

The Radical Scavenging Effects of Stilbene Glucosides from Polygonum multiflorum

Geonseek Ryu, Jeung Hoon Ju, Yong Ju Park, 1 Shi Yong Ryu, 2 Byoung Wook Choi, and Bong Ho Lee

Regional Research Center and Department of Chemical Technology, Hanbat National University, 16-1 Dukmyungdong, Yusung-ku, Daejon, 305-719, Korea, ¹Livechem, INC., Hanbat National University, 16-1 Dukmyung-dong, Yusung-ku, Daejon 305-719, Korea, and ²Korea Research Institute of Chemical Technology, P. O. Box 107, Yusung-ku, Daejon, 305-600, Korea

(Received May 20, 2002)

The extract of the root of *Polygonum multiflorum* exhibited a significant antioxidant activity assessed by the DPPH radical scavenging activity *in vitro*. The bioassay-guided fractionation of the extract yielded a stilbene glucoside, (*E*)-2,3,5,4'-tetrahydroxystilbene-2-O-O-d-glucopyranoside (1) as an active constituent responsible for the antioxidant property. Compound 1 demonstrated a moderate DPPH radical scavenging activity (IC₅₀, 40 μ M), while the corresponding deglucosylated stilbene 2 exhibited a much higher activity (IC₅₀, 0.38 μ M).

Key words: Stilbene glucoside, (*E*)-2,3,5,4'-Tetrahydroxystilbene-2-*O*-β-D-glucopyranoside, *Polygonum multiflorum*, DPPH Radical scavenging activity.

INTRODUCTION

Stilbene glucosides have attracted a great deal of interest because relatively high quantities are found in grapes, wine, and some medicinal plants (Goldberg, 1995; Mattivi et al., 1995; Frankel et al., 1993; Jang et al., 1997; Teguo et al., 1996). They seem to have a variety of biological activities such as antioxidant activity (Teguo et al., 1996; Teguo et al., 1996; Teguo et al., 1998), antiplatelet aggregation activity (Verotta et al., 1994; Orsini et al., 1997; Chen et al., 1994), coronary vasodilator activity (Inamori et al., 1989), antileukemic (Mannila et al., 1993), antifungal activity (Langcake, et al., 1979), and protein-tyrosine kinase (Jayatilake et al., 1993), monoamine oxidase A (Ryu et al., 1988) and gastric H⁺, K⁺-ATPase inhibitory activity (Murakami et al., 1992). Interestingly, many medicinal plants contain large amount of antioxidants other than vitamin C, vitamin E and carotenoids. The antioxidant effect is mainly due to the phenolic components such as flavonoids, phenolic acid, and phenolic diterpenes (Pietta et al., 1998). As part

of our continuous search for bioactive components from Korean medicinal plants, we have focused on the antioxidant effect of the methanol extract of *Polygonum multiflorum* (Polygonaceae). Bioassay-guided isolation of the extract afforded a stilbene glucoside 1 as an active compound of the antioxidant property.

MATERIALS AND METHODS

General experimental procedures

UV spectrum was recorded with a HP8453 UV/VIS spectrophotometer. IR spectrum was performed on a Perkin-Elmer model 1750 FT-IR spectrophotometer. MS spectra were measured on a JEOL JMX-SX 102 mass spectrometer. High resolution mass measurement was done with a JEOL AX-505H mass spectrometer at high resolution using NBA or glycerol as a matrix. ^1H NMR and ^{13}C NMR spectra were recorded in CD_3OD or CDCl_3 at 25°C on a Brucker ARX-400 NMR spectrometer. Chemical shifts (δ) are given relative to TMS, using the solvent peaks [CD_3OD (δ_{H} 3.30, δ_{C} 49.0) and CDCl_3 (δ_{H} 7.26, δ_{C} 77.1)] as the internal standards.

Plant materials

The root bark of *Polygonum multiflorum* was supplied from Hankook Sinyak Pharmaceutical Co., LTD, Taejon, Korea. A voucher specimen (KM-98203) was deposited at

Correspondence to: Byoung Wook Choi, Regional Research Center and Department of Chemical Technology, Hanbat National University, 16-1 Dukmyung-dong, Yusung-ku, Daejon 305-719, Korea

E-mail: choi@hanbat.ac.kr

the Laboratory of Natural Products Chemistry, Hanbat National University.

DPPH radical scavenging assay

Radical scavenging effects on DPPH (α , α' -diphenylpicrylhydrazyl) radical were determined by the method previously described (Ryu *et al.*, 2001).

Extraction and isolation

The dry root bark of *Polygonum multiflorum* (200 g) was extracted with MeOH at room temperature for a day. The MeOH extract (36 g) with the radical scavenging effect of IC₅₀, 26 mg/mL was partitioned between 30% MeOH and CHOL (4.8 g, 13.3% yield); the former layer was further partitioned between n-BuOH (8.3 g, 23%) and H₂O (23.4 g, 65%). The bioactive n-BuOH fraction was subjected to an ODS flash chromatography with aqueous MeOH [0, 20, 40, 60, 100%] to give five fractions. The most active 40% I/IeOH fraction was gel-filtered with MeOH to afford a crude phenolic glycoside fraction (IC₅₀, 4.6 μ g/mL). This fraction was finally purified by reversed-phase HPLC with 25% I/IeON to yield a glycoside 1 (74 mg, 0.89%).

(*E*)-2,3,5,4'-Tetrahydroxystilbene-2-*O*-β-D-glucopyranos de (1)

Brown powder; $[\alpha]_D^{23}$ +28.9° (c 1.2, MeOH); UV λ_{max} (log ϵ) 215 nm (4.16), 306 (4.35) and 318 (4.36); IR (film) v_{max} cm⁻¹ £650 (OH), 1630 and 1540 (aromatic); FABMS (pos) m/z 4.07 [M+H]⁺, 429 [M+Na]⁺; HRFABMS (pos) m/z $[M+H]^{+}$ 407.1353 (calcd 407.1345 for $C_{20}H_{23}O_{9}$); ¹H NMR (CD₂()D, 400 MHz) δ 7.70 (2H, d, 8.2 Hz, H-2' and H-6'), 7.45 (1H, d, 16.4 Hz, H-8), 6.91 (2H, d, 8.2 Hz, H-3' and H-5", 6.76 (1H, d, 16.4 Hz, H-7), 6.59 (1H, d, 1.7 Hz, H-6), 6 24 (1H, d, 1.7 Hz, H-4), 4.50 (1H, d, 7.8 Hz, H-1"), 3.77 (2H, m, H-6"), 3.57-3.54 (2H, overlapped, H-2" and H-4", 3.44 (1H, m, H-3"), 3.27 (1H, m, H-5"); ¹³C NMR (CD₂C)D, 100 MHz) δ 157.8 (C4'), 155.3 (C5), 152.2 (C3), 137.5 (C2), 132.9 (C1'), 130.0 (C8), 129.2 (C1), 128.5×2 (C2' and C6'), 121.0 (C7), 115.8 x 2 (C3' and C5'), 107.6 (C1", 103.1 (C4), 101.6 (C6), 77.5 (C5"), 77.3 (C3"), 74.8 (C2", 70.1 (C4"), 61.4 (C6").

Enzymatic hydrolysis of 1

Still ene glucoside **1** (20 mg) was dissolved in 3mL of AcOH-AcONa buffer solution (pH 5), to which β -glucosidase (10 mg) was added to lay aside at 37°C for 3 days. The reaction mixture was adjusted to neutral with AgNO3 and then filtered. The filtrate was partitioned between AcOEt (10 mL \times 3) and H₂O (10 mL). The organic layer was concentrated *in vacuo* to give (*E*)-2,3,5,4'-tetrahydroxystilbene (**2**, 8.2 mg), while the aqueous layer was further neutralized by passing through an ion-exchange resin (Amberlite MB-3) column, concentrated

(dried overnight), then treated with 1-(trimethylsilyl)imidazole at room temperature for 2 hours. After the excess reagent was decomposed with H_2O , the reaction product was extracted with n-hexane (2 mL \times 2). The TMSi derivative of the monosaccharide was identified to be d-glucose by co-GLC analysis with the TMSi derivative of a standard monosacchride.

2: colorless solid; 1 H NMR (CD₃OD, 400 MHz) δ 6.94 and 6.98 (2H, d, 16.4 Hz, H-7 and H-8), 7.51 (2H, d, 8.3 Hz, H-2' and H-6'), 6.82 (2H, d, 8.3 Hz, H-3' and H-5'), 6.45 (1H, d, 1.7 Hz, H-6), and 6.18 (1H, d, 1.7 Hz, H-4); FABMS (pos) m/z 245 (M+H)⁺ and 267 (M+Na).⁺

Acetylation of 1

A mixture of stilbene glucoside **1** (18 mg), Ac₂O (1.0 mL), and pyridine (1.0 mL) was stirred at room temperature overnight. The reagents were evaporated *in vacuo* and the residue was subjected to SiO₂ column chromatography [CH₂Cl₂/MeOH (95:5)] to yield a heptaacetyl stilbene glucoside **3** (28 mg).

3: white solid; ^1H NMR (CDCl $_3$, 400 MHz) δ 7.44 (2H, d, 8.2 Hz, H-2' and H-6'), 7.27 (1H, d, 16.3 Hz, H-8), 7.00 (2H, d, 8.2 Hz, H-3' and 5'), 6.93 (1H, d, 16.3 Hz, H-7), 6.77 (1H, br s, H-6), 6.49 (1H, br s, H-4), 5.22 (1H, t, 7.8 Hz, H-3"), 5.15 (2H, overlapped, H-2" and H-4"), 4.86 (1H, d, 7.9 Hz, H-1"), 4.22 and 3.85 (2H, br d, 12.6 Hz, H-6"); 3.54 (1H, m, H-5"), and 2.25, 2.22 × 2, 2.00, 1.95, 1.92, and 1.86 (7 singlet acyl methyls); ^{13}C NMR (CDCl $_3$, 100 MHz) δ 170.7-168.6 (7 carbonyls), 150.8, 147.6, 144.2, 142.6, 134.9, 133.8, 130.8, 130.3, 128.1 × 2, 122.4 × 2, 116.6 116.4, 101.6, 73.2, 72.4, 71.9, 68.1, 61.6, and 21.3-20.7 (7 acyl methyls); FABMS (pos) m/z 701 (M+H).*

RESULTS AND DISCUSSION

Compound **1** was obtained as a brown powder: $[\alpha]_D^{23}$ +28.9 (c 1.2, MeOH). A colorizing reaction of **1** with FeCl₃ showed dark blue and the UV spectrum of **1** exhibited bands at 214 nm (log ϵ 4.16), 306 (4.35) and 318 (4.36) to suggest the presence of a stilbene skeleton (Hata *et al.*, 1975). The ¹H NMR spectrum of **1** was similar to that of resveratrol glucoside **5** isolated from *Erythrophleum lasianthum* (Langcake *et al.*, 1979). Enzymatic hydrolysis of **1** yielded a glucose and compound **2**, which was identified as 2,3,5,4'-tetrahydroxystilbene by spectral analysis and chemical modification (compound **3**). Thus, **1** was identified as *trans*-2,3,5,4'-tetrahydroxystilbene-2-O-G-d-glucopyranoside, which was previously reported by Hata *et al.*

Stilbene glucose **1** is structurally similar to resveratrol glucoside **4** (Langcake *et al.*, 1979; Nyemba *et al.*, 1995) and piceatannol glucoside **5** (Geahlen *et al.*, 1989; Thakkar *et al.*, 1993; Oliver *et al.*, 1994). Compounds **1** and **5** exhibited moderate radical scavenging effect on

Table 1. Antioxidant activity of phenolic compounds

$$R_1$$
 R_2
 R_3
 R_4
 R_4
 R_4
 R_4
 R_4
 R_4

- 1 R₁=R₃=R₄=OH, R₂=O-β-D-Glu
- 2 R₁=R₂=R₃=R₄=OH
- 3 R₁=R₃=R₄=OAc, R₂=tetraacetyl-β-D-Glu
- 5 $R_1 = O \beta D Glu$, $R_2 = R_3 = R_4 = OH$
- 4 R₁=R₂= OH, R₃=H, R₄=O-β-D-Glu

Compound	IC ₅₀ (μM)
(E)-2,3,5,4'-tetrahydroxystilbene-2- <i>O</i> -β-D-glucopyranoside (1)	40
(E)-2,3,5,4'-tetrahydroxystilbene (2)	0.38
heptaacetyl stilbene glucoside (3)	> 100
resveratrol-4'-O-β-D-glucopyranoside (4)	1000 ± 95°
piceatannol-3-O-β-p-glucopyranoside (5)	29 ± 1.9^{a}
BHT	1.44
ascorbic acid	0.04
catechin	0.3

^aThese values were cited from a reference (Teguo et al., 1998).

DPPH radical with IC₅₀ values of 40 μ M and 29 μ M, respectively, while compound 4 exhibited a poor activity (IC₅₀, 1000 μM) (Table 1). Rice-Evans et al. asserted that the catechol moiety attached on stilbene skeleton is essential for antioxidant activity of stilbenes (Rice-Evans et al., 1995), and Merillon et al. reported that the glycosylation of stilbene reduces their activity compared to the corresponding aglycone, but stilbenes including the catechol structure dramatically increase antioxidant activity (Teguo et al., 1998). It has also been reported that the antioxidant activity of polyphenols is related to the position of hydroxyl groups. The presence of a second hydroxyl group in the ortho or para position is known to increase the antioxidant activity due to additional resonance stability and o-quinone or p-quinone formation (Chen and Ho, 1997; Bouchet et al., 1998). Their assertions were supported by the fact that deglucosylated stilbene 2 (IC₅₀, 0.38 μM), having a catechol moiety, was nearly a hundred times as potent as stilbene glucoside 1. As expected, acetylated stilbene 3 showed a poor antioxidant activity.

ACKNOWLEDGMENTS

This work was supported by the Korea Science and Engineering Foundation (KOSEF) through the Advanced Material Research Center for Better Environment at Hanbat National University. We thank Mr. D. Y. Hwang at Hankook Sinyak Pharmaceutical Co., LTD for supply of the plant materials and Dr. E. C. Lee, Korea Ginseng & Tobacco Research Institute for nuclear magnetic resonance and mass measurements.

REFERENCES

Aue, W. P., Bartholdi, E., Ernst, R. R., Two dimensional spectroscopy. Application to nuclear magnetic resonance, J. Chem. Phys., 64, 2229-2246 (1976).

A.; Subramanian, S., Sensitivity-enhanced dimensional hetero nuclear shift correlation **NMR** spectroscopy, J. Magn. Res., 67, 565-569 (1986).

Bax, A., Summer, M. F., Proton and carbon-13 assignments from multiple-bond connectivity by 2D multiple quantum NMR, J. Am. Chem. Soc., 108, 2093-2094 (1986).

Bouchet, N., Barrier, L., Fauconneau, B., Radical scavenging activity and antioxidant properties of tannins from Guiera senegalensis (Combretaceae), Phytother. Res., 12, 159-162 (1998)

Chen, C.-C., Wu, L.-G., Ko, F.-N., Teng, C.-M., Antiplatelet aggregation principles of Dendrobium Ioddigesii, J. Nat. Prod., 57, 1271-1274 (1994).

Chen, J. H., Ho, C.-T., Antioxidant activities of caffeic acid and its related hydroxycinnamic acid compounds, J. Agric. Food Chem., 45, 2374-2378 (1997).

Frankel, E. N., Waterhouse, A. L., Kinsella, J. E., Inhibition of human LDL oxidation by resveratrol, Lancet, 341, 1103-1104 (1993).

Geahlen, R. L., Piceatannol(3,4,3',5'-tetahydroxy-trans-stilbene) is a naturally occurring protein-tyrosine kinase inhibitor, Biochem. Biophys. Res. Commun., 165, 241-245 (1989).

Goldberg, D. M., Does wine work, Clini. Chem., 41, 14-16 (1995).

Hata, K., Kozawa, M., Baba, K., New stilbene glucose Chinese crude drug Heshouwu root of Poloygonum multiflorm, Yakugaku Zasshi, 95, 211-213 (1975).

Inamori, Y., Kubo, M., Tsujibo, H., Ogawa, M., Saito, Y., Miki, Y., Takemura, S., The ichthyotoxicity and cornary vasodilator action of 3,3'-diethylstilbene, Chem. Pharm. Bull., 35, 887-890 (1989).

Jang, M., Cai, L., Udeani, G. O., Slowing, K. V., Thomas, C. F., Beecher, C. W. W., Fong, H. S., Farnsworth, N. R., Kinghorn, A. D., Mehta, R. G., Moon, R. C., Pezzuto, J. M., Cancer chemopreventive activity of resveratrol, a natural product derived from grapes, Science, 275, 218-220 (1997).

Jayatilake, G. S., Jayasuriya, H., Lee, E.-S., Koonchanok, N. M., Geahlen, R. L., Ashendel, C. L., McLaughlin, J. L., Chang., C. J., Kinase inhibitors from Polygonum cuspidatum, J. Nat. Prod., 56, 1805-1810 (1993).

Langcake, P., Corngird, C. A., Pryce, R. J., Identification of pterostilbene as pyhtoalexin from Vitis vinifera leaves, Phytochem., 18, 1025-1027 (1979).

Mannila, E., Talvitie, A., Kolehmainen, E., Antileukemic compounds derived from stilbene in Picea abies bark, Phytochem., 33, 813-816 (1993).

Mattivi, F., Reniero, F., Korhammer, S., Isolation, characterization, and evolution in red wine vinification of resveratrol monomers.

- J. Agric. Food Chem., 43, 1820-1823 (1995).
- Murakami, S., Arai, I., Muramatsu, M., Otomo, S., Baba, K., Kido, T., Kozawa, M., Effect of stilbene derivatives on gastric H¹⁻⁻⁻-ATPase, *Biochem. Pharmacol.*, 44, 1947-1951 (1992).
- Nyemba, A. M., Mpondo, N. T., Kimbu, S. F., Connolly, J. D., Stirt ene glycosides from *Guibourtia Tessmannii, Phytochem.*, 39 395-898 (1995).
- Oliver, J. M., Burg, D. L., Wilson, B. S., McLaughlin, J. I., Inhibition of mast-cell FC-*e*-R1-mediated signaling and effector function by the SYK selective inhibitor, piceatannol, *J. ∃iol. Chem.*, 269, 29697-29703 (1994).
- Orsini, F., Pelizzoni, F., Verotta, L., Aburjai, T., Rogers, C. B., Ischation, synthesis, and antiplatelet aggregation activity of resveratrol-3-*O*-β-glucoside and related compounds, *J. Nat. Pro 1.*, 60, 1082-1087 (1997).
- Pietta, P., Simonetti, P., Mauri, P., Antioxidant activity of selected medicinal plants, *J. Agric. Food Chem.*, 46, 4487-4490 (1998).
- Rice-Evans, C. A., Miller, N. J., Bolwell, P. G., Brmaley, P. M., Pridham, J. B., The relative antioxidant activity of plant-derived polyphenolic flavonids, *Free Rad. Res.*, 22, 375-383 (1995).
- Ryu, G., Park, E. K., Joo, J. H., Lee, B. H., Choi, B. W., Jung, D. S. Lee, N. H., A new antioxidant monoterpene glycoside, ábenzoyloxypaeoniflorin from *Paeonia suffruticosa*, *Arch*.

- Pharm. Res., 24, 105-108 (2001).
- Ryu, S. Y., Han, Y. N., Han B. H., Monoamine oxidase-A inhibitors from medicinal plants, *Arch. Pharm. Res.*, 11, 230-239 (1988).
- Teguo, P. W., Decendit, A., Vercauteren, J., Deffieux, G., Merillon, J. M., Trans- resreratrol-3-O-β-glucose (piceid) in cell suspension cultures of *Vitis vinifera*, *Phytochem.*, 42, 1591-1593 (1996).
- Teguo, P. W., Decendit, A., Krisa, S., Vercauteren, J., Deffieux, G., Merillon, J. M., The accumulation of stilbene glycosides in Vitis vinifera cell suspension cultures of Vitis vinifera, J. Nat. Prod., 59, 1189-1191 (1996).
- Teguo, P. W., Deffieux, G., Vercauteren, J., Merillon, J. M., Isolation, identificaion, and antioxidant activity of three stilbene glucosides newly extracted from *Vitis vinifera* cell cultures, *J. Nat. Prod.*, 61, 655-657 (1998).
- Thakkar, K., Geahlen, R. L., Cushman, M., Synthesis and protein-tyrosine kinase inhibitory activity of polyhyroxylated stilbene analogs of piceatannol, *J. Med. Chem.*, 36, 2950-2955 (1993).
- Verotta, L., Rogers, B. C., Aburjai, T., Cignarella, A., Colli, S., Puglisi, L., Giornate di Chimica delle Sostanze Naturali, III convegno Nazionale, Amalfi (SA) 29 maggio-1 giugno, 3 (1994).