

## ***In Vitro* Studies on the Release of Intracellular Prolactin from Lymphocytes Using Stress Related Amines and Hormones**

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**ABSTRACT :** Circulating lymphocytes collected from control and heat-stressed buffaloes were subjected to *in vitro* culture with glucocorticoids, epinephrine or serotonin and their effect, if any, on the release of intracellular prolactin (PRL) was studied using ELISA and C-ELISA techniques. It was noted from the study that PRL level was higher in lymphocytes than in plasma of the control and heat-stressed animals, and that the PRL levels increased in the plasma of heat-stressed animals compared to that of non stressed animals with a significant decrease in lymphocytic PRL content by heat stress. Epinephrine and serotonin significantly increased the release of intracellular PRL from the lymphocytes of both in the control and the heat-stressed buffaloes but release of PRL from lymphocyte was not significantly changed by cortisol treatment in both control and heat-stressed buffaloes as compared to epinephrine and serotonin *in vitro*. When lymphocytes were incubated with serotonin, it caused drastic lysis of the lymphocytes but epinephrine and cortisol did not show any lysis. It may be concluded from this study that hormones like epinephrine or serotonin known to increase during stress, release intracellular PRL from lymphocytes, the satellite PRL storage/synthesizing organ of blood, although the mechanism of the release is different. (*Asian-Aus. J. Anim. Sci. 1999. Vol. 12, No. 7 : 1031-1034*)

**Key Words :** Buffalo, Serotonin, Epinephrine, Prolactin, Lymphocytes, *In Vitro*

### INTRODUCTION

Stress is a complex phenomenon which alters physiological, psychological and behavioral responses of animals. It involves areas of brain and endocrine system thus causing threat to homeostasis. Occurrence of diseases due to viral, bacterial or parasitic infection is common during this time. Different types of stress including heat stress have shown to increase plasma levels of prolactin (PRL), epinephrine (E) glucocorticoids, serotonin and  $\beta$  endorphin (Becker, 1987).

Besides pituitary, PRL has also been localized in the lymphocytes (Cevenger et al., 1991) and has been shown to bind and synthesize in these cells (Sharma et al., 1996). PRL helps in synthesis of IL (interleukin)-2 in B and T lymphocytes (Cevenger et al., 1991). IL-2 is both an autocrine/paracrine regulator of normal pituitary cell proliferation and a stimulator of the proliferation of a growth hormone (GH) producing pituitary adenoma cell line GH3 (Artsz, 1993). PRL antibody treatment to lymphocytes has shown to decrease IL-2 synthesis (Besedovsky et al., 1975).

So far no study has been conducted to examine effect of heat stress on the intracellular prolactin content of lymphocytes and the interaction of the stressors (epinephrine, glucocorticoids, serotonin) on intracellular prolactin content of lymphocyte. In the present study intracellular prolactin content of lymphocyte during prolonged heat stress and interaction between stressors viz. cortisol, epinephrine and

serotonin on *in vitro* release of PRL from lymphocytes have been studied.

### MATERIALS AND METHODS

Ten healthy buffaloes were selected for this study and were divided into two groups of five each. One group was kept in the climatic chamber and exposed to hot environmental conditions, i.e. 35-40°C 60% RH during the day; 30°C and 60-70% RH during night for two weeks. Animals were fed with concentrate, hay and water ad libitum. Control group was kept at ambient temperature i.e. 20-30°C (experiments were conducted during the month of December).

#### Preparation of bovine lymphocyte

For lymphocyte isolation, method of Boyum (1968) was followed. In brief blood was collected from all animals in tubes containing anticoagulant. This freshly collected blood was centrifuged at 1,800 rpm (600 g) for 30 minutes at room temperature to obtain a buffy coat. Buffy coat was collected and diluted 1:3 with RPMI-1640 medium (Sigma). It was centrifuged for 3 minutes at 1,500 rpm (400 g). Lymphocyte band was collected and pellet was suspended in Tris-NH<sub>4</sub>Cl (0.1 ml packed cells/ml Tris-NH<sub>4</sub>Cl) and were again centrifuged for 10 min. Cells were then washed two to three times with RPMI-1640 media, counted and put for culture as  $1 \times 10^6$  cells/ml.

#### Experimental design

Plasma collected by centrifugation was analysed by ELISA to estimate PRL levels. After preparation of lymphocytes, some cells were kept separately without

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any treatment for C-ELISA to check the cellular PRL. Rest of prepared lymphocytes from all animals of two groups were suspended in DPBS without serum so that each milliliter contains  $1 \times 10^6$  lymphocytes and used for the treatment with epinephrine, serotonin and cortisol dissolved in DPBS as follows :

Treatment	Dose	Time of incubation with lymphocytes
Epinephrine	10 $\mu$ g/ml	2 h
Serotonin	1 $\mu$ g/ml	2 h
Cortisol	0.16 nmol/ml	2 h
Control	-	-

After two hours of incubation at 37°C with 5% CO<sub>2</sub> in air, culture fluid was collected after centrifugation and the pellet was washed with PBS (0.01 M, pH 7.4). Cells were used for measuring PRL concentration by C-ELISA. Epinephrine, serotonin and cortisol were used as stressors in this study.

The cells ( $1 \times 10^6$ ) and culture fluids (100  $\mu$ l) were then coated in ELISA plate. PRL level was then determined according to Sharma et al. (1996) with an antibody of prolactin raised in the laboratory, antirabbit HRP conjugate purchased from Sigma, USA. Colour was developed by OPD. The reaction was blocked after 30 min. by 5N H<sub>2</sub>SO<sub>4</sub>. The OD was measured by an ELISA reader at 450 nm and data were calculated from the standard curve. The specific binding efficiency of prolactin antibody was found to be 90%. Statistical analysis was done using students' test according to Snedecor and Cochran (1967).

## RESULTS

Prolactin level of lymphocytes collected from the buffaloes which was maintained at ambient temperature (control) was found to be higher compared to the heat-stressed buffalo ( $53.60 \pm 23.13$  vs.  $31.80 \pm 9.34$   $\mu$ g). When freshly isolated lymphocytes from heat-stressed and non-stressed buffaloes were cultured in serum free DPBS for 2 h in presence and absence of epinephrine, serotonin or cortisol, all the three stressor released PRL from the lymphocytes as shown in table 1. PRL in supernatant was significantly higher in all the treatment groups compared to the control group. In the non-stressed animals  $48.20 \pm 8.73$ ,  $51.80 \pm 26.0$  and  $34.20 \pm 2.85$   $\mu$ g of PRL were measured in the supernatant of culture medium for epinephrine, serotonin and cortisol respectively as against  $7.40 \pm 1.24$   $\mu$ g PRL in the medium of cortisol group. Amongst the treatment groups cortisol did release intracellular PRL which is significantly higher than the control group but apparently it was lower than the PRL levels of epinephrine and serotonin group. In the heat-stressed animals intracellular PRL release by epinephrine, serotonin and cortisol was significantly higher than the non-stressed control group. Within the three treatment groups intracellular PRL release was significantly ( $p < 0.05$ ) higher by the epinephrine ( $41.50 \pm 10.3$   $\mu$ g) treatment than serotonin ( $29.00 \pm 1.70$   $\mu$ g) or cortisol ( $31.80 \pm 0.91$   $\mu$ g) treatment. *In vitro* culture of the lymphocytes with all the three stressors showed a decline in the intracellular PRL levels compared to the levels determined before treatment (table 1) and also after the incubation of lymphocytes

Table 1. *In vitro* effect of epinephrine, serotonin and glucocorticoids on intracellular release of PRL from lymphocytes ( $\mu$ g)

Groups	No treatment		Epinephrine		Serotonin		Glucocorticoids	
	Cells	Supernatant	Cells	Supernatant	Cells	Supernatant	Cells	Supernatant
Control	27 $\mu$ g	48 $\mu$ g	08 $\mu$ g	20 $\mu$ g	38 $\mu$ g	03 $\mu$ g	93 $\mu$ g	24 $\mu$ g
	34 $\mu$ g	10 $\mu$ g	10 $\mu$ g	63 $\mu$ g	28 $\mu$ g	30 $\mu$ g	30 $\mu$ g	36 $\mu$ g
	82 $\mu$ g	09 $\mu$ g	18 $\mu$ g	68 $\mu$ g	10 $\mu$ g	72 $\mu$ g	22 $\mu$ g	41 $\mu$ g
	61 $\mu$ g	04 $\mu$ g	18 $\mu$ g	20 $\mu$ g	15 $\mu$ g	28 $\mu$ g	31 $\mu$ g	39 $\mu$ g
	72 $\mu$ g	05 $\mu$ g	23 $\mu$ g	52 $\mu$ g	34 $\mu$ g	34 $\mu$ g	23 $\mu$ g	28 $\mu$ g
Mean $\pm$ SE	61.20 <sup>a</sup>	7.41 <sup>b</sup>	17.80 <sup>b</sup>	48.20 <sup>a</sup>	18.00 <sup>b</sup>	51.80 <sup>a</sup>	26.00 <sup>b</sup>	34.20 <sup>a</sup>
	$\pm 7.26$	$\pm 1.24$	$\pm 8.73$	$\pm 8.73$	$\pm 11.43$	$\pm 26.06$	$\pm 1.87$	$\pm 2.85$
Heat stressed	42 $\mu$ g	4 $\mu$ g	21 $\mu$ g	59 $\mu$ g	20 $\mu$ g	27 $\mu$ g	30 $\mu$ g	32 $\mu$ g
	28 $\mu$ g	3 $\mu$ g	22 $\mu$ g	38 $\mu$ g	13 $\mu$ g	25 $\mu$ g	22 $\mu$ g	30 $\mu$ g
	28 $\mu$ g	5 $\mu$ g	20 $\mu$ g	30 $\mu$ g	34 $\mu$ g	28 $\mu$ g	19 $\mu$ g	32 $\mu$ g
	32 $\mu$ g	7 $\mu$ g	18 $\mu$ g	44 $\mu$ g	20 $\mu$ g	35 $\mu$ g	21 $\mu$ g	35 $\mu$ g
	27 $\mu$ g	6 $\mu$ g	21 $\mu$ g	36 $\mu$ g	31 $\mu$ g	30 $\mu$ g	26 $\mu$ g	30 $\mu$ g
Mean $\pm$ SE	31.40 <sup>a</sup>	5.00 <sup>a</sup>	20.40 <sup>b</sup>	41.50 <sup>c</sup>	23.60 <sup>b</sup>	29.00 <sup>b</sup>	23.60 <sup>b</sup>	31.80 <sup>b</sup>
	$\pm 2.78$	$\pm 0.70$	$\pm 5.57$	$\pm 10.30$	$\pm 3.88$	$\pm 1.70$	$\pm 1.96$	$\pm 0.91$

Values with different superscript are significantly different ( $p < 0.05$ ) from each other.

for the similar time as was given for the treatment groups.

**Table 2.** Prolactin levels in the lymphocytes and plasma of heat-stressed and nonstressed buffaloes

Groups	Lymphocytes	Plasma
Control	30 $\mu$ g	15 $\mu$ g
	70 $\mu$ g	12 $\mu$ g
	90 $\mu$ g	14 $\mu$ g
	46 $\mu$ g	12 $\mu$ g
	32 $\mu$ g	10 $\mu$ g
Mean $\pm$ SE	53.6 <sup>a</sup> $\pm$ 23.13	12.60 <sup>b</sup> $\pm$ 1.74
Heat stressed	30 $\mu$ g	29 $\mu$ g
	24 $\mu$ g	26 $\mu$ g
	26 $\mu$ g	22 $\mu$ g
	29 $\mu$ g	30 $\mu$ g
	50 $\mu$ g	20 $\mu$ g
Mean $\pm$ SE	31.80 <sup>b</sup> $\pm$ 9.34	25.40 <sup>b</sup> $\pm$ 3.87

Values with different superscript are significantly different ( $p < 0.05$ ) from each other.

## DISCUSSION

It is evident from the results that serum of heat-stressed buffaloes were having significantly higher PRL than that of buffaloes maintained in ambient temperature. Similar observation has been made by other workers in the animals during heat stress (Vanjonack and Johnson, 1974; Kaprowski and Tucker, 1973), cold stress (Reid and Mills, 1962) and also psychological stress (Friend and Polan, 1974; Nienabar et al., 1974; Raud et al., 1971). The intracellular PRL level of lymphocytes, collected from heat-stressed buffaloes were significantly lower in comparison to non stressed buffaloes indicating that during heat stress either PRL liberated out of lymphocytes or that it did not bind to lymphocytes as PRL receptors have been localized on lymphocytes (Blalock, 1994). It is also possible that during heat stress the body temperature increases and PRL synthesis from lymphocytes decreases as lymphocytes also synthesise prolactin (Blalock, 1994). To understand the probable cause of increase in PRL level during stress we cultured lymphocytes collected from heat-stressed and non-stressed buffaloes in presence of cortisol, epinephrine and serotonin. It was noted from the results that all these three hormones enhanced PRL release from lymphocytes of both heat-stressed and non-stressed buffaloes showing that cortisol, epinephrine or serotonin that releases during stress (Becker, 1987) released PRL from lymphocytes. thus, if not total, surely partial increase in PRL level of plasma during stress is contributed due to PRL of lymphocytes. The release of PRL was higher in case of

lymphocytes of non-stressed buffaloes than those of heat-stressed buffaloes may be due to the fact that heat-stressed buffaloes continuously release PRL and have adapted to some extent with the changed environment while lymphocytes of non-stressed buffaloes experienced first time the effect of these stressors or that the heat-stressed buffalo lymphocytes contained less PRL than that of non-stressed buffaloes and thus release was comparatively less in lymphocytes heat-stressed buffalo than non-stressed ones.

There was more PRL release when incubated with epinephrine followed by serotonin and cortisol showing that epinephrine acts very quickly than that of cortisol. Epinephrine is also known to act very quickly in other body system and catabolised quickly (Guyton, 1995).

The lower effect of cortisol in case of heat-stressed lymphocytes on PRL release may be due to less receptors of cortisol available in the lymphocytes as Tornello et al. (1982) observed down regulation of corticosterone receptors and no receptor affinity during crowding stress in mice. Koch et al. (1983) also reported decreased glucocorticoids binding in cytosol and nuclear receptor under stress. During the experiment, a large number of cell lysis was noted in the group of lymphocytes cultured with serotonin though no such cell lysis was observed in case of epinephrine or cortisol group. This may be due to the higher dose of serotonin used in this experiment. Serotonin is known to increase maximally in the afternoon as associated with photoperiod (Aleandri et al., 1996). It is not known whether they have any role in elimination of lymphocytes *in situ*.

It may be concluded from the study that cause of hyperprolactinemia during stress if not fully but partially is due to PRL released from the lymphocytes under the influence of epinephrine, glucocorticoids, serotonin, the stressors released during the stress.

## REFERENCES

- Aleandri, V., V. Spina and Alberto Morini. 1996. The pineal gland and reproduction. *Human Repd. Update* 2(3):225-235.
- Artsz, E., R. Buric, G. Stelzer, J. Stalla, J. Sauer, U. Renner and G. K. Stalla. 1993. Interleukin involvement in anterior pituitary cell growth regulation; Effect of IL-2 and IL-6. *Endocrinology* 132:459-467.
- Becker, B. A. 1987. The phenomenon of stress: Concepts and mechanisms associated with stress induced responses of the neuroendocrine system. *Vet. Res. Commu.* 11:443-456.
- Besedovsky, H., E. Sorkin, M. Keller and J. Muller. 1975. Changes in blood hormone levels during the immune response. *Proc. Soc. Expo. Biol. Med.* 150:466-470.
- Blalock, J. E. 1994. The syntax of immune neuroendocrine communication. *Immunology Today* 15(11):504-511.

- Boyum, A. 1968. Isolation of mononuclear cells and granulocytes from human blood. *Scand. J. Clin. Lab. Invest.* 21:77-78.
- Cevenger, C. V., S. W. Altmann and M. B. Prystowsky. 1991. Requirement of nuclear prolactin for interleukin-2 stimulated proliferation of T lymphocytes. *Science.* 25: 77-79.
- Friends, T. H. and C. E. Polan. 1974. Social rank, feeding behaviour, and free stall utilization by dairy cattle. *J. Dairy Sci.* 57:1214.
- Kaprowski, J. A. and H. A. Tucker. 1973. Bovine serum growth hormones, corticoids and insulin during lactation. *Endocrinology.* 93:645.
- Nienaber, J. A., C. B. Gilbertson, T. J. Klopfenstein, S. D. Farlin and T. M. Mc Calla. 1994. Performance and lot surface conditions as affected by feedlot slope and animal density. pp. 324 in *Livestock Environment. Proceedings of the international Livestock Environment Symposium.*
- Raud, H. R., C. A. Kiddy and W. O. Odell. 1971. The effect of stress upon the determination of serum prolactin by radioimmunoassay. *Proc. Soc. Exp. Biol. Med.* 136:689.
- Reid, R. L. and J. C. Mill. 1962. Studies on the carbohydrate metabolism of sheep. x iv. The adrenal response of the climatic stresses of cold, wind and rain. *Aus. J. Agr. Res.* 13:296.
- Snedecor, G. W. and W. G. Cochran. 1967. *Statistical Methods*, 6th edn. Iowa State Univ. Press, Ames. I O.
- Vanjonack, W. J. and H. D. Johnson. 1974. Effect of sulphur dioxide and heat stressors on plasma glucocorticoids and thyroxine levels in mice. *Int. J. Biometeorol.* 18:301.
- Sharma, G. T., L. K. Gupta and A. C. Majumdar. 1996. Prolactin estimation in lymphocytes and serum using ELISA and C-ELISA techniques and its localization in WBCs by immunohistochemistry. *I. J. Phy & Allied Sci.* 50:98-102.