A New Cytotoxic Acyclic Diterpene from Carpesium divaricatum

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A new acyclic diterpene (1) and a known acyclic diterpene, 12(*S*)-hydroxygeranylgeraniol (2) were isolated from the aerial parts of *Carpesium divaricatum*. The structure of 1 was determined to be (2*E*,10*E*)-1,12-dihydroxy-18-acetoxy-3,7,15-trimethylhexadeca-2,10,14-triene (1) on the basis of spectroscopic studies. Compounds 1 and 2 exhibited cytotoxicity against cultured human tumor cell lines, A549, SK-OV-3, SK-MEL-2, XF498, and HCT15, with ED₅₀ values ranging from 4.3-10.2 µg/ml and 4.1-8.3 µg/ml, respectively.

Key words: Carpesium divaricatum, Acyclic diterpene, Cytotoxicity

INTRODUCTION

In our previous phytochemical studies of *Carpesium divaricatum S. et Z.* (Compositae), we reported four new cytotoxic sesquiterpene lactones (Kim *et al.*, 1997b) and two new and two known sesquiterpene lactones (Kim *et al.*, 1997a). Our continuing investigation on the constituents of this plant afforded two cytotoxic acyclic diterpenes, a new diterpene derivative (compound 1) and a known diterpene, 12(S)-hydroxy geranylgeraniol (2). This paper reports the isolation and structural elucidation of compound 1. Further, the cytotoxicity of compounds 1 and 2 was tested using cultured human tumor cell lines, A549, SK-OV-3, SK-MEL-2, XF498 and HCT15.

MATERIALS AND METHODS

General experimental procedures

Melting points were measured on a Gallenkamp melting point apparatus (uncorr.). ¹H- and ¹³C-NMR were recorded on a Bruker AMX-500 spectrometer. UV spectra were obtained on a Shimadzu UV₂₄₀ UV-Visible recording spectrophotometer. IR spectra were measured on Shimadzu IR-435 Infrared spectrophotometer. The EI-MS was measured on VG70-VSEQ and CI-MS was recorded with isobutane as reactant gas on Finnìgan/MAT HSQ/30 mass spectrometer.

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Plant material

Carpesium divaricatum was collected in August 1994 on Samyeong Mt., Kangwondo, Korea. A voucher specimen is deposited in the herbarium of College of Pharmacy, Sung Kyun Kwan University (SKKU-94-005).

Biological assay

Cytotoxicity of compounds **1** and **2** was measured against A549 (non small cell lung adenocarcinoma), SK-OV-3 (ovarian), SK-MEL-2 (skin melanoma), XF498 (CNS) and HCT15 (colon) cell lines *in vitro* using sulforhodamine B (SRB) assay (Giard *et al.*, 1973).

Extraction and isolation

The air-dried plant materials (3.5 kg) were finely ground and extracted at room temperature with MeOH for 2 weeks (3×2 L). The resultant MeOH extract (110 g) was suspended in H₂O (1 L), and partitioned with CH₂Cl₂ $(2\times1~L)$ and *n*-BuOH $(2\times1~L)$, successively, to give CH₂Cl₂ (30 g), n-BuOH (22 g) and water (50 g) soluble fractions. The CH₂Cl₂ soluble fraction was subjected to column chromatography over silica gel (800 g), eluted sequentially with *n*-hexane-EtOAc (5:1, 800 ml), n-hexane-EtOAc (3:1, 650 ml), n-hexane-EtOAc (1:1, 600 ml), and EtOAc (500 ml). The eluents were fractionated by TLC to yield fractions designated as MC1-MC5: void volume (450 ml), MC1 (520 ml), MC2 (350 ml), MC3 (410 ml), MC4 (380 ml), and MC5 (420 ml). The eluent (3.5 g) obtained from fraction MC4 was further chromatographed with silica gel (220 g) eluting with *n*-hexane-CH₂Cl₂-EtOAc (1:1:1) to give four subfractions (a-d). Subfraction c (1.6 g) was purified

Table I. ¹³C-(125 MHz) and ¹H-NMR (500 MHz) data for compound 1 (in CDCl.)

Position		DEPT	$\delta^{1}H^{a}$	¹H-¹H COSY	HMBC ^b
1	59.3	CH ₂	4.14 (d, 6.0)	2	C-2, 3
2	124.2	CH	5.41 (t, 6.0)	1	
3	138.5	C			
4	39.4	CH_2	obs*		
5	25.7	CH_2	obs		
6	32.4	CH_2	obs		
7	38.1	CH	1.55 (m)	19	
8	31.6	CH_2	obs		
9	25.6	CH_2	obs	10	
10	130.3	CH	5.40 (t, 6.0)	9	
11	135.2	C			
12	75.5	CH	3.39 (m)	13	
13	35.5	CH_2	obs	12, 14	
14	120.6	CH	5.17 (m)	16, 17	
15	134.1	C			
16	25.9	CH_3	1.74 (s)	14	C-14, 15, 17
17	18.0	CH_3	1.65 (s)	14	C-14, 15, 16
18	61.9	CH_2	4.58 (s)		C-10, 12, 1 ¹
19	15.5	CH_3	0.91 (d, 7.0)	7	C-6, 7, 8
20	16.2	CH_3	1.67 (s)		C-2, 3, 4
11	171.1	C			
2'	21.0	CH ₃	2.07 (s)		

^aChemical shifts (ppm), multiplicity and coupling constants (Hz in parentheses).

on Lobar[®]-A column (Merck) (CH₂Cl₂-EtOAc, 8:1) followed by reverse phase Lobar[®]-A column (MeOH-H₂O, 8:2) to yield **1** (30 mg) and **2** (20 mg), respectively.

(2*E*,10*E*)-1,12-dihydroxy-18-acetoxy-3,7,15-trimethylhexadeca-2,10,14-triene (1): an oil; $[α]_D^{2^4}$: +2.4 (MeOH, c 1.0); UV $λ_{max}^{MeOH}$ nm: 216; IR v_{max}^{Nujol} cm⁻¹: 3408 (OH), 1738 (C=O); El-MS m/z (rel. int. %): 348 (M⁺-H₂O, 2), 306 (4), 289 (24), 219 (77), 109 (98), 83 (100), 54 (97); Cl-MS (isobutane) m/z (rel. int. %): 367 (M⁺+1, 15), 289 (100); ¹H- and ¹³C-NMR data are presented in Table I.

RESULTS AND DISCUSSION

Compound **1** was obtained as an oil. Its IR spectrum showed hydroxyl and carbonyl absorption bands at 3410 cm⁻¹ and 1740 cm⁻¹, respectively. The highest mass spectral fragment of compound **1** in EI-MS appeared at m/z 348, corresponding to [M⁺-H₂O]. The molecular ion as M⁺+1 was confirmed by CI-MS (isobutane). From the DEPT (CH₃×5, CH₂×8, CH×5, C×4), ¹H-NMR, ¹³C-NMR (22C), and CI-MS (M⁺+1, 367) spectra, the molecular formula was deduced to be $C_{22}H_{38}O_4$. The formula showed four unsaturated-bond equivalents in the molecule. The ¹³C-NMR spectrum showed six olefinic carbons at δ 138.8, 135.2, 134.1, 130.3, 124.2, and 120.6 and a carbonyl carbon

Fig. 1. Structures of compounds 1-2

at 8171.1, thus accounting for four unsaturated-bond equivalents. Therefore, 1 was suggestive of an acyclic compound. The ¹H-NMR spectrum exhibited three olefinic proton signals at δ 5.41, 5.40, and 5.17, and three protons attached to the oxygenated carbons at $\delta4.58$, 4.14, and 3.39. Compound 1 also showed four singlet methyls at δ 2.07, 1.74, 1.67, and 1.65, and a doublet methyl δ 0.91 in the ¹H-NMR spectrum. These data suggested an acyclic diterpene skeleton for 1. The ¹H-NMR spectrum of 1 was very similar to that of compound 2 which was previously reported as an acyclic diterpene (Robert et al., 1986), apart from the replacement of a methine olefinic signal at δ 5.10 by signal at δ 1.55, attributable to the saturated aliphatic methine group, and the presence of an acetyl group at δ 2.07. The downfield shift of C-18 (δ 4.58) suggested that C-18 was possibly carrying an acetoxy group. The above evidence, together with the ¹H-¹H COSY, DEPT and HMQC spectra, led to the gross structure of 1. The position of the acetyl group was further confirmed by ${}^{1}H^{-13}C$ long-range correlation between δ 4.58 proton signal and δ 171.1 carbon signal in the HMBC spectrum. The trans (E) configurations of the double bonds at C-2 and C-10 were established on the basis of the chemical shifts of the vinyl methyl groups in the ¹³C-NMR spectrum (Breitmaier et al., 1990) and by comparison of its spectral data with those of compound 2 (Robert et al., 1986). Finally, compound 1 was characterized as (2E,10E)-1,12-dihydroxy-18acetoxy-3,7,15-trimethyl-hexadeca-2,10,14-triene (18acetoxy-12-hydroxy-6,7-dihydrogeranyl geraniol). The compound 1 is optically active ($[\alpha]_D^{24}+2.4$), but the configurations at C-7 and C-12 remain uncertain.

The structure of an acyclic known diterpene, 12(*S*)-hydroxy geranylgeraniol (**2**) (Robert *et al.*, 1986) was established by comparing its UV, IR, NMR spectral data and physical properties with those reported in the literature.

Acyclic diterpenes 1 and 2 are cytotoxic in vitro in

^bMajor ¹H-¹³C long range correlations determined from HMBC experiment.

^{*}Obscure

Table 2. Cytotoxicity of compounds 1 and 2*

cancer cell lines	A-549	SK-OV-3	SK-MEL-2	XF-498	HCT-15
compound 1	9.74	10.20	6.57	7.04	4.38
compound 2	8.20	8.33	7.16	8.26	4.17

^{*}ED₅₀ value of compounds against each cancer cell line, which was defined as a concentration ($\mu g/ml$) that caused 50% inhibition of cell growth *in vitro*.

cultured human tumor cellular models, A549, SK-OV-3, SK-MEL-2, XF498, and HCT15, as determined by sulforhodamine B (SRB) assay (Giard *et al.*, 1973). The cytoxicity of these compounds is shown as ED_{50} values in Table II.

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