

## Cyclooxygenase-2 Inhibitor from *Evodia rutaecarpa*

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**Abstract** – By bioassay guided fractionation followed by chromatographic separation of the MeOH extract from the fruit of *Evodia rutaecarpa* (Juss.) Benth. (Rutaceae), a new cyclooxygenase-2 inhibitor was isolated and identified as an alkaloid, rutaecarpine. Other alkaloids such as evodiamine and dehydroevodiamine together with limonoids were also isolated and characterized.

**Key words** – *Evodia rutaecarpa*, Rutaceae, alkaloid, rutaecarpine, cyclooxygenase-2 inhibitory activity.

### Introduction

The *Evodia* fruit has been used in traditional Chinese prescriptions and there were many previous studies on the *Evodia* fruit (Tang and Eisenbrand, 1992). A number of indolopyridoquinazoline alkaloids such as evodiamine, rutaecarpine and dehydroevodiamine have been isolated from the fruits of *E. rutaecarpa* (Tang *et al.*, 1997; Shoji *et al.*, 1988; Danieli *et al.*, 1979). In addition, some quinolone alkaloids (Shin *et al.*, 1998; Kamikado *et al.*, 1976; Sugimoto *et al.*, 1988a), other nitrogen-containing compounds (Shoji *et al.*, 1988; Takagi *et al.*, 1979), limonoids (Sugimoto *et al.*, 1988a; 1988b) and flavonoids (Kang *et al.*, 1997) were also isolated. And some of them were reported to possess a multifaceted biological/pharmacological activities *in vitro* and *in vivo* (Chiou *et al.*, 1997; Haji *et al.*, 1994; Itokawa *et al.*, 1990; Kano *et al.*, 1991; Kim *et al.*, 1998; King *et al.*, 1980; Matsuda *et al.*, 1997; 1998a,b; Shoji *et al.*, 1986; Yang *et al.*, 1990; Yamahara *et al.*, 1989). Recently we also reported that dehydroevodiamine showed anticholinesterase and antiemetic activities (Park *et al.*, 1996). During the screening programme of Korean medicinal plants for anti-inflammatory activity we have found considerable inhibitory activity of cyclooxygenase-2 (COX-2) in MeOH extract of *E. rutaecarpa* (Moon *et al.*, 1998). From this extract, bioassay guided fractionation followed by chromatographic separation of the extract led to the isolation of

a new cyclooxygenase-2 inhibitor, rutaecarpine.

This paper describes the isolation and structure elucidation of the alkaloids and limonoids from *E. rutaecarpa* and the inhibitory activity of cyclooxygenase-2 by rutaecarpine.

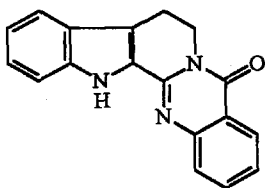
### Experimental

**General experimental procedures and Plant material** – General experimental procedures and plant materials were described in a previous paper of Kang *et al.*(1997).

**Extraction and isolation** – The fruits of *E. rutaecarpa* (10 kg) were extracted three times with 80% MeOH under reflux. The MeOH extract was evaporated under reduced pressure to dryness. The dried extract was partitioned between H<sub>2</sub>O and CH<sub>2</sub>Cl<sub>2</sub> and gave 240 g of the CH<sub>2</sub>Cl<sub>2</sub> extract. The CH<sub>2</sub>Cl<sub>2</sub> fraction was subjected to silica gel (Merck, No. 7734, 3 kg) column chromatography eluting with CHCl<sub>3</sub>, CHCl<sub>3</sub>-acetone (30 : 1, 20 : 1, 10 : 1) and then CHCl<sub>3</sub>-MeOH (10 : 1, 1 : 1) to give 20 subfractions. Subfraction 6 was recrystallized from MeOH to yield compound **1** (0.9 g) as pale yellowish needles. Subfractions 9 and 17 were treated with the same manner as subfraction 6 to give compound **2** (1.2 g) and **3** (0.63 g) as pale yellowish plate and as pale yellowish powder, respectively. Subfraction 11 was rechromatographed on silica gel (Merck, No. 7729) with hexane-EtOAc (5:8) to afford compound **4** (30 mg) as an amorphous powder. Subfraction 12 was recrystallized from MeOH to give compound **5** (150 mg) as needles.

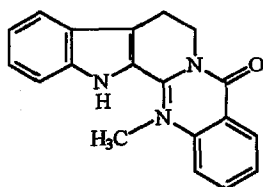
\*Author for correspondence.

**Compound 1** – mp 261~262°; UV,  $\lambda_{\max}$  (MeOH) nm 240 (sh), 288, 330, 343, 360; IR,  $\nu_{\max}$  (KBr) 3345, 1655, 1599, 1471, 1327, 1229, 731  $\text{cm}^{-1}$ ; MS,  $m/z$  (rel. int.) 287  $[\text{M}]^+(100)$ , 259  $[\text{M}-\text{CO}]^+(13.3)$ , 258  $[\text{M}-\text{CHO}]^+(14.8)$ , 144 (25.1), 141 (10.2), 130 (26.5), 77 (23.0);  $^1\text{H-NMR}$  (300 MHz,  $\text{DMSO-d}_6$ )  $\delta$ : 3.17 (2H, t,  $J=6.9\text{Hz}$ , H-6), 4.45 (2H, t,  $J=6.9\text{Hz}$ , H-5), 7.09 (1H, brt,  $J=7.7\text{Hz}$ , H-10), 7.26 (1H, dt,  $J=0.9$ , 7.0 Hz, H-11), 7.46 (1H, dt,  $J=0.9$ , 7.5Hz, H-12), 7.49 (1H, brd,  $J=7.9$  Hz, H-18), 7.64 (1H, brd,  $J=7.9$  Hz, H-9), 7.68 (1H, brd,  $J=7.9\text{Hz}$ , H-16), 7.80 (1H, dt,  $J=1.3$ , 7.9 Hz, H-17), 8.16 (1H, dd,  $J=1.3$ , 7.9Hz, H-19), 11.83 (1H, s, NH);  $^{13}\text{C-NMR}$  (75.5MHz,  $\text{DMSO-d}_6$ )  $\delta$ : 18.88, 40.76, 112.51, 117.80, 119.67, 119.86, 120.66, 124.66, 124.85, 125.89, 126.38, 126.52, 127.04, 134.32, 138.63, 145.22, 147.32, 160.54.



Rutaeacarpine (1)

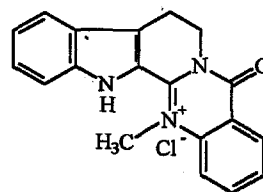
**Compound 2** – mp 274 ~8°; UV,  $\lambda_{\max}$  (MeOH) nm 225, 268, 282, 291; IR,  $\nu_{\max}$  (KBr) 3234, 1630, 1607, 1512, 1310, 1281, 1233, 1167, 747, 725 $\text{cm}^{-1}$ ; MS,  $m/z$  (rel. int.) 303  $[\text{M}]^+(35.9)$ , 302  $[\text{M}-\text{H}]^+(25.1)$ , 288  $[\text{M}-\text{CH}_3]^+(8.8)$ , 274  $[\text{M}-\text{CHO}]^+(19.2)$ , 170 (30.9), 169 (100), 134 (81.8), 133 (13.1);  $^1\text{H-NMR}$  (300 MHz,  $\text{DMSO-d}_6$ )  $\delta$ : 2.80 (1H, dd,  $J=5.1$ , 15.5Hz, H-6a), 2.88 (3H, s,  $\text{CH}_3$ ), 2.90 (1H, ddd,  $J=1.6$ , 5.1, 15.5Hz, H-6b), 3.20 (1H, ddd,  $J=1.6$ , 5.5, 13.0Hz, H-5a), 4.65 (1H, ddd,  $J=1.6$ , 5.5, 13.0Hz, H-5b), 6.11 (1H, s, H-3), 6.97 (1H, dt,  $J=1.3$ , 7.8Hz, H-18), 7.01 (1H, brt,  $J=7.8\text{Hz}$ , H-11), 7.04 (1H, brt,  $J=8.1\text{Hz}$ , H-12), 7.10 (1H, dd,  $J=1.1$ , 8.1Hz, H-10), 7.37 (1H, brd,  $J=8.1\text{Hz}$ , H-9), 7.46 (1H, dd,  $J=1.7$ , 8.0Hz, H-16), 7.48 (1H, dd,  $J=1.8$ , 7.8Hz, H-17), 7.82 (1H, dd,  $J=1.5$ , 7.8Hz, H-19), 11.00 (1H, s, NH);  $^{13}\text{C-NMR}$  (75.5MHz,  $\text{DMSO-d}_6$ )  $\delta$ : 19.43 ( $\text{CH}_2$ ), 36.37 ( $\text{CH}_3$ ),



Evodiamine (2)

40.77 ( $\text{CH}_2$ ), 69.68 (CH), 111.45 (C), 111.58 (CH), 117.42 (CH), 118.13 (CH), 118.82 (CH), 119.26 (C), 120.21 (CH), 121.78 (CH), 125.90 (C), 127.92 (CH), 130.50 (C), 133.33 (CH), 136.44 (C), 148.74 (C), 164.14 (C).

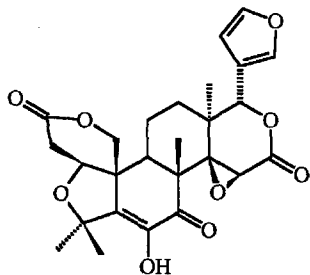
**Compound 3** – mp 249~251°; UV,  $\lambda_{\max}$  (MeOH) nm (log  $\epsilon$ ) 246 (4.08), 305 (sh, 3.64), 314 (3.71), 365 (4.36); IR,  $\nu_{\max}$  (KBr) 3437, 3235, 1709, 1611, 1557, 1501, 1350, 1339, 1285, 1219, 1103, 766, 721, 687  $\text{cm}^{-1}$ ; MS,  $m/z$  (rel. int.) 302  $[\text{M}-\text{Cl}]^+(100)$ , 287  $[\text{302}-\text{CH}_3]^+(83.6)$ , 273  $[\text{302}-\text{CHO}]^+(43.7)$ , 258 (23.8), 246 (35.3), 156 (9.7), 77 (45.2);  $^1\text{H-NMR}$  (300MHz,  $\text{DMSO-d}_6$ )  $\delta$ : 3.35 (2H, t,  $J=6.9\text{Hz}$ , H-6), 4.47 (2H, t,  $J=6.9\text{Hz}$ , H-5), 4.37 (3H, s,  $\text{CH}_3$ ), 7.29 (1H, brt,  $J=7.6\text{Hz}$ , H-10), 7.54 (1H, brt,  $J=7.6\text{Hz}$ , H-11), 7.69 (1H, brd,  $J=8.6\text{Hz}$ , H-12), 7.81 (1H, dt,  $J=1.2$ , 8.5Hz, H-18), 7.89 (1H, brt,  $J=8.2\text{Hz}$ , H-9), 8.14 (1H, dt,  $J=1.1$ , 8.6Hz, H-17), 8.20 (1H, brd,  $J=8.3\text{Hz}$ , H-16), 8.36 (1H, brd,  $J=7.8\text{Hz}$ , H-19), 12.33 (1H, s, NH);  $^{13}\text{C-NMR}$  (75.5MHz,  $\text{DMSO-d}_6$ )  $\delta$ : 18.44 (C-6), 40.64 ( $\text{CH}_3$ ), 42.02 (C-5), 113.37 (C-12), 118.44 (C-16), 118.65 (C-20), 120.05 (C-2), 121.50 (C-10), 121.56 (C-9), 123.26 (C-8), 127.60 (C-19), 128.55 (C-18), 128.75 (C-11), 130.38 (C-7), 136.57 (C-17), 139.62 (C-15), 141.30 (C-13), 149.94 (C-3), 158.09 (CO).



Dehydroevodiamine (3)

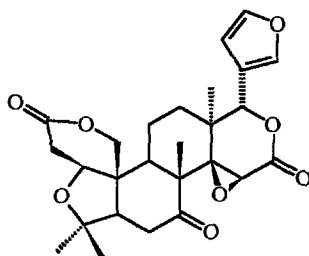
**Compound 4** – UV,  $\lambda_{\max}$  (MeOH) nm (log  $\epsilon$ ) 276 (3.59); IR,  $\nu_{\max}$  (KBr) 3437, 1748, 1686, 1655, 1358, 1287, 1032, 914, 876  $\text{cm}^{-1}$ ; MS,  $m/z$  484  $[\text{M}]^+$ ;  $^1\text{H-NMR}$  (300MHz,  $\text{CDCl}_3$ )  $\delta$ : 1.04 (3H, s, H-18), 1.15 (3H, s, H-30), 1.49 (3H, s, H-28), 1.54 (3H, s, H-29), 2.67 (1H, dd,  $J=1.7$ , 12.9Hz, H-9), 2.83 (1H, dd,  $J=4.4$ , 18.1Hz, H-2a), 2.97 (1H, dd,  $J=1.6$ , 18.1Hz, H-2b), 4.07 (1H, brt,  $J=3.2\text{Hz}$ , H-1), 4.12 (1H, s, H-15), 4.63 (2H, brs, H-19), 5.43 (1H, s, H-17), 6.33 (1H, d,  $J=1.6\text{Hz}$ , H-22), 7.33 (1H, d,  $J=3.0\text{Hz}$ , H-23), 7.34 (1H, d,  $J=3.0\text{Hz}$ , H-21);  $^{13}\text{C-NMR}$  (75.5MHz,  $\text{CDCl}_3$ )  $\delta$ : 79.12 (C-1), 34.77 (C-2), 169.07 (C-3), 81.79 (C-4), 139.45 (C-5), 140.68 (C-6), 195.19 (C-7), 48.34 (C-8), 46.31 (C-9), 46.82 (C-10), 20.41 (C-11), 31.63 (C-12), 37.33 (C-13), 65.25 (C-14), 52.11 (C-15), 166.43 (C-16), 77.68 (C-17), 68.55 (C-19),

119.73 (C-20), 141.06 (C-21), 109.60 (C-22), 143.29 (C-23), 18.10, 20.18, 25.19, 25.63 (CH<sub>3</sub>).



Evodol (4)

**Compound 5** – mp 255–9°; UV,  $\lambda_{\max}$  (MeOH) nm (log  $\epsilon$ ) 206 (4.09), 2.37 (sh, 3.33), 334 (2.78); IR,  $\nu_{\max}$  (KBr) 3437, 1757, 1709, 1287, 1263, 1165, 1030, 916, 876, 762, 602 cm<sup>-1</sup>; MS,  $m/z$  470 [M]<sup>+</sup>; <sup>1</sup>H-NMR (300MHz, CDCl<sub>3</sub>)  $\delta$ : 1.08 (3H, s, 30-CH<sub>3</sub>), 1.18 (6H, s, 18, 29-CH<sub>3</sub>), 1.29 (3H, s, 28-CH<sub>3</sub>), 2.24 (1H, dd,  $J=3.3, 15.9$ Hz, H-5), 2.46 (1H, dd,  $J=3.2, 14.4$ Hz, H-6), 2.69 (1H, dd,  $J=1.7, 16.6$ Hz, H-2), 2.87 (1H, t,  $J=15.1$ Hz, H-6), 2.97 (1H, dd,  $J=3.6, 16.6$ Hz, H-2), 4.05 (2H, s, H-1, 15), 4.47 (1H, d,  $J=13.2$ Hz, H-19), 4.77 (1H, d,  $J=13.2$ Hz, H-19), 5.47 (1H, s, H-17), 6.35 (1H, brs, H-22), 7.40 (1H, d,  $J=2.6$ Hz, H-23), 7.41 (1H, d,  $J=2.6$ Hz, H-21); <sup>13</sup>C-NMR (75.5MHz, CDCl<sub>3</sub>)  $\delta$ : 79.11 (C-1), 35.63 (C-2), 169.20 (C-3), 80.29 (C-4), 60.45 (C-5), 36.36 (C-6), 206.16 (C-7), 51.29 (C-8), 48.06 (C-9), 45.91 (C-10), 18.86 (C-11), 30.74 (C-12), 37.95 (C-13), 65.73 (C-14), 53.85 (C-15), 166.68 (C-16), 77.81 (C-17), 65.34 (C-19), 119.97 (C-20), 141.10 (C-21), 109.66 (C-22), 143.20 (C-23), 17.60, 20.63, 21.35, 30.12 (CH<sub>3</sub>).



Limonin (5)

**COX-2 inhibition test** – The inhibitory activity of the isolated compounds on COX-2 was measured using aspirin-treated mouse bone marrow derived mast cells (BMMC) as described previously (Moon *et al.*, 1998; Son *et al.*, 1998). In brief, BMMC from male BALB/cJ mice were cultured for up to 10 weeks in 50% enrichment

medium (RPMI 1640 containing antibiotics, 2 mM L-glutamine, 0.1 mM nonessential amino acid and 10% FBS) and 50% WEHI-3 cell conditioned medium as a source of IL-3. After 3 weeks, BMMC were suspended in enrichment medium and preincubated with 10 /ml aspirin for 2 hr in order to inactivate preexisting COX-1. The cells were activated with KL (100 ng/ml), IL-10 (100 U/ml) and LPS (100 ng/ml) in the presence/absence of plant extract or the isolated compounds for 8 hr. Media were collected and PGD<sub>2</sub> concentrations were measured using a PGD<sub>2</sub> assay kit (Amersham, Buckinghamshire, UK).

## Results and Discussion

Since methanol extract of *E. rutaecarpa* showed strong inhibitory activity of PGD<sub>2</sub> formation (Table 1), further fractionation and chromatographic separation were carried out leading to 3 alkaloids and 2 limonoids. These compounds were identified as rutaecarpine (1), evodiamine (2), dehydroevodiamine (3), evodol (4) and limonin (5). The spectroscopic measurements of all isolates were consistent with the data obtained in literatures (Haji *et al.*, 1994; Itokawa *et al.*, 1990; Sugimoto *et al.*, 1988a). Although the spectroscopic data of these alkaloids from this plant were well documented, the <sup>1</sup>H signals of these compounds were not fully assigned. The <sup>1</sup>H-<sup>1</sup>H COSY experiments allowed unambiguous assignment of all proton signals for 1, 2 and 3 as indicated in Experimental. Among these isolates, rutaecarpine showed strong COX-2 inhibitory activity with an IC<sub>50</sub> value of 80 ng/ml, while two other alkaloids also possessed inhibitory activity of PGD<sub>2</sub> formation by COX-2 (Table 1). The alkaloids from *E. rutaecarpa* showed a wide variety of biological properties. Recent investigations have found that some alkaloids from *E. rutaecarpa* showed inhibition of nitric oxide production

**Table 1.** Inhibition of PGD<sub>2</sub> formation of the activated BMMC by the extract and isolates from *E. rutaecarpa*

Extract/isolates	% Inhibition of PGD <sub>2</sub> formation
MeOH extract	64.5
CH <sub>2</sub> Cl <sub>2</sub> fraction	76.2
Evodiamine (2)	66.3
Evodol (4)	28.7
* Rutaecarpine (1)	100.0

All compounds and the extract were tested at 2.5  $\mu$ g/ml.

and antinociceptive actions (Chiou *et al.*, 1997; Matsuda *et al.*, 1998a). These biological activities along with our findings from this study may contribute to the anti-inflammatory activity of *E. rutaecarpa* used in Chinese medicine. The results of further biological evaluation including *in vivo* study of rutaecarpine will be the subject of a separate report.

### Acknowledgments

This work was supported by New Drug Development Program of the Korea Ministry of Health and Social Affairs, 1996-1999.

### References

- Chiou, W.-F., Sung, Y.-J., Liao, J.-F., Shum, A.Y.-C., and Chen, C.-F., Inhibitory effect of dehydroevodiamine and evodiamine on nitric oxide production in cultured murine macrophages. *J. Nat. Prod.* **60**, 708-711 (1997).
- Danieli, B., Lesma, G., and Palmisano, G., A new tryptophan derived alkaloid from *Evodia rutaecarpa* (Juss.) Benth. et Hook. *Experientia* **35**, 156 (1979).
- Haji, A., Momose, Y., Takeda, R., Nakanishi, S., Horiuchi, T., and arisawa, M., Increased feline cerebral blood flow induced by dehydroevodiamine hydrochloride from *Evodia rutaecarpa*. *J. Nat. Prod.*, **57**, 387-389 (1994).
- Itokawa, H., Inamatsu, M., and Takeya, K., A cytotoxic principle from *Evodia rutaecarpa*. *Shoyakugaku Zasshi* **44**, 135-137 (1990).
- Kamikado, T., Chang, C.-F., Murakoshi, S., Sakurai, A., and Tamura, S., Isolation and structure elucidation of three quinolone alkaloids from *Evodia rutaecarpa*. *Agric. Biol. Chem.* **40**, 605-609 (1976).
- Kang, S. S., Um, B. H., Kim, J. S., and Ahn, B. T., Isolation of flavonoids from *Evodia Fructus*. *Kor. J. Pharmacogn.* **28**, 9-14 (1997).
- Kano, Y., Zong, Q., and Komatsu, K.-I., Pharmacological properties of galenic preparation. XIV. Body temperature retaining effect of the Chinese traditional medicine, "Goshuyu-to" and component crude drugs. *Chem. Pharm. Bull.* **39**, 690-692 (1991).
- Kim, Y. C., Ki, N. Y., Jeong, S. J., Sohn, D.-H., Miyamoto, T., and Higuchi, R., biologically active quinolone alkaloids from *Evodia rutaecarpa* on *Artemia salina*. *Planta Med.* **64**, 490 (1998).
- King, C. L., Kong, Y. C., Wong, N. S., Teung, H. W., Fong, H. H. S., and Sankawa, U., Uterotonic effect of *Evodia rutaecarpa* alkaloids. *J. Nat. Prod.* **43**, 577-582 (1980).
- Matsuda, H., Wu, J.-X., Tanaka, T., Iinuma, M., and Kubo, M., Antinociceptive activities of 70% methanol extract of *Evodia Fructus* and its alkaloidal components. *Biol. Pharm. Bull.* **20**, 243-248 (1997).
- Matsuda, H., Yoshikawa, M., Ko, S.-K., Iinuma, M., and Kubo, M., Antinociceptive and antiinflammatory activities of evodiamine and rutaecarpine. *Nat. Med.* **52**, 203-208 (1998a).
- Matsuda, H., Yoshikawa, M., Iinuma, M., and Kubo, M., Antinociceptive and antiinflammatory activities of limonin from the fruits of *Evodia rutaecarpa* var. *bodinieri*. *Planta Med.* **64**, 339-342 (1998b).
- Moon, T. C., Chung, K. C., Son, K. H., Kim, H. P., Kang, S. S., Chang, H. W., Screening of Cyclooxygenase-2 (COX-2) Inhibitors from Natural Products. *Yakhak Hoeji* **42**, 214-219 (1998).
- Park, C. H., Kim, S.-H., Choi, W., Lee, Y.-J., Kim, J. S., Kang, S. S., and Suh, Y. H., Novel anticholinestrase and anti-amnesic activities of dehydroevodiamine, a constituent of *Evodia rutaecarpa*. *Planta Med.* **62**, 405-409 (1996).
- Shin, H.-K., Do, J.-C., Son, J.-K., Lee, C.-S., Lee, C.-H., Cheong, C. J., Quinoline alkaloids from the fruits of *Evodia officinalis*. *Planta Med.* **64**, 764-765 (1998).
- Shoji, N., Umeyama, A., Takemoto, T., Kajiwara, A., and Ohizumi, Y., Isolation of evodiamine, a powerful cardiotonic principle, from *Evodia rutaecarpa*. Bentham (Rutaceae). *J. Pharm. Sci.* **75**, 612-613 (1986).
- Shoji, N., Umeyama, A., Iuchi, A., Saito, N., Takemoto, T., Nomoto, K., and Ohizumi, Y., Isolation of a new alkaloid from *Evodia rutaecarpa*. *J. Nat. Prod.* **51**, 791-792 (1988).
- Son, K. H., Kwon, S. Y., Kim, H. P., Chang, H. W., Kang, S. S., Constituents from *Syzygium aromaticum* Merr. et Perry. *Nat. Prod. Sci.* **4**, 263-267 (1998).
- Sugimoto, T., Miyase, T., Kuroyanagi, M., Ueno, A., Limonoids and quinolone alkaloids from *Evodia rutaecarpa*. Bentham. *Chem. Pharm. Bull.* **36**, 4453-4461 (1988a).
- Sugimoto, T., Ueno, A., Kadota, S., Cui, C., and Kikuchi, T., New 5-H limonoids from *Evodia rutaecarpa*. Bentham. *Chem. Pharm. Bull.* **36**, 1237-1240 (1988b).
- Takagi, S., Kinoshida, T., Sameshima, M., Akiyama, T., Kobayashi, S., and Sankawa, U., Isolation of synephrine from *Evodia* fruits. *Shoyakugaku Zasshi* **33**, 35-37 (1979).
- Tang, W., and Eisenbrand, G., Chinese Drugs of Plant Origin, Springer-Verlag, Berlin, pp. 509-519 (1992).

Tang, Y.-Q., Feng, X.-Z., and Huang, L., Studies on the Chemical Constituents of *Evodia rutaecarpa* [Juss] Benth. *J. Chinese Pharm. Sci.*, **6**, 65-69 (1997).

Yamahara, J., Yamada, T., Kitani, T., Naitoh, Y., and Fujimura, H., Antianoxic action and active constituents of *Evodiae Fructus*. *Chem. Pharm. Bull.* **37**, 1820-1822 (1989).

Yang, M.C.M., Wu, S.-L., Kuo, J.-S., Chen, C.-F., The hypotensive and negative chronotropic effects of dehydroevodiamine. *Eur. J. Pharmacol.* **182**, 537-542 (1990).

(Accepted February 9, 1999)