.* **16**: 16-21
- Abdel-Hafez, S. I. I., El-Said, A. H. M. and Maghraby, T. A. 1995. Studies on fungi isolated from skin diseases and associated fungi of students in Qena and Red Sea Governorates, *Egypt. Bull. Fac. Sci., Assiut Univ.* **24**(2-D): 181-209.
- Afzal, M. M. and Cgaudhary, M. I. 1991. Effect of carbon sources on the production of alkaline protease by *Aspergillus oryzae*. *Pak J. Sci. Ind. Res.* **34**: 91-94.
- Al-Sogair, S. M., Al-Humaidan, Y. M. and Moawad, M. K. 1989. Scalp fungus infection in the eastern province of Saudi Arabia. *Annals of Saudi Medicine* **9**: 259-262.
- Ali-Shtayeh, M. S., Arda, H. M., Hassouna, M. and Shaheen, S. F. 1988a. Keratinophilic fungi on the hair of goats from the West Bank of Jordan. *Mycopathologia* **104**: 103-108.
- _____, _____, _____ and _____. 1988b. Keratinophilic fungi on the hair of cows, donkeys, rabbits, cats and dogs from the West Bank of Jordan. *Mycopathologia* **104**: 109-121.
- Carmichael, J. W. 1962. *Chrysosporium* and some others Alerui-
osporic Hypomycetes. *Canada. J. Bot.* **40**: 1137-1172.
- Chadegani, M., Moment, A., Shadzi, S. and Amin Javaheri, M. 1987. A study of dermatophytoses in Esfahan (Iran). *Mycopathologia* **98**: 101-104.
- Chopra, S. and Mehta, P. 1985. Influence of various nitrogen and carbon sources on the production of pectolytic, cellulytic and proteolytic enzymes by *Aspergillus niger*. *Folia Microbiol.* **30**: 117-125.
- Cohen, B. L. 1973. Regulation of intracellular and extra cellular neutral and acid protease in *Aspergillus nidulans*. *J. Gen. Microbiol.* **79**: 311-320.
- Domsch, K. H., Gams, W. and Anderson, T. H. 1980. Compendium of soil fungi. Acad. Press, London, p. 859.
- Drucker, H. 1972. Regulation of exocellular protease in *Neurospora crassa*: Induction and repression of enzyme synthesis. *J. Bacteriol.* **110**: 1041-1049.
- Ekanem, L. S. and Gugnani, H. C. 1987. Etiology of dermatophytoses among school children in Cross River State of Nigeria. *Mykosen* **30**: 493-498.
- El-Benhawi, M. O., Fathy, S., Moubasher, A. H. and Alem, N. S. 1991. Mycologic study of tinea capitis in Qatar. *Int. J. Dermatol.* **30**: 204-205.
- El-Gendy, Z. K. A. 1988. Studies on dermatophytes in Minia Governorate, M. Sc. Thesis. Bot. Dept. Fac. Sci. Minia Univ., Egypt.
- El-Shanawany, A. A. 1993. Human dermatophytes in Assiut and New Valley Governorates, Ph. D. Thesis, Bot Dept., Fac. Sci., Assiut Univ., Egypt, p. 151.
- Elghoul, M. T., Safi, M., Joshi, R. M. and Mashina, H. 1989. Frequency of dermatophytes causing tinea corporis in Tripoli, Libya. *Trans. royal Soc. Tropical Medicine and Hygien* **83**: 418.
- Frey, D., Oldfield, R. J. and Bridger, R. C. 1979. A colour atlas of pathogenic fungi. Wolfe Medical publications Ltd., Smeets-Weert, Holland, p. 168).
- Groninger, H. S. and Eklund, M. W. 1966. Characteristics of a proteinase of a *Trichosporon* species isolated from Dungeness crab meat. *Applied Microbiol.* **14**: 110-114.
- Gugnani, H. C., Wattal, B. L. and Sandhu, R. S. 1975. Dermatophytes and other keratinophilic fungi recovered from small mammals in India. *Mykosen* **18**: 529-538.
- Hubalek, Z. and Hornick, M. 1977. Experimental infection of white mouse with *Chrysosporium* and *Paeecilomyces*. *Mycopath. Mycol. Appl.* **62**: 173-178.
- Imwidthaya, S., Tianprasit, M. and Srimuang, S. 1989. A study of

- pityriasis versicolor in Bangkok (Thailand). *Mycopathologia* **105**: 157-161.
- Katoh, T., Nishioka, K. and Sano, T. 1991. Isolation of *Microsporum canis* from clinically normal skin. *Japanese Journal of Medical Mycology* **32**: 127-131.
- Lee, M. M., Diven, D. G., Smith, E. B. and Pupo, E. B. 1990. Onychomycosis. *Archives of Dermatology* **126**: 402.
- Lenney, J. R. and Dabec, J. M. 1967. Purification and properties of two proteases from *Saccharomyces cerevisiae*. *Archives of Biochemistry and Biophysics* **120**: 42-48.
- Mahmoud, A. L. E. 1988. Some physiological studies on fungi isolated from Poultry feedstuffs. M. Sc. Thesis, Bot. Dept. Fac. Sci., Assiut Univ., Egypt, p. 161.
- _____. 1991. Some physiological and biochemical studies of fungi isolated from skin of mammals. Ph.D. Thesis. Bot Dept. Fac. Sci., Assiut Univ., Egypt, p. 173.
- Meevootisom, V. and Niederpruem, D. J. 1979. Control of extracellular proteases in dermatophytes and especially in *Trichophyton rubrum*. *Sabouraudia* **17**: 91-106.
- Mohamed, A. K. A. and Turner, A. G. 1983. Proteolytic activity of *Nomuraea rileyi* and host insect cuticle. *Mycopathologia* **82**: 13-15.
- Nichoils, D. S. H. and Midgley, G. 1989. Onychomycosis caused by *Trichophyton equinum*. *Clinical and Experimental Dermatology* **14**: 464-465.
- Ogbonna, C. I. C., Robinson, R. O. and Abubakar, J. M. 1985. The distribution of ringworm infections among primary school children in Jos, Plateau State of Nigeria. *Mycopathol.* **89**: 101-106.
- Olutiola, P. O. and Nwaogwu, R. I. 1982. Growth, Saporulation and production of maltase and proteolytic enzymes in *Aspergillus aculeatus*. *Trans. Br. Mycol. Soc.* **78**: 105-113.
- Pitt, J. I. 1985. A laboratory guide to common *Penicillium* species. Commonwealth Scientific and Industrial Research Organization, Division of Food Research, p. 184.
- Raper, K. B. and Fennell, D. J. 1965. The genus *Aspergillus*. Williams and Wilkins, Baltimore, USA, p. 686.
- Rhode, B. and Hartmann, G. 1980. Introducing mycology by examples. Hamburg, presented by Schering Aktiengesellschaft.
- Rick, W. 1963. Methods of enzymatic analysis. Academic Press Inc., New York. Trypsin pp. 807-818. In H. U. Bergmeyer (ed.).
- Sanyal, A. K., Das, S. K. and Banerjee, A. B. 1985. Purification and partial characterization of an extracellular protease from *T. rubrum*. *Sabouraudia* **20**: 281-288.
- Shekelakov, N. D., Z. V. Stepanova, Klimova, I. Y. and Mosalova, N. A. 1984. Role of humans as the source of infection in Zooanthropotic, microsporiosis caused by *M. canis*. *Vestnik Dermatologii. I. Venerologii.* **12**: 29-30.
- Takiuchi, I., Higuchi, D., Sei, Y. and Koga, M. 1982. Isolation of an extracellular proteinase, Keratinase from *M. canis*. *Sabouraudia* **20**: 281-288.
- Velez, H. and Diaz, F. 1985. Onychomycosis due to saprophytic fungi. *Mycopathol.* **91**: 87-92.
- Wadhvani, K. and Srivastava, A. K. 1985. Some cases of onychomycosis from North India indifferent working environments. *Mycopathol.* **92**: 149-155.
- Zaini, F. and Ghagari, A. 1989. Epidemiological and mycological studies on tinea capitis at nurserie and schools of Bander Chabhar. *Iranian Pub. Health* **18**: 39.
- Zohdi, H. A. A., Youssef, Y. A., Abdel-Moneim, M. M. A., Farghaly, M. S., Emam, F. M. and Abdallah, M. A. 1988. Study of dermatophytes and dermatophytosis. *Egypt. J. Derm. & Ven.* **8**: 41-50.