

Austropeplea ollula (Pulmonata: Lymnaeidae):
A new molluscan intermediate host of
a human intestinal fluke, *Echinostoma cinetorchis*
(Trematoda: Echinostomatidae) in Korea

Pyung-Rim CHUNG*, Younghun JUNG, Yun-Kyu PARK and Myung-Ki HWANG

Department of Parasitology, Inha University College of Medicine, Jungsuck B/D B Dong,
7-241, 3 RD. Shinheung-Dong, Choong-Gu, Incheon 400-712, Korea

Abstract: Three freshwater snail species of the family Lymnaeidae have been reported from Korea, *Radix auricularia coreana*, *Austropeplea ollula* and *Fossaria truncatula*. Out of 3 lymnaeid snail species, *A. ollula* was naturally infected with the *Echinostoma cinetorchis* cercariae (infection rate = 0.7%). In the experiments with the laboratory-bred snails, *F. truncatula* as well as *A. ollula* was also susceptible to the *E. cinetorchis* miracidia with infection rates of 25% and 40%, respectively. All of three lymnaeid snail species exposed to the *E. cinetorchis* cercariae were infected with the *E. cinetorchis* metacercariae. It is evident that *A. ollula* acts as the first molluscan intermediate host of *E. cinetorchis* in Korea, and *F. truncatula* may be a possible candidate for the first intermediate host of this intestinal fluke. Also, three lymnaeid snail species targeted were experimentally infected with *E. cinetorchis* metacercariae.

Key words: *Austropeplea ollula*, *Echinostoma cinetorchis*, susceptibility

INTRODUCTION

Life cycle studies of *E. cinetorchis* Ando and Ozaki, 1923 (Trematoda: Echinostomatidae), first described as a new species by Ando and Ozaki (1923), were carried out in Korea mostly after Seo and his colleagues (1980) reported a human case of echinostomiasis cinetorchis (Miki, 1923; Seo et al., 1980, 1984; Lee SH et al., 1988; Lee et al., 1990). This human intestinal fluke is morphologically characterized by a head crown with 37-38 collar spines and

especially with 6 spines on the ventral lobe (Ando and Ozaki, 1923; Seo et al., 1964; Lee et al., 1992). A total of 6 human cases of infection with this trematode have been reported in Korea (Seo et al., 1980; Ryang et al., 1986; Lee SK et al., 1988; Ryang, 1990; Son et al., 1994).

Three freshwater lymnaeid snail species, such as *R. auricularia coreana*, *A. ollula*, and *F. truncatula*, belonging to the family Lymnaeidae have been reported so far in Korea. However, only a planorbid snail, *Hippeutis cantori* was listed to be naturally infected with the cercariae of *E. cinetorchis* (Ahn et al., 1989), and was also examined experimentally as the first and second intermediate hosts of this trematode (Lee et al., 1990). As the second intermediate hosts of this trematode, one of the loaches named

• Received 13 August 2001, accepted after revision 28 August 2001.

• This study was supported, in part, by the 2000 Research Fund of Inha University.

*Corresponding author (e-mail: chungpr@inha.ac.kr)

Misgurnus anguillicaudatus (Koga, 1938) and several freshwater snail species such as *H. cantori*, *Radix auricularia coreana*, *Physa acuta* and *Cipangopaludina chinensis malleata* were listed by the Korean parasitologists (Lee SH et al., 1988; Ahn et al., 1989; Chung and Jung, 1999). Recently, *Segmentina hemisphaerula* was confirmed naturally and experimentally as the first and second intermediate hosts of this trematode (Chung et al., 2001).

The present study was carried out to observe the natural infection of the cercariae of *E. cinetorchis* from the lymnaeid snails collected from the local sites, and to test the susceptibility of laboratory-bred Korean lymnaeid snails to infection with the miracidia and cercariae of *E. cinetorchis*.

MATERIALS AND METHODS

Cultivation of the snails

Several local collections of three lymnaeid snail species were mainly used in the culture studies. The snails were maintained mainly in conventional aquaria. Initially, tap water was conditioned before use simply by adding 4-5 drops of 5% sodium thiosulfate solution to each aquarium for dechlorination (pH, 6.5-7.0). This water was aerated for one week prior to use. The aquaria were placed on shelves in a shelving unit and exposed alternately to 12 hr of artificial light provided by a 15-watt cool, white fluorescent tube and 12 hr of darkness. An automatic timer controlled the light cycle. The water of all the glass aquaria or the plastic trays for the juvenile snails was continuously aerated by bubbling air from an air compressor through the water. The water temperature for the aquaria ranged from 25-27°C. Lettuce and Tetra SML flakes (Tetra SML®, Tetra Co., D-452 Melle, West Germany) were supplied to the lymnaeid snails twice a week (Chung, 1984).

Observation of naturally emerging cercariae from snails

Three species of lymnaeid snails collected from various local areas were first examined for the emergence of trematode cercariae. For the observation of trematode cercariae from snails, each snail was placed in a well of 24-

well plastic plate (Emerald Biostructures Inc., Bainbridge Island, WA, USA) containing the appropriate amount of aerated tap water, kept under a fluorescent light for 2 hr and examined for the presence of shedding cercariae. Later, the snails were crushed for further examination of cercariae inside the snails.

Cercarial infection in the second intermediate host

Three species of laboratory-bred lymnaeid snails infected with cercariae, were examined in order to obtain metacercariae of *E. cinetorchis*. A total of 50 cercariae shed from lymnaeid snails were exposed to each experimental snail of *S. hemisphaerula*, known as the second intermediate host (Chung et al., 2001) and the infected specimens were examined for the detection of metacercariae. After cracking the snail shell and removing the pieces of shell, the animal bodies were dissected and digested in artificial gastric juice for 1 hr at 37°C. The metacercariae were collected from snails every week from two weeks after the cercarial challenge. The number of *Echinostoma* metacercariae per known volume was counted under the low power (x 100) of a light microscope.

Metacercarial infection in the final host

A total of 50 metacercariae was fed orally using a tuberculin syringe connected with a plastic tube to each rat (Sprague-Dawley strain, 120 g of body weight). Rat feces was examined daily for eggs of *E. cinetorchis*. Immediately after finding the eggs, the rats were killed by cervical dislocation and dissected. The small intestines of the rats were removed. The worms were collected under the dissecting microscope and their numbers were determined. Worms were fixed with 10% formalin under the cover slip pressure and stained with Semichon's acetocarmine. The stained worms were observed for morphological comparisons with the standard parasitic specimens.

Miracidial and cercarial infection in the laboratory-bred snails

Parasite-free snails were reared in the

laboratory to obtain their laboratory-bred offsprings. The snail eggs of each species were collected and cultured to get laboratory-bred adult snails in a batch which were employed for the susceptibility experiments to the target parasite *E. cinetorchis*. The infected rats were used as the source of eggs for the susceptibility studies of snails. For the susceptibility experiments of *E. cinetorchis* miracidia to the laboratory-bred snails, eggs (Fig. 1B) were collected by tearing the worms. The eggs were rinsed 3 times with distilled water and transferred to the petri dishes containing conditioned water with a few drops of

Fungizon solution (Gibco Life Technologies Inc., Grand Island, New York). The eggs were incubated in the conditioned water at 26°C in a dark incubator with aeration. Some hatched miracidia were used to challenge the three lymnaeid snail species targeted in this study. Each snail was exposed to a dose of twenty miracidia hatched from the eggs of *E. cinetorchis*. Twenty days after miracidial infection, the release of cercariae was examined from the snails kept under the fluorescent illumination (700 Lux) for 2 hr. The cercariae released from laboratory-bred snails were also exposed to the three lymnaeid

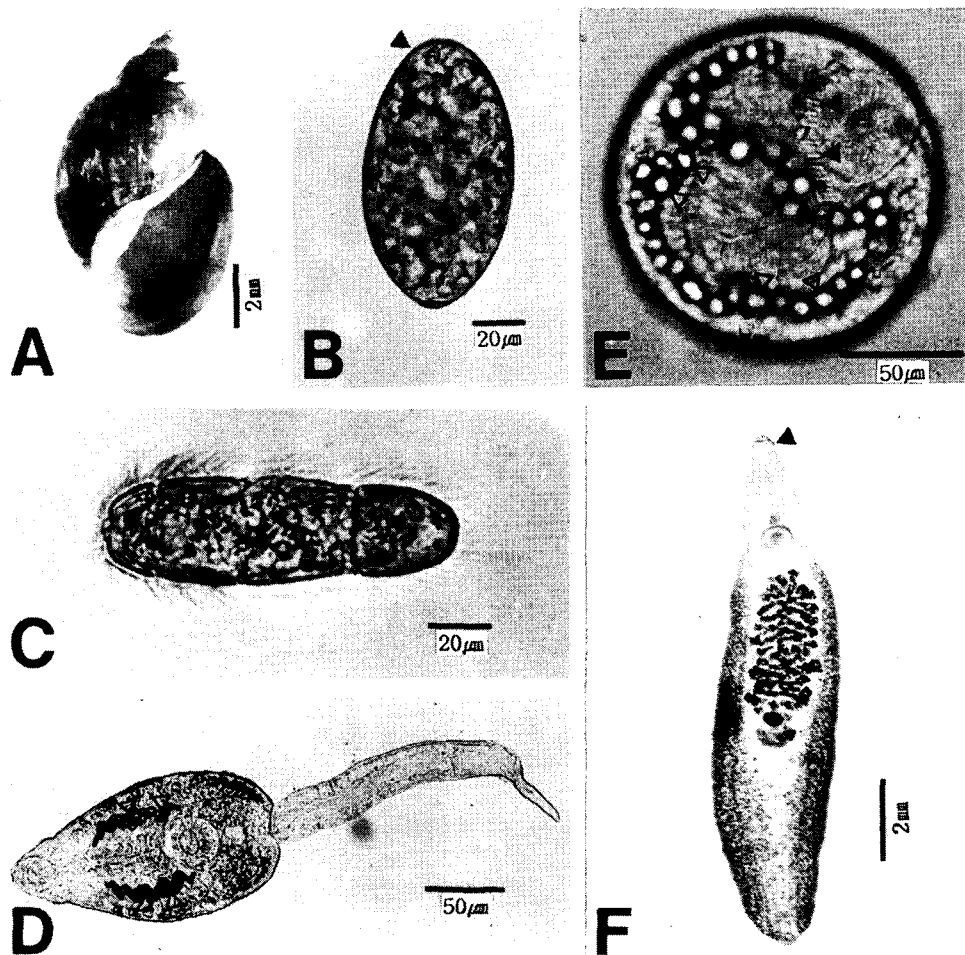


Fig. 1. Experimental subjects used in this study. A, A shell of *Austropeplea ollula*; B, An egg with an inconspicuous operculum (arrow head); C, A miracidium of *Echinostoma cinetorchis* ($\times 100$); D, A cercaria of *Echinostoma cinetorchis*. Note the excretory granules; E, A metacercaria of *Echinostoma cinetorchis*, characterized by the 37-38 spines (black arrow heads) and the excretory granules (white arrow heads); F, An adult worm of *Echinostoma cinetorchis*. A head crown (arrow head) is noticeable.

species targeted for observing the metacercariae of *E. cinetorchis* as described before.

RESULTS

Cercarial infectivity of field-collected snails

Natural infections of *E. cinetorchis* in the three lymnaeid snail species collected from several local areas are shown in Table 1. In field-collected lymnaeid snails, only *A. ollula* (Fig. 1A) was found to be infected with cercariae of *E. cinetorchis* (infection rate: 0.7%), while none of *R. auricularia coreana* and *F. truncatula* were found shedding the cercariae.

The metacercariae of *E. cinetorchis* were recovered in all *S. hemisphaerula* (100%) experimentally infected with echinostome cercariae from field-collected *Austropeplea* snails.

Adult worms were also recovered from the ileo-cecal region of the rats infected with the metacercariae from experimental *S. hemis-*

sphaerula (Fig. 1F). The recovery rate of 20% was recorded in the rats infected with metacercariae three weeks after metacercarial infection.

Miracidial and cercarial infection to the laboratory-bred snails

When the petri dishes containing embryonated eggs were placed under the light sources (17,000 Lux), almost 90% of the miracidia hatched on the 21-22nd day after incubation at 26°C. Miracidia (Fig. 1C) hatched from eggs of *E. cinetorchis* were exposed to the laboratory-bred snails of three lymnaeid species suspected as the first intermediate hosts in this study.

Out of 20 snails of each species infected with miracidia, 8 *A. ollula* and 5 *F. truncatula* (infection rates, 40% and 25%, respectively) shed a large number of echinostome cercariae (Fig. 1D). The snails of *R. auricularia coreana* were not infected experimentally with miracidia of *E. cinetorchis* (Table 2). An average of 1,249 cercariae per each *A. ollula* snail infected with *E. cinetorchis* miracidia were

Table 1. Natural infectivity of *Echinostoma cinetorchis* cercariae in the lymnaeid snails collected from several localities in Korea

Species of snails	Local area collected	Date collected	No. of snails examined	No of positive snails	Infection rate (%)
<i>Radix auricularia coreana</i>	Koyang, Kyunggi	Apr. 29, 1998	70	0	0
	Kimpo, Kyunggi	May 29, 1998	270	0	0
		Sub-total	340	0	0
<i>Austropeplea ollula</i>	Sohre, Inchon	May 29, 1998	720	0	0
		Jun. 24, 1998	470	0	3.8
	Kanghwa, Inchon	Jul. 7, 1998	560	18	2.3
		May 29, 1998	520	13	0
		Jun. 24, 1998	360	0	0.6
	Ansung, Kyunggi	May 29, 1998	650	2	0
	Jinchon, Chungbuk	May 29, 1998	640	0	0
	Umsung, Chungbuk	May 29, 1998	150	0	0
	Wonju, Kangwon	May 29, 1998	870	0	0
May 29, 1998		50	0	0	
	Sub-total	4,990	33	0.7	
<i>Fossaria truncatula</i>	Sohre, Inchon	Jun. 25, 1998	360	0	0
		Jul. 7, 1998	430	0	0
		Sub-total	790	0	0

Table 2. Susceptibility of laboratory-bred lymnaeid snails to the miracidia^{a)} of *Echinostoma cinetorchis*

Snail species	No. of snails examined	No. of snails shedding cercariae	Infection rate (%)
<i>Radix auricularia coreana</i>	20	0	0
<i>Austropeplea ollula</i>	20	8	40
<i>Fossaria truncatula</i>	20	5	25

^{a)}Twenty miracidia of *E. cinetorchis* were exposed to each experimental snail.

Table 3. Cercarial emergence from laboratory-bred snails of *Austropeplea ollula* infected with *Echinostoma cinetorchis* miracidia^{a)}

No. of snails	Days taken for cercarial emergence	Days observed for cercarial emergence	Accumulated No. of cercariae/snail
1	21	18	1,170
2	21	21	1,384
3	22	18	1,020
4	22	19	1,305
5	22	20	1,366
Average	21.6	19.2	1,249

^{a)}Twenty miracidia of *E. cinetorchis* were exposed to each experimental snail.

Table 4. Infection rate of metacercariae in three species of laboratory-bred lymnaeid snails exposed to the *Echinostoma cinetorchis* cercariae^{a)}

Snail species	Days after cercarial exposure	No. of snails examined	No. of snails infected	Infection rate (%)
<i>Radix auricularia coreana</i>	14	10	4	40
<i>Austropeplea ollula</i>	14	10	10	100
<i>Fossaria truncatula</i>	14	10	6	60

^{a)}A total of 50 cercariae of *E. cinetorchis* was exposed to each experimental snail.

released during the 19-day period of observation after the 21st day after miracidial infection (Table 3).

The metacercariae in three laboratory-bred lymnaeid snail species infected with *E. cinetorchis* cercariae from experimentally infected *A. ollula* and *F. truncatula* were also found at the 14th day after cercarial challenge. All three experimental lymnaeid snail species became infected with infection rate of 40%, 100% and 60%, respectively (Table 4 and Fig. 1E).

DISCUSSION

Life-cycle studies of an intestinal trematode,

E. cinetorchis in Korea were carried out mainly with planorbid snails (Ahn et al., 1989; Lee et al., 1990; Chung et al., 2001). Out of 3 lymnaeid snail species occurring in Korea, only *R. auricularia coreana* have been experimentally examined and confirmed that these snails act as the second intermediate host of *E. cinetorchis* with the metacercarial infection rates of 37-66% (Ahn et al., 1989). However, randomly collected field specimens were all employed for the cercarial susceptibility tests. In the present study, the parasite-free, same-aged, laboratory-bred lymnaeid snails were used for testing their susceptibilities to the *E. cinetorchis* miracidia and cercariae. Although we observed that *R. auricularia coreana* were

not susceptible to the *E. cinetorchis* miracidia as confirmed by Ahn et al. (1989), those snails also acted as the second intermediate host of *E. cinetorchis* with the metacercarial infection rate of 40%.

Lymnaea pervia sharing the breeding niches with *H. cantori* in Korea was considered to be a possible second intermediate host of *E. cinetorchis* (Lee et al., 1990). The taxonomic position of this lymnaeid species is confused with that of *A. ollula*, because Lee et al. (1990) listed *L. pervia* without any malacological accounts. On the other hand, *A. ollula* occurring in Korea is very similar to *Lymnaea viridis* from Australia. Pace (1973) reviewed the taxonomic remarks on *A. ollula*, mentioning that *L. ollula* (= *A. ollula*) and *L. pervia* are synonyms of *L. viridis*. Although there is some taxonomic confusion between *Lymnaea* spp. and *A. ollula*, Dr. Morrison of the U. S. National Museum mentioned that *L. pervia* is a junior synonym of *L. (= Austropeplea) ollula* Gould (Burch et al., 1964). However, we have malacological evidence of Korean populations of *A. ollula* different from Australian *L. viridis*. The lymnaeid species employed in this study were identified on the basis of taxonomic keys according to Yoo (1976), Burch et al. (1987) and Kwon (1990). It is the first time that *A. ollula* was confirmed as the first and second gastropod intermediate hosts of *E. cinetorchis* in the present study.

Fossaria truncatula, one of the Korean lymnaeid species, is also a possible gastropod to serve as the first and second intermediate hosts of *E. cinetorchis* based on the experimental results of this study. However, the natural shedding of the *E. cinetorchis* cercariae was not observed in this study. While two lymnaeid species, *R. auricularia coreana* and *A. ollula*, are common mainly in the standing water bodies such as rice fields and pools nationwide in Korea, *F. truncatula* as an amphibious species is uncommon and with less distribution. Unfortunately, the present authors collected *F. truncatula* specimens only from a single locality (Sohre, Incheon) because the ecological habitats of *F. truncatula* were quite limited during the survey period of this study. Therefore, the present authors presume that the natural infection of the *E. cinetorchis*

cercariae from *Fossaria* snails could be confirmed if more snail populations from various collecting sites are investigated in future studies.

ACKNOWLEDGEMENTS

Authors express their appreciation to Prof. Bernard Fried, Department of Biology, Lafayette College, Easton, Pennsylvania for reading the manuscript.

REFERENCES

- Ahn YK, Ryang YS, Chai JY, Sohn WM (1989) Cercarial shedding of *Echinostoma cinetorchis* and experimental infection of the cercariae to several kinds of snails. *Korean J Parasitol* **27**: 203-211 (in Korean).
- Ando R, Ozaki Y (1923) On four new species of trematodes of the family Echinostomatidae. *Dobutsugaku Zasshi* **35**: 108-119 (in Japanese).
- Burch JB, Chung PR, Jung YH (1987) A guide to the freshwater snails of Korea. *Walkerana* **2**: 195-232.
- Burch JB, William JE, Hishinuma Y, Natarajan R (1964) Chromosomes of some Japanese freshwater snails (Basommatophora: Branchiopulmonata). *Malacologia* **1**: 403-415.
- Chung PR (1984) A comparative study of three species of Bithyniidae (Mollusca: Prosobranchia): *Parafossarulus manchouricus*, *Gabbia misella* and *Bithynia tentaculata*. *Malacological Review* **17**: 1-66.
- Chung PR, Jung Y (1999) *Cipangopaludina chinensis malleata* (Gastropoda: Viviparidae): A new second molluscan intermediate host of a human intestinal fluke, *Echinostoma cinetorchis* (Trematoda: Echinostomatidae) in Korea. *J Parasitol* **85**: 963-964.
- Chung PR, Jung Y, Park YK (2001) *Segmentina hemisphaerula*: A new molluscan intermediate host for *Echinostoma cinetorchis* in Korea. *J Parasitol* (in press).
- Koga M (1938) Addition of second intermediate host of *Echinostoma cinetorchis*. *Nippon Kiscichu Gakkai Kiji* **10**: 83.
- Kwon OK (1990) Illustrated encyclopedia of fauna and flora of Korea. Vol. 32, Mollusca (I), Ministry of Education, Seoul, pp. 1-466.
- Lee SH, Chai JY, Hong ST, Sohn WM (1990) Experimental life history of *Echinostoma cinetorchis*. *Korean J Parasitol* **28**: 39-44.

- Lee SH, Jun HS, Sohn WM, Chai JY (1992) Tegumental ultrastructure of juvenile and adult *Echinostoma cinetorchis*. *Korean J Parasitol* **30**: 65-74 (in Korean).
- Lee SH, Sohn WM, Hong ST, Hong SJ, Chai JY (1988) Metacercariae of *Echinostoma cinetorchis* encysted in the freshwater snail, *Hippeutis (Helicorbis) cantori*, and their development in rats and mice. *Korean J Parasitol* **26**: 189-197.
- Lee SK, Chung NS, Ko IH, Ko HI, Sohn WM (1988) A case of natural human infection by *Echinostoma cinetorchis*. *Korean J Parasitol* **26**: 61-64 (in Korean).
- Miki T (1923) On the second intermediate host of *Echinochasmus cinetorchis*. *Aichi Igakkai Zasshi* **30**: 499-504.
- Pace GL (1973) The freshwater snails of Taiwan (Formosa). *Malacological Review Suppl.* **1**: 1-118.
- Ryang YS (1990) Studies on *Echinostoma* spp. in the Chungju Reservoir and upper streams of the Namhan River. *Korean J Parasitol* **28**: 221-233 (in Korean).
- Ryang YS, Ahn YK, Kim WT, Shin KC, Lee KW, Kim TS (1986) Two cases of human infection by *Echinostoma cinetorchis*. *Korean J Parasitol* **24**: 71-76 (in Korean).
- Seo BS, Cho SY, Chai JY (1980) Studies on intestinal trematodes in Korea. I. A human case of *Echinostoma cinetorchis* infection with an epidemiological investigation. *Seoul J Med* **21**: 21-29.
- Seo BS, Park YH, Chai JY, Hong SJ, Lee SH (1984) Studies on intestinal trematodes in Korea. XIV. Infection status of loaches with metacercariae of *Echinostoma cinetorchis* and their development in albino rats. *Korean J Parasitol* **22**: 181-189 (in Korean).
- Seo BS, Rim HJ, Lee CW (1964) Studies on the parasitic helminths of Korea. I. Trematodes of rodents. *Korean J Parasitol* **2**: 20-26.
- Son WY, Huh S, Lee SU, Woo HC, Hong SJ (1994) Intestinal trematode infections in the villagers in Koje-myon, Kochang-gun, Kyongsangnam-do, Korea. *Korean J Parasitol* **32**: 149-155.
- Yoo JS (1976) Korean shells in colour. Il Ji Sa Publishing Co., Seoul, pp. 1-199.