RAPD Variation and Genetic Distances among Tibetan, Inner Mongolia and Liaoning Cashmere Goats

Shilin Chen¹, Menghua Li, Yongjun Li², Shuhong Zhao*, Chuanzhou Yu, Mei Yu, Bin Fan and Kui Li Laboratory of Molecular Biology & Animal Breeding, School of Animal Science and Veterinary Medicine Huazhong Agricultural University, Wuhan, 430070, P. R. China

ABSTRACT: Relationship among Tibetan cashmere goats, Inner Mongolia cashmere goats and Liaoning cashmere goats was studied using the technique of random amplified polymorphic DNA (RAPD). One primer and four primer combinations were screened. With the five primers and primer combinations, DNA fragments were amplified from the three breeds. Each breed has 28 samples. According to their RAPD fingerprint maps, the Nei's (1972) standard genetic distance was: 0.0876 between Tibetan cashmere goats and Inner Mongolia cashmere goats, 0.1601 between Tibetan cashmere goats and Liaoning cashmere goats, 0.0803 between the Inner Mongolia cashmere goats and Liaoning cashmere goats it coincides with their geographic location. The genetic heterogeneity of Tibetan cashmere goats, Inner Mongolia cashmere goats and Liaoning cashmere goats is 0.3266, 0.2622 and 0.2475 respectively. It is also consistent with their development history. (Asian-Aust. J. Anim. Sci. 2001. Vol 14, No. 11: 1520-1522)

Key Words: Tibetan Cashmere Goats, Inner Mongolia Cashmere Goats, Liaoning Cashmere Goats, RAPD, Genetic Heterogeneity, Genetic Distance

INTRODUCTION

Cashmere goats originate from Tibet, Middle Asia and other inner Asian areas (Seng, 1997; Jia et al., 1997). They exist extensively in the area of 'Three North' in China. The output of Chinese cashmere down is the highest and the quality is the best of the cashmere downs in the world (Chang, 1996). The Tibet, Inner Mongolia and Liaoning cashmere goats are the most popular in the Chinese cashmere goats. Understanding the germplasm characters and genetic background of the Tibet, Inner Mongolia and Liaoning cashmere goats at the molecular level is becoming increasingly important.

RAPD is a new technique based on PCR (Williams et al., 1990; Welsh and McClelland, 1990). It is a convenient, efficient and sensitive genetic marker of detecting the polymorphism of genome DNA. 10 base (GC-rich) oligonucleotide primers of arbitrary sequence were used in the PCR amplification. And it can get amplification results of several loci. The RAPD technique is extensively used since it appears (Welsh et al., 1991; Kemp and Wolferen, 1992; Rothuizen et al., Mohd-Azmi et al., 2000, 1994; Hwang et al., 2001).

MATERIALS AND METHODS

Sample collecting

Blood samples were collected from three goats breeds:

Received January 20, 2001; Accepted July 10, 2001

The Tibetan were from the Baitu county cashmere goats keeping farm in Ali territory, Tibetan; The Inner Mongolia were from the Inner Mongolia cashmere goats keeping farm; The Liaoning were from the Liaoning cashmere goats keeping farm in Gaizhou city, Liaoning province. All blood samples are unrelated in two generations. Genomic DNA was prepared from peripheral blood leukocyte cells using the procedure described by Li (1997).

PCR amplification

PCR reactions performed in a 25 µl volume containing 50 ng template DNA, 10×Buffer 2.5 μl, 25 mM MgCl₂2.0 μl, 2 mM of each dNTP 2 µl, 5 U/µl Taq DNA polymerase 0.3 μl, 10 μM primer 1 μl. each reaction was overlaid with 20 µl mineral oil to prevent evaporation. DNA was amplified in a DNA-Thermocycler programmed for 3 min predenaturation at 95°C, followed by 40 cycles. Including 45 seconds at 94°C, 1 min cycle at 37°C, 2 min cycle at 72°C, and then followed by a 5 min extension at 72°C. Blank control was set up for each PCR reaction. All the random primers were recommended by the Operon Company and synthetised by the Shanghai Institute of Cell, the Chinese Academy of Sciences. With the five screened primers and primer combinations, DNA fragments were amplified from the three breeds. Each breed had 28 samples. All the amplification products separated by electrophoresis in 1.5% agarose gels (Tonghai Company, Shanghai) containing 0.05% ethidium bromide, and photographed under UV light. Reproductibility of results was evaluated by replicating the RAPD analysis on DNA samples of all 84 individuals with primers RA59, RA03+RA35, RA04+RA23, RA09+RA35 and RA20+RA33.

^{*} Corresponding Author: Shuhong Zhao, Tel: +86-27-87281306,

Fax: +86-27-87280408, E-mail:zhaoshuhong@yahoo.com

Address reprint request to Shilin Chen. Tel: +86-27-87281311,
E-mail: chen shilin@263.net

²Department of Animal Science, Jilin Agricultural University, Changchun, 130118, P. R. China.

Data analysis

Only clear bands of RAPD products on agarose gels were scored. Nei's (1972) standard genetic distance (D) between breeds and genetic heterogeneity within breeds (H) were calculated according to the following equation.

$$D = -\ln \left(\frac{\sum \sum P_{1mi} P_{2mi}}{\sqrt{\sum \sum P_{1mi}^2} \sqrt{\sum \sum P_{2mi}^2}} \right)$$

Where m is number of loci, i is the ith locus, P_{1mi} and P_{2mi} are the band frequencies of the ith loci in the first and the second population respectively.

$$H = 1 - \sum \frac{2N_{ab}}{(N_a + N_b)}$$

Where N_a and N_b are number of bands for individual a and individual b respectively, N_{ab} is the number of shared bands for individual a and individual b.

RESULTS

Primer screening

There DNA samples of Tibet cashmere goats were used as the amplification templates to carry out the primer screening. Of the 37 arbitrary primers used, the primer of RA59 yield more than four polymorphism bands. Four combination (RA03+RA35, RA04+RA23, primer RA09+RA35, RA20+RA33), whose polymorphism bands are more than four, are screened from 55 primer combinations, which are formed by 11 primers (RA03, RA04, RA09, RA20, RA22, RA23, RA33, RA35, RA41, RA45, RA50) whose amplification bands are more than 8. Those 92 primers and primer combinations yield 560 clear banding pattern of amplified fragments ranging from 300 bp to 5000 bp. Table 1 shows the screening results of the five primers and primer combinations.

PCR amplification

61 clear-banding pattern of amplified fragment were

produced. 55 (99.16%) of them were polymorphic. Six loci were common. 2,422 (82.7%) were polymorphic in 2926 fragments amplified by the five primer and primer combinations. The frequency of these polymorphic bands ranged from 0.0952 to 0.9048.

Genetic heterogeneities of Tibetan, Inner Mongolia and Liaoning cashmere goats are 0.3266, 0.2622 and 0.2475 respectively. The genetic distances are: 0.0876 between Tibetan cashmere goats and Inner Mongolia cashmere goats, 0.1601 between Tibetan cashmere goats and Liaoning cashmere goats, 0.0803 between the Inner Mongolia cashmere goats and Liaoning cashmere goats and Liaoning cashmere goats.

DISCUSSION

Polymorphism of RAPD fingerprint

Goats are highly resistant to inbreeding depression. The study demonstrates that the frequency of polymorphic bands is too low (0.0833) or too high (0.9167). In order to detect the polymorphism of RAPD markers between goats, a method of combined primers was used. It can meet the requirements of RAPD polymorphic fingerprint. However, it often results in reducing amplification amount. The reason may be the length of the amplified fragments becoming shorter or may be the reduced amount of primers to the amplified fragments. The flaw can be easily ameliorated by increasing the amount of amplified products in electrophoresis.

Genetic variation of cashmere goats

The genetic heterogeneity in the populations of Tibetan, Mongolia and liaoning cashmere goats were 0.3266, 0.2622 and 0.2475 respectively. The results reflected that the genetic variations maybe consistent with their selection history. The selection and breeding of Inner Mongolian cashmere goats and Liaoning cashmere goats have been carried out since the 1980s. So genetic variation is lower. On the other hand, the breeding population of Tibetan cashmere goat was formed in 1994, so the selection and

Table 1. The screening results of the five primers and primer combinations

Primer primer combinations	Nucleotides Sequence (5'—3')	Number of RAPD Markers	
		Total number	Variable number
RA59	CGGGCAACGT	11	5
RA03+RA35	CGATCGAGGA	11	4
	AAGCTCCCCG		
RA04+RA35	GCAGAGCATC	11 .	5
	CTAGCTGACG		
RA09+RA35	ACTCCGCAGA	14	6
	AAGCTCCCCG		
RA20+RA33	AATCGATACG	11	4
	TGCGGACGTC		

1522 CHEN ET AL.

breeding history of it is short and the genetic variation of it is higher (0.3266).

Genetic distance between cashmere breeds

In china, Inner Mongolia is located in the middle of Tibetan and Liaoning province. The genetic distances between Tibetan and Inner Mongolia cashmere goats, Tibetan and Liaoning cashmere goats, Inner Mongolia and Liaoning cashmere goats are 0.0816, 0.1601 and 0.0803 respectively. They coincide with their geographic location. The result conforms that goats originate from Tibetan, Middle Asia and other inner Asian areas. Furthermore, we can conclude that the goat's index of Genetic distance and the time of migration from Tibet to the around areas is consistent with its migration distance.

The results described in this paper demonstrate that the technique of RAPD is short of stability, but it is a simple and convenient method. Producing much polymorphic information of target genomes with more primers can compensate the shortcoming of this method. The technique of RAPD is still a good method of studying genetic variation with breed and genetic distance among breeds.

ACKNOWLEDGEMENTS

This study was financially supported by the International Foundation for Science, the National Natural Science Foundation of China to Shuhong Zhao, the Key Scientific Project of Tibet to Yongjun Li and Key Projects of State Basic Research and Developmental Plan of China to Kui Li. The authors acknowledge Pinglei Zhou, Tongan Xun, and Zhengfang Wu for their technical assistance during these trials.

REFERENCES

- Chang, Q. 1996. Introduction of cashmere goat and goat down. J. Qinhai Anim. Husb. and Veteri. 26:34-40.
- Jia, Q., H. Chang and Q. Zhang. 1997. Origination, Domestication and Breeds Formation of Goat. J. Hebei Agr. Uni. 20(2):68-71.
- Hwang, K. C., K. D. Song, T. H. Kim, D. K. jeong, S. H. Sohn, H. S. Litlehoj and J. Y. Han. 2001. Genetic Linkage Mapping of RAPD Markers Segregating in Korean Ogd Chicken-White Lehorn Backcross Populatioin. Asian-Aust. J. Anim. Sci. 14:302-306.
- Kemp, S. J. and A. J. Teale. 1992. Random amplified DNA polymorphisms (RAPDs) and pooled DNA in bovine genetic studies. Anim. Genet. 23(Supp. 1):62.
- Li, X., Y. Z. Gong, S. H. Zhao, K. Li and Z. Z. Peng 1997. A whole blood lyses method for the isolation of porcine genomic DNA in pig farm. J. Hebei Agri. Uni. 20(4):84-86.
- Li, Y. J., S. L. Chen and L. Ma. 1998. Random amplified polimorphic DNA study of Tibetan cashmere goat. Proceedings of the 6th World Congress on Genetics Applied to Livestock Production, Armidale, 24:103-106.
- Mohd-Azmi M. L., A. S. Ali and W. K. Kheng. 2000. DNA Fingerprinting of Red Jungle Foul, Village Chicken and Broilers. Asian-Aust. J. Anim. Sci. 13:1040-1043.
- Nei. M. 1972. Genetic distance between populations. Ame. Nat. 106: 283-292.
- Rothuizen, J. and M. V. Wolferen. 1994. Random amplified DNA polymorphisms in dogs are reproducible and display Mendelian transmission. Anim. Genet. 25:13-18.
- Seng, J. M. 1997. Origination and development of goat. J. Chinese Sheep and Goat 1997(1):6-8.
- Welsh, J. and M. McClelland. 1990. Fingerprinting genomes using PCR with arbitrary primers. Nuc. Aci. Res. 18:7213-7218.
- Welsh, J., C. Petersen and M. McClelland. 1991. Polymorphisms generated by arbitrary primed PCR in the mouse application to strain identification and genetic mapping. Nuc. Aci. Res. 19:303-306.
- Williams, G. K., A. R. Kubelik, K. J. Livak, J. A. Rafalski and S. V. Tingey. 1990. DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. Nuc. Aci. Res. 18:6531-6535.