

## In Vitro Reaction for Calcium Phosphate Ceramics

Koji Ioku, Hiroyuki Toya, Hirotaka Fujimori and Seishi Goto

Department of Advanced Materials and Engineering, Faculty of Engineering, Yamaguchi University, 2-16-1 Tokiwadai, Ube, Yamaguchi 755-8611, Japan

(Received February 24, 2000)

Hydroxyapatite (HA) and  $\beta$ -tricalcium phosphate ( $\beta$ -TCP) are bio-compatible materials with bones and teeth. HA has been widely applied as bone substitutes because of chemical stability *in vivo*, while  $\beta$ -TCP has higher resorbability than HA when the material is implanted in a bone defect. In the present study, both HA and  $\beta$ -TCP porous ceramics were soaked in the simulated body fluid in order to investigate the reaction between the materials and the fluid. After the soaking test, carbonate hydroxyapatite was formed on HA surface at 1 week, and then the amount of precipitates increased with increasing period of the soaking test. While  $\beta$ -TCP was not dissolved in the fluid, carbonate hydroxyapatite was also formed on  $\beta$ -TCP surface after 12 weeks, and the amount of precipitates was less than that on HA. *In vitro* behavior of HA was similar to that *in vivo*, but *in vitro* behavior of  $\beta$ -TCP was not similar to that *in vivo*.

**Key words:** Bioceramics, Hydroxyapatite,  $\beta$ -TCP, Calcium phosphate

### I. Introduction

Hydroxyapatite ( $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$ ; HA) is inorganic principal constituent of hard tissue such as bones or teeth. There are reports that HA is the most bio-compatible materials because of chemical stability *in vivo* and the new bone is formed in direct contact with the HA surface.<sup>1,2)</sup> While  $\beta$ -tricalcium phosphate ( $\beta$ - $\text{Ca}_3(\text{PO}_4)_2$ ;  $\beta$ -TCP) is also the bio-compatible material with bones and teeth, however,  $\beta$ -TCP show the different behavior in comparison with HA *in vivo*. When they are implanted into bones, the resorption of HA is hardly noticed. In the contrast,  $\beta$ -TCP has higher resorbability than HA.<sup>3)</sup> The authors have reported quantitative analysis of the new bone formation for HA and  $\beta$ -TCP in the distal femurs of rabbits and the resorption of HA and  $\beta$ -TCP in the bones and muscles of rabbits.<sup>4-8)</sup> In the present study, in order to appreciate inorganic chemical reaction *in vivo*, HA and  $\beta$ -TCP were analyzed after soaking test in the simulated body fluid with the mineral constituent of human blood plasma without organic matter.

### II. Experimental Method

#### 2.1. Materials

The porous HA used in this study was prepared by sintering at 900°C, provided by Mitsubishi Materials Co., Ltd., Japan. This porous HA is the bone substitute material which has been already used for clinical applications. The Ca/P molar ratio of this HA is 1.67 with stoichiometric composition of HA. The content of impurities is listed in Table 1. The porous  $\beta$ -TCP used in this study was prepared from the HA. In order to obtain fully crystallized pure  $\beta$ -TCP with

the same pore structure as the HA, the porous HA was soaked in diammonium hydrogen phosphate ( $(\text{NH}_4)_2\text{HPO}_4$ ) aqueous solution and then sintered at 900°C for 3 h.<sup>4)</sup> Porous materials of HA and  $\beta$ -TCP had the cylindrical shape of 5 mm in diameter and 5 mm in length. Both materials had the porosity of 60±5%, pore size from 150 to 400  $\mu\text{m}$  with interconnecting pore structure.

#### 2.2. Soaking test with simulated body fluid

The ion concentrations and pH of the simulated body fluid are almost equal to those of the human blood plasma, i. e.,  $\text{Na}^+$  142.0,  $\text{K}^+$  5.0,  $\text{Mg}^{2+}$  1.5,  $\text{Ca}^{2+}$  2.5,  $\text{Cl}^-$  148.8,  $\text{HCO}_3^-$  4.2 and  $\text{HPO}_4^{2-}$  1.0 mM and buffered at pH 7.25 with the trishydroxymethylaminomethane. The solution was prepared according to previous method by Kokubo *et al.*<sup>9)</sup> This solution was used to investigate the chemical reaction between materials and the solution.

The materials were soaked in 100  $\text{cm}^3$  of the simulated body fluid at 37°C for 1-24 weeks. When the soaking test was carried out over 4 weeks, the solution was exchanged every for 4 weeks. After the soaking test, the specimens were taken out from the solutions, washed with very dilute ammonia solution, and dried at 105°C.

#### 2.3. Analysis

The weight of samples was measured before and after

**Table 1.** Impurity Content of the Porous Hydroxyapatite used in the Present Study

(mass %)						
MgO	SiO <sub>2</sub>	Al <sub>2</sub> O <sub>3</sub>	Fe <sub>2</sub> O <sub>3</sub>	SO <sub>3</sub>	Na <sub>2</sub> O	K <sub>2</sub> O
0.270	<0.001	0.020	0.010	0.010	<0.001	<0.001

soaking test. The produced phases were identified by powder X-ray diffractometry with graphite monochromated  $\text{CuK}\alpha$  radiation, operating at 40 kV and 20 mA (XRD; Mac Science MXP<sup>8</sup>, Japan). In order to analyze the deposit on samples, the precise angle of  $2\theta$  and full width of half maximum intensity (FWHM) were measured by accurate XRD measurement with using the internal standard of Si (99.99% pure; Kinzoku Kagaku Co. Ltd., Japan). In addition, the samples after soaking test were subjected to Fourier transform infrared spectroscopy (FT-IR; Perkin-Elmer Spectrum 2000, USA), and thermogravimetry and differential thermal analysis were carried out at a heating rate of  $10^\circ\text{C}/\text{min}$  (TG-DTA; Seiko Instruments TG/DTA32, Japan). The morphology of samples was observed by scanning electron microscopy (SEM; JEOL JSM25S, Japan).

### III. Results and Discussion

#### 3.1. Soaking test with simulated body fluid for HA

Weight change of porous HA after soaking test with the simulated body fluid is shown in Fig. 1. The weight of porous HA increased obviously at 1 week, and then the weight increased linearly with soaked period up to 12 weeks, and slightly increased between 12 and 24 weeks. The weight of the materials was about 15 mass% increase at 24 weeks.

No other phases than hydroxyapatite were revealed by XRD in any products studied. The line broadening was observed for the samples after soaking test over 1 week. Full width at half maximum of intensity for the HA line of  $2\theta=31.8^\circ$  increased with increasing soaking period (Fig. 2). The weight increase mentioned above and the line broadening of XRD mean that the precipitation on the porous HA must be hydroxyapatite with small crystallite size and/or with defective lattice structure. The ions of  $\text{Ca}^{2+}$ ,  $\text{PO}_4^{3-}$  and  $\text{OH}^-$  of the solution are supersaturated with respect to the hydroxyapatite, therefore growth of the hydroxyapatite was achieved on the porous HA.

Fig. 3 shows infrared spectra of HA after soaking test with

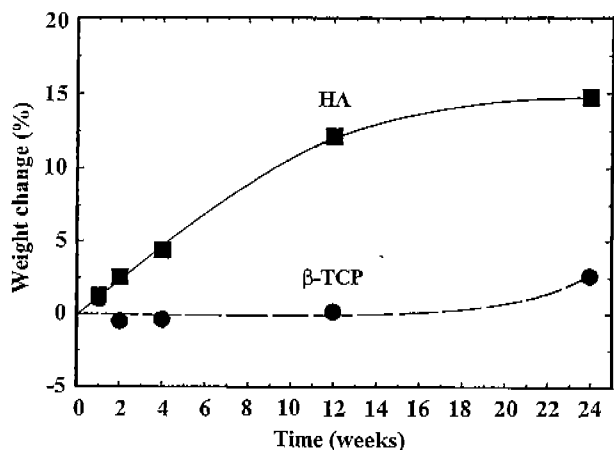


Fig. 1. Weight change of porous HA and porous  $\beta$ -TCP after soaking test with simulated body fluid.

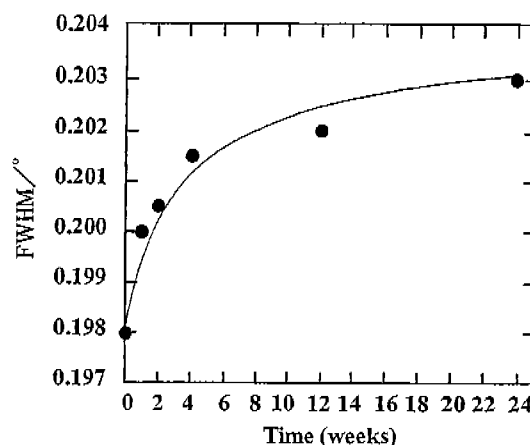


Fig. 2. Full width of half maximum intensity (FWHM) of XRD for HA after soaking test with simulated body fluid.

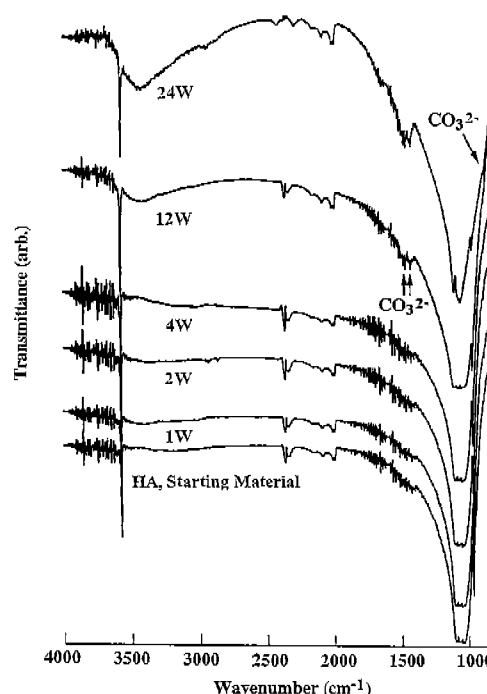


Fig. 3. Infrared spectra of HA after soaking test for 1, 2, 4, 12 and 24 weeks.

the simulated body fluid. The main spectra were assigned as the typical spectra of hydroxyapatite. In the spectra of specimens after soaking at 4 and 12 weeks, carbonate ions in hydroxyapatite were recognized at 1455, 1415 and  $880\text{ cm}^{-1}$ , and they were more clearly observed at 24 weeks. According to the previous investigations for carbonate hydroxyapatite,<sup>4,10</sup> the apatite formed on the starting HA must be  $\text{OH}^-$  and  $\text{PO}_4^{3-}$  carbonated hydroxyapatite. Thus the increase in weight of HA after soaking test was due to the carbonate-containing hydroxyapatite.

Thermogravimetric curves of the porous HA before and after soaking test are shown in Fig. 4. The weight of samples after soaking test decreased more than that before

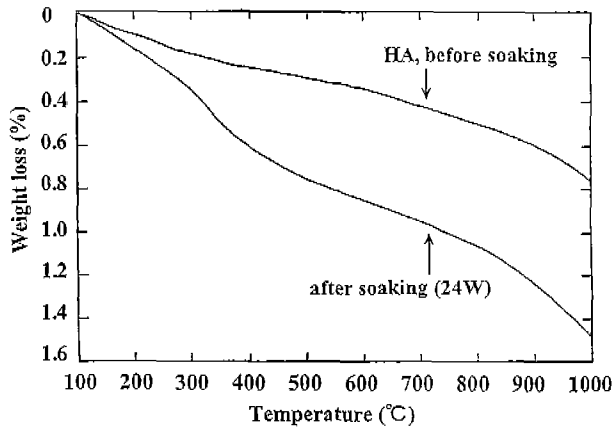


Fig. 4. TG curves of HA before soaking test and after soaking test for 24 weeks.

soaking test. From FT-IR measurement, the spectra of  $\text{CO}_3^{2-}$  in the sample at 24 weeks was disappeared by heating at about  $1000^\circ\text{C}$  (Fig. 5) because of elimination of  $\text{CO}_2$  from carbonated hydroxyapatite. This result was corresponded to the report of Monma *et al.*<sup>11)</sup> According to XRD,  $\beta$ -TCP was recognized from the heated sample after the thermal analysis. It is known that stoichiometric hydroxyapatite with Ca/P molar ratio of 1.67 and  $\beta$ -TCP with Ca/P=1.50 are formed by decomposition of calcium deficient hydroxyapatite with Ca/P molar ratio less than 1.67 with dehydration on heating above  $800^\circ\text{C}$ .<sup>12)-14)</sup> Porous HA of

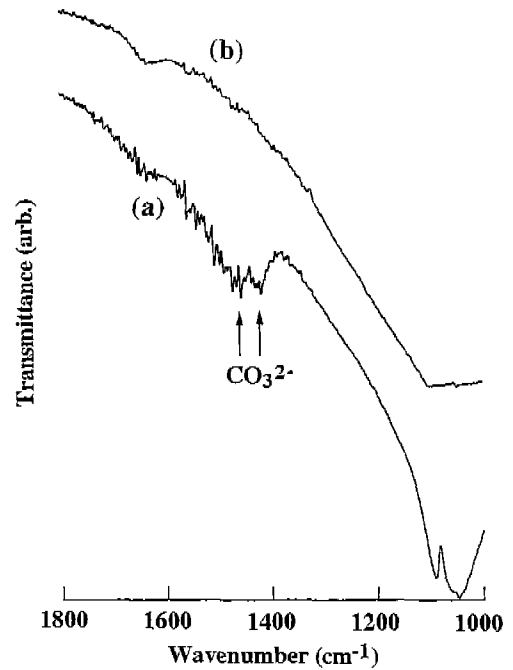


Fig. 5. Infrared spectra of HA; (a) after soaking test for 24 weeks, and (b) after heating of sample (a) at  $1000^\circ\text{C}$  in air.

starting material was stoichiometric hydroxyapatite, so it was thought that precipitates on the surface of porous HA after soaking test were calcium deficient hydroxyapatite. From these results, it was considered that carbonate-con-

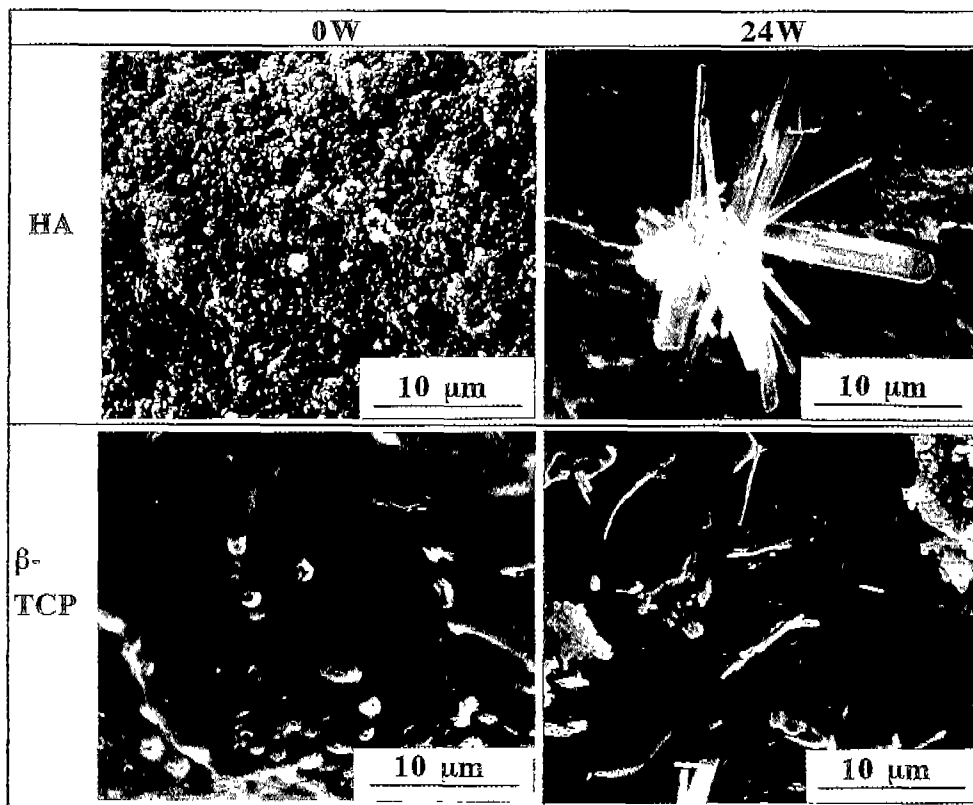


Fig. 6. SEM photographs of porous HA and porous  $\beta$ -TCP before soaking test and after soaking test for 24 weeks.

taining calcium deficient hydroxyapatite was precipitated on porous HA in the simulated body fluid.

After soaking test, formed apatite on the surface of porous HA was observed by SEM. At 2 weeks, rod-like particles were observed on some part of the surfaces and they were more clearly observed at 24 weeks (Fig. 6). The gradual increase of the weight after soaking in the simulated body fluid was due to the carbonate-containing hydroxyapatite precipitation.

When porous HA was implanted in muscles of rabbits, the weight of samples increased because of formation of carbonate-containing hydroxyapatite.<sup>4)</sup> Therefore, the results from the experiment *in vitro* using simulated body fluid was similar to the results from implantation tests in the muscle.

### 3.2. Soaking test with simulated body fluid for $\beta$ -TCP

The weight of porous  $\beta$ -TCP was scarcely changed up to 12 weeks and slightly increased at 24 weeks (Fig. 1). X-ray diffraction patterns were a little affected by soaking in the simulated body fluid (Fig. 7), and a little changes of infrared spectra of the samples after soaking test were observed. The spectrum of hydroxyl stretch was confirmed faintly at about  $3570\text{ cm}^{-1}$ , and the spectrum of carbonate ions was observed slightly in the region of  $1400$  to  $1550\text{ cm}^{-1}$ , because carbonate ions occupied two different sites, that is,  $\text{PO}_4$  site and OH site in hydroxyapatite.<sup>15,16)</sup> It was suggested that carbonate-containing hydroxyapatite was deposited on the surface of porous  $\beta$ -TCP. At 12 weeks, some fine rod-like particles were observed by SEM on the surfaces of porous  $\beta$ -TCP and they were more clearly observed at 24 weeks (Fig. 6). The morphology of particles on  $\beta$ -TCP was extremely similar to the carbonate-containing hydroxyapatite precipitated on HA. Thus these results indicate that the carbonate-containing hydroxyapatite is deposited on the surface of

porous  $\beta$ -TCP, as well as porous HA. Precipitation of carbonate-containing hydroxyapatite on porous  $\beta$ -TCP was later than that on porous HA, because porous HA must be the nuclear for carbonate-containing hydroxyapatite but  $\beta$ -TCP must not be the apatite nuclear.

When porous  $\beta$ -TCP was implanted in muscles of rabbits, resorption of the material observed.<sup>3-5)</sup> In this study, however,  $\beta$ -TCP not dissolved by chemical reaction with the solution. On the contrary, the carbonate-containing hydroxyapatite is deposited on the surface of porous  $\beta$ -TCP, as well as porous HA. From these result, it was thought that  $\beta$ -TCP was bio-degraded by the lowering in pH value by a function of cells such as osteoclast and macrophage *in vivo*.

## IV. Summary

Porous ceramics of HA and  $\beta$ -TCP were soaked in the simulated body fluid with pH 7.25 at  $37^\circ\text{C}$  for 1-24 weeks. On the surface of porous HA, the rod-like carbonate-containing calcium deficient hydroxyapatite precipitated at 1 week and the amount of precipitates increased with increasing soaking period. This result was similar to that from implantation tests in muscles of rabbits. While  $\beta$ -TCP was not dissolved in the solution and the rod-like carbonate-containing hydroxyapatite also precipitated on the surface of porous  $\beta$ -TCP as well as porous HA. The amount of precipitates on porous  $\beta$ -TCP was less than that on porous HA. The soaking test with simulated body fluid for  $\beta$ -TCP was not similar to the results from implantation tests in muscles of rabbits.

## Acknowledgement

This work was supported by a Grant-in-Aid for Encouragement of Young Scientists (No. 09780806), The Ministry of Education, Science and Culture, Japan. The authors also wish to thank Mitsubishi Material Co. Ltd. Japan for the sample preparation.

## References

1. H. Aoki, "Medical Applications of Hydroxyapatite," Ishiyaku EuroAmerica, Tokyo and St. Louis, 1994.
2. T. Iwano, H. Kurosawa, K. Murase, H. Takeuchi and Y. Ohkubo, "Tissue Reaction to Collagen-Coated Porous Hydroxyapatite," *Clin. Orthop.*, (268), 243-252 (1991).
3. P. S. Egli, W. Muller and R. K. Schenk, "Porous Hydroxyapatite and Tricalcium Phosphate Cylinders with Two Different Pore Size Ranges Implanted in the Cancellous Bone of Rabbits," *Clin. Orthop.*, (232), 127-138 (1988).
4. K. Ioku, K. Yanagisawa, N. Yamasaki, H. Kurosawa, K. Shibuya and H. Yokozeki, "Preparation and Characterization of Porous Apatite Ceramics Coated with  $\beta$ -Tricalcium Phosphate," *Bio-Med. Mater. Eng.*, 3(3), 137-145 (1993).
5. H. Yokozeki, H. Kurosawa, K. Shibuya, K. Ioku, T. Hayashi and H. Kawahara, "Reaction of Porous Hydroxyapatite,  $\beta$ -TCP and  $\beta$ -TCP Coated Hydroxyapatite in the Bone." pp.

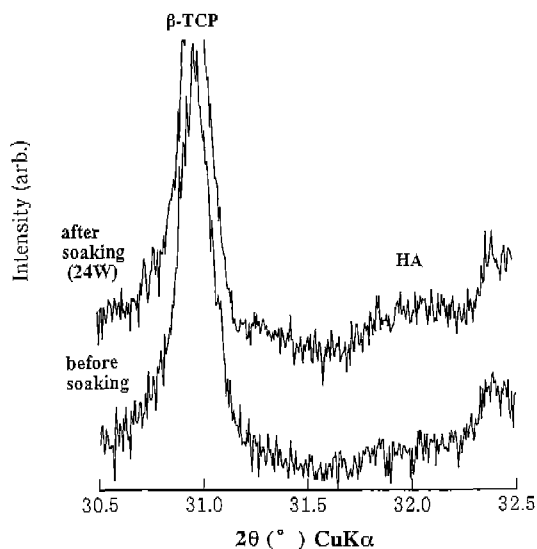


Fig. 7. XRD patterns of  $\beta$ -TCP after soaking test for 24 weeks.

- 172-175 in *Bioceramics* Vol. 7, Ed. by O.H. Andersson, R-P. Happonen, A. Yli-Urpo, Butterworth-Heinemann, Finland, 1994.
6. K. Ioku, S. Goto, H. Kurosawa, K. Shibuya, H. Yokozeki, T. Hayashi and T. Nakagawa, "Reaction of Porous  $\beta$ -TCP with Different Microstructure *in Vivo*," pp. 201-204 in *Bioceramics* Vol. 9, Ed. by T. Kokubo, T. Nakamura, F. Miyaji, Elsevier Science, Japan, 1996.
  7. H. Yokozeki, H. Kurosawa, K. Shibuya, K. Ioku, T. Hayashi and T. Nakagawa, "Reaction of Porous  $\beta$ -TCP with Different Microstructure in the Bone," pp. 101-104 in *Apatite* Vol. 2, Ed. by T. Umegaki, H. Monma, M. Akao, A. Ito, M. Ohgaki, S. Nakamura, Jpn. Assoc. Apatite Sci., Tokyo, 1997.
  8. H. Yokozeki, T. Hayashi, T. Nakagawa, H. Kurosawa, K. Shibuya and K. Ioku, "Influence of Surface Microstructure on the Reaction of the Active Ceramics *in Vivo*," *J. Mater. Sci.: Mater. Med.*, **9**(7), 381-384 (1998).
  9. T. Kokubo, H. Kushitani, S. Sakka, T. Kitsugi and T. Yamamuro, "Solutions Able to Reproduce *in Vivo* Surface-structure Changes in Bioactive Glass-ceramic A-W," *J. Biomed. Mater. Res.*, **24**(6), 721-734 (1990).
  10. M. Sato, S. Ishii and Y. Kato, "On the  $\text{CO}_3$  Site in the Carbonate Apatite," pp. 71-76 in *Apatite* Vol. 1, Ed. by H. Aoki, M. Akao, N. Nagai, T. Tsuji, Jpn. Assoc. Apatite Sci., Tokyo, 1992.
  11. H. Monma and T. Takahashi, "Preparation and Thermal Changes of Carbonate-Containing Apatite," *Gypsum & Lime*, (210), 287-291 (1987).
  12. T. Kanazawa, T. Umegaki and H. Monma, "Apatite, New Inorganic Materials," *Ceramics Japan*, **10**(7), 461-468 (1975).
  13. K. Ioku, T. Murakami, Y. Ikuma and M. Yoshimura, "Preparation of Microstructure-Controlled Porous Hydroxyapatite- $\beta$ -Tricalcium Phosphate Composites by Reaction Sintering," *J. Ceram. Soc. Japan*, **100**(8), 1015-1019 (1992).
  14. K. Ishikawa, P. Ducheyue and S. Radin, "Determination of the Ca/P Ratio in Calcium-deficient Hydroxyapatite Using X-ray Diffraction Analysis," *J. Mater. Sci.: Mater. Med.*, **4**(2), 165-168 (1993).
  15. D. G. A. Nelson and B. E. Williamson, "Low-temperature Laser Raman Spectroscopy of Synthetic Carbonated Apatites and Dental Enamel," *Aust. J. Chem.*, **35**, 715-727 (1982).
  16. I. Rehman and W. Bonefield, "Characterization of Hydroxyapatite and Carbonated Apatite by Photo Acoustic FTIR Spectroscopy," *J. Mater. Sci.: Mater. Med.*, **8**(1), 1-4 (1997).