

I. 14~16) plasma (titanium plasma spray; TPS)  
 Hahn Palich<sup>16)</sup>  
 1970 . TPS

. 1950  
 Brånemark<sup>3,4)</sup> 1,2),  
 (osseointegration) (removal torque value; RTV)  
 Albrektsson<sup>5)</sup> 17,18) - 19). Buser<sup>20)</sup>  
 5  
 가  
 , hydroxyapatite (HA)  
 sandblasting  
 6~11). 가 가  
 21). HA HA가  
 sandblasting , HCl/H<sub>2</sub>SO<sub>4</sub>  
 가  
 plasma  
 6,12,13). 20,21),  
 20 blasting 22) sand -  
 (sand -

blasted larger - grit acid - etched; SLA)  
가 23,24). Buser  
25), Simpson Snetivy24)

U.S.A.) 9 ,  
( 4.0mm, 8.5mm,  
Osseotite , Implant Innovations, Inc.,  
U.S.A.) 9 , SLA ( 4.1mm,  
8mm, ITI , Straumann, Ger -  
many) 9 4 , 12 3

SLA  
, TPS

SLA  
가 2.

23~26).

TPS  
3 - 8  
( ) 2mg/kg

가 27), 23~26)  
SLA

가 1 , 1

20,22,24,25)

, )  
. No. 15  
가

가  
가 Steri - Oss surgical  
kit, 3i surgical kit, ITI surgical kit

3

가 3

II.

1.

1

가 15kg sandblasting 2 ,  
6 3

( 3.8mm,  
8mm, Steri - Oss , Bausch & Lim Co.,

가

3 - 0  
(Mersilk , Ethicon Co., U.K.)

lation Inc., U.S.A.)

7 ( 4 12 3 - 5  
, 500mg/day) 2 1/3 3

2)

가

. 4

2

5)

12

9

calcein (Sigma Co., U.S.A.) , 3  
oxytetracycline  
HCl( , ) . 12  
2 calcein,  
4 oxytetracycline HCl, 3  
alizarin red (Juncei  
Chemical Co., Japan) kg 20mg

handpiece disk  
torque driver  
가 ' - '  
Tohnichi torque driver (Tohnichi Mfg. Co.,  
Japan) ( :Ncm)

3)

3 4 , 12  
가

6)

70%

7

2

methylmethacrylate ,  
8 - 10 $\mu$ m  
Hema -

, SAS program  
one - way ANOVA Duncan

toxylin - Eosin

III.

4)

1.

Global Lab Image  
Analysis (Data Translation Inc., U.S.A.)  
(Olympus BH - 2,  
Olympus Co., Japan) CCD (ITC -  
47, Ikegami Tsushinki Co., Ltd., Japan)  
(1DT - 55, Data Trans -

4 가 2 3  
12 가 3  
0.12% 2

2.

1) 1  
4

2) 2  
4 가 1

가 1 가

(Figure 3).

12 1 가  
1 가 가  
1 12

4  
(Figure 4).

가  
(Figure 1).  
12

4

가 3) 3  
4 가 1  
, 2 가  
1, 2

가 4 가 1  
가 4 가 1, 2

가 12  
(Figure 2).

가 12  
(Figure 5).

Table 1. Implant bone contact ratios for 3 different implant types in the canine mandibles at 4 and 12 weeks of healing (n=3).

Group	Weeks of healing		significance level for the difference	
	4 weeks	12 weeks		
Group	* [ *	54.3 ± 2.5	64.3 ± 1.9	P<0.05
Group		57.7 ± 2.1	66.7 ± 2.2	P<0.05
Group		66.2 ± 2.1	71.2 ± 2.5	N S

Values are mean ± SE (%).

Group : Titanium machined implant

Group : Titanium implant with acid - etched surface

Group : Titanium implant with SLA surface

Asterisk (\*) means statistically significant difference among 3 groups by one - way ANOVA and Duncan's mul -

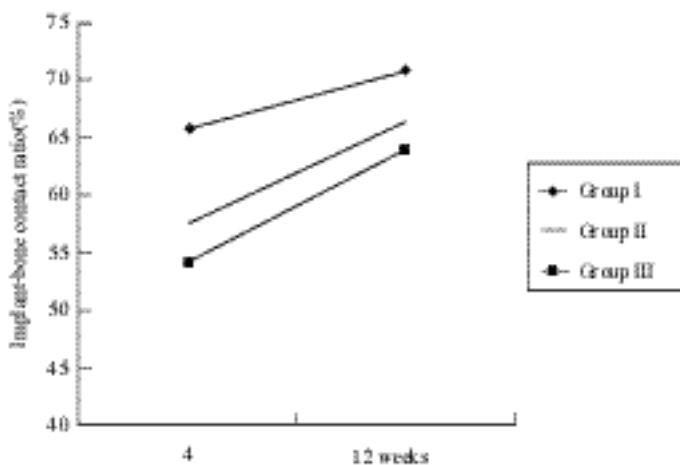


Figure 7. Implant bone contact ratios for 3 different implant types in the canine mandibles at 4 and 12 weeks of healing (n=3).

Group : Titanium machined implant

Group : Titanium implant with acid - etched surface

Group : Titanium implant with SLA surface

Table 2. Removal torque values for 3 different implant types in the canine mandibles at 12 weeks of healing (n=3).

Group	RTV (Ncm)	Duncan Grouping
Group	77.1 ± 0.6	B*
Group	81.6 ± 0.5	A B
Group	90.9 ± 0.4	A

The values are mean ± SE (Ncm).

Group I : Titanium machined implant.

Group II : Titanium implant with acid - etched surface.

Group III : Titanium implant with SLA surface.

Asterisk (\*) means with the same letter are not significantly different by Duncan grouping(p<0.05).

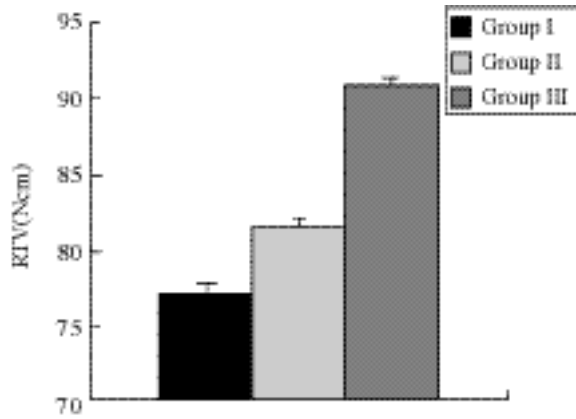


Figure 8. Removal torque values for 3 different implant types in the canine mandibles at 12 weeks of healing (n=3).

- Group I : Titanium machined implant
- Group II : Titanium implant with acid - etched surface
- Group III : Titanium implant with SLA surface

4 1 2 가 (p<0.05), (Table 1, Figure 7).

가 1 가 4. 가 4 가 12 . 1 77.1 Ncm, 2 81.6 Ncm, 3 90.9 Ncm 3 ,

가 (Figure 6). 가 3 1 (p<0.05), 1 2 , 2

3. 3 (Table 2, Figure 8).

4 1 IV. 54.3%, 2 57.7%, 3 66.2% 3 가 2 3 가 1 2 가 1~5,28~30). , 1 3 가 (p<0.05). 12 4 가 1 .



54.3%, 2 57.7 %, 3 66.2% . Buser <sup>25)</sup> 가

1 2 3 (p<0.05). 12 1 SLA 가

64.3%, 2 66.7%, 3 71.2% 가 가

3 가 4

12 가 가

1 2 가

가 (p<0.05), 3 4 가 1

12 TPS SLA 151.5 mm<sup>2</sup>, 2 158.5 mm<sup>2</sup>, 3 135.8 mm<sup>2</sup>

3 SLA 1 50.9 Ncm, 2 51.9 Ncm, 3 66.6 Ncm 3 가

Cochran <sup>35)</sup> SLA (p<0.05)

가 SLA 가 22,40,42) 가

가 가

Johansson <sup>37)</sup> SLA 가 가

가 4

Torque gauge SLA TPS

Anusavice <sup>41)</sup> 가 SLA TPS

1 77.1 Ncm, 2 81.6 Ncm, 3 90.9 Ncm 3 1, 2 가 Martin <sup>26)</sup>

SLA 24,25,34)

SLA 가

Simpson Snetivy<sup>24)</sup> Buser <sup>25)</sup> 가 가 가 SLA



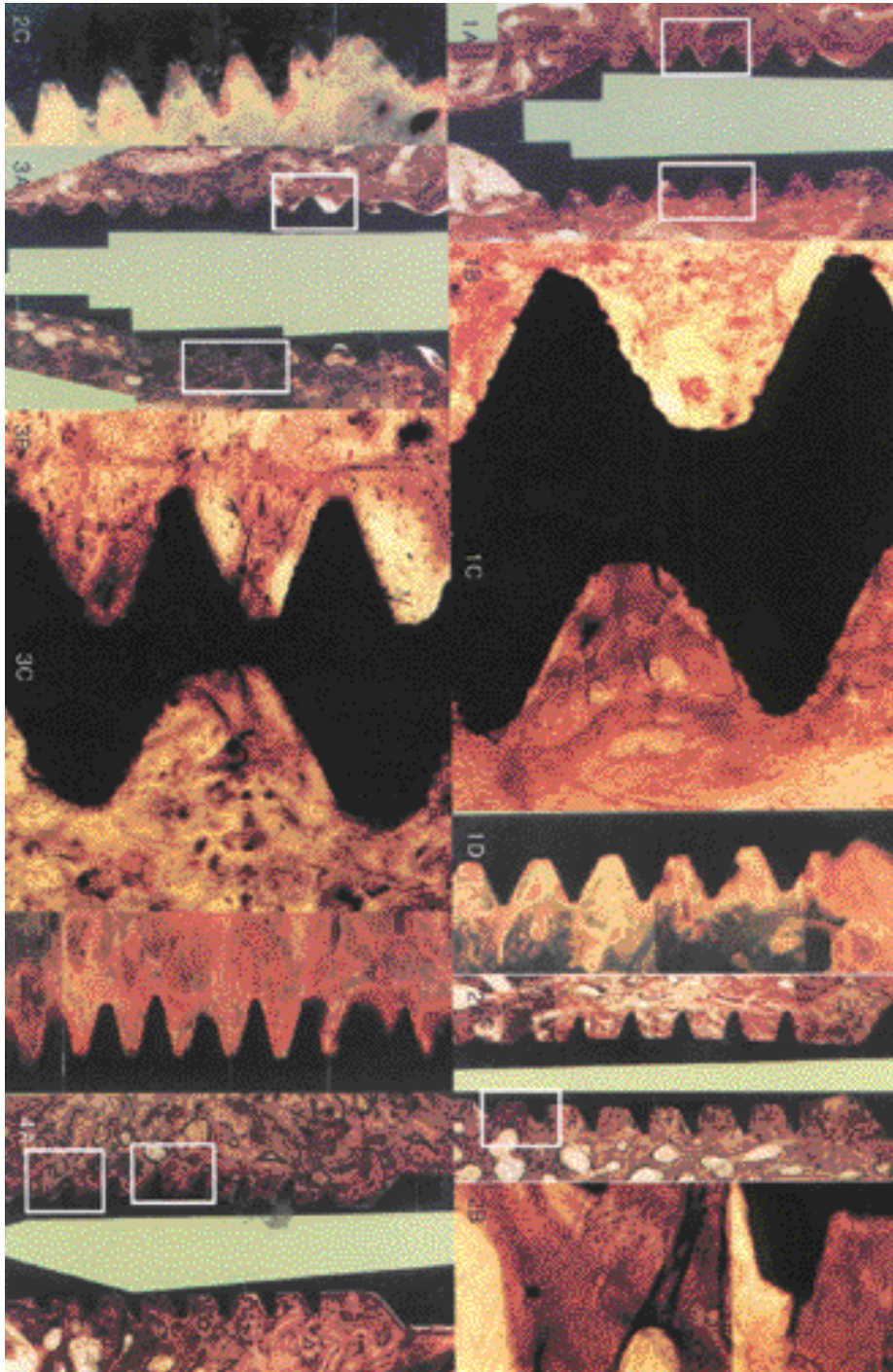


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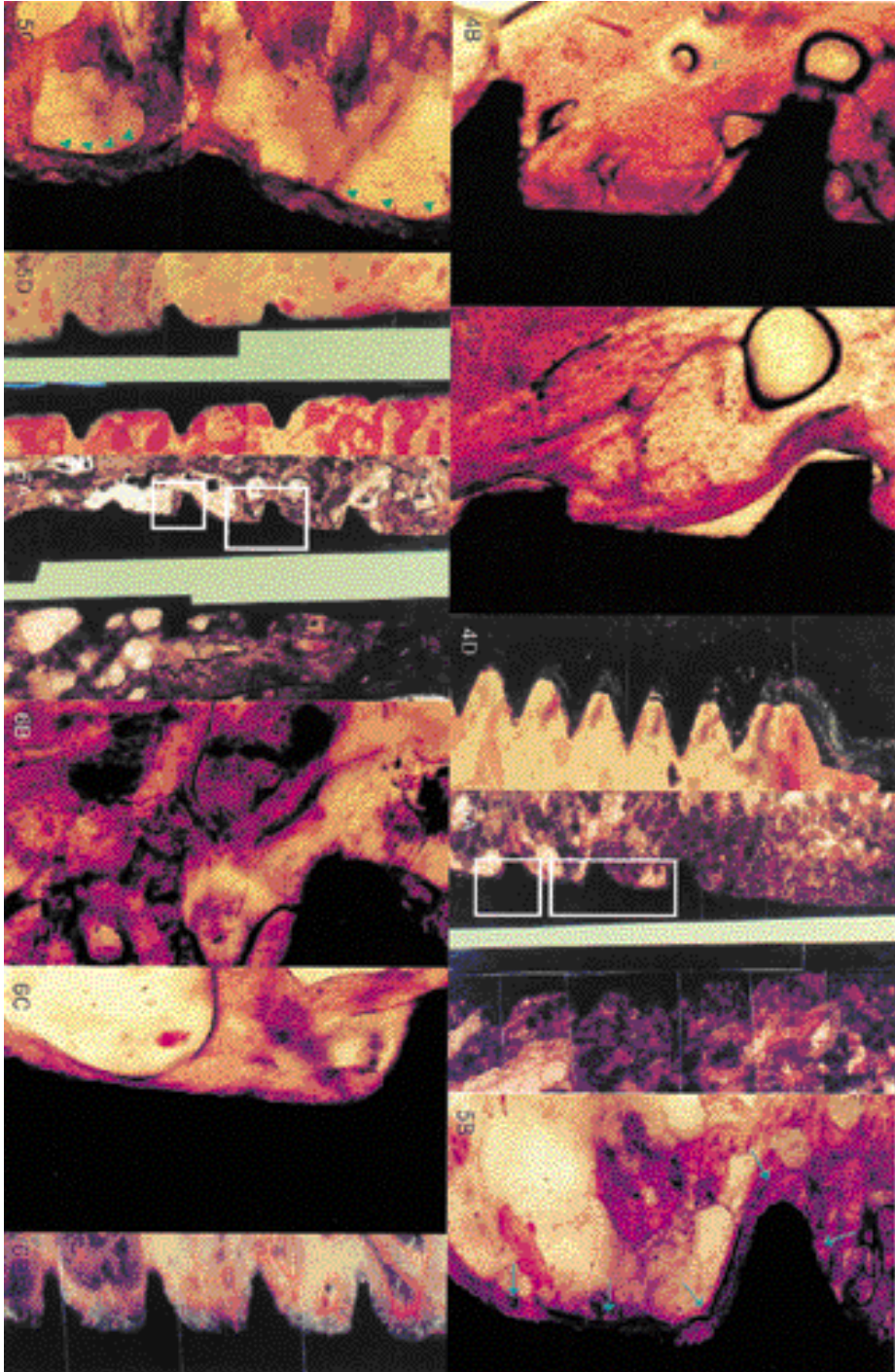
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(1)



( II )



1981.

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Figure 1. Bucco - lingual ground section at 4 weeks after implantation (Group I).  
Soft tissue (fibrous tissue) ingrowth is found on top of the implant, but limited to the neck (arrow heads). The implant threads are in contact with newly formed immature bone, but some threads are not contacted with immature bone (H - E stain, A  $\times$  40, B and C  $\times$  100).

Under the fluorescent microscope, irregular yellow fluorescence is observed in some threads (arrow heads) and illustrated bone apposition. Irregular elliptic shaped lines of green - yellow color are observed adjacent to the implant threads (D  $\times$  40).

Figure 2. Bucco - lingual ground section at 12 weeks after implantation (Group I).  
The surrounding bone of implant is more mature than at 4 weeks. The implant is well connected with the mature lamellar bone (A  $\times$  40, B  $\times$  100).

Under the fluorescent microscope, inner yellow and outer red fluorescent bands are seen adjacent to the implant (C  $40\times$ ).

Figure 3. Bucco - lingual ground section at 4 weeks after implantation (Group II).  
There are direct apposition of new woven bone to implant threads (arrow heads), but some threads do not contact with immature bone (H - E stain, A  $\times$  40, B and C  $\times$  100). Under the fluorescent microscope, irregular green and yellow fluorescent lines are observed in some threads, which illustrated bone apposition (D  $\times$  40).

Figure 4. Bucco - lingual ground section at 12 weeks after implantation (Group II).  
The surrounding bone of implant is more mature than at 4 weeks. The trabecular pattern appears thicker and compacter than at 4 weeks. The implant is well connected with the mature lamellar bone. In areas adjacent to the implant surface, lamellation (L) of the newly formed bone would found (H - E stain, A  $\times$  40, B and C  $\times$  100).

Under the fluorescent microscope, yellow fluorescent areas are surrounded by red fluorescent lines in bony trabeculae (D  $\times$  40).

Figure 5. Bucco - lingual ground section at 4 weeks after implantation (Group III).  
There are extensive direct apposition of new woven bone to implant surface, seen as

the darker red stained areas (arrow heads). Osteoblasts (arrow heads) are lined along the newly formed bone (H - E stain, A × 40, B and C × 100).

Under the fluorescent microscope, thick regular green fluorescent lines are observed along the threads (arrow heads) which illustrated early bone apposition (D × 40).

Figure 6. Bucco - lingual ground section at 12 weeks after implantation (Group III).

The surrounding bone of implant is more mature than at 4 weeks after implantation. The trabecular pattern appears thicker and more compact than at 4 weeks after implantation. The implant is well connected with the mature lamellar bone. In areas closer to the implant thread, lamellation (L) of the newly formed bone would found (H - E stain, A × 40, B and C × 100).

Under the fluorescent microscope, thick regular green fluorescent lines are observed along the threads. Inner yellow and outer red fluorescent bands are seen adjacent to the implant (D × 40).

- Abstract -

## Bone Healing around Screw - shaped Titanium Implants with Three Different Surface

## Topographies

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It is well known that the apposition of bone at implant surface would be influenced by the microstructure of titanium implants. The purpose of this study was to compare bone healing around the screw - shaped titanium implant with three different surface topographies in the canine mandibles by histological and biomechanical evaluation.

All mandibular premolars of six mongrel dogs were extracted and implants were placed one month later. The pure titanium implants had different surface topographies: smooth and machined (Steri - Oss : Group I); acid - etched (Osseotite : Group II); sandblasted and acid - etched (ITI , SLA: Group ) surface. The fluorescent dyes were injected on the 2nd (calcein), 4th (oxytetracycline HCl) and 12th (alizarin red) weeks of healing. Dogs were sacrificed at 4 and 12 weeks after implantation. The decalcified and undecalcified specimens were prepared for histological and histometrical evaluation of implant - bone contact. Some specimens at 12 weeks after implantation were used for removal torque testing.

Histologically, direct bone apposition to implant surface was found in all of the treated groups. More mature and dense



bone was observed at the implant - bone interface at 12 weeks than that at 4 weeks after implantation. Under the fluorescent microscope, thick regular green fluorescent lines which mean early bone apposition were observed at the implant - bone interface in Group III, while yellow and red fluorescent areas were found at the implant - bone interface in Group I and II. The average implant - bone contact ratios at 4 weeks of healing were 54.3% in Group I, 57.7% in Group II and 66.2% in Group III. In Group I, implant - bone contact ratio was significantly lower than Group II and III ( $p < 0.05$ ). The average implant - to - bone contact ratios at 12 weeks after implantation were 64.3% in Group I, 66.7% in Group II and 71.2% in Group III. There was no significant difference among the three groups. In Group I and II, the implant - bone contact ratio at 12 weeks increased significantly in comparison to ratio at 4 weeks ( $p < 0.05$ ). The removal torque values at 12 weeks after implantation were 90.9 Ncm in Group I, 81.6 Ncm in Group II and 77.1 Ncm in Group III, which were significantly different ( $p < 0.05$ ).

These results suggest that bone healing begin earlier and be better around the surface - treated implants compared to the smooth surface implants. The sandblasted and acid - etched implants showed the most favorable bone response among the three groups during the early healing stage and could reduce the waiting period prior to implant loading.