# Effects of Rutin on Adhesion Molecules Expression and NO Production Induced by γ-irradiation in Human Endothelial cells

Eun-Wha Son, Kang-Ro Lee, Dong-Kwon Rhee and Suhkneung Pyo

Division of Immunopharmacology, College of Pharmacy, Sungkyunkwan University, Suwon, Kyunggi-do, Korea, 440-746

(Received September 3 2001; accepted September 21, 2001)

Abstract – Inflammation is a frequent radiation-induced following therapeutic irradiation. Treatment of human umbilical endothelial cells (HUVEC) with  $\gamma$ -irradiation ( $\gamma$ IR) induces the expression of adhesion proteins such as intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1), and E-selectin. Since the upregulation of these proteins on endothelial cell surface has been known to be associated with inflammation, interfering with the expression of adhesion molecules is an important therapeutic target. In the present study, we demonstrate that bioflavonoid rutin inhibits  $\gamma$ IR induced expression of ICAM-1, VCAM-1, and E-selectin on HUVEC in a dose- and time dependent manner. Rutin also inhibited  $\gamma$ IR induced production of NO. These data suggest that rutin has therapeutic potential for the treatment of various inflammatory disorder associated with an increase of endothelial leukocyte adhesion molecules.

**Key words** Rutin, γ-irradiation, Endothelial cells, ICAM-1, VCAM-1, E-selectin, NO

The flavonoids exhibit a wide variety of biological activities, including antiviral, antibacterial, anti-inflammatory, anticarcinogenic, and antioxidant actions (Hertog et al, 1993; Formica et al. 1995). Among them, rutin, a non-toxic flavonoid glycoside with P vitamin activity, is able to suppress various free radical-mediated processes such as in vitro lipid peroxidation (Kozlov et al. 1994), the mutagenic effect of asbestos fibers on human lymphocytes (Korkina et al. 1992), and the overproduction of free radicals in iron-overloaded rats (Afana'ev et al. 1995). Recently, rutin has been shown to have the inhibitiory effects on lung inflammation and interstitial fibrosis in rats induced by bleomycine (Afanas'ev et al. 2001).

Ionizing radiation damage is partially characterized by the generation and the maintenance of an inflammatory reaction (Hruza and Pentland, 1993). An important event in this inflammatory response is the localization of leukocytes at the sites of inflammatory lesions through a multistep process. The endothelial cell adhesion molecules such as E-selectin, intercellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1), play an important role in leukocyte adhesion and transendothelial migration at sites of inflammation. Endothelial expression of these molecule has been known to be elevated in both a temporal and spatial associa-

Since rutin has anti-inflammatory activity and expression of adhesion molecules play an important role in inflammation, we determined whether rutin modulates the expression of adhesion proteins and NO release in irradiated-human umbilical vein endothelial cells (HUVEC). The results of the present study suggest that rutin inhibits the upregulation of  $\gamma$ IR-induced adhesion protein expression. We also observed that rutin blocked the production of NO induced by  $\gamma$ IR.

## MATERIALS AND METHODS

#### Reagents

Rutin was purchased from Sigma Chemical Co. (St Louis,

tion with inflammatory cell infiltrates (Carlos et al, 1994). When activated by inflammatory cytokines, endothelial cells exhibit an upregulation of specific adhesion molecules on their surface, the ligands for which are borne on circulation leukocyte (Bevilacqua et al. 1985; Springer et al, 1990). A logical target for new drug development would be the design of compounds that would interfere with adhesion molecule interactions. It has been suggested that various small molecules, such as glucocorticoids, aspirin and pentoxifylline inhibit the upregulation of adhesion protein expression and have a protective effect on inflammatory diseases (Brostjan et al, 1997; Weber et al, 1995; Neuner et al, 1997).

<sup>\*</sup>To whom correspondence should be addressed.

USA). Anti-ICAM-1 (BBA3), anti-VCAM-1 (BBA6) and anti-E-selectin (BBA1) antibodies were purchased from R & D Systems, USA. Anti-mouse IgG-HRP and p-nitorphenyl phosphate were purchased from Sigma Chemical Co. Fetal bovine serum was purchased from Gibco, USA.

#### Cells and Cell culture

HUVEC were purchased from Clonetics (San Diego, CA) and were grown in EGM-2 meidium (Clonetics) in gelatin coated tissue culture flasks. For subculturing, the cells were detached using 0.125% trypsin containing 0.01 M EDTA. Cells used in this study were from the first to third passage.

#### y-irradiation on cells

Cells were irradiated by a previous method of Gaugler *et al.* (1997). Just before irradiation, the medium of confluent cells was replaced with new medium. The cells were then uniformly irradiated at room temperature with various doses of a  $^{137}$ Cs  $\gamma$ -source (dose rate of 5.94 Gy/min) (IBL 437 C type H, CIS Biointernational, France). The culture medium was renewed after irradiation. For each dose, control cells were simultaneously exposed to sham irradiation.

## Cytotoxicity evaluation

Rutin and γIR at concentrations presented herein were shown to be non-toxic by the following procedure. Viability of HUVEC treated with rutin and/or γIR was determined using MTT assay. Irradiated HUVEC were cultured in gelatin coated 96-well microplate (Costar Products, Cambridge, MA) until confluent. Cells were treated with or without rutin in quadruplicate for indicated time and subsequently 3-(4,5-dimethylthiazol-2-yl)2,5-diphenyl tetrazolium bromide (MTT) was added for 4 h. Also, a visual inspection of the cells was performed by trypan blue exclusion staining under inverted microscope.

### ELISA for measurement of adhesion molecules

The cell surface expression of adhesion molecules on endothelial monolayers was quantified using ELISA by modification of the methods described previously (Gupta and Ghosh, 1999). After irradiation, HUVEC were seeded at a concentration of 2×10<sup>4</sup> cells/well in 96-well, flat bottom, gelatin-coated plates (Nalge Nunc International, IL). The cells were incubated without or with various doses of rutin for the time indicated in the text to measure ICAM-1, VCAM-1 and E-selectin expression. Following incubation, the cells were

washed with phosphate buffer saline pH 7.4 (PBS) and fixed with 10% glutaraldehyde for 30 min at 4°C. Bovine serum albumin (1.0% in PBS) was added to the cells to reduce nonspecific binding. Cells were incubated with anti-ICAM-1, anti-VCAM-1 and anti-E-selectin monoclonal antibody or isotype matched control antibody (0.25 g/ml, diluted in blocking buffer) overnight at 4°C, washed with PBS followed by incubation with alkaline phosphatase-conjugated goat antimouse secondary antibody (1 g/ml, diluted in PBS). The cells were then washed with PBS and exposed to the phosphatase substrate (p-nitorphenyl phosphate 1 mg/ml in 0.1 M glycin buffer, pH 10.4 containing 1 mM MgCl<sub>2</sub> and 1 mM ZnCl<sub>2</sub>). Absorbance was determined at 405 nm by a Molecular device microplate reader (Menlo Park, CA). The absorbance values of isotype matched control antibody were taken as blank and subtracted from the experimental values.

#### Nitrite determination

Irradiated HUVEC were treated with various doses of rutin for times indicated in the text and the accumulation of nitrite in culture supernatant was measured using the assay system described by Ding et al. (1998). Briefly, 100 µl supernatant was removed from each well into an empty 96-well plate. After the addition of 100 µl Griess reagent to each well, absorbance at 540 nm was measured using an a Molecular device microplate reader. Nitrite concentration was calculated from a NaNO<sub>2</sub> standard curve. The levels of absorbance are indicative of NO production. Griess reagent was prepared by mixing one part 0.1% naphthylethylenediamine dihydrochloride in distilled water plus one part 1% sulfanilamide in 5% concentrated H<sub>3</sub>PO<sub>4</sub>.

#### Statistical anaylsis

Each data were given as means ± S.E.M. Statistical difference between groups was determined by one-way analysis of variance (ANOVA) and significant values were represented by an asterisk (\*p<0.05, \*\*p<0.01).

#### RESULTS

# Endothelial cell viability and growth after AR

Although the effect of radiation on endothelial cells has been extensively reported (Rubin *et al.* 1989; Eissner *et al.* 1995; Gaugler *et al.* 1997), the radiosensitivity of HUVEC was assessed in our culture conditions. Viability of the adherent endothelial cells was >95% at the different times tested

158 Eun-Wha Son et al.

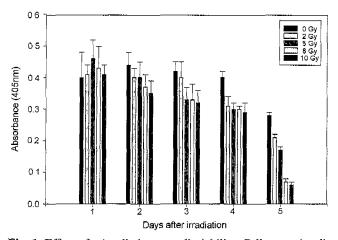


Fig. 1. Effect of  $\gamma$ -irradiation on cell viability. Cells were irradiated with various doses of  $\gamma$ -ray for various times. Each experiment was performed in quadruplicate. The results are mean  $\pm$  S.E.M from three independent experiments.

\*p<0.05, \*\*p<0.01; significantly different from control (shamirradiated cells).

after exposure. However, the number of irradiated cells was decreased with time after exposure, while the number of non-irradiated cells was stable. 3 days after a 8-Gy irradiation, the number of irradiated cells represented 80% of the number of control cells. This percentage was decreased to 10% by 5 days after irradiation (Fig. 1). Therefore, in subsequent studies 8-Gy irradiation was used.

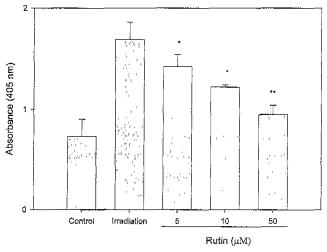


Fig. 2. Dose dependent inhibition of  $\gamma$ -irradiation induced ICAM-1 expression by rutin. Three independent experiments were performed in which confluent HUVEC were sham- or 8 Gy-irradiated without or with indicated concentrations of rutin and the expression of ICAM-1 measured by ELISA. The results are mean  $\pm$  S.E.M from three independent experiments.

\*p<0.05, \*\*p<0.01; significantly different from irradiation-treated control.

# Rutin inhibits \( \gamma \text{IR-induced ICAM-1 expression on endothelial cells in a dose dependent manner} \)

To examine the effect of rutin, HUVEC were incubated without or with various concentrations of rutin for 3 days after  $\gamma$ IR. The time of incubation and concentration of rutin used in these experiments had no effect on the viability as determined by trypan blue staining and morphology of the endothelial cells (data not shown). These concentrations are in complete agreement with the optimum concentrations of rutin needed in other system (Gerritsen *et al*, 1995). As detected by ELISA, ICAM-1 was expressed at low levels on unstimulated endothelial cells and was significantly induced by  $\gamma$ IR (Fig. 2). Rutin did not have effect on the basal level of ICAM-1 expression, whereas it led to a reduction in the  $\gamma$ IR-

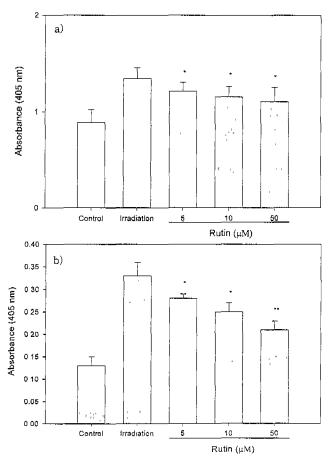


Fig. 3. Effect of rutin on  $\gamma$ -irradiation induced VCAM-1 (a) and E-selectin (b) expression by HUVEC. Three independent experiments were performed in which confluent HUVEC were shamor 8 Gy-irradiated without or with indicated concentrations of rutin and the expression of VCAM-1 or E-selectin measured by ELISA. The results are mean S.E.M from three independent experiments.

\*p<0.05, \*\*p<0.01; significantly different from irradiation-treated control.

induced ICAM-1 expression in a dose dependent manner (Fig. 2).

# Rutin also inhibits $\gamma IR$ -induced VCAM-1 and E-selectin expression

In addition to ICAM-1, γIR also induced VCAM-1 and E-selectin expression in endothelial cells. To examine the effect of rutin on γIR-induced expression of E-selectin and VCAM-1, HUVEC were incubated with various concentrations of rutin for 6 h for the expression of E-selectin and for 3 days for VCAM-1. As measured by ELISA, the unstimulated cells expressed detectable amounts of E-selectin and VCAM-1 (Fig. 3). Upon induction with γIR, a significant increase in the expression of E-selectin was observed. Treatment with rutin inhibited slightly but significantly the expression of VCAM-1 induced by γIR (Fig. 3A). γIR-induced E-selectin expression was also inhibited by rutin in a dose dependent manner (Fig. 3B). Taken together, these results suggest that rutin is effective in blocking the induced level of expression of ICAM-1, VCAM-1 and E-selectin.

#### Rutin inhibits AR-induced NO production

Since NO has been known to be an important modulator of the inflammatory response to various stimuli, we determined the effect of rutin on NO production in endothelial cells. As

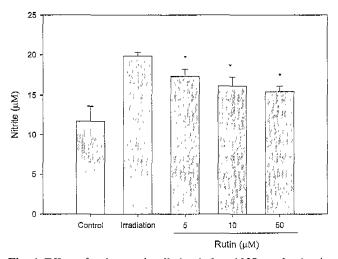


Fig. 4. Effect of rutin on  $\gamma$ -irradiation induced NO production in HUVEC. Three independent experiments were performed in which confluent HUVEC were sham- or 8 Gy-irradiated without or with indicated concentrations of rutin. The conditioned medium was collected, and the nitrite concentration was determined by means of the Griess reagent. The results are mean S.E.M from three independent experiments.

\*p<0.05; significantly different from irradiation-treated control.

shown in Fig. 4, treatment of HUVEC with  $\gamma$ IR resulted in an increase of NO release that was inhibited by rutin in a dose dependent manner.

#### DISCUSSION

Recently, rutin has been known to have an inhibitory effect on lung inflammation and interstitial fibrosis (Afanas'ev *et al.* 2001). Although rutin is found to have anti-inflammatory properties, very little is known in regard to its effect on the induction of cell adhesion molecules by  $\gamma$ IR. In the present study, rutin blocked  $\gamma$ IR-induced expression of leukocyte adhesion molecules, ICAM-1, VCAM-1 and E-selectin. Thus, rutin posseses anti-inflammatory effect on the expression of adhesion protein induced by radiation. This compound also inhibited the NO production in  $\gamma$ -irradiated HUVEC.

Radiation has been shown to induce the expression of a number of genes that participate in the inflammatory response. These include TNF- $\alpha$  and IL-1 which have been known to induce the expression of adhesion molecules such as E-selection when added to endothelial cells in culture (Sherman *et al*, 1991; Brach *et al*, 1993; Bevilacqua *et al*, 1989). When the effect of  $\gamma$ IR on cytokine production was examined in HUVEC, it did not induce the production of TNF- $\alpha$  in irradiated cells (data not shown). Recently, Hallahan *et al*. (1995) demonstrated that E-selectin gene induction by ionizing radiation is independent of cytokine induction. In accordance with their report, our data confirmed that adhesion protein expression did not require cytokine synthesis.

Nitric oxide is a biologically active gas that is synthesized by a variety of cells, including vascular endothelium, from the guanido group of L-arginine. This reactive nitrogen molecule has been invoked as a mediator of vascular phenomena such as arteriolar dilation, platelet aggregation, and platelet-leukocyte adhesion (Moncada, 1992). It has been suggested that NO is able to inhibit LPS-induced ICAM-1 expression (Spiecker et al, 1998). In addition, Kawachi et al. (1999) demonstrated that iNOS-/- mice injected with TNF-α showed an enhancement of VCAM-1 expression in 50% of all tissues compared to wild-type controls. Based on these findings NO could inhibit the expression of adhesion molecules. Our data showed that YIR induces the production of NO and rutin inhibits NO release. Recently, it has been shown that UVB radiation acts as a potent stimulator of NO in human endothelial cells and NO is involved in skin erythema and inflammation (Deliconstantinos et al, 1996). Thus, a role could be proposed for NO 160 Eun-Wha Son et al.

either in inhibiting or promoting inflammation. At present the precise mechanisms accounting for these modulation are unknown. It may be possible, however, to override inhibiting by enhancing inflammation. Moreover, the involvement of NO in the modulation of adhesion molecules expression in inflammation may be dependent on the source of NO, the cells involved and type of stimulus used to induce inflammation.

During severe injury, infection, or ischemia and reperfusion damage, spillover of chemoactivators in the systemic circulation results in cellular activation, leading to the release of injurious agents which damage host tissues. These inflammatory mediators can alter the functional integrity of the vascular system which may be due to the upregulation of expression of cell adhesion molecules. Thus various strategies, such as monoclonal antibodies against adhesion molecules, soluble receptors, soluble counterreceptors, peptides derived from adhesion molecules to prevent receptor-ligand interactions, and antisense oligonucleotides have been employed to inhibit the cell adhesion molecules (Weiser et al, 1997). Glucorticoid, bensothiophene-carboxamide and vitamin A have been shown to inhibit cytokine-or irradiation-induced cell adhesion molecule expression (Brostjan et al. 1997; Cobb et al, 1996; Redlich et al, 1998). Here we demonstrated that rutin effectively blocked the expression of leukocyte adhesion molecules. These studies suggest that rutin may serve as a potential therapeutic tool toward radiation-induced inflammation. Further studies are needed to clarify how this modulation occurs and to what extent it occurs in vivo.

#### **ACKNOWLEDGEMENT**

This work was supported by a National Nuclear R & D Program from the Ministry of Science and Technology of Korea

#### REFERENCES

- Bevilacqua, M. P., Pober, J. S., Wheeler, M. E., Cotran, R. S. and Gimbrone, M. A. Jr. (1985). Interleukin 1 acts on cultured human vascular endothelium to increase the adhesion of polymorphonuclear leukocytes, monocytes and related cell lines. *J. Clin. Invest* **76**, 2003-2009.
- Bevilacqua, M. P., Stengelin, S., Gimbrone, M. A. Jr, and Seed B. (1989). Endothelial leukocyte adhesion molecule 1: an inducible receptor for neutrophils related to complement regulatory proteins and lectins. *Science* **243**, 1160-1165.

- Brach, M. A., Gruss, H. J., Kaisho, T., Asano, Y., Hirano, T. and Herrmann, F. (1993). Ionizing radiation induce expression of interleukin 6 by human fibroblasts involving activation of nuclear factor-kappa B. *J. Biol. Chem.* **268**, 8466-8472.
- Brostjan, C., Anrather, J., Csizmadia, V., Natrajan, G., and Winkler, H. (1997). Glucocorticoid inhibit E-selectin expression by targeting NF-kB and not ATF/c-jun. *J. Immunol.* **158**, 3836-3844.
- Carlos, T. and Harlan, J. M. (1994). Leukocyte-endothelial interactions. *Blood* **84**, 17656-1792.
- Cobb, R. R., Felts, K. A., Mckenzie, T. C., Parry, G. and Mackman, N. A (1996). Benxothiophene-carboxamide is a potent inhibitor of IL-1β induced VCAM-1 gene expression in human endothelial cells. *FEBS Lett.* **382**, 323-326.
- Ding, A. H., Nathan, C. F. and Stuehr, D. J. (1988). Release of reactive nitrogen intermediates and reactive oxygen intermediated from mouse peritoneal macrophages. Comparison of activationg cytkines and evidence for independent production. J. Immunol. 141, 2407-2412.
- Eissner, G., Kohlhuber, F., Grell, M., Ueffing, M., Scheurich., P., Hieke, A., Milthoff, G., Bornkamm, G. W. and Holler, E. (1995). Critical involvement of transmembrane tumor necrosis factor-α in endothelial programmed cell death mediated by ionizing radiation and bacterial endotoxin. *Blood* **86**, 4184-4193.
- Formica, J. V. and Regelson, W. (1995). Review of the biology of quercetin and related bioflavonoids. *Food Chem. Toxicol.* **33**, 1061-1080.
- Gaugler, M. H., Squiban, C., Van der Meeren, A., Bertho, J. M., Vandamme, M. and Mouthon, M. A. (1997). Late and persistent up-regulation of intercellular adhesion molecule-1 (ICAM-1) expression by ionizing radiation in human endothelial cells *in vitro*. *Int. J. Radiat. Biol.* **72**. 201-209.
- Gerritsen, M. E., Carley, W. W., Ranges, G. E., Shen, C., Phan, S. A., Ligon, F. G., and Perry, C. A. (1995). Flavonoids inhibit cytokine-induced endothelial cell adhesion protein gene expression. Am. J. Pathol. 147, 278-292.
- Gupta, B. and Ghosh, B. (1999). Curcuma longa inhibits TNF-α induced expression of adhesion molecules on human umbilical vein endothelial cells. Int. J. Immunopharmac. 21, 745-757.
- Hertog, M. G. L., Hollman, P. C. H., Katan, M. B., and Kromhout, D. (1993). Intake of potentially anticarcinogenic flavonoids and their determinants in adults in the Netherlands. *Nutr. Cancer* 20, 21-29.
- Hruza, L. L. and Pentland, A. P. (1993). Mechanisms of UVinduced inflammation. J. Invest. Dermatol. 100, 35S-41S.
- Kawachi, S., Cockrell, A., Laroux, F. S., Gray, L., Granger, D. N., Van Der Heyde, H. C., and Grisham, M. B. (1999). Role of inducible nitric oxide synthase in the regulation of VCAM-1 expression in gut inflammation. *Am. J. Physiol.* 277, G572-G576.
- Moncada, S. (1992). The L-arginine:nitric oxide pathway. *Acta Physiol. Scand.* **145**, 201-227.
- Neuner, P., Klosner, G., Pourmojib, M., Knobler, R., and Schwarz, T. (1997). Pentoxifylline in vivo and in vitro

- down-regulates the expression of the intercellular adhesion molecule-1 in monocytes. *Immunology* **90**, 435-439.
- Redlich, C. A., Rockwell, S., Chung, J. C., Sikora, A. G., Kelley, M., and Mayne, S. T. (1998). Vitamin A inhibits radiation-induced pneumonitis in rats. J. Nutr. 128. 1661-1664.
- Rubin, P., Johnston C. J., Williams, J. P., McDonald, S. and Finkelstein, J. N. (1995). A perpetual cascade of cytokines postirradiation leads to pulmonary fibrosis. *Int. J. Radiat. Biol.* 33, 99-109.
- Sherman, M. L., Datta, R., Hallahan, D. E., Weicheslbaum, R. R. and Kufe, D. W. (1991). Regulation of tumor necrosis factor gene expression by ionizing radiation in human myeloid leukemia cells and peripheral blood monocytes. *J. Clin. Invest.* 81, 1506-1510.
- Spiecker, M., darius, H., kaboth, K., Hubner, F., and Liao, J. K. (1998). Differential regulation of endothelial cell adhesion molecule expression by nitric oxide donors and antioxidants. *J. Leuk. Biol.* 63, 732-739.
- Springer, T. A. (1990). Adhesion receptors of the immune system. *Nature* **346**, 425-434.
- Weber, C. Erl, W., Pietsch, A. and Weber, P. C. (1995). Aspirin inhibits nuclear factor-kB mobilization and monocyte adhesion in stimulated human endothelial cells. *Circulation* 91, 1914-1917.
- Weiser, M. R., Gibbs, S., and Hechtman, H. B. (1997). Strategies to inhibit cellular adhesion molecules: In Paul, L.C., and Issekutz, T. (Eds.) Adhesion molecules in Health and Disease. Marcel Dekker Inc., New York, pp. 55-86.