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< Review >

## The Chemistry of Secondary Products from *Acanthopanax* Species and their Pharmacological Activities

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**Abstract** – The chemistry of secondary products from *Acanthopanax* species and their pharmacological activities were reviewed. A nitrogenous compound, a furan compound, a quinoid, benzoids, coumarins, phenylpropanoids, lignans, flavonoids, terpenoids, phytosterols, polyacetylenes, a pyrimidine, cyclitols, monosaccharides and an aliphatic alcohol have been isolated from *Acanthopanax* species and have been shown to have various levels of activities such as anti-bacterial, anti-cancer, anti-gout, anti-hepatitis, anti-hyperglycemic, anti-inflammatory, anti-leishmanicidic, anti-oxidant, anti-pyretic, anti-xanthine oxidase, choleretic, hemostatic, hypocholesterolemic, immunostimulatory and radioprotectant effects, etc.

Keywords - Acanthopanax species, Araliaceae, secondary products, pharmacological activities

## Introduction

Acanthopanax species that belongs to the family Araliaceae is known to be native to Asia, the Malay peninsula, Polynesia, Europe, North Africa and the Americas (Wielgorskaya and Takhtajan, 1995), and about 15 species of Acanthopanax are found in eastern Asia. Among Acanthopanax species growing in the Korean peninsula, A. senticosus, A. chiisanensis and A. sessiliflorus are most abundant species.

Acanthopanax species have traditionally been used as a tonic and a sedative as well as in the treatment of rheumatism and diabetes. Regular use was said to restore vigor, appetite, memory, impotence and increase longevity.

A nitrogenous compound, a furan compound, a quinoid, benzoids, coumarins, phenylpropanoids, lignans, flavonoids, terpenoids, phytosterols, polyacetylenes, a pyrimidine, cyclitols, monosaccharides and an aliphatic alcohol have been isolated from *Acanthopanax* species and have been shown to have various levels of activities such as anti-bacterial, anti-cancer, anti-gout, anti-hepatitis, anti-hyperglycemic, anti-inflammatory, anti-leishmanicidic, anti-oxidant, anti-pyretic, anti-xanthine oxidase, choleretic, hemostatic, hypocholesterolemic, immunostimulatory and radioprotectant effects, etc.

This paper reviewed the chemistry of secondary products from *Acanthopanax* species and their pharmacological activities.

A nitrogenous compound – Sessiline [1], a new nitrogenous compound, was isolated from the fruits of *A. sessiliflorus* 

(Lee *et al.*, 2002a). There has been no previous report on the isolation of a nitrogenous compound from *Acanthopanax* species. To our knowledge, this is the first report on a nitrogenous compound from *Acanthopanax* species.

A furan compound – 5-Hydroxymethylfurfural (HMF) [2] was isolated from the fruits of A. sessiliflorus (Lee et al., 2002b). There has been no previous report on a furan compound from Acanthopanax species. To our knowledge, this is the first report on a furan compound from Acanthopanax species. Shimizu et al. (1993) reported the isolation of HMF having aldose reductase inhibitory activity from Hachimi-jio-gan (Kampo medicine). HMF, occurs in many foods, does not pose a serious health risk, even though the highest concentrations in specific foods approach the biologically effective concentration range in cell systems (Janzowski et al., 2000), and the physiological effects of HMF on Saccharomyces cerevisiae CBS 8066 has been studied. Addition of HMF caused a decrease in the carbon dioxide evolution rate (Taherzadeh et al., 2000). HMF from the roasted fruits of Prunus mume had been demonstrated anthelmintic activity against Clonorchis sinensis (Kwak et al., 1985).

**A quinoid** – 2,6-Dimethoxy-*p*-benzoquinone [3] was isolated from the stem barks of *A. senticosus* (Nishibe *et al.*, 1990).

**Benzoids** – Protocatechuic acid (3,4-dihydroxybenzoic acid, DBA) [4] was isolated from the roots and the stem barks of *A. senticosus* (Yun-Choi *et al.*, 1986) and from the fruits of *A. sessiliflorus* (Lee *et al.*, 2002b). DBA from the stem barks of *A. senticosus* had an anti-platelet aggregatory

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activity (Yun-Choi *et al.*, 1986; Yun-Choi *et al.*, 1987). DBA from the flowers of *Hibiscus sabdariffa* L. (Malvaceae) exhibited a protection against *tent*-butylhydroperoxide-induced hepatotoxicity by its anti-oxidant and anti-inflammatory characteristics accompanied by blocking of stress signal transduction (Liu *et al.*, 2002). Overdoses of DBA, a naturally occurring simple phenolic anti-oxidant in dietary plant foodstuff, can disturb the detoxication of other electrophilic toxicants including ultimate carcinogens (Nakamura *et al.*, 2001). DBA from the BuOH fraction of *Polygonum bistorta* (Polygonaceae) showed an anti-inflammatory activity, its IC<sub>50</sub> value, being 165.27 μg/ml (Ahn *et al.*, 1999).

Ethyl 3,4-dihydroxybenzoate [5] was isolated from the roots of *A. senticosus* (Yun-Choi *et al.*, 1986). It has been demonstrated to possess an anti-platelet aggregatory activity (Yun-Choi *et al.*, 1986).

**Coumarins** – Isofraxidin [6] was isolated from *A. senticosus* (Wagner *et al.*, 1982; Bladt *et al.*, 1990) and from the stem barks of *A. senticosus* (Nishibe *et al.*, 1990). It showed cytotoxicity in lymphocytic leukemia in mice and stimulated bile as well (Borris *et al.*, 1980), and exhibited a choleretic effect when administered orally at 25 mg/kg

6 R<sub>1</sub>: H, R<sub>2</sub>: OCH<sub>3</sub> 7 R<sub>1</sub>: Glc, R<sub>2</sub>: OCH<sub>3</sub> 8 R<sub>1</sub>: CH<sub>3</sub>, R<sub>2</sub>: H

(Danielak et al., 1973).

Isofraxidin-7-*O*-β-D-glucoside (eleutheroside B<sub>1</sub>) [7] was isolated from *A. senticosus* (Wagner *et al.*, 1982; Bladt *et al.*, 1990; Wagner and Wurmböck, 1977), from the roots of *A. senticosus* (Slacanin *et al.*, 1991) and from the stem barks of *A. senticosus* (Nishibe *et al.*, 1990).

Scoparone [8] was isolated from the fruits of *A. sessiliflorus* (Lee *et al.*, 2002b). Overdoses of this compound can disturb the detoxification of other electrophilic toxicants including ultimate carcinogens (Nakamura *et al.*, 2001). After the treatment with this compound, hepatic microsomal UDP glucuronyltransferase activity (Huh *et al.*, 1987) and hepatic cytosolic sulfotransferase activity (Huh *et al.*, 1990) were increased in dose-dependent manner. This compound from *Artemisia capillaris* caused a reduction of cold ischemic injury in liver transplantation (Cho *et al.*, 2000). Differential oxidation of scoparone can be used as a sensitive indicator for distinguishing between different cytochrome P<sub>450</sub> isoforms (Meyer *et al.*, 2001).

**Phenylpropanoids** – Caffeic acid [9] was isolated from *A. senticosus* (Bladt *et al.*, 1990). Caffeic acid showed an inhibition of xanthine oxidase activity and a nitric oxide production in C<sub>6</sub> astrocyte cells (Soliman and Mazzio, 1998), inhibited the toxic action of aflatoxin from *Aspergillus parasiticus* (Aziz *et al.*, 1998), showed an anti-gout and an anti-hepatitis activity (Chan *et al.*, 1995) and inhibited tumor promotion *in vivo* and *in vitro* in murine peritoneal macrophages treated with tumor promoters, and it also produced superoxide anions (Kaul and Khanduja, 1998).

Caffeic acid ethylester [10] was isolated from *A. senticosus* (Wagner *et al.*, 1982; Bladt *et al.*, 1990). It showed a protectant activity against single stranded DNA breaks caused by hydrogen peroxide in Chinese hamster (V79 cells) (Nakayama *et al.*, 1996).

Coniferin [11] was isolated from the root barks of *A. koreanum* (Kim *et al.*, 1988) and from the roots of *A. sesnticosus* (Slacanin *et al.*, 1991). Coniferin exhibited dual inhibitory effects, since it produced reduction in the generation of both cyclooxygenase and 5-lipoxygenase metabolites (Díaz Lanza *et al.*, 2001). Coniferin produced concentration-dependent contractions in rat aortic rings

(Deliorman et al., 2000).

Coniferylaldehyde [12] was isolated from the *A. sesnticosus* (Wagner *et al.*, 1982). Coniferylaldehyde showed a protectant activity against DNA breaks caused by UV light-derived hydroxyl radicals (Taira *et al.*, 1992).

Coniferylaldehyde glucoside [13] was isolated from *A. sesnticosus* (Slacanin *et al.*, 1991).

*p*-Coumaric acid [14] and *p*-coumaric acid methylester [15] were isolated from the leaves of *A. sciadophylloides* (Kitajima *et al.*, 1989). *p*-Coumaric acid showed synergistic properties in reinforcing the anti-oxidant activity of lactoferrin in lipid systems containing iron (Medina *et al.*, 2002).

Sinapaldehyde glucoside [16] was isolated from the roots of *A. senticosus* (Slacanin *et al.*, 1991).

Sinapylalcohol [17] was isolated from *A. senticosus* (Wagner *et al.*, 1982; Bladt *et al.*, 1990).

Syringin (eleutheroside B) [18] was isolated from A. koreanum (Hahn et al., 1985; Wagner et al., 1982; Wagner and Wurmböck, 1977), from the root bark of A. koreanum (Chung and Kim, 1986), from the roots of A. senticosus (Li et al., 2001; Slacanin et al., 1991), from the roots and rhizomes of A. sesnticosus (Yat et al., 1998) and from the stem barks of A. koreanum (Nishibe et al., 1990) as one of main constituents. Syringin isolated from the roots of A. senticosus protected the animals from the stress-induced decreases in sex behaviors and in rectal temperature (Nishiyama et al., 1985). Syringin functioned to prevent the stress-induced decreases in grip tone and exploratory movement, and to accelerate recovery from the decreases in grip tone, exploratory movement and spontaneous movement (Takasugi et al., 1985). Syringin showed a protectant activity against damage from radiation. Fewer deaths occurred in mice after X-ray irradiation (400 rads). This compound decreased leucopoenia, and improved white blood cell count and thrombocyte level in human workers after exposure to unspecified radioactive substances (Ruijun et al., 1990), and inhibited immunohaemolysis of antibody-coated sheep erythrocytes by guinea pig serum (Kapil and Sharma, 1997).

Eugenyl  $\beta$ -rutinoside [19] and sasanquin [20] were isolated for the first time from the stem barks of *A. setchuenensis* (Zhao *et al.*, 1999).

Chlorogenic acid [21] was isolated from the leaves of *A. divaricatus* forma *nambunensis* (Cho *et al.*, 1999), from the roots of *A. sesnticosus* (Slacanin *et al.*, 1991), from the root barks of *A. sesnticosus* (Fujikawa *et al.*, 1996), from the stem barks of *A. sesnticosus* (Nishibe *et al.*, 1990) and from the leaves of *A. trichodon* (Miyakoshi *et al.*, 1997b). Chlorogenic acid showed a significant inhibitory effect on gastric ulcer by 21.4% (Fujikawa *et al.*, 1996).

1,5-Di-O-caffeoylquinic acid [22] was isolated from the

$$R_2$$
 $R_3$ 

9 R1: OH, R2: OH, R3: H, R4: COOH

10 R<sub>1</sub>: OH, R<sub>2</sub>: OH, R<sub>3</sub>: H, R<sub>4</sub>: COOCH<sub>2</sub>CH<sub>3</sub>

11 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc, R<sub>3</sub>: H, R<sub>4</sub>: CH<sub>2</sub>OH

12 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: OH, R<sub>3</sub>: H, R<sub>4</sub>: CHO

13 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc, R<sub>3</sub>: H, R<sub>4</sub>: CHO

14 R<sub>1</sub>: H, R<sub>2</sub>: OH, R<sub>3</sub>: H, R<sub>4</sub>: COOH

15 R<sub>1</sub>: H, R<sub>2</sub>: OH, R<sub>3</sub>: H, R<sub>4</sub>: COOCH<sub>3</sub>

16 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc, R<sub>3</sub>: OCH<sub>3</sub>, R<sub>4</sub>: CHO

17 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: OH, R<sub>3</sub>: OCH<sub>3</sub>, R<sub>4</sub>: CH<sub>2</sub>OH

18 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc, R<sub>3</sub>: OCH<sub>3</sub>, R<sub>4</sub>: CH<sub>2</sub>OH

19 R: Rha-Glc

20 R: Xyl-Glc

21 COOH HO

HO

22

roots of A. senticosus (Slacanin et al., 1991).

**Lignans** – Acanthoside B [(+)-syringaresinol-O-β-D-glucoside, eleutheroside E<sub>1</sub>] [23] was isolated from the root barks of *A. setchuenensis* (Zhao *et al.*, 1999) and from the

roots of A. senticosus (Li et al., 2001).

Acanthoside D [(+)-syringaresinol-O- $\beta$ -D-diglucoside, eleutheroside E] [24] was isolated from the stem barks of A. senticosus (Nishibe et al., 1990), from the root cortex of A. chiisanensis (Kim and Hahn, 1981), from the fruits of A. chiisanensis (Shin et al., 1992), from the root barks of A. divaricatus (Yook et al., 1996), from the roots of A. koreanum (Hahn et al., 1985; Kim et al., 1985; Chung and Kim, 1986), from the root barks of A. setchuenensis (Zhao et al., 1999), from the barks of A. senticosus forma inermis (Yook et al., 1991), from the root barks of A. senticosus (Fujikawa et al., 1996), from the roots and rhizomes of A. senticosus (Yat et al., 1998) and from the roots of A. sesnticosus (Slacanin et al., 1991; Li et al., 2001). Acanthoside D isolated from the root barks of A. koreanum has been found to have s-GPT and s-GOT lowering effect, BSP-retention rate and survival rate in the toxic state through the bio-pharmacological experiments (Hahn et al., 1985) were significantly increased. Among eleutherosides B<sub>1</sub>, C and E in stress, eleutheroside E exhibited the most pronounced protective effect against stress (Brekhman and Dardymov, 1969). Acanthoside D isolated from the stem barks of A. senticosus had the pharmacological effect in chronic swimming stressed rats (Nishibe et al., 1990). Acanthoside D from the root barks of A. senticosus showed a significant inhibition against gastric ulcer by 51.3% (Fujikawa et al., 1996). Acanthoside D isolated from A. senticosus protected the animals from the stress-induced decreases in sex behaviors and in rectal temperature, from the stress-induced failure of retrieval of memory and from the stress-induced enlargement of adrenal gland (Nishiyama et al., 1985). Acanthoside D also functioned to prevent the stress-induced decreases in spontaneous movements and to accelerate recovery from the decreases in grip tone, exploratory movement and spontaneous movement (Takasugi et al., 1985).

Eleutheroside E octaacetate [25] was isolated from A. koreanum (Chung and Kim, 1986).

Liriodendrin [26] was isolated from the cortex of *A. sessiliflorus* forma *chungbunensis*. Liriodendrin stimulated the incorporation of <sup>14</sup>C-leucine into mouse liver protein (Ro *et al.*, 1977).

(+)-Medioresinol-O- $\beta$ -D-diglucoside [27], (+)-pinoresinol-O- $\beta$ -D-glucoside [28] and (+)-pinoresinol-O- $\beta$ -D-diglucoside [29] were isolated from the stem bark of *A. senticosus* (Nishibe *et al.*, 1990).

Syringaresinol [30] was isolated from the root barks of *A. setchuenensis* (Zhao *et al.*, 1999).

Sesamin (eleutheroside B<sub>4</sub>) [31] was isolated from the roots and the stem barks of *A. chiisanensis* (Jang, 1970),

23 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc, R<sub>3</sub>: OCH<sub>3</sub>, R<sub>4</sub>: OCH<sub>3</sub>, R<sub>5</sub>: OH, R<sub>6</sub>: OCH<sub>3</sub>
24 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc, R<sub>3</sub>: OCH<sub>3</sub>, R<sub>4</sub>: OCH<sub>3</sub>, R<sub>5</sub>: O-Glc, R<sub>6</sub>: OCH<sub>3</sub>
25 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc<sub>Ac</sub>, R<sub>3</sub>: OCH<sub>3</sub>, R<sub>4</sub>: OCH<sub>3</sub>, R<sub>5</sub>: O-Glc<sub>Ac</sub>, R<sub>6</sub>: OCH<sub>3</sub>
26 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc, R<sub>3</sub>: OCH<sub>3</sub>, R<sub>4</sub>: OCH<sub>3</sub>, R<sub>5</sub>: O-Glc, R<sub>6</sub>: OCH<sub>3</sub>
27 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc, R<sub>3</sub>: OCH<sub>3</sub>, R<sub>4</sub>: H, R<sub>5</sub>: O-Glc, R<sub>6</sub>: OCH<sub>3</sub>
28 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc, R<sub>3</sub>: H, R<sub>4</sub>: H, R<sub>5</sub>: OH, R<sub>6</sub>: OCH<sub>3</sub>
29 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: O-Glc, R<sub>3</sub>: H, R<sub>4</sub>: H, R<sub>5</sub>: O-Glc, R<sub>6</sub>: OCH<sub>3</sub>
30 R<sub>1</sub>: OCH<sub>3</sub>, R<sub>2</sub>: OH, R<sub>3</sub>: OCH<sub>3</sub>, R<sub>4</sub>: OCH<sub>3</sub>, R<sub>5</sub>: OH, R<sub>6</sub>: OCH<sub>3</sub>
31 R<sub>1</sub>, R<sub>2</sub>: OCH<sub>2</sub>O, R<sub>3</sub>: H, R<sub>4</sub>, R<sub>5</sub>: OCH<sub>2</sub>O, R<sub>6</sub>: H

from the roots of A. chiisanensis (Lee et al., 2002c), from the stem barks of A. divaricatus (Yook et al., 1996), from the roots of A. divaricatus (Miyakoshi et al., 1995; Yook et al., 1996), from the root barks of A. divaricatus var. albeofructus (Oh et al., 2000), from the fruits of A. sessiliflorus (Lee et al., 2002b), from the root barks of A. senticosus (Bo et al., 1998), from the barks of A. senticosus forma inermis (Yook et al., 1991) and from the root barks of A. sessiliflorum (Yook et al., 1977). Sesamin induced hypocholesterolemia especially low-density lipoproteins and cholesterol, which are risk factors for human atherosclerosis (Hirata et al., 1996), showed 36% reduction in 7,12-dimethylbenz[a]anthracene-induced mammary cancer in female rats when measured at 12 weeks after uptake (Hirose et al., 1992), decreased liver enlargement caused by excessive alcohol intake and increased the concentration of immunoglobulin G (Nonaka et al., 1997), and improved impaired liver function in rodents caused by 1% EtOH or CCl<sub>4</sub> (100 mg/kg) (Akimoto et al., 1993). Sesamin is one of the most abundant lignans in sesame seeds. The dietary sesamin-dependent decrease in lipogenic enzyme gene expression is due to the suppression of the gene expression of the sterol regulatory element binding protein-1 as well as the proteolysis of the membranebound precursor form of this transcriptional factor to generate the mature form (Ide et al., 2001). The chronic ingestion of sesamin from sesame oil attenuated each of elevation in blood pressure, oxidative stress and thrombotic tendency, suggesting that these treatments might be beneficial in the prevention of hypertension and stroke (Noguchi et al., 2001). Dietary sesamin significantly increased the

activities of hepatic mitochodrial and peroxisomal fatty acid oxidation enzymes such as mitochondrial carnitine acyltransferase, acyl-CoA dehydrogenase and peroxisomal acyl-CoA oxidase (Umeda-Sawada *et al.*, 2001). A consumption of sesamin rich in lignans resulted in physiological activity to alter lipid metabolism in a potentially beneficial manner (Sirato-Yasumoto *et al.*, 2001). Sesamin exhibited significant anti-feedant activity and moderate growth inhibition towards 4<sup>th</sup> instar larvae of *Spilarctia oblique* (Srivastava *et al.*, 2001). Sesamin suppressed the growth and induced apoptosis in the cells (Hibasami *et al.*, 2000).

Eleutheroside  $E_2$  [32] was isolated for the first time from the roots of *A. senticosus* (Li *et al.*, 2001).

Ariensin [33] was isolated from the root barks of *A. koreanum* (Chung and Kim, 1986; Kim *et al.*, 1988).

Helioxanthin [34] were isolated from the roots of *A. chiisanensis* (Lee *et al.*, 2002c) and from the roots of *A. divaricatus* (Miyakoshi *et al.*, 1995).

Taiwanin C [35] was isolated from the roots of *A. chiisanensis* (Lee *et al.*, 2002c). Taiwanin C from the roots of *A. chiisanensis* has anti-inflammatory activity (Ban *et al.*, 2002; Lee *et al.*, 2002c).

3-(3,4-Dimethoxybenzyl)-2-(3,4-methylenedioxybenzyl)

**34** R<sub>1</sub>: H, R<sub>2</sub>,R<sub>3</sub>: OCH<sub>2</sub>O, R<sub>4</sub>: H, R<sub>5</sub>: =O **35** R<sub>1</sub>,R<sub>2</sub>: OCH<sub>2</sub>O, R<sub>3</sub>: H, R<sub>4</sub>: =O, R<sub>5</sub>: H

butyrolactone [36] was isolated from the roots of *A. chiisanensis* (Lee *et al.*, 2002c).

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Matairesinoside [37] was isolated from the roots of *A. sieboldianus* (Miyakoshi *et al.*, 1995).

Savinin (taiwanin B) [38] was isolated from the roots of *A. chiisanensis* (Lee *et al.*, 2002c), from the root barks of *A. divaricatus* var. *albeofructus* (Oh *et al.*, 2000), from the fruits of *A. chiisanensis* (Shin *et al.*, 1992), from the root barks of *A. senticosus* (Bo *et al.*, 1998) and from the barks of *A. senticosus* forma *inermis* (Yook *et al.*, 1991). Savinin exhibited potent spermicidal and significant insecticidal activities (Nissanka *et al.*, 2001).

**Flavonoids** – Delphinidin 3-lathyroside (delphinidin 3-xylosylgalactoside) [**39**] was isolated from the fruits of *A. divaricatus* (Ishikura, 1975).

Afzelin (kaempferol 3-*O*-rhamnoside) [40] was isolated from the leaves of *A. sciadophylloides* (Kitajima *et al.*, 1989).

Antoside [41] was isolated from the leaves of *A. sciadophylloides* (Yasue *et al.*, 1968).

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40 R<sub>1</sub>: H, R<sub>2</sub>: Rha, R<sub>3</sub>: H

41 R<sub>1</sub>: Rha, R<sub>2</sub>: Glc, R<sub>3</sub>: OH

42 R<sub>1</sub>: Rha, R<sub>2</sub>: Gal, R<sub>3</sub>: H

43 R1: H, R2: Glc, R3: OH

44 R1: H, R2: Gal, R3: OH

45 R<sub>1</sub>: Rha, R<sub>2</sub>: Rha, R<sub>3</sub>: OH

46 R<sub>1</sub>: H, R<sub>2</sub>: H, R<sub>3</sub>: H

47 R<sub>1</sub>: H, R<sub>2</sub>: Glc-Rha, R<sub>3</sub>: H

48 R<sub>1</sub>: Rha, R<sub>2</sub>: H, R<sub>3</sub>: H

49 R<sub>1</sub>: Rha, R<sub>2</sub>: H, R<sub>3</sub>: OH

50 R<sub>1</sub>: Rha, R<sub>2</sub>: Rha, R<sub>3</sub>: OH

51 R<sub>1</sub>: H, R<sub>2</sub>: Rha, R<sub>3</sub>: OH

52 R<sub>1</sub>: H, R<sub>2</sub>: Glc-Rha, R<sub>3</sub>: OH

3-*O*-β-D-Galactopyranosyl-kaempferol 7-rhamnoside [**42**] was isolated from the leaves of *A. sciadophylloides* (Kitajima *et al.*, 1989).

Isoquercitrin (hirsutrin) [43] was isolated from the leaves of *A. sciadophylloides* (Yasue *et al.*, 1969; Kitajima *et al.*, 1989).

Hyperin [44] was isolated from the leaves of *A. divaricatus* (Matsumoto *et al.*, 1987; Shirasuna *et al.*, 1997), from the leaves of *A. divaricatus* forma *nambunensis* (Cho *et al.*, 1999) and for the first time from the fruits of *A. sessiliflorus* (Lee *et al.*, 2002b). Hyperin had no significant effects on the resting Ca<sup>2+</sup>, markedly inhibited the increase of Ca<sup>2+</sup> evoked by K<sup>+</sup> in a concentration-dependent manner and inhibited the increase of Ca<sup>2+</sup> induced by norepinephrine. Also hyperin markedly attenuated 5-hydroxytryptamine and L-glutamic acid-induced increase of Ca<sup>2+</sup>. As a result, hyperin possessed an inhibitory effect on influx of Ca<sup>2+</sup> in the neonatal rat brain cells (Chen and Ma, 1999).

Kaempferitrin (lespedin) [45] was isolated from the leaves

of A. sciadophylloides (Yasue et al., 1968; Kitajima et al., 1989), from the leaves of A. senticosus var. subinermis (Chang, 1990), and from the leaves of A. sessiliflorum (Kim, 1985).

Kaempferol [46] was isolated from the leaves of *A. sciadophylloides* (Kitajima *et al.*, 1989).

Kaempferol 3-O-rutinoside [47] was isolated from the leaves of A. sieboldianus (Sawada et al., 1993).

Kaempferol 7-O-rhamnoside [48] was isolated from the leaves of *A. sciadophylloides* (Yasue *et al.*, 1969; Kitajima *et al.*, 1989; Sawada *et al.*, 1993).

Quercetin 7-O-rhamnoside (vincetoxicoside B) [49] was isolated from the leaves of A. sciadophylloides (Yasue et al., 1969).

Quercetin 3,7-*O-bis*-α-L-rhamnopyranoside [**50**] was isolated from the leaves of *A. sciadophylloides* (Kitajima *et al.*, 1989).

Quercitrin [51] was isolated from the leaves of *A. divaricatus* (Shirasuna *et al.*, 1997).

Rutin [52] was isolated from the leaves of *A. koreanum* (Chung and Hahn, 1991).

Hesperidin [53] was isolated from the root barks of *A. setchuenensis* (Zhao *et al.*, 1999).

Sesquiterpenoids –  $\beta$ -Caryophyllene [54] was isolated from the leaves of *A. sciadophylloides* (Kitajima *et al.*, 1989). It markedly inhibited the growth of tested Gram-(+) and Gram-(-) bacteria except for *Pseudomonas aeruginosa* (Haznedaroglu *et al.*, 2001).

Farnesol [55] was isolated from the roots of *A. divaricatus* (Miyakoshi *et al.*, 1995).

β-Farnesene [56] was isolated from the leaves of A. sciadophylloides (Kitajima et al., 1989).

**Diterpenoids** – Phytol [57] was isolated from the leaves of *A. sciadophylloides* (Kitajima *et al.*, 1989).

Acanthoic acid [(-)-pimara-9(11),15-dien-19-oic acid] [58] was isolated from the root barks of *A. koreanum* (Yook *et al.*, 1996; Kang *et al.*, 1996; Kim and Chung, 1988). Acanthoic acid has a potent anti-inflammatory and anti-fibrosis effect by reducing IL-1 and TNF-α production (Kang *et al.*, 1996).

(-)-Pimara-9(11),15-dien-19-ol [59] and *iso*-pimara-9(11), 15-dien-19-ol [60] were isolated for the first time from the root barks of *A. koreanum*. (-)-Pimara-9(11),15-dien-19-ol 19-acetate [61] and (-)-pimara-9(11),15-diene [62] were isolated from the root barks of *A. koreanum* (Kim and Chung, 1988).

Sumogaside [63] was isolated from the root barks of *A. koreanum* (Kim *et al.*, 1990)

ent-Kaur-16-en-19-oic acid [64] was isolated from the root barks of *A. koreanum* (Kim and Chung, 1988) and from the leaves of *A. trichodon* (Miyakoshi et al., 1997b).

ent-16β,17-Dihydroxy-(-)-kauran-19-oic acid [65] was isolated from the root barks of A. koreanum (Kim and

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58 R<sub>1</sub>: COOH, R<sub>2</sub>: CHCH<sub>2</sub>
59 R<sub>1</sub>: CH<sub>2</sub>OH, R<sub>2</sub>: CHCH<sub>2</sub>
61 R<sub>1</sub>: CH<sub>2</sub>OAc, R<sub>2</sub>: CHCH<sub>2</sub>
62 R<sub>1</sub>: CH<sub>3</sub>, R<sub>2</sub>: CHCH<sub>2</sub>

63 R<sub>1</sub>: COO-Glc, R<sub>2</sub>: CHOHCH<sub>2</sub>OH

Chung, 1988)

ent-16b,17-iso-Valerate-kauran-19-oic acid [66] and ent-16 $\beta$ H,17-methyl butanoate-kauran-19-oic acid [67] were isolated for the first time from the stem bark of *A. koreanum* (Kim et al., 1995).

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64 R: O

**65** R: α-OH, β-CH<sub>2</sub>OH

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**Oleane triterpenoids** – Ciwujianoside  $D_1$  [68] was isolated from the aerial parts of A. senticosus (Umeyama et al., 1992). It strongly inhibited histamine release in a concentration-dependent manner in rat peritoneal mast cells induced by anti-immunoglobulin E (Umeyama et al., 1992).

Eleutherosides I [69], K [70], L [71] and M (hederasaponin B) [72] were isolated from *A. senticosus* (Wagner and Wurmböck, 1977). Hederasaponin B [eleutheroside M] showed antileishmanicidic effect but could not be confirmed (Majester-Savornin *et al.*, 1991).

Hederagenin [73] was isolated from leaves of *A. hypoleucus* (Kohda *et al.*, 1990).

Hypoleucosides A [74] and B [75] were isolated for the first time from the leaves of *A. hypoleucus* (Kohda *et al.*, 1990).

**68** R<sub>1</sub>: O-Ara, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 69 R<sub>1</sub>: O-Rha(1→4)-Ara, R<sub>2</sub>: H, R<sub>3</sub>: H, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> **70** R₁: O-Rha(1→2)-Ara, R₂: H, R₃: H, R₄: CH₃, R₅: CH₃, R₆: CH₃ 71 R<sub>1</sub>: O-Rha(1→4)-Ara, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 72 R<sub>1</sub>: O-Rha(1→2)-Ara, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 73 R<sub>1</sub>: H, R<sub>2</sub>: H, R<sub>3</sub>: H, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> **74** R<sub>1</sub>: O-Glc, R<sub>2</sub>: OCH<sub>3</sub>, R<sub>3</sub>: Glc, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 75 R<sub>1</sub>: O-Glo-Ara-Glo, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 76 R<sub>1</sub>: =O, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 77 R<sub>1</sub>: β-O-Glc, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 78 R<sub>1</sub>:  $\beta$ -O-Glc, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: CHO, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 79 R<sub>1</sub>: β-OH, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>2</sub>OH 80 R<sub>1</sub>:  $\beta$ -OH, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: OH 81 R<sub>1</sub>:  $\beta$ -OH, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 82 R<sub>1</sub>: O-Ara-Rha, R<sub>2</sub>: H, R<sub>3</sub>: Gle-Gle-Rha, R<sub>4</sub>:  $CH_2OH$ , R<sub>5</sub>:  $CH_3$ , R<sub>6</sub>:  $CH_3$ 83 R<sub>1</sub>: O-Ara-Rha, R<sub>2</sub>: H, R<sub>3</sub>: H, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 84  $R_1$ : O-Ara-Rha-Ara,  $R_2$ : H,  $R_3$ : H,  $R_4$ :  $CH_2OH$ ,  $R_5$ :  $CH_3$ ,  $R_6$ :  $CH_3$ 85 R<sub>1</sub>: O-Ara-Rha-Xyl, R<sub>2</sub>: H, R<sub>3</sub>: H, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 86 R<sub>1</sub>: O-Ara-Rha-Xyl, R<sub>2</sub>: H, R<sub>3</sub>: H, R<sub>4</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 67 R1: O-Ara-Rha-Xyl, R2: H, R3: Glc-Glc-Rha, R4: CH2OH, R5: CH3, R6: CH3 88 R $_1$ : O-Ara-Rha-Xyl, R $_2$ : H, R $_3$ : Glo-Glo-Rha, R $_4$ : CH $_3$ , R $_5$ : CH $_3$ , R $_6$ : CH $_3$ 89 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: H, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 90 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: CHO, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: OH 91 R<sub>1</sub>: O-Ara-Glo, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 92  $R_1$ : O-Ara-Rha,  $R_2$ : H,  $R_3$ : Glc-Glc-Rha,  $R_4$ :  $CH_3$ ,  $R_5$ :  $CH_3$ ,  $R_6$ :  $CH_2OH$ 93 R<sub>1</sub>: O-Ara, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc<sub>Ac</sub>-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>2</sub>OH 94 R<sub>1</sub>: O-Ara-Glc, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc<sub>Ac</sub>-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>2</sub>OH 95 R<sub>1</sub>: O-Ara, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 96 R<sub>1</sub>: O-Ara-Rha, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc<sub>Ac</sub>-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub> 97 R<sub>1</sub>: O-Ara, R<sub>2</sub>: H, R<sub>3</sub>: Glc-Glc<sub>Ac</sub>-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>3</sub>

Nipponosides A [76], B [77], C [78], D [79] and E [80] were isolated for the first time from the leaves of *A. nipponicus* (Miyakoshi *et al.*, 1999). Niponoside E was isolated from the leaves of *A. japonicus* (Park *et al.*, 2002).

Kalopanax saponin G [81] was isolated from the leaves of *A. nipponicus* (Miyakoshi *et al.*, 1999).

Kalopanax saponins A [82], B [83], saponin A [84], sapindoside B [85] and CP<sub>3</sub> [86] were isolated from the leaves of *A. sieboldianus* (Sawada *et al.*, 1993).

Sieboldianosides A [87] and B [88] were isolated for the first time from the leaves of *A. sieboldianus* (Sawada *et al.*, 1993).

Oleanolic acid [89] was isolated from the barks of A.

senticosus forma inermis (Yook et al., 1991), and from A. divaricatus (Yook et al., 1996; Kohda et al., 1990; Bladt et al., 1990). This compound from Fabiana patagonica showed diuretic activity (Alvarez et al., 2002).

Acanjaposide C [90] was isolated for the first time from the leaves of *A. japonicus* (Park *et al.*, 2002).

Ciwujianosides  $A_1$  [91],  $A_3$  [92],  $D_3$  [93] and  $A_4$  [94] were isolated for the first time from the leaves of A. senticosus (Shao et al., 1989). Ciwujianosides  $C_3$  [95],  $C_4$  [96] and  $D_1$  [97] were isolated for the first time from the leaves of A. senticosus (Shao et al., 1988).

Spinosides  $D_1$  [98],  $D_2$  [99],  $D_3$  [100]  $C_1$  [101],  $C_4$  [102] and  $C_5$  [103] were isolated for the first time from the leaves of *A. spinosus* (Miyakoshi *et al.*, 1993a; Miyakoshi *et al.*, 1993b). Spinosides  $C_2$  [104],  $C_3$  [105],  $C_6$  [106] and  $C_7$  [107] were isolated from the leaves of *A. spinosus* (Miyakoshi *et al.*, 1997a).

Ciwujianoside  $C_1$  [108] was isolated from the aerial parts of *A. senticosus* (Umeyama *et al.*, 1992). It strongly inhibited histamine release in a concentration-dependent manner in rat peritoneal mast cells induced by anti-immunoglobulin E (Umeyama *et al.*, 1992).

Acanjaposides A [109] and B [110] were isolated for the first time from the leaves of *A. japonicus* (Park *et al.*, 2002).

Ciwujianoside  $A_2$  [111] was isolated for the first time from the leaves of A. senticosus (Shao et al., 1989). Ciwujianosides B [112],  $C_1$  [113],  $C_2$  [114],  $D_2$  [115] and E [116] were isolated for the first time from the leaves of A. senticosus (Shao et al., 1988).

3b-{O-β-D-Glucopyranosyl-( $1 \rightarrow 3$ )-O-β-D-galactopyranosyl-( $1 \rightarrow 4$ )-[O-α-L-rhamnopyranosyl-( $1 \rightarrow 2$ )]-O-β-D-glucuronopyranosyl}-16 $\alpha$ -hydroxy-13 $\beta$ ,28-epoxyoleanane [117] and 3 $\beta$ -{O-α-L-rhamnopyranosyl-( $1 \rightarrow 4$ )-O-α-L-rhamnopyranosyl-( $1 \rightarrow 4$ )-O-b-D-glucopyranosyl-( $1 \rightarrow 4$ )-O-β-D-glucuronopyranosyl}-16 $\alpha$ -hydroxy-13 $\beta$ ,28-epoxyoleanane [118] were isolated for the first time from the roots of A. senticosus (Segiet-Kujawa and Kaloga, 1991).

**Taraxane triterpenoids** – Taraxerol [119] was isolated from the leaves of *A. sciadophylloides* (Yasue *et al.*, 1969; Kitajima *et al.*, 1989; Yasue *et al.*, 1970; Chen *et al.*, 1972a).

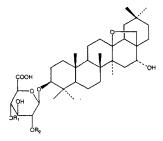
Taraxeryl acetate [120] was isolated from the leaves of *A. sciadophylloides* (Chen *et al.*, 1972a).

**Ursane triterpenoids** – Bauerenyl acetate [121] was isolated from the leaves of *A. trichodon* (Miyakoshi *et al.*, 1997b).

Ursolic acid (UA) [122] was isolated from the fruits of *A. sessiliflorus* (Lee *et al.*, 2002b). There have been no previous reports of an ursane triterpenoid from *Acanthopanax* species. To our knowledge, this is the first report of an ursane triterpenoid from *Acanthopanax* species. UA showed anti-

98 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: COOH
99 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CHO, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: COOH
100 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: COOH
101 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>2</sub>OH
102 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CHO, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>2</sub>OH
103 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: CH<sub>2</sub>OH
104 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Rha-Glo-Glo, R<sub>4</sub>: CHO, R<sub>5</sub>: CH<sub>2</sub>OH, R<sub>6</sub>: CH<sub>3</sub>
105 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Rha-Glo-Glo, R<sub>4</sub>: CH<sub>3</sub>, R<sub>5</sub>: CH<sub>3</sub> R<sub>6</sub>: OH
106 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Rha-Glo-Glo, R<sub>4</sub>: CH<sub>2</sub>OH, R<sub>5</sub>: CH<sub>3</sub>, R<sub>6</sub>: OH

108 R<sub>1</sub>: O-Ara, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CH<sub>3</sub>
109 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: COOH
110 R<sub>1</sub>: OH, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CHO
111 R<sub>1</sub>: O-Ara-Glo, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CH<sub>3</sub>
112 R<sub>1</sub>: O-Ara-Rha, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CH<sub>3</sub>
113 R<sub>1</sub>: O-Ara-Rha, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo-Rha, R<sub>4</sub>: CH<sub>3</sub>
114 R<sub>1</sub>: O-Ara-Rha, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo<sub>Ao</sub>-Rha, R<sub>4</sub>: CH<sub>3</sub>
115 R<sub>1</sub>: O-Ara-Rha, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo<sub>Ao</sub>-Rha, R<sub>4</sub>: CH<sub>3</sub>
116 R<sub>1</sub>: O-Ara-Rha, R<sub>2</sub>: H, R<sub>3</sub>: Glo-Glo<sub>Ao</sub>-Rha, R<sub>4</sub>: CH<sub>3</sub>



117 R<sub>1</sub>: O-β-D-Glc-(1 $\to$ 3)-β-D-Gal, R<sub>2</sub>:  $\alpha$ -L-Rha 118 R<sub>1</sub>: O- $\alpha$ -L-Rha-(1 $\to$ 4)-O- $\alpha$ -L-Rha-(1 $\to$ 4)-[O- $\alpha$ -L-Rha-(1 $\to$ 2)]-β-D-Glc, R<sub>2</sub>: H

**119** R: OH **120** R: OAc

tumor effects and chemopreventive properties in normal cells (Novotný et al., 2001) and changes in tumor growth,

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O<sub>2</sub> consumption, and tumor interstitial fluid pressure (Lee *et al.*, 2001). UA stimulates NO and TNF- $\alpha$  release and is able to up regulate iNOS and TNF- $\alpha$  expression through NF- $\kappa$ B transactivation in the resting macrophages (You *et al.*, 2001). The treatment of UA from *Origanum majorana* L. inhibited the Abeta-induced neurotoxic effect (Heo *et al.*, 2002).

**Lupane triterpenoids** – Acantrifoside A [123] was isolated for the first time from the leaves of *A. koreanum* (Chung and Hahn, 1991) and from the leaves of *A. trifoliatus* and *A. koreanum* (Yook *et al.*, 1998).

Acankoreosides A [124], B [125], C [126] and D [127] were isolated from the leaves of *A. koreanum* (Chang *et al.*, 1998; Chang *et al.*, 1999).

 $3\alpha$ ,  $11\alpha$ -Dihydroxylup-20(29)-en-28-oic acid (impressic acid) [128],  $3\alpha$ ,  $11\alpha$ , 23-trihydroxylup-20(29)-en-28-oic acid [129] and  $3\alpha$ ,  $11\alpha$ -dihydroxy-23-oxo-lup-20(29)-en-28-oic acid [130] were isolated for the first time from the leaves of *A. trifoliatus* (Ty *et al.*, 1984a; Ty *et al.*, 1984b).

Acanthodiol [131] was isolated from the leaves of *A. koreanum* (Chung and Hahn, 1991; Kim, 1988).

Acanthodiol glycoside [132] was isolated from the leaves of *A. koreanum* (Chung and Hahn, 1991) and from the leaves of *A. trifoliatus* forma *tristigmatis* (Yook *et al.*, 1999).

Methyl betulin [133] was isolated from the fruits of *A. chiisanensis* (Shin *et al.*, 1992).

Protochiisanoside [134] was isolated from the leaves of *A. divaricatus* (Shirasuna *et al.*, 1997).

24-nor- $3\alpha$ ,  $11\alpha$ -Dihydroxylup-20(29)-en-28-oic acid [135] was isolated for the first time from the leaves of A.

123 R<sub>1</sub>: OH, R<sub>2</sub>: OH, R<sub>3</sub>: CH<sub>3</sub>, R<sub>4</sub>: COO-Glc-Glc-Rha
124 R<sub>1</sub>: H, R<sub>2</sub>: OH, R<sub>3</sub>: COOH, R<sub>4</sub>: COO-Glc-Glc-Rha
125 R<sub>1</sub>: OH, R<sub>2</sub>: OH, R<sub>3</sub>: CH<sub>2</sub>OH, R<sub>4</sub>: COO-Glc-Glc-Rha
126 R<sub>1</sub>: OH, R<sub>2</sub>: O-Glc, R<sub>3</sub>: CH<sub>3</sub>, R<sub>4</sub>: COO-Glc-Glc-Rha
127 R<sub>1</sub>: OH, R<sub>2</sub>: OH, R<sub>3</sub>: CHO, R<sub>4</sub>: COO-Glc-Glc-Rha
128 R<sub>1</sub>: OH, R<sub>2</sub>: OH, R<sub>3</sub>: CH<sub>3</sub>, R<sub>4</sub>: COOH
129 R<sub>1</sub>: OH, R<sub>2</sub>: OH, R<sub>3</sub>: CH<sub>2</sub>OH, R<sub>4</sub>: COOH
130 R<sub>1</sub>: OH, R<sub>2</sub>: OH, R<sub>3</sub>: CHO, R<sub>4</sub>: COOH
131 R<sub>1</sub>: OH, R<sub>2</sub>: β-OH, R<sub>3</sub>: CH<sub>3</sub>, R<sub>4</sub>: COOH
132 R<sub>1</sub>: OH, R<sub>2</sub>: β-OH, R<sub>3</sub>: CH<sub>3</sub>, R<sub>4</sub>: COO-Glc-Glc-Rha
133 R<sub>1</sub>: H, R<sub>2</sub>: β-OCH<sub>3</sub>, R<sub>3</sub>: CH<sub>3</sub>, R<sub>4</sub>: CH<sub>2</sub>OH

134

R<sub>1</sub>

COOR<sub>3</sub>

**135** R<sub>1</sub>: α-OH, β-H, R<sub>2</sub>: α-OH, β-H, R<sub>3</sub>: H **136** R<sub>1</sub>: α-OH, β-H, R<sub>2</sub>: O, R<sub>3</sub>: H

trifoliatus (Lischewski et al., 1985; Kutschabsky et al., 1985). 24-nor-11α-Hydroxy-3-oxo-lup-20(29)-en-28-oic acid [136] was isolated for the first time from the leaves of *A. trifoliatus* (Lischewski et al., 1985).

**Seco-lupane triterpenoids** – Chiisanogenin [137] was isolated from the root barks of *A. divaricatus* var. *albeofrutus* 

137 R<sub>1</sub>: OH, R<sub>2</sub>: CH<sub>3</sub>, R<sub>3</sub>: H, R<sub>4</sub>: H 138 R<sub>1</sub>: OH, R<sub>2</sub>: CH<sub>3</sub>, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: H 139 R<sub>1</sub>: H, R<sub>2</sub>: CH<sub>3</sub>, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: H 141 R<sub>1</sub>: OH, R<sub>2</sub>: CH<sub>2</sub>OH, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: H 143 R<sub>1</sub>: OH, R<sub>2</sub>: CH<sub>3</sub>, R<sub>3</sub>: Glc-Glc-Rha, R<sub>4</sub>: OH 144 R<sub>1</sub>: OH, R<sub>2</sub>: CH<sub>3</sub>, R<sub>3</sub>: Glc-Glc, R<sub>4</sub>: H

**140** R₁: Glc-Glc-Rha, R₂: H, R₃: H **145** R₁: Glc-Glc-Rha, R₂: H, R₃: OH **146** R₁: Glc-Glc-Rha, R₂: CH₃, R₃: OH

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(Oh et al., 2000).

Chiisanoside [138] was isolated for the first time from the leaves of *A. chiisanensis* (Kim and Hahn, 1980). Chiisanoside was isolated from the barks of *A. senticosus* forma *inermis* (Yook *et al.*, 1991), from the leaves of *A. senticosus* forma *inermis* (Park *et al.*, 2000b), from the leaves and stem barks of *A. chiisanensis* (Hahn *et al.*, 1984; Kasai *et al.*, 1986), from the fruits of *A. chiisanensis* (Shin *et al.*, 1992), from the leaves of *A. divaricatus* (Matsumoto *et al.*, 1987; Yook *et al.*, 1996; Shirasuna *et al.*, 1997), from the root barks of *A.* 

divaricatus var. albeofrutus (Oh et al., 2000) and from the leaves of A. divaricatus forma nambunensis (Cho et al., 1999). Chiisanoside exhibited non-toxic effects and significant anti-histaminic activities. It was found that it showed the anti-diabetic activities against epinephrine- and alloxaninduced diabetes, decreased the toxicities by ephedrine hydrochloride and promoted the elimination of chloramphenicol from blood. It also increased the survival rate in rats intoxicated by carbon tetrachloride and led to the re-establishment of normal enzymatic function. In the histopathological studies, it improved fatty degeneration and parenchymal cell necrosis of the liver induced by carbon tetrachloride in rats (Kim and Hahn, 1980). An anti-cancer activity and an antinephrotoxicity were tested by MTT assay. An anti-cancer effect of chiisanoside from A. divaricatus was much lower than that of cisplatin (Yook et al., 1996). Chiisanoside decrease the clearance rate of carbon (Lee *et al.*, 1987).

1-Deoxychiisanoside [139], 11-deoxyisochiisanoside [140], 24-hydroxychiisanoside [141] and inermoside [142] were isolated for the first time from the leaves of *A. senticosus* forma *inermis* (Park *et al.*, 2000a). 22a-Hydroxychiisanoside [143] was isolated from the leaves of *A. divaricatus* (Shirasuna *et al.*, 1997) and the leaves of *A. senticosus* forma *inermis* (Park *et al.*, 2000b).

Divaroside [144] was isolated from the leaves of *A. divaricatus* (Matsumoto *et al.*, 1987) and the leaves of *A. senticosus* forma *inermis* (Park *et al.*, 2000b).

iso-Chiisanoside [145] was isolated from the leaves and stem barks of *A. chiisanensis* (Kasai *et al.*, 1986), from the leaves of *A. senticosus* forma *inermis* (Park *et al.*, 2000b) and from the leaves of *A. divaricatus* (Matsumoto *et al.*, 1987; Shirasuna *et al.*, 1997).

iso-Chiisanoside methylester [146] was isolated from the leaves and stem barks of *A. divaricatus* (Kasai *et al.*, 1986) and the leaves of *A. senticosus* forma *inermis* (Park *et al.*, 2000b).

Sachunoside [147] was isolated from the leaves of *A. divaricatus* var. *sachunensis* (Park *et al.*, 2001).

**Phytosterols** – Campesterol [148] was isolated from the root barks of *A. divaricatus* (Yook *et al.*, 1996) and the root barks of *A. sessiliflorus* (Yook *et al.*, 1977).

β-Sitosterol [149] was isolated from the barks of A. senticosus forma inermis (Yook et al., 1991), from the root barks of A. divaricatus (Yook et al., 1996), from the root barks of A. sessiliflorus (Yook et al., 1977), from the leaves of A. trifoliatus (Chen et al., 1972a; Chen et al., 1972b; Chen et al., 1973) and from the root barks of A. setchuenensis (Zhao et al., 1999). β-Sitosterol inhibited growth of human colon cancer (ht-29) by activating sphingomyelin cycle (Awad et al., 1998), showed anti-inflammatory and anti-

148 R<sub>1</sub>: OH, R<sub>2</sub>: CH<sub>3</sub> 149 R<sub>1</sub>: OH, R<sub>2</sub>: CH<sub>2</sub>CH<sub>3</sub>

150 R<sub>1</sub>: O-Glc, R<sub>2</sub>: CH<sub>2</sub>CH<sub>3</sub>

**151** R: OH **152** R: O-Glo

pyretic effect (Gupta *et al.*, 1980), reduced dietary cholesterol absorption in humans (Heinemann *et al.*, 1993) and reduced levels of insulin (Ivorra *et al.*, 1988).

Daucosterol ( $\beta$ -sitosterol glucoside, eleutheroside A) [150] was isolated from the leaves of *A. trifoliatus* (Chen *et al.*, 1972b; Chen *et al.*, 1973) and from the root barks of *A. setchuenensis* (Zhao *et al.*, 1999). Daucosterol decreased vascular permeability and showed hemostatic effect (Sugiyama and Seki, 1991).

Stigmasterol [151] was isolated from the root barks of *A. divaricatus* (Yook *et al.*, 1996), from the root barks of *A. sessiliflorus* (Kim, 1985; Yook *et al.*, 1977), from the leaves of *A. trifoliatus* (Chen *et al.*, 1972b; Chen *et al.*, 1973) and from the barks of *A. senticosus* forma *inermis* (Yook *et al.*, 1991).

Stigmasterol glucoside [152] was isolated from the leaves of *A. trifoliatus* (Chen *et al.*, 1972b; Chen *et al.*, 1973).

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**Polyacetylenes** – Falcarinol [153] and falcarindiol [154] were isolated from the root barks of *A. koreanum* (Kim *et al.*, 1988) and from the roots of *A. divaricatus* (Miyakoshi *et al.*, 1995). Falcarindiol was isolated from the root barks of *A. koreanum* (Chung and Kim, 1986).

**A pyrimidine** – Thymidine [155] was isolated from the roots of *A. senticosus* (Li *et al.*, 2001).

**Cyclitols** – Myo-inositol [156] and scyllo-inositol [157] were isolated from the leaves of *A. sciadophylloides* (Yasue *et al.*, 1968) and from the leaves of *A. trifoliatus* (Chung *et al.*, 1972a).

**Monosaccharides** – Isomaltol 3-O- $\alpha$ -glucoside [158] was isolated for the first time from the roots of A. senticosus (Li et al., 2001).

Eleutheroside C [159] was isolated from the roots of *A. senticosus* (Brekhman and Dardymov, 1969).

An aliphatic alcohol – 1-Dotriacontanol [160] was isolated from the leaves of *A. sciadophylloides* (Kitajima *et al.*, 1989).

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